

MINIREVIEW

View Article Online
View Journal | View Issue

Cite this: *Nanoscale Adv.*, 2021, **3**, 6040

Received 29th June 2021
Accepted 6th September 2021

DOI: 10.1039/d1na00508a

rsc.li/nanoscale-advances

Recent progress in cellulose-based electrospun nanofibers as multifunctional materials

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Cellulose, the most abundant natural polymer, has good biocompatibility, biodegradability, and non-toxicity, which make it and its derivatives promising candidates for the fabrication of multifunctional materials, while maintaining sustainability and environmental friendliness. The combination of electrospinning technology and cellulose (and its derivatives) provides a feasible approach to produce nanostructured porous materials with promising functionalities, flexibility, renewability and biodegradability. At the same time, it enables value-added applications of cellulose and its derivatives that are derived from nature or even biomass waste. This review summarizes and discusses the latest progress in cellulose-based electrospun nanofibers, including their construction methods and conditions, various available raw materials, and applications in multiple areas (water treatment, biomaterials, sensors, electro-conductive materials, active packaging, and so on), which are followed by the conclusion and prospects associated with future opportunities and challenges in this active research area.

1. Introduction

Electrospinning is a facile and efficient method for preparing fibers with nano-scaled diameters, which have a large specific surface area, good interconnectivity, and structural stability. Meanwhile owing to the random arrangement of nanofibers prepared by electrospinning, a large number of isotropic pores evenly distribute among fibers.^{1,2} These characteristics make electrospun nanofibers useful in a wide range of applications, including filtration & adsorption materials, “smart” materials, catalytic materials, and so on.^{3–7} However, the existing nanofibers fabricated by electrospinning are usually based on non-degradable polymers, which may cause environmental issues after disposal.

As the most abundant biodegradable polymer on the earth, cellulose possesses many fascinating properties, such as biocompatibility, environmental friendliness, and inexhaustible renewability.⁸ Taking advantage of the reactive, numerous, and regularly arranged hydroxyl groups along cellulose chains, a wide variety of cellulose derivatives have been developed by simple reactions between functional substituents and active hydroxyls.⁹ The derivatives possess interesting properties inherited from both cellulose and modified functional groups, including good solubility and processability, excellent flexibility, high mechanical strength, and so on.^{10,11} Hence, the

combination of electrospinning technology and cellulose (and its derivatives) provides a feasible approach to produce nanostructured porous materials with desirable properties. At the same time, it enables value-added applications of cellulose and its derivatives that are derived from nature or even biomass waste.

Overviewing the research work published in last 3 years (2018–2021), various electrospun nanofibers have been successfully prepared from cellulose and cellulose derivatives in different solvent systems.^{12–17} These nanofibers demonstrated promising applications in the fields of filtration, tissue engineering and biomedical engineering.^{18–22} According to Sci-Finder, 17 literature reviews with the keywords “cellulose” and “electrospin” have been published during 2018–2021. It is certain that this is an active research area. However, all these reviews focus on either a single compound (*e.g.*, cellulose acetate and nanocellulose)/single solvent system (*e.g.*, ionic liquids) or a single application (*e.g.*, supercapacitors, adsorbents, and wound healing materials). Therefore, this review will provide the researchers with a holistic view of recently reported cellulose-based electrospun nanofibers, including their raw materials (cellulose, cellulose derivatives, and nanocellulose), construction methods (electrospinning solvents and conditions), and applications (water treatment, biomaterials, sensors, electro-conductive materials, active packaging, *etc.*).

2. Cellulose-based electrospun nanofiber construction

Cellulose can be directly electrospun into nanofibers, but the process is limited by the poor solubility of cellulose in most

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Table 1 Typical cellulose-based electrospun nanofibers and their production conditions^a

Raw materials	Solvents	Electrospinning conditions (concentrations, flow rates, voltages, and receiving distances)	Ref.
Wood pulp cellulose	DMAc/LiCl	3 wt%; 1 mL h ⁻¹ ; 15 kV; 15 cm	23
Bacterial cellulose	TFA	2–5 wt%; 0.5 mL h ⁻¹ ; 20 kV; 20 cm	24
Cellulose acetate	Acetic acid	19 wt%; 0.75 mL h ⁻¹ ; 20 kV; 15 cm	25
Cellulose acetate	Acetone/DMF	5 wt%; 35 μ L min ⁻¹ ; 16 kV; 10 cm	26
Cellulose acetate	Acetone/DMAc	16 wt%; 0.3 mL h ⁻¹ ; 11 kV; 15 cm	13
Hydroxypropyl methylcellulose	Ethanol	1–6 wt%; 0.5–1 mL h ⁻¹ ; 10 kV; 15 cm	27
Ethyl(hydroxyethyl) cellulose	DMF	5 wt%; 0.01–3.0 mL h ⁻¹ ; 7–14 kV; 10–20 cm	28
Ethyl(hydroxyethyl) cellulose	THF	1 wt%; 0.2 mL h ⁻¹ ; 27 kV; 15 cm	29
Ethyl cellulose	Ethanol	2 wt%; 0.3 mL h ⁻¹ ; 10–13 kV; 13 cm	30
Aldehyde cellulose	Water	7.5 wt%; 0.01 mL min ⁻¹ ; 10 kV; 20 cm	31
PVA/CNCs	Water	7 wt%; 0.5 mL h ⁻¹ ; 22 kV; 10 cm	32
PLA/CNCs	CHCl ₃ /DMF	10 wt%; 16 kV; 15 cm	33
PVDF/CNCs	Acetone/DMF	12 wt%; 0.02 mL min ⁻¹ ; 12 kV; 18 cm	34
PAN/CNFs	DMF	1–10 wt%; 15 μ L min; 20 kV; 20 cm	35

^a Note: PVA, poly(vinyl alcohol); CNCs, cellulose nanocrystals; PLA, poly(lactic acid); PVDF, polyvinylidene difluoride; PAN, poly(acrylonitrile); CNFs, cellulose nanofibrils; DMAc, dimethylacetamide; TFA, trifluoroacetic acid; DMF, *N,N*-dimethylformamide; THF, tetrahydrofuran.

commonly used solvents. Therefore, many research studies reported the electrospinning of cellulose derivatives. For different raw materials, optimized electrospinning conditions are essential for the successful fabrication of nanofibers. Typical examples of cellulose-based electrospun nanofibers and their production conditions published in the last three years are summarized in Table 1.

2.1 Electrospinning of cellulose

The most effective way is to directly convert cellulose into nanofibers; however, raw materials have to be dissolved or melted for electrospinning. Although several cellulose solvent systems have been developed, such as ionic liquids,³⁶ *N*-methylmorpholine-*N*-oxide (NMMO),³⁷ LiCl/DMAc,³⁸ tetra(*n*-butyl) ammonium hydroxide/dimethyl sulfoxide (TBAH/DMSO),³⁹ alkali/urea aqueous solution,⁴⁰ and sulphuric acid solution,⁴¹ most of them are not suitable for electrospinning due to their high boiling point or high salt concentration. LiCl/DMAc has been proved as an effective solvent, and recent research focused

on improving the current process and looking for new solvent systems. Shu *et al.*²³ reported that wood pulp cellulose nanofibers were successfully prepared by the design of a new electrospinning receiving device and optimization of process parameters in a LiCl/DMAc dissolution system (Fig. 1). Compared to using a sink as the receiver, a novel drum-type receiving device was able to deal with high boiling-point solvents and obtain nanofibers with uniform diameters. The diameters of the resultant nanofibers were in the range of 200–550 nm; the porosity of the nanofiber mat was 76.5%; and there was no LiCl residue in the nanofibers. The breaking strength of the nanofibers could reach 148.2 cN. T. Jayani *et al.*²⁴ developed a new approach to electrospin bacterial cellulose in a TFA/PVA solvent system. Without the addition of PVA as a co-solvent, only beaded particles were collected, and there was no nanofiber formation, irrespective of the solution concentration, voltage, and flow rate. Continuous nanofibers were formed due to the excellent spinnability of PVA and the hydrogen bonding interactions between PVA and bacterial cellulose, which exhibited an average breaking strength of 448 g F, breaking elongation of 10%, surface area of 4.248 m² g⁻¹, average pore volume of 0.005 cm³ g⁻¹, and pore diameter of 1.72 nm.

2.2 Electrospinning of cellulose derivatives

Compared to the poor solubility of cellulose, cellulose derivatives have greatly improved solubility in many organic and inorganic solvents due to changes in the structure of cellulose. Cellulose acetate (CA) is one of the most commonly used cellulose derivatives for the production of electrospun nanofibers because CA can achieve continuous and controllable electrospinning in several conventional solvents, such as acetic acid,²⁵ acetone/DMAc solvent,¹³ and acetone/DMF/water,²⁶ and the obtained CA nanofibers can be easily hydrolysed into cellulose nanofibers. The utilization of other cellulose derivatives has also been reported recently. For example, P. M. Silva *et al.*²⁷ produced electrospun nanofibers from hydroxypropyl

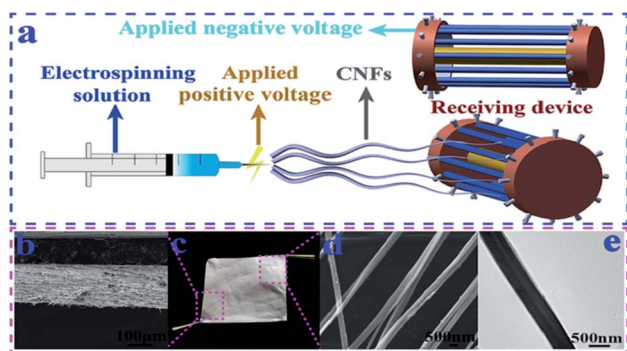


Fig. 1 (a) Improved electrospinning receiving device. (b) SEM image of the cross section of the cellulose nanofiber mat. (c) Photo of a piece of the cellulose nanofiber mat. (d) SEM image of CNFs. (e) TEM image of CNFs (reprinted with permission,²³ Elsevier publishing).



methylcellulose (HPMC) with different molecular weights. High molecular weight HPMC formed beaded fibers at a concentration of 1% (w/v) and smooth fibers when the concentrations were between 1.5 and 2.25% (w/v). However, HPMC with a low molecular weight could not form nanofibers but just particles. Electrospun nanofibers were also fabricated from ethyl cellulose and modified ethyl cellulose in DMF, THF, ethanol solutions for fluorescent materials, drug carriers, and food preservation, respectively, since ethyl cellulose has high versatility, biocompatibility, and excellent mechanical properties.^{28–30} At the same time, it was noticed that some water-soluble cellulose derivatives were employed for electrospinning in order to avoid the use of unfavourable organic solvents. Zhang *et al.*³¹ prepared water-soluble aldehyde cellulose by periodate oxidation, which was capable of forming nanofibers with a smooth surface and uniform size. The nanofiber diameters decreased from 833 nm to 303 nm with the increase of the oxidation degree from 34% to 91%. The prepared fibrous mats showed a moderate wet mechanical strength of around 1 MPa, and were able to absorb water equal to 30 times their own weight. These aldehyde cellulose nanofibers were particularly designed for biomedical fields, such as wound healing and tissue regeneration, and displayed a Murine L929 fibroblast cell viability of higher than 90%, suggesting good biocompatibility. However, their water-solubility limited the applications in high humidity or water environments.

2.3 Electrospinning of nanocellulose

Generally, the term “nanocellulose” encompasses rigid, rod-shaped cellulose nanocrystals (CNCs) and flexible, fiber-like cellulose nanofibrils (CNFs).⁴² Nanocellulose is distinguished by high mechanical strength, structurally configurable chemistry, a high aspect ratio, and biodegradability and has been applied as a sustainable “green” filler in many industrial applications.⁴³ Due to its nanoscale dimensions, nanocellulose can be easily incorporated into electrospun nanofibers by dispersing in spinning solutions rather than dissolving, so as to maintain the nanostructure and special features. It is worth noting that nanocellulose can be modified to be dispersed in different organic and inorganic solutions. Recently, nanocellulose has been combined with various polymers (*i.e.*, PLA,³³ PVDF,³⁴ and PAN³⁵) to prepare composite nanofibers. For example, Zhang *et al.*³² fabricated electrospun PVA/CNC nanofibers for the filtration of particulate matter. The addition of nanocellulose might increase the diameter of nanofibers by interfering with the interactions of the original matrix⁴⁴ or reduce the nanofiber diameter by increasing the charge density of electrospinning solution.³²

3. Cellulose-based electrospun nanofiber applications

3.1 Water treatment

With the rapid development of urbanization and industrialization, wastewater discharge from various sources (agricultural wastes, biomedical industry, food industry, petroleum industry,

manufacturing industry, *etc.*) poses a significant threat to the sustainability of the ecosystem and human health. Cellulose and its derivatives are widely involved in the treatment of contaminated water in the form of electrospun nanofibers, not only due to their intrinsic nanostructures but also owing to their active functional groups and subsequently good surface chemistry. For example, Phan *et al.*⁴⁵ reported electrospun chitosan/cellulose nanofibers for the effective adsorption of heavy metal ions, where arsenic(v), lead(II), and copper(II) were chosen as representatives. It was illustrated that the hydrogen bonding in the structure of cellulose contributed to the high water stability of nanofibers, hindering the swelling behavior of chitosan in water to enable high adsorption capacity.

CA can be easily dissolved in conventional solvents, making it attractive as a carrier for loading other polymers and functional fillers with desired properties. Therefore, CA was of great significance in the field of water treatment in the last three years. For example, to tackle the aggregation issue of hydroxyapatite nanoparticles (a mineral fixator), Hamad *et al.*⁴⁶ impregnated them into the CA matrix through the electrospinning technique. Hydroxyapatite particles with concentrations of up to 3 wt% could be well dispersed in the spinning solution, enabling the improvement of the mechanical strength and adsorption performance of nanofibers. The strong hydrogen bonding interactions between the nanoparticles and CA matrix also contributed to the composite integrity and stability during the adsorption process. Other modifications of electrospun CA nanofibers, such as the addition of thiol groups,⁴⁷ citric acid,⁴⁸ and polyvinylamine,¹³ have also been applied for the removal of Cu(II), Cr(VI), and organoarsenic contaminants, respectively. Photocatalysts such as ZnO⁴⁹ and metal–organic frameworks⁵⁰ are another group of functional fillers that are usually incorporated into electrospun nanofibers for degrading organic contaminants like effluent dye molecules. Besides the most widely used CA, cellulose acetate butyrate was also employed as a low-cost and accessible bearer for the immobilization of photocatalysts, which could provide a smooth and uniform membrane with an average fiber diameter of 1 μm for growing ZnO nanocrystals.⁴⁹

The release of petroleum hydrocarbons from oil spills has serious environmental impacts on marine ecosystems. Cellulose has been electrospun into nanofibers for the application of oil/water separation, delivering extraordinary separation efficiencies of up to 99.5% for the emulsion of vacuum pump oil and water.⁵¹ A water film was generated immediately on the surface of the electrospun cellulose membrane owing to the existence of hydroxyl groups, while the oil droplets were extruded by the membrane due to their relatively larger diameters compared to the porous fiber network (Fig. 2). Electrospun cellulose nanofibers are usually obtained by deacetylating CA fibers.^{52,53} The incorporation of CNCs into a PVDF nanofibrous membrane also contributed to the oil/water separation by forming convex and concave structures on the nanofiber surface.³⁴ The 4 wt% CNC/PVDF membrane showed the highest flux of water-in-toluene emulsion ($5842 \text{ L m}^{-2} \text{ h}^{-1}$) and separation efficiency (97%), which were due to the larger water contact angle along with the increased roughness, higher



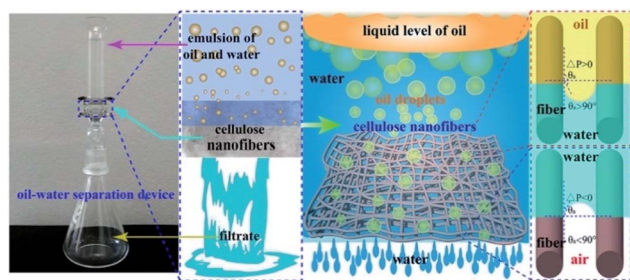


Fig. 2 Illustration of the oil–water separation process and mechanisms by using electrospun cellulose nanofibers (reprinted with permission,⁵¹ Elsevier publishing).

porosity, and larger average pore sizes. However, the water contact angle started to drop when the amount of CNCs increased to 6 wt%, owing to the presence of a high number of hydrophilic hydroxyl groups.

3.2 Biomaterials

Electrospun nanofibers from cellulose and their derivatives have been widely used in the fields of biomedical materials and tissue engineering. For example, the requirements for wound healing materials such as biocompatibility, oxygen permeability, water absorption, and non-toxicity can be fulfilled by electrospun cellulose-based fabrics. Electrospun CA/collagen nanofibers were incorporated with latex (L) from *C. Procera* (medicinal plant) for wound healing, and it was found that CA played an important role in restricting the swelling behaviour and strengthening the fabrics.⁵⁴ Similar electrospun CA nanofibers loaded with gelatin and *Zataria multiflora* (antibacterial plant) could absorb accumulated wound exudates and exhibited good wound healing capacity and antibacterial properties.⁵⁵ A considerably smaller wound area on the burned upper back of rats was observed compared to the commercial gauze treatment after 22 days (Fig. 3). Moreover, electrospun ethyl cellulose⁵⁶ and cellulose diacetate⁵⁷ nanofibers with antibacterial properties were prepared by doping with silver sulfadiazine and protoporphyrin, respectively. The former showed good inhibition activities against *Bacillus* (9.71 ± 1.15 mm) and *E. coli* (12.46 ± 1.31 mm),⁵⁶ while the protoporphyrin IX-embedded cellulose diacetate membranes exhibited 99.8% reduction in Gram-positive *S. aureus* after illumination.⁵⁷

Bioactive compounds have also been loaded into electrospun cellulose-based nanofibers to increase their stability in harsh environments and controlled release at the destinations where they are typically absorbed. Recently, flufenamic acid (an anti-inflammatory drug) was encapsulated in electrospun carboxymethyl cellulose/PVA nanofibers for prolonged drug release.⁵⁸ Covalent bonds between the amine groups of the drug and the carboxyl groups of carboxymethyl cellulose were established to extend the biological half-life of the drug. Electrospun CNC/polycaprolactone nanofibers for the release of tetracycline were reported by Hivechi *et al.*⁵⁹ The presence of CNCs resulted in improved mechanical strength, degradability, and controlled release. The nanofibers were broken in the second week, and

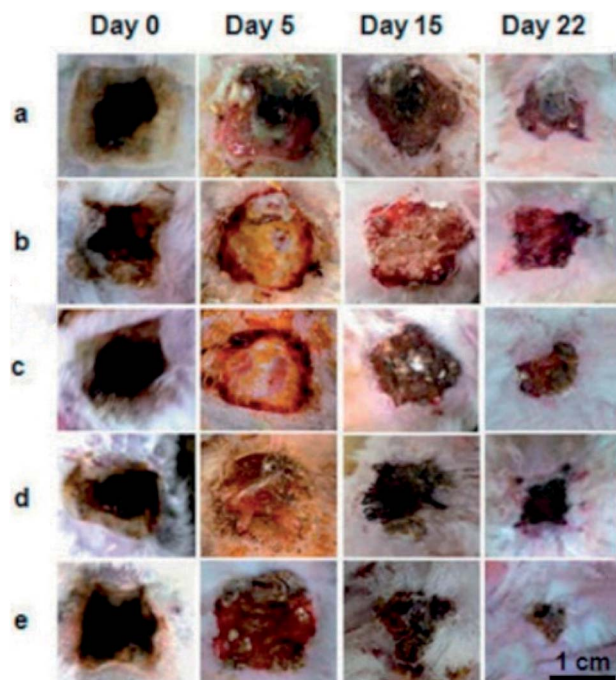


Fig. 3 Representative images of the burn wound healing process treated with (a) commercial gauze, (b) drug-free nanofiber CA/Gel = 100 : 0, (c) drug-free nanofiber CA/Gel = 50 : 50, (d) drug-loaded nanofiber CA/Gel = 100 : 0, and (e) drug-loaded nanofiber CA/Gel = 50 : 50 (reprinted with permission,⁵⁵ Elsevier publishing).

the number of broken fibers increased when the CNC content was up to 1 wt%.

3.3 Sensors

Electrospun nanofibers have a large surface area to volume ratio and excellent pore-interconnectivity, which are particularly important for initiating the signalling pathways for targeted compound detection in a short period of time. Moreover, compared to free nanoparticles, electrospun nonwoven fabrics have fewer safety issues owing to the immobilization of responsive compounds on the fibers, and better handling properties enabling easy application in various devices. Ethyl(-hydroxyethyl)cellulose (EHEC) was functionalized by anchoring 4-(2-(pyridine-4-yl)vinyl)phenol (PBM) and 4-[4-(dimethylamino)styryl]pyridine, and then mixed with poly(methyl methacrylate) (PMMA) to prepare electrospun nanofibers for the detection of CN^- in water.⁶⁰ EHEC endowed the entire matrix with high hydrophilicity even at a low concentration (PMMA/EHEC 20 : 1 wt/wt), so as to have a high water retention capacity for application in fluorogenic optical devices and achieve the detection and quantification limits of 2.15×10^{-5} and $7.17 \times 10^{-5} \text{ mol L}^{-1}$, respectively.

More research work was based on electrospun CA matrices. For instance, core-shell ferrous nanoparticles functionalized with Rhodamine B were embedded into electrospun CA nanofibers for gas ammonia sensing.⁶¹ It was revealed that the encapsulation of $\gamma\text{-Fe}_2\text{O}_3/\text{SiO}_2/\text{Rhodamine B}$ nanoparticles impeded the release of fluorescent molecules to a significant



extent and ensured the stability of sensing materials. Moreover, the good spinnability of CA enabled the loading of a high amount of sensing elements, and the large surface-to-volume ratio of CA nanofibers contributed to a high sensitivity in the response concentration range of 200 ppm to 12 750 ppm and a quick response time (30 s). Jia *et al.*⁶² exploited electrospun CA-based ratiometric fluorescent materials for monitoring the freshness of seafood through ammonia sensing (Fig. 4). The fluorescent unit, protoporphyrin IX, was covalently bonded to CA, which secured food safety by preventing migration. An instant fluorescent color-response (<1 s) of the prepared nanofibrous membrane to a drop of ammonia (concentration as low as 5.0 ppm) was demonstrated. Similar approaches were applied to synthesize CA-based nanofiber sensors for detecting mercury(II) and lead(II) ions, hydrogen chloride vapors, and gas ammonia.^{63,64} All of them exhibited fast detection of targeted substances.

3.4 Electro-conductive materials

A flexible support material with a three-dimensional porous structure is essential for developing conducting electrodes. Wu *et al.*⁶⁵ reported electrospun polyindole/carbon nanotube/bacterial cellulose electrodes for energy storage devices. Its outstanding specific capacitance was ascribed to the three-dimensional hierarchical network formed by bacterial cellulose and polyindole for effective charge transfer. An electrospun polyacrylonitrile/cellulose nanofibril electrode carbon material has been developed³⁵ and displayed improved mechanical strength and electrical conductivity compared to a neat polyacrylonitrile membrane. Cellulose nanofibrils also accelerated the stabilization process by reducing the required energy for nitrile-based cleavage in the carbonization reaction.

An interesting and novel study was reported by Lyu *et al.*,⁶⁶ who utilized the porous structure of electrospun CA fabrics to construct moisture-induced electricity generators. The hydroxyl and carbonyl groups of CA dissociated and formed negatively charged surfaces when the nanofibers were exposed to moisture. Due to the accumulation of moisture on one side of the

fabrics, an imbalanced distribution of protons occurred, and streaming potential was generated consequently (Fig. 5c). The potential was primarily impacted by the porosity and average pore size of electrospun nanofabrics that could be tuned by varying the annealing time. The generation of electricity in response to the change of the moisture content is shown in Fig. 5a and b. Water molecules moved along the 3D nano-channels with charged surfaces, generating a voltage output of up to 0.3 V and a short-circuit current density of 80 nA cm⁻².

3.5 Active packaging

Cellulose and many of its derivatives, such as CMC, CA, ethyl cellulose, *etc.*, have been regarded as safe food substances according to the U.S. Department of Agriculture and considered as food additives by the FDA regulations. Therefore, they are promising raw materials for the preparation of advanced food packaging. Zaitoon *et al.*⁶⁷ fabricated electrospun ethyl cellulose/poly(ethylene oxide) hybrid nanofibers encapsulated with ethyl formate for the preservation of strawberries (Fig. 6). The efficacy was confirmed visually, where the strawberries treated with 2 mg ethyl formate released from electrospun nanofibers were not spoiled for up to 10 days, and the untreated strawberries showed microbial growth on day 8. Nevertheless, the stability of the prepared nanofibers was not good at 100% relative humidity, which might limit its applications under high moisture conditions. Niu *et al.*⁶⁸ incorporated cinnamaldehyde essential oil (CEO) into electrospun zein/ethyl cellulose nanofibers for improved water resistance of food. Hydrogen bonds formed between the hydroxyl groups of ethyl cellulose and the amino groups of zein thus reduced the presence of free hydrophilic groups and improved the water-resistance remarkably, resulting in the controlled release of CEO during the storage period of fresh produce for achieving a prolonged shelf-life. At the same time, Yi *et al.*⁶⁹ constructed electrospun anthraquinone-2-carboxylic acid grafted silk fibroin (G-SF)/CA nanofibers with antibacterial activity against *E. coli*, where CA contributed to the handling properties and the reinforcement of tensile strength from 34.52 cN mm⁻² to 88.92 cN mm⁻².

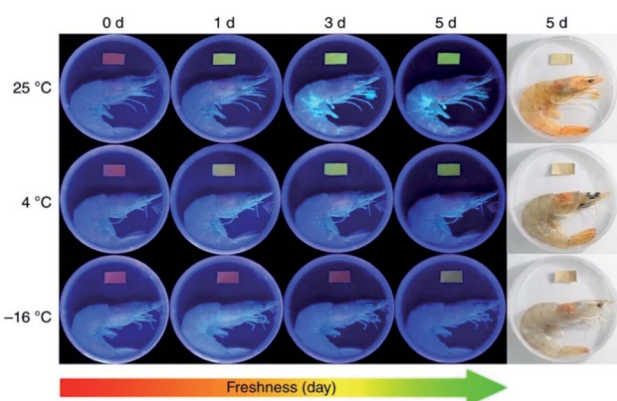


Fig. 4 Electrospun cellulose acetate-based ratiometric fluorescent material with red initial fluorescence for monitoring the freshness of shrimps stored under different conditions (reprinted with permission,⁶² Springer Nature publishing).

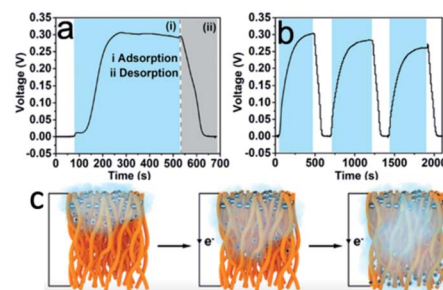


Fig. 5 (a) Electricity generation as a function of moisture change based on the CA nanofiber with an annealing time of 5 min. The two shaded areas correspond to the (i) adsorption (blue zone) and (ii) desorption (grey zone) of moisture. (b) Electricity generation based on the CA nanofiber with an annealing time of 5 min in response to the change of RH ($\Delta RH = 85\%$). (c) Illustration of electrospun cellulose acetate nanofibers exposed to the moist flow (reprinted with permission,⁶⁶ ACS publishing).



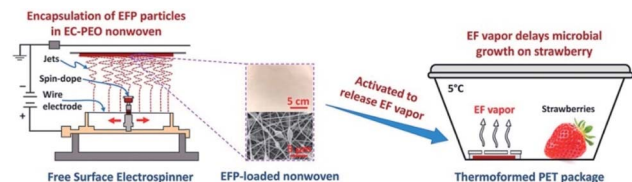


Fig. 6 Ethyl formate-encapsulated electrospun ethyl cellulose/poly(ethylene oxide) nanofibers for strawberry preservation (reprinted with permission,⁶⁷ Elsevier publishing).

3.6 Other applications

Based on the unique structure and properties of electrospun cellulose-based nanofibers, some other applications have been proposed in recent years. An interesting study was reported by Knapczyk-Korczak *et al.*, who fabricated electrospun CA/polystyrene nanofibers for the collection of water from fog (Fig. 7).⁷⁰ Due to the hydrophobicity of the CA/polystyrene matrix, water vapor was repelled from the fiber surface. The existence of CA reduced the roughness and contact angle of nanofibers, allowing an easier and free drainage pathway. Also, CA was proved to strengthen the entire matrix, which is a prominent property of water collectors.

Electrospun cellulose nanofibers have played a role in energy conservation, such as dye-sensitized solar cells⁷¹ and solar thermal applications.⁷² Electrospun CA and deacetylated CA nanofibers were used as electrolyte membranes in dye-sensitized solar cells, which showed higher open circuit voltages (780 ± 10 mV and 780 ± 10 mV) compared to the reference cell (771 ± 9 mV).⁷¹ It was attributed to the more uniform distribution of the electrolyte within the electrospun membranes in contact with the electrode interface. At the same time, electrospun CA nanofibers loaded with thermal conductive cadmium selenide nanoparticles displayed high solar-thermal conversion potential and efficiency, which reached 40°C under 1 sun (100 mW cm^{-2}) illumination.⁷²

Besides water treatment, electrospun nanofibers were also applied for air purification.⁷³ With the aim of toluene (volatile organic compound) adsorption, CNCs were filled in electrospun polyamide 6 nanofibers to improve adsorption efficiency and mechanical strength, owing to their high hydrophilicity, large active surface area, and crystalline structure.⁷⁴ In the meantime,

electrospun PAN-based activated carbon and PVA nanofibers incorporated with CNCs were used for the adsorption of cyclohexane⁷⁵ and filtration of particular matter.^{32,76} It was worth noting that various other functional fillers, such as silica nanoparticles,⁷⁷ silica zeolites,⁷⁸ iron oxide nanoparticles,⁷⁹ and $\text{Ti}_3\text{C}_2\text{T}_x$ (metal carbide),⁸⁰ were doped in cellulose-based nanofibers for desalination, adsorption of ammonia vapor, tunable saturation magnetization, and electromagnetic interference shielding, respectively.

4. Conclusion and future prospects

Recent progress in cellulose-based electrospun nanofiber production and applications is summarized. In one aspect, research work focused on exploring new cellulose solvents and cellulose derivatives that are suitable for electrospinning and improving electrospinning apparatus for better fiber quality; in another, various applications in water treatment, biomaterials, sensors, electro-conductive materials, active packaging, and so on have been proposed to make full use of these renewable, biodegradable and nontoxic nanofibrous networks with a large surface area and interconnected porous structure. Different forms of cellulose (wood pulp, nanocellulose, *etc.*) and derivatives (CA, hydroxypropyl methylcellulose, carboxymethyl cellulose, azido-cellulose, aldehyde cellulose, ethyl cellulose, *etc.*) have been employed as raw materials for electrospinning. However, the optimized electrospinning conditions of all these materials have relatively low polymer concentrations in spinning solutions and low flow rates during spinning, resulting in low productivity. Therefore, future research on new solvents and cellulose derivatives is still required to address these issues and improve productivity. It is worth noting that the electrospinning technology improved largely in the past 15 years; for example, advanced electrospinning apparatuses allow the production of nanofabrics with multiple structures (aligned, multi-layered, composite, *etc.*). However, current research work mainly focuses on the formulations and electrospinning conditions of nanofibers, and not many novel structures have been reported. Hence, the rational design of electrospun nanofibers with better performance is expected in future research, which should be based on the understanding of the structure–property relationship and linked tightly with the targeted application.

Author contributions

Yirong Zhang: writing-original draft preparation, Cunzhi Zhang: writing-original draft preparation, and Yixiang Wang: supervision, writing-reviewing and editing.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

We would like to acknowledge financial support from the Fonds de Recherche du Quebec-Nature et Technologies (266796),

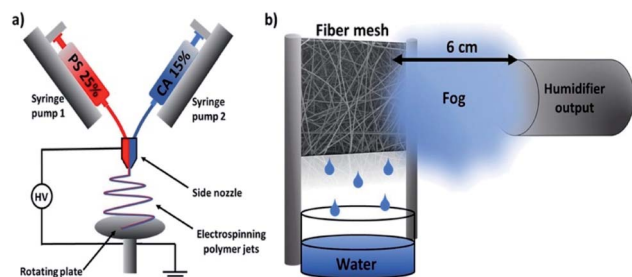


Fig. 7 Schematic diagrams of electrospun cellulose acetate/poly-styrene nanofibers for water harvesting (reprinted with permission,⁷⁰ ACS publishing).



Natural Sciences and Engineering Research Council of Canada (250374), Natural Sciences and Engineering Research Council of Canada Discovery Launch Supplement (250531), and McGill University Academic Startup Grant (130209). C. Z. would like to thank the China Scholarship Council (CSC No. 202006150146) for financial support.

References

- 1 J. Xue, T. Wu, Y. Dai and Y. Xia, *Chem. Rev.*, 2019, **119**, 5298–5415.
- 2 Y. Zhang, F. Wang and Y. Wang, *Mater. Today Commun.*, 2021, **27**, 102272.
- 3 S. Lee, S. Franklin, F. A. Hassani, T. Yokota, M. O. G. Nayeem, Y. Wang, R. Leib, G. Cheng, D. W. Franklin and T. Someya, *Science*, 2020, **370**, 966–970.
- 4 X. Sun, L. Bai, J. Li, L. Huang, H. Sun and X. Gao, *Carbon*, 2021, **182**, 11–22.
- 5 J. Yan, Y. Wang, Y. Zhang, S. Xia, J. Yu and B. Ding, *Adv. Mater.*, 2021, **33**, e2007525.
- 6 K. S. Han, S. Lee, M. Kim, P. Park, M. H. Lee and J. Nah, *Adv. Funct. Mater.*, 2019, **29**, 1903633.
- 7 C. Chen, W. Zhang, H. Zhu, B.-G. Li, Y. Lu and S. Zhu, *Nano Res.*, 2020, **14**, 1465–1470.
- 8 Q. Xia, C. Chen, Y. Yao, J. Li, S. He, Y. Zhou, T. Li, X. Pan, Y. Yao and L. Hu, *Nat. Sustainability*, 2021, **4**, 627–635.
- 9 M. Beaumont, P. Jusner, N. Gierlinger, A. W. T. King, A. Potthast, O. J. Rojas and T. Rosenau, *Nat. Commun.*, 2021, **12**, 2513.
- 10 M. C. Li, Q. Wu, R. J. Moon, M. A. Hubbe and M. J. Bortner, *Adv. Mater.*, 2021, **33**, e2006052.
- 11 R. M. O'Dea, J. A. Willie and T. H. Epps, *ACS Macro Lett.*, 2020, **9**, 476–493.
- 12 S. R. Djafari Petroudy, S. Arjmand Kahagh and E. Vatankhah, *Carbohydr. Polym.*, 2021, **251**, 117087.
- 13 K. Liu, Z. Huang, J. Dai, Y. Jiang, G. Yang, Y. Liu, C. Lin, Y. Lv and M. Liu, *Chem. Eng. J.*, 2020, **382**, 122775.
- 14 Q. Cao, M. Zhu, J. Chen, Y. Song, Y. Li and J. Zhou, *ACS Appl. Mater. Interfaces*, 2020, **12**, 1210–1221.
- 15 T. Pirzada, Z. Ashrafi, W. Xie and S. A. Khan, *Adv. Funct. Mater.*, 2019, **30**, 1907359.
- 16 S. Hell, K. Ohkawa, H. Amer, A. Potthast and T. Rosenau, *Nanomaterials*, 2020, **10**, 671.
- 17 X. Miao, J. Lin and F. Bian, *J. Bioresour. Bioprod.*, 2020, **5**, 26–36.
- 18 N. Tang, Y. Li, J. Ge, Y. Si, J. Yu, X. Yin and B. Ding, *ACS Appl. Mater. Interfaces*, 2020, **12**, 31852–31862.
- 19 S. Zheng, M. Du, W. Miao, D. Wang, Z. Zhu, Y. Tian and L. Jiang, *Adv. Funct. Mater.*, 2018, **28**, 1800832.
- 20 M. A. Teixeira, M. C. Paiva, M. T. P. Amorim and A. H. P. Felgueiras, *Nanomaterials*, 2020, **10**, 557.
- 21 T. Lu, J. Cui, Q. Qu, Y. Wang, J. Zhang, R. Xiong, W. Ma and C. Huang, *ACS Appl. Mater. Interfaces*, 2021, **13**, 23293–23313.
- 22 M. Zhang, W. Ma, J. Cui, S. Wu, J. Han, Y. Zou and C. Huang, *J. Hazard. Mater.*, 2020, **383**, 121152.
- 23 D. Shu, P. Xi, B. Cheng, Y. Wang, L. Yang, X. Wang and X. Yan, *Int. J. Biol. Macromol.*, 2020, **162**, 1536–1545.
- 24 T. Jayani, B. Sanjeev, S. Marimuthu and S. Uthandi, *Carbohydr. Polym.*, 2020, **250**, 116965.
- 25 N. Olaru, N. Anghel, P. Pascariu and G. Ailiesei, *J. Appl. Polym. Sci.*, 2019, **136**, 47772.
- 26 W. Chen, H. Ma and B. Xing, *Int. J. Biol. Macromol.*, 2020, **158**, 1342–1351.
- 27 P. M. Silva, C. Prieto, J. M. Lagarón, L. M. Pastrana, M. A. Coimbra, A. A. Vicente and M. A. Cerqueira, *Food Hydrocolloids*, 2021, **118**, 106761.
- 28 J. P. Dreyer, R. I. Stock, L. G. Nandi, I. C. Bellettini and V. G. Machado, *Carbohydr. Polym.*, 2020, **236**, 115991.
- 29 A. Wali, Y. Zhang, P. Sengupta, Y. Higaki, A. Takahara and M. V. Badiger, *Carbohydr. Polym.*, 2018, **181**, 175–182.
- 30 B. Niu, L. Zhan, P. Shao, N. Xiang, P. Sun, H. Chen and H. Gao, *Int. J. Biol. Macromol.*, 2020, **142**, 592–599.
- 31 H. Zhang, Y. Liu, S. Cui, Y. Zhou, J. Hu, J. Ma and Y. Liu, *Cellulose*, 2020, **27**, 8695–8708.
- 32 Q. Zhang, Q. Li, T. M. Young, D. P. Harper and S. Wang, *ACS Sustainable Chem. Eng.*, 2019, **7**, 8706–8714.
- 33 D. K. Patel, S. D. Dutta, J. Hexiu, K. Ganguly and K. T. Lim, *Int. J. Biol. Macromol.*, 2020, **162**, 1429–1441.
- 34 X. Wang, W. Cheng, D. Wang, X. Ni and G. Han, *J. Membr. Sci.*, 2019, **575**, 71–79.
- 35 W. Xu, B. Xin and X. Yang, *Cellulose*, 2020, **27**, 3789–3804.
- 36 B. Araldi da Silva, R. de Sousa Cunha, A. Valério, A. De Noni Junior, D. Hotza and S. Y. Gómez González, *Eur. Polym. J.*, 2021, **147**, 110283.
- 37 R. Protz, A. Lehmann, J. Ganster and H. P. Fink, *Carbohydr. Polym.*, 2021, **251**, 117027.
- 38 J. Garemark, X. Yang, X. Sheng, O. Cheung, L. Sun, L. A. Berglund and Y. Li, *ACS Nano*, 2020, **14**, 7111–7120.
- 39 X. Chen, X. Chen, X.-M. Cai, S. Huang and F. Wang, *ACS Sustainable Chem. Eng.*, 2018, **6**, 2898–2904.
- 40 H. Tu, M. Zhu, B. Duan and L. Zhang, *Adv. Mater.*, 2021, **33**, 2000682.
- 41 C. Zhou and Y. Wang, *J. Appl. Polym. Sci.*, 2021, **138**, 51255.
- 42 N. Peng, D. Huang, C. Gong, Y. Wang, J. Zhou and C. Chang, *ACS Nano*, 2020, **14**, 16169–16179.
- 43 M. E. Pasaoglu and I. Koyuncu, *Chemosphere*, 2021, **269**, 128710.
- 44 Y. Wang and L. Chen, *ACS Appl. Mater. Interfaces*, 2014, **6**, 1709–1718.
- 45 D.-N. Phan, H. Lee, B. Huang, Y. Mukai and I.-S. Kim, *Cellulose*, 2019, **26**, 1781–1793.
- 46 A. A. Hamad, M. S. Hassouna, T. I. Shalaby, M. F. Elkady, M. A. Abd Elkawi and H. A. Hamad, *Int. J. Biol. Macromol.*, 2020, **151**, 1299–1313.
- 47 H. Y. Choi, J. H. Bae, Y. Hasegawa, S. An, I. S. Kim, H. Lee and M. Kim, *Carbohydr. Polym.*, 2020, **234**, 115881.
- 48 D. Zhang, W. Xu, J. Cai, S.-Y. Cheng and W.-P. Ding, *Int. J. Biol. Macromol.*, 2020, **149**, 459–466.
- 49 P. Pascariu, N. Olaru, A. Rotaru and A. Airinei, *Nanomaterials*, 2020, **10**, 1873.
- 50 W. Lu, C. Duan, Y. Zhang, K. Gao, L. Dai, M. Shen, W. Wang, J. Wang and Y. Ni, *Carbohydr. Polym.*, 2021, **258**, 117676.
- 51 D. Shu, P. Xi, B. Cheng, Y. Wang, L. Yang, X. Wang and X. Yan, *Int. J. Biol. Macromol.*, 2020, **162**, 1536–1545.



- 52 H. P. Karki, L. Kafle, D. P. Ojha, J. H. Song and H. J. Kim, *Sep. Purif. Technol.*, 2019, **210**, 913–919.
- 53 W. Wang, J. Lin, J. Cheng, Z. Cui, J. Si, Q. Wang, X. Peng and L.-S. Turng, *J. Hazard. Mater.*, 2020, **385**, 121582.
- 54 G. Ramanathan, L. S. Seleenmary Sobhanadhas, G. F. Sekar Jeyakumar, V. Devi, U. T. Sivagnanam and P. Fardim, *Biomacromolecules*, 2020, **21**, 2512–2524.
- 55 H. Farahani, A. Barati, M. Arjomandzadegan and E. Vatankhah, *Int. J. Biol. Macromol.*, 2020, **162**, 762–773.
- 56 S. Ahmadian, M. Ghorbani and F. Mahmoodzadeh, *Int. J. Biol. Macromol.*, 2020, **162**, 1555–1565.
- 57 T. Wang, H. Ke, S. Chen, J. Wang, W. Yang, X. Cao, J. Liu, Q. Wei, R. A. Ghiladi and Q. Wang, *Mater. Sci. Eng., C*, 2021, **118**, 111502.
- 58 A. Allafchian, H. Hosseini and S. M. Ghoreishi, *Int. J. Biol. Macromol.*, 2020, **163**, 1780–1786.
- 59 A. Hivechi, S. H. Bahrami and R. A. Siegel, *Mater. Sci. Eng., C*, 2019, **94**, 929–937.
- 60 J. P. Dreyer, R. I. Stock, L. G. Nandi, I. C. Bellettini and V. G. Machado, *Carbohydr. Polym.*, 2020, **236**, 115991.
- 61 A. Petropoulou, S. Kralj, X. Karagiorgis, I. Savva, E. Loizides, M. Panagi, T. Krasia-Christoforou and C. Riziotis, *Sci. Rep.*, 2020, **10**, 367.
- 62 R. Jia, W. Tian, H. Bai, J. Zhang, S. Wang and J. Zhang, *Nat. Commun.*, 2019, **10**, 795.
- 63 S. Ahmadian-Fard-Fini, D. Ghanbari, O. Amiri and M. Salavati-Niasari, *Carbohydr. Polym.*, 2020, **229**, 115428.
- 64 M. Kim, H. Lee, M. Kim and Y. C. Park, *Nanomaterials*, 2021, **11**, 222.
- 65 J. Wu, Z. Du, P. Xiong and Z. Cai, *Cellulose*, 2020, **27**, 6353–6366.
- 66 Q. Lyu, B. Peng, Z. Xie, S. Du, L. Zhang and J. Zhu, *ACS Appl. Mater. Interfaces*, 2020, **12**, 57373–57381.
- 67 A. Zaitoon, L.-T. Lim and C. Scott-Dupree, *Food Hydrocolloids*, 2021, **112**, 106313.
- 68 B. Niu, L. Zhan, P. Shao, N. Xiang, P. Sun, H. Chen and H. Gao, *Int. J. Biol. Macromol.*, 2020, **142**, 592–599.
- 69 S. Yi, Y. Wu, Y. Zhang, Y. Zou, F. Dai and Y. Si, *ACS Sustainable Chem. Eng.*, 2020, **8**, 16775–16780.
- 70 J. Knapczyk-Korczak, J. Zhu, D. P. Ura, P. K. Szewczyk, A. Gruszczyński, L. Benker, S. Agarwal and U. Stachewicz, *ACS Sustainable Chem. Eng.*, 2021, **9**, 180–188.
- 71 J. J. Kaschuk, K. Miettunen, M. Borghei, E. Frollini and O. J. Rojas, *Cellulose*, 2019, **26**, 6151–6163.
- 72 N. Angel, S. N. Vijayaraghavan, F. Yan and L. Kong, *Nanomaterials*, 2020, **10**, 1329.
- 73 D. Lv, G. Tang, L. Chen, M. Zhang, J. Cui, R. Xiong and C. Huang, *ACS Appl. Polym. Mater.*, 2020, **2**, 5686–5697.
- 74 E. Buyukada-Kesici, E. Gezmis-Yavuz, D. Aydin, C. E. Cansoy, K. Alp and D. Y. Koseoglu-Imer, *Mater. Sci. Eng., B*, 2021, **264**, 114953.
- 75 R. Awad, A. Haghighat Mamaghani, Y. Boluk and Z. Hashisho, *Chem. Eng. J.*, 2021, **410**, 128412.
- 76 J. Cui, Y. Wang, T. Lu, K. Liu and C. Huang, *J. Colloid Interface Sci.*, 2021, **597**, 48–55.
- 77 N. Dizge, E. Shaulsky and V. Karanikola, *J. Membr. Sci.*, 2019, **590**, 117271.
- 78 A. Ojstršek, D. Fakin, S. Hribernik, T. Fakin, M. Bračič and M. Kurečič, *Carbohydr. Polym.*, 2020, **236**, 116071.
- 79 G. Papaparaskeva, M. M. Dinev, T. Krasia-Christoforou, R. Turcu, S. A. Porav, F. Balanean and V. Socoliuc, *Nanomaterials*, 2020, **10**, 517.
- 80 C. Cui, C. Xiang, L. Geng, X. Lai, R. Guo, Y. Zhang, H. Xiao, J. Lan, S. Lin and S. Jiang, *J. Alloys Compd.*, 2019, **788**, 1246–1255.

