

MINIREVIEW

[View Article Online](#)
[View Journal](#) | [View Issue](#)Cite this: *Chem. Sci.*, 2020, **11**, 3390 All publication charges for this article have been paid for by the Royal Society of Chemistry

Photo-controllable bioorthogonal chemistry for spatiotemporal control of bio-targets in living systems

Jinbo Li,  Hao Kong,  Chenghong Zhu and Yan Zhang  *

The establishment of bioorthogonal chemistry is one of the most significant advances in chemical biology using exogenous chemistry to perturb and study biological processes. Photo-modulation of biological systems has realized temporal and spatial control on biomacromolecules in living systems. The combination of photo-modulation and bioorthogonal chemistry is therefore emerging as a new direction to develop new chemical biological tools with spatiotemporal resolution. This minireview will focus on recent development of bioorthogonal chemistry subject to spatiotemporal control through photo-irradiation. Different strategies to realize photo-control on bioorthogonal bond-forming reactions and biological applications of photo-controllable bioorthogonal reactions will be summarized to give a perspective on how the innovations on photo-chemistry can contribute to the development of optochemical biology. Future trends to develop more optochemical tools based on novel photochemistry will also be discussed to envision the development of chemistry-oriented optochemical biology.

Received 27th December 2019

Accepted 7th March 2020

DOI: 10.1039/c9sc06540g

rsc.li/chemical-science

1. Introduction

Molecular events in living systems are usually dynamic and therefore difficult to decipher. Technologies with spatiotemporal resolution have found profound applications in exploring dynamic biological processes.¹ Photo-irradiation as a conventional external stimulus can be tuned with excellent spatiotemporal resolution as well as biocompatibility.² Optogenetics that uses light to modulate intracellular molecular events in a spatiotemporal manner has been considered as a revolutionary technique in neuroscience.³ However, limited types of genetically encoded proteins that are able to respond rapidly to light irradiation by changing conformation to alter cell behaviour restricted the broad application of optogenetics. In addition, optogenetic technology is not applicable to biomacromolecules that are not able to be genetically engineered. Alternatively, optochemical methods using photo-controllable chemical moieties integrated with various types of biomacromolecules provide promising spatiotemporal methods to explore dynamic biological systems.⁴

Three types of photo-induced reactions have been applied to optochemical biology to develop molecular tools with spatiotemporal resolution. Photo-isomerization has been applied to

the development of photo-switches to reversibly regulate a wide range of biological targets including transmembrane proteins, lipid membranes, nucleic acids, and so on.⁵ Photo-induced bond cleavage reactions have been applied to photo-uncaging for transient activation of specific bioactivity of “caged” biomolecules modified with photo-removable groups.^{6–8} Photo-induced bond-forming reactions as a major class of photoreactions of small molecules have rarely been used to develop molecular tools in optochemical biology.^{9,10} Efforts to revisit photo-induced bond-forming reactions under bio-mimetic conditions have demonstrated great potential in the development of biocompatible or bioorthogonal tools for spatiotemporal labelling of bio-targets.¹¹

Bioorthogonal chemistry has presented an unprecedented approach for the *in situ* labelling and modulation of bio-targets in living systems without disturbing other “innocent” biomolecules.^{12–14} The development of bioorthogonal reactions to expand the bioorthogonal tool box has been of intense research interest.^{15–18} However, it remains a great challenge to establish new bioorthogonal reactions that are orthogonal to the most commonly used bioorthogonal reactions such as azide alkyne cycloaddition (AAC) catalysed by Cu (CuAAC) or promoted by strain (SPAAC).^{19,20} Inverse electron-demand Diels–Alder cycloaddition (IEDDAC) reactions using various strained alkenes have been established as new bioorthogonal tools that are orthogonal to current AAC chemistry. The importance of photo-controllable bioorthogonal chemistry has encouraged the development of UV or visible light-triggered

State Key Laboratory of Analytical Chemistry for Life Sciences, Jiangsu Key Laboratory of Advanced Organic Materials, School of Chemistry and Chemical Engineering, Chemistry and Biomedicine Innovation Center (ChemBIC), Nanjing University, Nanjing 210023, China. E-mail: njuzy@nju.edu.cn



cycloadditions as biocompatible or bioorthogonal reactions. In this review, we will focus on the recent progress of photo-triggered bioorthogonal chemistry, including different strategies to realize photo-control of bioorthogonal bond-forming reactions and the biological applications of photo-controllable bioorthogonal reactions.

2. Strategies to realize photo-control on bioorthogonal bond-forming reactions

Typical bioorthogonal bond-forming reactions that proceed *via* distinct mechanisms are listed in Fig. 1. Some of the catalyst-free bioorthogonal reactions, including SPAAC and IEDDAC, have been applied for the *in situ* labelling of bio-targets in living cells or even in mice models.^{21,22} However, the precise control of these bioorthogonal bond-forming reactions at single cell or subcellular levels is difficult to achieve, because biomacromolecules modified with bioorthogonal functionalities were hard to introduce into a population with single cellular or subcellular resolution. One potential approach to define where the bioorthogonal reaction will occur, such as at single-cellular or subcellular levels, is to use an external stimulus with spatial resolution to trigger the bioorthogonal reaction inside a specific area of living subjects. Light irradiation is a typical external stimulus with high spatiotemporal resolution and minimal damage to living subjects. Therefore, different strategies to trigger bioorthogonal reactions with light irradiation have been developed, which will be introduced in detail in the following sections.

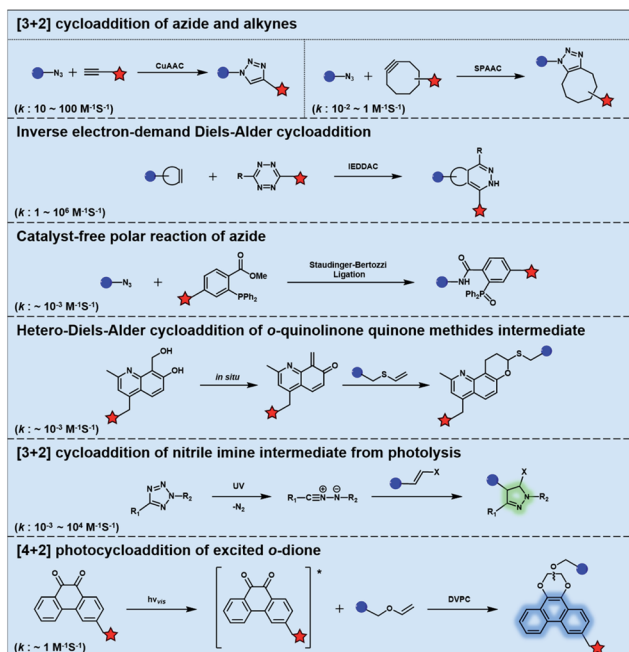


Fig. 1 Typical bioorthogonal bond-forming reactions that proceed *via* distinct mechanisms.

2.1 Bioorthogonal dione–vinyl ether photocycloaddition (DVPC) reaction triggered by visible light

To provide bond-forming bioorthogonal reactions under photo-control, the most straightforward strategy is to use photo-excitable bioorthogonal functionality that reacts with its partner *via* a specific photoreaction pathway leading to a single ligation product under biocompatible conditions. However, photoreactions that proceed *via* photo-induced electron transfer (PeT) from the high-energy excited state to form highly reactive radical species usually lead to a complex mixture of products. To develop new types of photo-controllable bioorthogonal reactions *via* PeT, the key challenge is to realize the control of both the PeT process of high-energy excited states and highly reactive radical species under complex biological environments.

Revisiting classical photoreactions such as the Paterno–Büchi reaction involving alkenes or alkynes provides a feasible approach to develop a prototype of photo-triggered bioorthogonal reactions. Inspired by the photoreactions of excited *o*-diones with π -bonds,²³ we recently established visible-light driven dione–vinyl ether photocycloaddition (DVPC), which is the first bioorthogonal reaction *via* the PeT pathway (Fig. 2A).²⁴ The photo-controllable bioorthogonal functionality in DVPC is the *o*-dione moiety such as 9,10-phenanthrenequinone (PQ). PQ has an absorption peak between 400 and 500 nm and is able to be efficiently excited by focused visible light from light sources such as LED lamps. The two excited states of PQ, $n\pi^*$ and $\pi\pi^*$ excited states, are energetically close and have a similar conformational structure.²⁵ In protic solvents such as water, the first excited state of PQ (PQ*) transits from $n\pi^*$ to $\pi\pi^*$. The $\pi\pi^*$ excited state of PQ* is inert to various reactive species abundant in biological systems. It is therefore possible to use water that is highly abundant in biological systems as a “solvent cage” to block side reactions that proceed *via* the excited states of *o*-diones, provided that there is any photocycloaddition substrate undergoing efficient PeT with the $\pi\pi^*$ excited state of PQ*. With calculations on the Gibbs free energy on PeT of PQ* in $\pi\pi^*$ with various substrates, we found that the highly electron-rich alkene vinyl ether (VE) has a strong tendency to proceed *via* PeT with PQ* in $\pi\pi^*$ in aqueous solution. As shown in Fig. 2, the reaction of PQ* with VE proceeded *via*

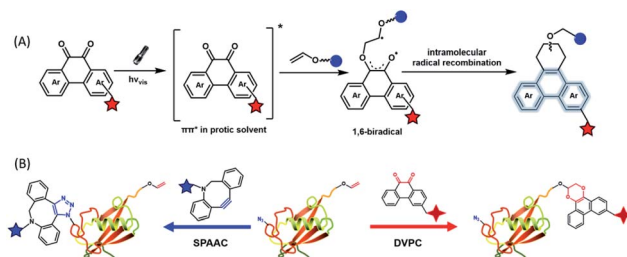


Fig. 2 (A) Visible-light driven bioorthogonal dione–vinyl ether photocycloaddition (DVPC) reaction *via* the PeT pathway; (B) orthogonal protein labelling achieved by SPAAC and DVPC that are orthogonal to each other.



a rapid PeT process to form the first covalent bond with high specificity. The resulting biradical intermediate then proceeded exclusively *via* an intramolecular radical recombination to form the [4 + 2] photo-cycloadduct with two regioisomers. With this strategy, we addressed the challenge to control both the PeT process of excited states and the following radical-involved pathways under complex biological environments.

The successful establishment of the visible-light driven DVPC reaction added a new type of bioorthogonal bond-forming reaction to the current bioorthogonal tool-kit. While highly electron-rich alkenes such as VE reacted with the highest reaction rate with excited PQ* in aqueous solution, strained alkenes such as *trans*-cyclooctene also showed modest reactivity. However, neither terminal alkynes nor strained alkynes were able to undergo PeT with PQ*. DVPC and SPAAC were therefore able to be used together as orthogonal bioorthogonal reactions for orthogonal labelling of biomacromolecules such as proteins (Fig. 2B). Biomacromolecules on live cells can be labelled with high spatiotemporal resolution using the DVPC reaction driven by visible light with a tunable wavelength, intensity and irradiation area. Because light irradiation can be precisely confined at the single-cell or even the subcellular level, single-cellular or subcellular level labelling of targeted biomacromolecules is thus feasible using the bioorthogonal DVPC reaction.

2.2 Photo-release of bioorthogonal functionalities or nitrile imine for spatial control on the following reactions

Photo-responsive precursors of established bioorthogonal functionalities have been designed and optimized to realize photo-control on several well-established bioorthogonal reactions such as SPAAC and Staudinger ligation. For example, photochemical generation of reactive dibenzocyclooctynes was developed by Popik *et al.* from the known photochemical decarbonylation of thermally stable diarylsubstituted cyclopropenones.²⁶ Pioneered by this work, various photo-labile precursors of strained alkynes have been explored for photo-controllable generation of strained alkynes as the bio-orthogonal functionality.^{27–30} Visible-light triggered active alkyne generation from cyclopropenones was established with the aid of visible light-responsive photocatalysts.³¹ Caged phosphines have also been designed and prepared for UV or even visible light triggered generation of active phosphine for light-controllable Staudinger–Bertozzi ligation.^{32,33} Photo-caging of cyclopropenes has also been reported to provide different versions of cyclopropenes with different ligation kinetics with s-tetrazines.^{34,35} Table 1 summarizes typical photo-labile precursors that have been reported to generate known bioorthogonal functionalities under different conditions for photolysis, with general comments on the advantages and limits for biological applications.

Table 1 Precursors and photo-released bioorthogonal functionalities to develop photo-controllable bioorthogonal reactions

Bioorthogonal functionality obtained after photolysis	Photo-labile precursor	Typical conditions for photolysis	Advantages/limits for biological applications
		1–3 min' irradiation in a mini-Rayonet photoreactor equipped with multiple 350 nm fluorescent tubes ^{26,27}	Photo-SPAAC enabled photo-control on labeling azido-bearing biomolecules in live cells, with concern on diffusion of the <i>in situ</i> formed strained alkyne due to fast kinetics in photolysis but a relatively slow rate of SPAAC
		5 min' irradiation at 365 nm from a UV lamp VL-215.L (ref. 28)	Photo-IEDDAC reactions with fast kinetics enabled rapid labeling of tetrazine-bearing proteins within minutes at low μ M concentrations. Future applications on mammalian cells need more biocompatible light than UV
		Pulsed laser at 375 nm (ref. 29)	Ultra-fast and efficient photo-release of dibenzosilacyclohept-4-yne achieved well-balanced combination of reactivity and stability in aqueous solutions. Photo-stability of the released alkyne exposed to a laser at the irradiation wavelength was a concern
		Femtosecond laser for two-photon (690 nm) or three-photon (1050 nm) excitation ³⁰	Multiphoton activation of strained alkynes allowed MP-SPAAC with low phototoxicity and deep penetration in biological systems, with a limit in the region of interest irradiated with a two-photon or three-photon laser
		30–180 min' UV light irradiation with a 350 W medium pressure Hg arc lamp ³²	Simple uncaging approach helped to realize photo-control on the highly specific Staudinger–Bertozzi ligation, with concern on the biocompatibility of the light irradiation conditions with live cells
		20–40 min' irradiation with broadband visible light (>420 nm) from a 500 W Hg lamp ³³	Photo-uncaging quantum efficiency of Staudinger ligation triggered by visible light is lower than UV uncaging
		Hours of irradiation under UV ³⁴	Module photo-cage of cyclopropane needs to be further optimized for real biological applications



Photolysis or uncaging of the listed precursors together with the subsequent bioorthogonal reactions such as SPAAC or IEDDAC has been termed photo-SPAAC, photo-IEDDAC, *etc.* Photo-triggered release of the bioorthogonal functionality helps to control when and where the bioorthogonal partners react with each other. If the photolysis proceeds fast enough to generate the reactive bioorthogonal group, diffusion of molecules bearing the photo-released bioorthogonal functionalities would not jeopardize the spatial resolution of the following bioorthogonal reaction in living systems. The photophysical and photochemical properties of these precursors have been investigated extensively to optimize the photolysis step.^{26,29,30,32,33} The absorption wavelength and quantum yield are two important parameters that have been considered for the purpose of photo-release of bioorthogonal functionalities, because light with a longer wavelength and lower intensity is more compatible with living systems. Fast rates of photolysis and the following bioorthogonal ligation process are important for the spatiotemporal resolution of a photo-controlled reaction.^{28,29,34,35} It should be noted that this type of photo-controllable bioorthogonal reaction is only a modified version of known bioorthogonal reactions. Cross-talk of the original version and photo-controllable version of the same bioorthogonal reactions is therefore not avoidable.

Nitrile imines were reported by Lin *et al.* to react rapidly with alkenes *via* 1,3-dipolar cycloaddition in aqueous solution.³⁶ Tetrazole-ene photo-click chemistry (TEPC) was therein developed using mild UV light to trigger the fast generation of nitrile imines from 2,5-biaryltetrazoles. TEPC has been tested in living cells with unique fluorescence turn-on accompanied by the formation of the pyrazoline product.¹¹ To make the reaction more bio-compatible, tetrazoles substituted with thiophene³⁷ or naphthalene³⁸ had been used to realize visible light triggered TEPC. Other than tetrazoles, diarylsydones were also reported to generate nitrile imines upon light irradiation.^{39,40} To address the issue of competitive reactions of nitrile imine intermediates with carboxylic acid and hydroxyl groups in living systems,^{41,42} efforts have been made to design and prepare tetrazole with bulky substitution groups to minimize side reactions in competition with TEPC (Fig. 3).⁴³ Lin *et al.* designed and prepared di(*o*-2'-*N*-Boc-pyrrole)-substituted tetrazole and found that it upon UV irradiation formed nitrile imines with an extraordinarily long half-life (up to 120 s). The sterically shielded nitrile imine showed significantly improved selectivity between 1,3-dipolar addition with alkenes and nucleophilic addition (Fig. 3). The TEPC reaction of this bulky tetrazole was successfully applied to rapid (~1 min) bioorthogonal labeling of the glucagon receptor in mammalian cells. Other than nitrile imine, nitrile ylide generated by azirine through UV irradiation has also been reported to undergo 1,3-dipolar cycloaddition with alkenes,⁴⁴ which has been applied to protein bioconjugation.

2.3 Photocatalytic activation of biocompatible or bioorthogonal reactions

Light with longer wavelengths is more compatible with biological systems, and it is therefore attractive to use visible, red or near infrared (IR) light to control bioorthogonal reactions.^{38,45}

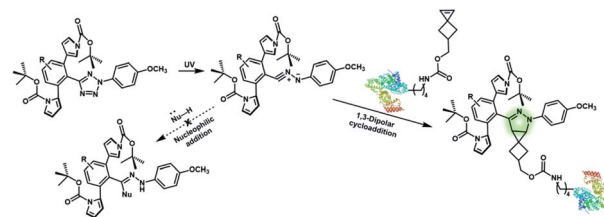


Fig. 3 Strategy to improve the bioorthogonal TEPC reaction to block the side reaction of nucleophilic addition to nitrile imine by using a sterically shielded tetrazole substrate.



Fig. 4 Photocatalytic activation of the IEDDAC reaction.

Photocatalysts sensitive to red light have been utilized to trigger the formation of bioorthogonal substrates such as tetrazine for the subsequent bioorthogonal IEDDAC reactions (Fig. 4).⁴⁶ Visible-light catalysis has attracted great research interest in synthetic chemistry in recent years,⁴⁷ which resulted in a large pool of visible light-catalysed bond-forming reactions. Because visible light photo-redox catalysis based on ruthenium complexes has been applied to various chemical biological studies,⁴⁸ the adaptability of this large family of organic reactions to bioorthogonal reactions is worth exploring.^{49–51} Fluorescent dyes such as fluorescein and rhodamine derivatives have been reported to catalyse visible light driven reactions in mammalian cells including neurons.⁵² They may also be applied to photocatalytic activation of bioorthogonal reactions.

3 Biological applications of photo-controllable bioorthogonal reactions

Most of the naturally occurring photo-switches in biological systems are based on photo-isomerization, which has been reviewed elsewhere.⁵³ Except for photo-switches based on photo-isomerizable double bonds, bioorthogonal molecular tools based on bioorthogonal bond-forming as well as bond-cleavage reactions have also been utilized to explore biological systems.⁴ Small bioorthogonal handles have been successfully inserted into endogenous bio-targets using chemical biological techniques, such as genetic engineering with unnatural amino acid⁵⁴ and metabolic incorporation.¹³ Spatiotemporal labelling or modulation of these bio-targets has been achieved by using photo-controllable bioorthogonal reactions.

3.1 Photo-controlled labelling of specific bio-targets in live cells

In situ labelling of specific bio-targets in live cells or in mice has been achieved by using bioorthogonal reactions such as SPAAC and IEDDAC.^{55,56} With the photo-controllable version of these



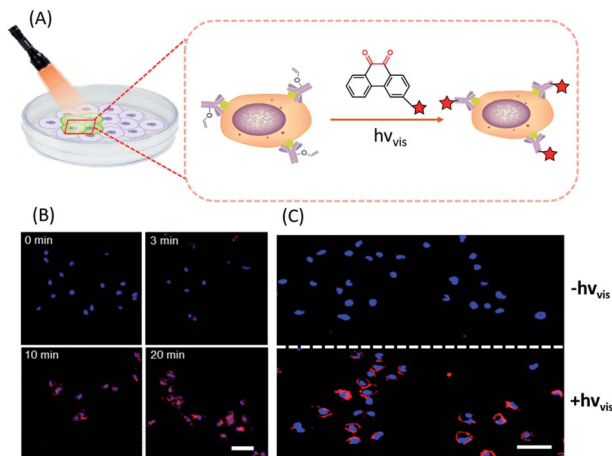


Fig. 5 Spatiotemporal labelling of live cells realized by the visible-light driven DVPC reaction (scale bar: 50 μm).

bioorthogonal reactions, *in situ* labelling of bio-targets in live cells has been achieved with spatiotemporal resolution. By genetically introducing a tetrazine group into the outer-membrane OmpC protein, Lang *et al.* used a photo-IEDDAC of a cyclopropanone-caged dibenzoannulated bicyclo[6.1.0]nonyne probe to realize UV-controllable labelling of tetrazine-bearing OmpC in live *E. coli*.⁵⁷ Carrico *et al.* applied a photo-Staudinger–Bertozzi reaction for spatial labelling of cell-surface azidoglycoproteins in fixed cells and zebrafish.³² We utilized the visible light-triggered DVPC reaction to label the epidermal growth factor receptor (EGFR) protein on a live cell membrane bound with VE modified Cetuximab (Fig. 5).²⁴ The labelling process was spatiotemporally performed on living A549 cells by changing the irradiation time and the area of live cells exposed to a focused visible light beam from a hand-held LED lamp.

Nucleic acids inside live cells have also been spatiotemporally labelled by various photo-controllable ligation reactions. Azide-modified nucleosides were metabolically incorporated into nascent RNAs in subpopulations of live cells. Selective UV irradiation of cyclopropanone-caged oxadibenzocyclooctynes induced photo-SPAAC reactions with azide-modified RNAs.²⁷ Tetrazole-modified microRNAs after being transfected into live cells were found to be functional to suppress target gene expression.⁵⁸ We therefore applied intramolecular TEPC reactions for spatiotemporal labelling of microRNAs inside cells by controlling the UV irradiated area and time on live cells.⁵⁹ Our attempts to trigger intermolecular TEPC reactions of tetrazole-modified microRNAs with monomethyl fumarate in live cells did not result in intracellular fluorescence turn-on, which indicated that the efficiency of the intermolecular TEPC reaction in live cells was much lower than that of intramolecular TEPC.

3.2 Spatiotemporal regulation of intracellular biological functions

Manipulating the biological function of intracellular biomolecules with spatiotemporal resolution is pivotal to deciphering dynamic molecular events occurring within a specific region of

live cells. Various biological activities, such as gene expression, protein modification and cell signalling, have been regulated with spatiotemporal resolution using optochemical tools based on bioorthogonal photo-uncaging or photo-isomerization reactions. Because Deiters and co-workers have summarized the significant advances of optochemical control of various biological processes in cells and animals reported before 2018,⁶⁰ we will briefly introduce the recent advances achieved thereafter in this section.

Site-specific incorporation of photo-removable blocking groups into active residues of a target protein has been achieved *via* unnatural amino acid insertion. This approach, combined with bioorthogonal photo-uncaging, has become a canonical strategy to spatiotemporal regulation of protein functions. While the key residues of most proteins are not amenable to genetic expanding strategies, Chen *et al.* recently reported a computationally aided and genetically encoded proximal decaging (CAGE-prox) strategy to block residues close to active sites for subsequent photo-activation.⁶¹ Time-resolved activation of a broad range of proteins was achieved by this strategy using UV irradiation to remove blocking groups in proximity to the functional residue of specific proteins. Spatiotemporal control on the activity of Cas9 protein for gene editing has been demonstrated by Choudhary *et al.* They fused Cas9 with the destabilized domains of dihydrofolate reductase (DHFR) and used a photo-caged ligand for DHFR, trimethoprim, to realize precise control on the activity of Cas9 through light irradiation.⁶² Small molecule ligands that can degrade target proteins *via* the proteolysis targeting chimera (PROTAC) strategy have also been used together with photo-decaging or photo-isomerization strategies for light-induced or reversible regulation of protein expression.^{63,64}

3.3 Spatiotemporal modulation of microenvironments to trigger cellular responses

Cells are known to respond to dynamic changes of microenvironments. Photo-modulation of cellular microenvironments engineered with photo-controllable units may trigger cellular responses.⁶⁵ To mimic native cellular microenvironments, cells have been encapsulated in three dimensional microenvironments that are supramolecular or polymer hydrogels engineered with photo-controllable bioorthogonal functionalities.^{66,67} A UV-triggered intramolecular TEPC reaction has been adopted to modulate the mechanical properties of supramolecular hydrogel encapsulating stem cells.⁶⁶ Photo-induced bioorthogonal bond-cleavage reactions has been used together with bioorthogonal ligation reactions such as SPAAC to construct a 3D cellular microenvironment.⁶⁷ Photo-reversible protein-patterning of the 3D cellular microenvironment to guide the stem cell fate has been demonstrated.⁶⁸ Recently, photo-reversible immobilization for the 4D patterning of cellular microenvironments has been realized by using site-specifically modified proteins bearing bioorthogonal functionalities.⁶⁵ Photo-release of soluble proteins in engineered cellular microenvironments promoted canonical EGFR activation and associated membrane endocytosis of the encapsulated cells.



4. Future outlook

Nature has developed precise mechanisms for living systems to sense and utilize sunlight. Through learning from mother nature how to use biologically occurring photo-switches, optogenetics has made significant contributions to decipher molecular events in complex and dynamic biological systems. With the development of versatile photochemical tools, a new era of photochemical biology is emerging. Based on diversified photochemistry of small molecules in combination with chemical biological methods, the development of photo-controllable bioorthogonal chemistry is of great importance in photochemical biology. With the establishment of bioorthogonal photocycloadditions *via* controllable excited states induced by visible light, a variety of photochemical molecular tools can be developed to explore biological processes at a single cellular or even at a subcellular level. Meanwhile, how to use light with a long wavelength to control bioorthogonal reactions is worth exploring because of potential applications in deep tissues of living animals.

5. Conclusions

The recent development of bioorthogonal chemistry subject to photo-control to realize spatiotemporal resolution was reviewed. Different strategies to develop photo-controllable bioorthogonal bond-forming reactions were summarized. A visible-light driven DVPC reaction that proceeds *via* the excited state of *o*-dione with the electron-rich alkene vinyl ether was introduced in detail. The successful establishment of a bioorthogonal reaction *via* PeT of *o*-dione at $\pi\pi^*$ excited states to its partner bioorthogonal functionality represented a new direction to develop light-driven bioorthogonal reactions. Alternative ways to provide bioorthogonal bond-forming reactions under photo-control, such as using photo-uncaging or photo-activation for *in situ* formation of known bioorthogonal functionalities, were also introduced such as photo-SPAAC, photo-IEDDAC, *etc.* Recent examples of photochemical tools based on photo-controllable bioorthogonal chemistry were briefly mentioned with respect to their applications in the spatiotemporal labelling or regulation of bio-targets in living systems. A future outlook on the development of new photo-controllable bioorthogonal chemistry as well as new photochemical tools envisioned the capacity of chemistry-oriented photochemical biology to create powerful spatiotemporal molecular tools for precise modulation of biological systems. Moreover, significant challenges including shifting the activation wavelength of photo-controllable bioorthogonal chemistry from the UV region to the visible light region and extending their applications into intracellular microenvironments in living cells are yet to be addressed.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We would like to acknowledge financial support from the National Natural Science Foundation of China (91753116, 21977043, and 21877058).

References

- 1 S. Aubert, M. Bezagu, A. C. Spivey and S. Arseniyadis, *Nat. Rev. Chem.*, 2019, **3**, 706–722.
- 2 K. Glusac, *Nat. Chem.*, 2016, **8**, 734–735.
- 3 K. Deisseroth, *Nat. Methods*, 2011, **8**, 26–29.
- 4 N. Ankenbruck, T. Courtney, Y. Naro and A. Deiters, *Angew. Chem., Int. Ed.*, 2018, **57**, 2768–2798.
- 5 K. Hüll, J. Morstein and D. Trauner, *Chem. Rev.*, 2018, **118**, 10710–10747.
- 6 H. Yu, J. Li, D. Wu, Z. Qiu and Y. Zhang, *Chem. Soc. Rev.*, 2010, **39**, 464–473.
- 7 P. Klán, T. Šolomek, C. G. Bochet, A. Blanc, R. Givens, M. Rubina, V. Popik, A. Kostikov and J. Wirz, *Chem. Rev.*, 2013, **113**, 119–191.
- 8 M. J. Hansen, W. A. Velema, M. M. Lerch, W. Szymanski and B. L. Feringa, *Chem. Soc. Rev.*, 2015, **44**, 3358–3377.
- 9 G. W. Prestonab and A. J. Wilson, *Chem. Soc. Rev.*, 2013, **42**, 3289–3301.
- 10 H. Wu and J. Kohler, *Curr. Opin. Chem. Biol.*, 2019, **53**, 173–182.
- 11 C. P. Ramil and Q. Lin, *Curr. Opin. Chem. Biol.*, 2014, **21**, 89–95.
- 12 E. M. Sletten and C. R. Bertozzi, *Acc. Chem. Res.*, 2011, **44**, 666–676.
- 13 B. Cheng, R. Xie, L. Dong and X. Chen, *ChemBioChem*, 2016, **17**, 11–27.
- 14 H. W. Shih, D. N. Kamber and J. A. Prescher, *Curr. Opin. Chem. Biol.*, 2014, **21**, 103–111.
- 15 M. F. Debets, S. S. van Berkel, J. Dommerholt, A. J. Dirks, F. P. J. T. Rutjes and F. L. van Delft, *Acc. Chem. Res.*, 2011, **44**, 805–815.
- 16 F. Liu, Y. Liang and K. N. Houk, *Acc. Chem. Res.*, 2017, **50**, 2297–2308.
- 17 E. Kim and H. Koo, *Chem. Sci.*, 2019, **10**, 7835–7851.
- 18 D. N. Kamber, S. S. Nguyen, F. Liu, J. S. Briggs, H. W. Shih, R. D. P. J. M. Johnson, A. Halpin, T. Morizumi, V. I. Prokhorenko, O. P. Ernst and R. J. Dwayne Miller, *Nat. Chem.*, 2015, **7**, 980–986.
- 19 D. M. Patterson and J. A. Prescher, *Curr. Opin. Chem. Biol.*, 2015, **28**, 141–149.
- 20 R. D. Row and J. A. Prescher, *Acc. Chem. Res.*, 2018, **51**, 1073–1081.
- 21 H. J. Jeonga, R. J. Yoob, J. K. Kima, M. H. Kimb, S. H. Parka, H. Kima, J. W. Lima, S. H. Doc, K. C. Leeb, Y. J. Leeb and D. W. Kim, *Biomaterials*, 2019, **199**, 32–39.
- 22 N. K. Devaraj, *ACS Cent. Sci.*, 2018, **4**, 952–959.
- 23 L. Wang, Y. Huang, Y. Liu, H. Fun, Y. Zhang and J. Xu, *J. Org. Chem.*, 2010, **75**, 7757–7768.
- 24 J. Li, H. Kong, L. Huang, B. Cheng, K. Qin, M. Zheng, Z. Yan and Y. Zhang, *J. Am. Chem. Soc.*, 2018, **140**, 14542–14546.



- 25 V. R. Kumar, N. Rajkumar, F. Ariese and S. Umapathy, *J. Phys. Chem. A*, 2015, **119**, 10147–10157.
- 26 A. A. Poloukhine, N. E. Mbua, M. A. Wolfert, G. Boons and V. V. Popik, *J. Am. Chem. Soc.*, 2009, **131**, 15769–15776.
- 27 S. Nainar, M. Kubota, C. McNitt, C. Tran, V. V. Popik and R. C. Spitale, *J. Am. Chem. Soc.*, 2017, **139**, 8090–8093.
- 28 S. V. Mayer, A. Murnauer, M. K. von Wrisberg, M. L. Jokisch and K. Lang, *Angew. Chem., Int. Ed.*, 2019, **58**, 15876–15882.
- 29 M. Martinek, L. Filipová, J. Galeta, L. Ludvíková and P. Klán, *Org. Lett.*, 2016, **18**, 4892–4895.
- 30 C. D. McNitt, H. Cheng, S. Ullrich, V. V. Popik and M. Bjerknes, *J. Am. Chem. Soc.*, 2017, **139**, 14029–14032.
- 31 K. Mishiro, T. Kimura, T. Furuyama and M. Kunishima, *Org. Lett.*, 2019, **21**, 4101–4105.
- 32 L. Shah, S. T. Laughlin and I. S. Carrico, *J. Am. Chem. Soc.*, 2016, **138**, 5186–5189.
- 33 P. Hu, K. Berning, Y. Lam, I. H. Ng, C. Yeung and M. H. Lam, *J. Org. Chem.*, 2018, **83**, 12998–13010.
- 34 P. Kumar, O. Zainul, F. M. Camarda, T. Jiang, J. A. Mannone, W. Huang and S. T. Laughlin, *Org. Lett.*, 2019, **21**, 3721–3725.
- 35 P. Kumar, T. Jiang, S. Li, O. Zainul and S. T. Laughlin, *Org. Biomol. Chem.*, 2018, **16**, 4081–4085.
- 36 A. Herner and Q. Lin, *Top. Curr. Chem.*, 2016, **374**, 1–31.
- 37 P. An, Z. Yu and Q. Lin, *Chem. Commun.*, 2013, **49**, 9920–9922.
- 38 Z. Yu, T. Y. Ohulchanskyy, P. An, P. N. Prasad and Q. Lin, *J. Am. Chem. Soc.*, 2013, **135**, 16766–16769.
- 39 L. Zhang, X. Zhang, Z. Yao, S. Jiang, J. Deng, B. Li and Z. Yu, *J. Am. Chem. Soc.*, 2018, **140**, 7390–7394.
- 40 X. Zhang, X. Mu, S. Jiang, J. Gao, Z. Yao, J. Deng, L. Zhang and Z. Yu, *Chem. Commun.*, 2019, **55**, 7187–7190.
- 41 Z. Li, L. Qian, L. Li, J. C. Bernhammer, H. V. Huynh, J. S. Lee and S. Yao, *Angew. Chem., Int. Ed.*, 2016, **55**, 2002–2006.
- 42 Y. Tian, M. P. Jacinto, Y. Zeng, Z. Yu, J. Qu, W. R. Liu and Q. Lin, *J. Am. Chem. Soc.*, 2017, **139**, 6078–6081.
- 43 P. An, T. M. Lewandowski, T. G. Erbay, P. Liu and Q. Lin, *J. Am. Chem. Soc.*, 2018, **140**, 4860–4868.
- 44 R. K. V. Lim and Q. Lin, *Chem. Commun.*, 2010, **46**, 7993–7995.
- 45 R. J. Mart and R. K. Allemann, *Chem. Commun.*, 2016, **52**, 12262–12277.
- 46 H. Zhang, W. S. Trout, S. Liu, G. A. Andrade, D. A. Hudson, S. L. Scinto, K. T. Dicker, Y. Li, N. Lazouski, J. Rosenthal, C. Thorpe, X. Jia and J. M. Fox, *J. Am. Chem. Soc.*, 2016, **138**, 5978–5983.
- 47 Q. Liu and L. Wu, *Natl. Sci. Rev.*, 2017, **4**, 359–380.
- 48 S. Angerani and N. Winssinger, *Chem.–Eur. J.*, 2019, **25**, 6661–6672.
- 49 H. Huang, G. Zhang, L. Gong, S. Zhang and Y. Chen, *J. Am. Chem. Soc.*, 2014, **136**, 2280–2283.
- 50 C. Hu and Y. Chen, *Tetrahedron Lett.*, 2015, **56**, 884–888.
- 51 J. Yang, J. Zhang, L. Qi, C. Hua and Y. Chen, *Chem. Commun.*, 2015, **51**, 5275–5278.
- 52 H. Wang, W. Li, K. Zeng, Y. Wu, Y. Zhang, T. Xu and Y. Chen, *Angew. Chem., Int. Ed.*, 2019, **58**, 561–565.
- 53 P. J. M. Johnson, A. Halpin, T. Morizumi, V. I. Prokhorenko, O. P. Ernst and R. J. Dwayne Miller, *Nat. Chem.*, 2015, **7**, 980–986.
- 54 Z. Hao, S. Hong, X. Chen and P. R. Chen, *Acc. Chem. Res.*, 2011, **44**, 742–751.
- 55 K. Lang and J. W. Chin, *ACS Chem. Biol.*, 2014, **9**, 16–20.
- 56 K. Lang and J. W. Chin, *Chem. Rev.*, 2014, **114**, 4764–4806.
- 57 S. V. Mayer, A. Murnauer, M. K. von Wrisberg, M. L. Jokisch and K. Lang, *Angew. Chem.*, 2019, **131**, 16023–16029.
- 58 J. Li, L. Huang, X. Xiao, Y. Chen, X. Wang, Z. Zhou, C. Zhang and Y. Zhang, *J. Am. Chem. Soc.*, 2016, **138**, 15943–15949.
- 59 L. Huang, Y. Chen, L. Chen, X. Xiao, X. Wang, J. Li and Y. Zhang, *Chem. Commun.*, 2017, **53**, 6452–6455.
- 60 A. Bardhan and A. Deiters, *Curr. Opin. Struct. Biol.*, 2019, **57**, 164–175.
- 61 J. Wang, Y. Liu, Y. Liu, S. Zheng, X. Wang, J. Zhao, F. Yang, G. Zhang, C. Wang and P. R. Chen, *Nature*, 2019, **569**, 509–513.
- 62 D. Manna, B. Maji, S. A. Gangopadhyay, K. J. Cox, Q. Zhou, B. K. Law, R. Mazitschek and A. Choudhary, *Angew. Chem., Int. Ed.*, 2019, **58**, 6285–6289.
- 63 G. Xue, K. Wang, D. Zhou, H. Zhong and Z. Pan, *J. Am. Chem. Soc.*, 2019, **141**, 18370–18374.
- 64 P. Pfaff, K. T. G. Samarasinghe, C. M. Crews and E. M. Carreira, *ACS Cent. Sci.*, 2019, **5**, 1682–1690.
- 65 J. A. Shadish, G. M. Benuska and C. A. DeForest, *Nat. Mater.*, 2019, **18**, 1005–1014.
- 66 M. He, J. Li, S. Tan, R. Wang and Y. Zhang, *J. Am. Chem. Soc.*, 2013, **135**, 18718–18721.
- 67 C. A. DeForest and K. S. Anseth, *Nat. Chem.*, 2011, **3**, 925–931.
- 68 C. A. DeForest and D. A. Tirrell, *Nat. Mater.*, 2015, **14**, 523–531.

