

RESEARCH ARTICLE

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Cite this: *Inorg. Chem. Front.*, 2020, 7, 4062

A bioorthogonal time-resolved luminogenic probe for metabolic labelling and imaging of glycans†

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The use of bioorthogonal fluorogenic probes is superior to labelling and imaging of biomolecules in live cells and organisms, although overcoming the limitation of autofluorescence is still a challenge for current probes to achieve high illumination resolution of the target of interest. We herein demonstrate a functionalized terbium complex **Tb-1** that is stable and biocompatible to enable bioorthogonal ligation with engineered cell-surface glycans for providing responsive luminescence. A luminescence resonance energy transfer (LRET) quencher with bioorthogonal properties is strategically incorporated into a tripodal terbium complex with low toxicity, which can undergo a click-cycloaddition reaction with a cyclooctene to completely change the electronic structure of the quencher, resulting in a much less efficient LRET but a 5-fold enhancement in the long-lived terbium emission intensity. This work therefore establishes a time-resolved platform that enables labelling and imaging of the biomolecules of interest.

Received 21st June 2020,
Accepted 31st August 2020

DOI: 10.1039/d0qi00728e

rsc.li/frontiers-inorganic

Introduction

Bioorthogonal chemistry, pioneered by Prescher and Bertozzi, refers to chemical reactions within cells that do not affect the physiological processes in place.^{1,2} The ideal visualization of various biological processes taking place in living systems should be performed without any interference from the imaging probes. By metabolic engineering,³ a chemical reporter, which is usually a small functional group that causes minimal structural changes to the substrate and does not interfere with its functions, can be incorporated into the target substrate, such as a glycan, in its biosynthetic pathways.⁴ Glycans existing mostly in glycoproteins or glycolipids *via* metabolic engineering play vital roles in various biological events.^{5–8} Subsequently, a fluorogenic labeling or an imaging probe carrying another functional group capable of reacting specifically with the chemical reporter would be introduced,

and the cell surface glycans can be labeled without any interference or distress to the physiological environment.⁹ There are reports of various types of fluorogenic bioorthogonal reporters,^{10,11} such as tetrazine,^{12,13} azide,^{14,15} alkenes,^{16,17} cycloalkynes^{18,19} and cyclopropenes,²⁰ that exhibit specific and click reactions to perform bioorthogonal ligation. However, the major drawback associated with conventional fluorescent dyes lies in the interference of autofluorescence.

Various imaging techniques have been utilized in the imaging of glycans, including stochastic optical reconstruction microscopy (STORM) imaging,²¹ magnetic resonance (MR) imaging²² and fluorescence imaging.^{23–26} The development of both bioorthogonal chemical reporters and labelling agents has gathered pace in the past decade as researchers hope to increase their specificity and reactivity by introducing new pairs of bioorthogonal conjugators and to gain better resolution in the visualization of the substrate.

Trivalent lanthanides, in particular europium(III) and terbium(III), have been commonly used as emission centers due to their characteristic emission profiles and long emission lifetimes (μs–ms time frame).^{27–30} The fingerprint emission bands allow differentiation from the broad and structureless emission bands of organic chromophores and convincingly distinguish lanthanide emissions from short-lived biological autofluorescence (ns-based) using time-resolved measurements.^{31–33} The technique of time-resolved spectroscopy makes use of a time-gated system to delay data acquisition of pulse-excited emission signals so that the detection window is only opened after the unwanted emissions have faded, and the desired longer-lived emission(s) are distinctly

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†Electronic supplementary information (ESI) available. See DOI: 10.1039/d0qi00728e

detected.^{34–36} The lanthanide probes have gained great momentum for application in biological detection and imaging, especially the “switch-off” strategy based on the principle of photoinduced electron transfer or antenna-analyte electron transfer.^{27,37} Recently, lanthanide complexes incorporated with several functional groups have also been covalently linked to metabolically engineered substrates *via* bioorthogonal ligation to perform time-resolved imaging and labelling of cell surface glycans in order to obtain more information on how glycans regulate progression.^{38–40} However, these complexes with “always-on” luminescence signals suffer from the interference of unremoved complexes. Therefore, to obtain high signal/background (S/B) ratios, the development of a luminogenic lanthanide probe that can be activated by a bioorthogonal reaction is highly desirable. Furthermore, time-resolved luminescence spectroscopy allows the probing of the excited state dynamics and deactivation pathways of our designed imaging complexes, offering complementary evidence for our bioorthogonal ligation mechanism.

To improve the imaging efficacy *via* combining bioorthogonal chemistry with lanthanide emission, herein, we report the design and synthesis of a luminogenic terbium(III) probe (**Tb-1**, Scheme 1). **Tb-1** was found to be activated *via* a bioorthogonal tetrazine ligation and exhibited dramatic enhancement of long-lived luminescence (5-fold) upon cycloaddition reaction with a strained *trans*-cyclooctene (TCO). Notably, **Tb-1** could enable robust and sensitive detection of cyclooctene-modified glycome under no-wash conditions in living cells and zebrafish.

Results and discussion

As the absorption of tetrazine overlaps well with the emission of Tb³⁺ (Fig. S1†), we rationally hypothesized that the tetrazine-functional group could quench the luminescence of terbium *via* luminescence resonance energy transfer (LRET) between the terbium compound and the tetrazine moiety. To confirm the hypothesis, we also synthesized and characterized a reference complex **Tb-M** without the tetrazine moiety together with the probe **Tb-1** based on a 7-substituted quinolinone antenna (Fig. 1 and Fig. S2–S13†).

Typically, **Tb-M** and **Tb-1** exhibited a similar quinolinone absorption band, located at 348 and 346 nm, respectively

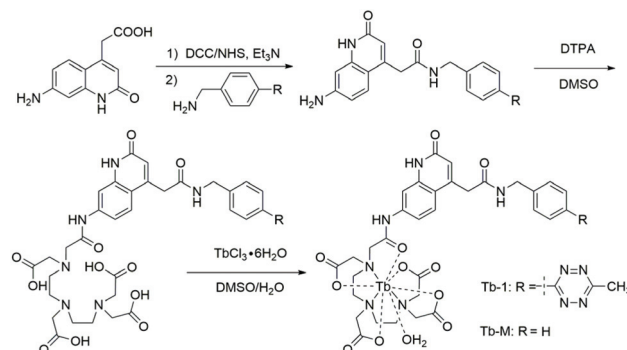


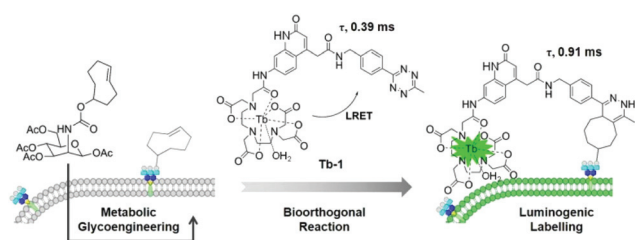
Fig. 1 The synthesis route to **Tb-M** and **Tb-1**.

Table 1 Photophysical parameters of the Tb(III) complexes

	λ_{abs}^a / nm	ϵ^b / M ⁻¹ cm ⁻¹	$\tau(\text{H}_2\text{O})^c$ / ms	$\tau(\text{D}_2\text{O})^c$ / ms	$\Phi^d/\%$
Tb-M	348	12 113	1.06	1.32	10.7 ± 0.6
Tb-1	346, 522	10 188, 1132	0.39	0.89	1.0 ± 0.8
Tb-1 + TCO ^e	347	10 312	0.91	—	9.0 ± 0.5

^a Measured in DI H₂O. ^b Absorption coefficient, 298 K. ^c Tb emission decay ($\lambda_{\text{em}} = 545$ nm, $^5\text{D}_4 \rightarrow ^7\text{F}_5$, $\lambda_{\text{ex}} = 340$ nm, 5 mM); hydration number of Tb(III) complexes, $q = 1.2 \times [k(\text{H}_2\text{O}) - k(\text{D}_2\text{O}) - 0.25]$, $k = \tau^{-1}$.^{41,42} ^d Overall Tb emission quantum yield in H₂O, determined by integrating sphere.⁴³ ^e With the addition of 20 equivalents TCO.

(Table 1). Unlike the colorless **Tb-M**, the pale red **Tb-1** exhibited an absorption band centered at 522 nm,^{44,45} which is attributed to the tetrazine moiety (Table 1). On the other hand, there were no energy transfer pathways between the tetrazine moiety and antenna ligand (Fig. S14†). The emission quantum yield ($\lambda_{\text{ex}} = 340$ nm) of **Tb-1** showed a dramatic difference to that of **Tb-M**. **Tb-M** exhibited a strong green emission with a quantum yield of 10.7% in aqueous solution, while **Tb-1** showed only very weak Tb emission under identical experimental conditions ($\Phi = 1.0\%$). The room temperature emission spectra of **Tb-M** and **Tb-1** under the same excitation wavelength as the antenna ligand are given in Fig. S15†, which shows the characteristic $^5\text{D}_4 \rightarrow ^7\text{F}_j$ ($J = 3, 4, 5, 6$) transitions of Tb³⁺. The ratios of the $^5\text{D}_4 \rightarrow ^7\text{F}_j$ emissions from **Tb-M** and **Tb-1** indicate that both complexes have similar coordination geometry. As for **Tb-1**, there were two steps in the energy transfer pathways including the energy transferred from antenna ligands to Tb³⁺, and further from Tb³⁺ to tetrazine. On the basis of the reported methods,^{29,46} the sensitization and LRET efficiency of these was obtained as 63.21% and 46.5%, respectively. By measuring the phosphorescence spectra of **La-H** and **La-1** obtained at 77 K, we found that the two complexes exhibited a similar phosphorescence band of the antenna ligand at 445 nm (Fig. S16†), indicating that the energy level of the T₁ excited states of the antenna chromophores are around 22 472 cm⁻¹ and fall in the optimum energy transfer range of the $^5\text{D}_4$ excited state of Tb³⁺ (20 400 cm⁻¹). The quenched Tb³⁺ emission in **Tb-1** is thereby attributed to LRET energy transfer



Scheme 1 Schematic illustration of the bioorthogonal time-resolved luminogenic probe **Tb-1** for metabolic labelling and imaging of glycans.

from the excited Tb^{3+} state to the excited state of tetrazine (19 157 cm^{-1}). Once the tetrazine binds with TCO, the excited state level of tetrazine–TCO would be higher than that of tetrazine, which could block the LRET process, resulting in enhancement of the Tb^{3+} luminescence emission. And the proposed energy transfer mechanism of **Tb-1** is summarized in Fig. S17†.

Considering that the cycloadduct of tetrazine has almost no absorption in the region of interest, the LRET degree should be controlled by the bioorthogonal reaction of tetrazine and TCO (Fig. 2a). The absorption of **Tb-1** in the range of 500–600 nm was then found to disappear upon titration, and simultaneously, the solution turned from red to colorless, indicating the loss of the tetrazine adduct, further confirming that the quenching of luminescence was attributed to LRET (Fig. 2b and Fig. S18†). The time-gated spectra of **Tb-1** upon titrating with TCO clearly show that the probe responded well and achieved a 5-fold increase in luminescence intensity (Fig. 2c). The luminescence lifetime was extended from 0.39 to 0.91 ms in PBS buffer solution, which is sufficiently long to eliminate background fluorescence (Fig. 2d, Table 1 and Fig. S19†) and the number of metal-bound water molecules in solution can be calculated to be one *via* reported methods, which was further confirmed by the Fourier transform infrared (FTIR) spectrum, demonstrating that there is no difference in

solvent quenching (Fig. S20†).⁴⁷ A linear relationship between the emission intensity at 545 nm and the concentration of TCO from 0–4 μM was observed, and the response limit of **Tb-1** for TCO in solution was then determined to be 19.6 nM by means of the reported method (Fig. S21†).⁴⁸

Next, the sensing of **Tb-1** towards TCO was measured in a physiological environment with the pH ranging from 8.0 to 5.0 by monitoring the Tb^{3+} luminescence (Fig. 2e). The results showed that **Tb-1** showed excellent responsiveness to TCO in such a physiological environment, despite the fact that pH-dependent emission was observed below 5.0 and over 8.0 due to the protonation and hydrolysis of the organic antenna.⁴⁹ Meanwhile, the biostability of **Tb-1** was also evaluated by measuring its luminescence in PBS solution under various conditions: PBS solution containing different serum concentrations (from 0 to 50%) were kept for different times (from 1 to 24 h). Persistent luminescence of the **Tb-1** under these conditions was observed (Fig. S22†), indicating its high stability in the simulated physiological environments. Besides, the interference from unsaturated fatty acids including oleic acid or linoleic acid was studied by recording the luminescence intensity changes. And only the addition of TCO resulted in a prominent increase in the luminescence emission intensity within 60 min (Fig. S23†), which was because of the faster kinetics of reaction between tetrazine and TCO ($3910 \text{ M}^{-1} \text{ s}^{-1}$, Fig. S24†) compared with the tetrazine and alkene.⁵⁰ These performances strongly suggest that the bio-stable probe **Tb-1** could be used as a preferable tool with higher sensitivity and a larger turn-on luminescence response to TCO.

Having shown that **Tb-1** responds well to TCO *in vitro*, we then evaluated the capability of the probe in the labelling of specific biomolecules, glycans, in the living cells. The cytotoxicity of **Tb-1** was firstly tested on different cell lines (such as A549 and EMT 6 or LO2 cells) by using the standard cell counting kit 8 (CCK-8) assays (Fig. S25†). The complex appeared to be quite nontoxic, with IC_{50} values of *ca.* 1 mmol: a level far above the normal working concentration of a probe. Then, A549 cells were treated with 50 μM peracetylated *N*-TCO-mannosamine ($\text{Ac}_4\text{ManNTCO}$) that would be metabolically converted into the corresponding alkynylsialic acid bearing TCO groups on the cell surface for 72 h. Control cells were incubated with peracetylated mannosamine ($\text{Ac}_4\text{ManNH}_2$). After this, A549 cells pretreated with the $\text{Ac}_4\text{ManNTCO}$ or $\text{Ac}_4\text{ManNH}_2$ were incubated with **Tb-1** for 3 h, and then imaging by fluorescence microscopy was performed without washing. The two-photon photophysical properties of some quinolinone-sensitized Tb complexes have been studied previously, further providing a means of reducing the interference from other biological matter by utilizing low energy near infrared (NIR) excitation.^{51–54} Two-photon excitation of **Tb-1** with a NIR laser can also induce the typical **Tb-1** emission in the presence of TCO at 298 K (Fig. S26†). Compared with Fig. 2c, a broad band arising from the intraligand transition is observed, and this band overlaps with the standard set of well-resolved narrow emission bands owing to the Tb^{3+} centered transitions due to the quinolinone fluorescence or phosphorescence at

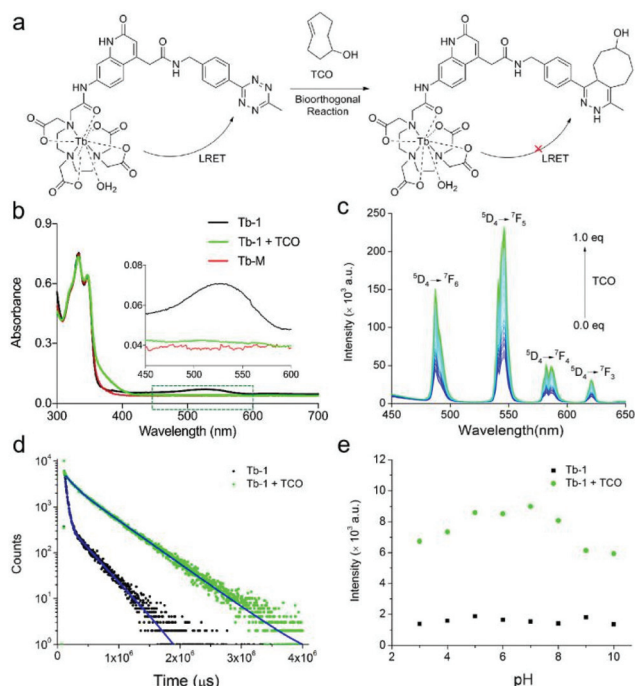


Fig. 2 (a) Schematic of sensing bioorthogonal reaction of luminogenic **Tb-1** with TCO based on the LRET mechanism. (b) The absorbance spectra of **Tb-M**, and **Tb-1** in the absence or presence of TCO. (c) Time-gated luminescence spectra of **Tb-1** titrating with TCO ($\lambda_{\text{ex}} = 340 \text{ nm}$). (d) Decay time curves of the $^5\text{D}_4 \rightarrow ^7\text{F}_5$ emission of Tb^{3+} (10 μM) in the absence and presence of TCO (20 μM). (e) The luminescence intensity ($\lambda_{\text{em}} = 545 \text{ nm}$) of complex **Tb-1** reacted with TCO in various pH buffers.

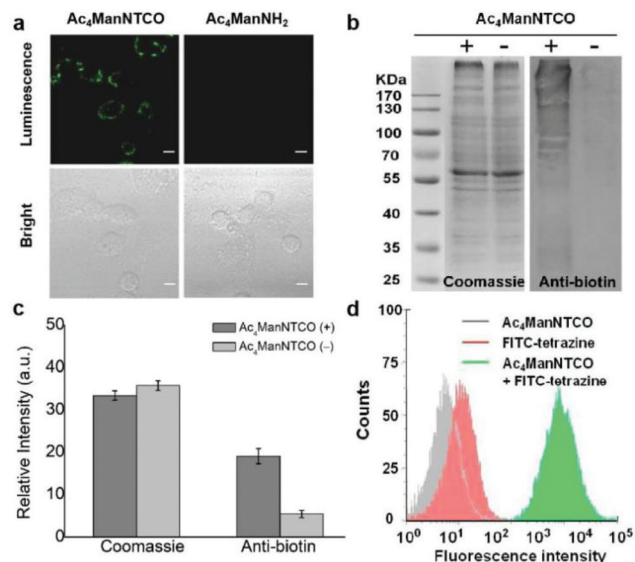


Fig. 3 (a) Two-photon luminescence images of A549 cells after treatment with Ac₄ManNTCO or Ac₄ManNH₂ (50 μM) for three days and further incubation with Tb-1 (50 μM) for 3 h. (b) Western blot analysis of lysates from A549 cells treated with Ac₄ManNTCO or Ac₄ManNH₂ (50 μM). (c) Corresponding quantified intensity from (b). (d) Flow cytometry analysis of A549 cells pretreated with Ac₄ManNTCO or not, followed by incubation with FITC-tetrazine.

298 K, which would only be partially quenched in the emission spectrum recorded at 77 K.⁴⁷ As shown in Fig. 3a, A549 cells treated with Ac₄ManNTCO and Tb-1 displayed a strong luminescence on the cell surface as observed by two-photon ($\lambda_{\text{ex}} = 730$ nm) confocal microscopy, while the control cells labelled with Ac₄ManNH₂ exhibited only a very weak emission. The expression of TCO groups on sialoglycoproteins was then studied by Coomassie Brilliant Blue staining and western blot analysis (Fig. 3b and c). The results revealed that abundant TCO groups were generated *via* metabolic glycoengineering of proteins on the cell surface. The significant cell membrane labelling and imaging over the background is attributed to the good luminogenic behavior of Tb-1. Flow cytometry analysis of the A549 cells treated with Ac₄ManNTCO and fluorescein isothiocyanate (FITC)-tetrazine further confirmed that the cells modified with TCO groups can be labelled and light-up dramatically (Fig. 3d).

Next, our tetrazine-functionalized luminogenic probe Tb-1 was further employed for glycome imaging in living systems, and zebrafish was selected as a typical proof-of-concept visualization model due to its advantages of well characterized morphology, optical transparency and high homology with mammals.⁵⁵ In order to label sialylated glycans in zebrafish, its embryos were microinjected with 10 nL medium containing 5 mM Ac₄ManNTCO for 72 h post-fertilization, resulting in metabolic labelling of O-linked sialylated glycans with TCO. We then incubated the embryos with Tb-1 for 3 h and imaged them by confocal microscopy (Fig. 4a and b). Notably, a very sharp luminescence signal was observed in the enveloping cell

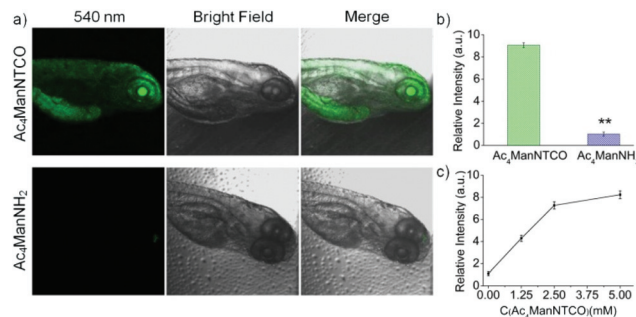


Fig. 4 (a) Two-photon emission images of the Ac₄ManNTCO (top row) or Ac₄ManNH₂ (bottom row) labelled zebrafish embryo at 72 h after fertilization. (b) Corresponding quantified intensity from (a). (c) Plots of the dose-dependent generation of TCO groups on the zebrafish after treatment with varied concentrations of Ac₄ManNTCO.

membranes of zebrafish pre-labeled with TCO-sialic acid, whereas there was no significant emission in control groups treated with Ac₄ManNH₂. And dose-dependent staining was observed in fishes treated with 0–5 mM Ac₄ManNTCO (Fig. 4c and Fig. S27†). As a consequence, Tb-1 could be used as a valuable and efficient luminogenic probe for TCO-labelled glycomes *in vivo* *via* a two-step imaging procedure.

Conclusions

In conclusion, we have presented a terbium(III) complex Tb-1 as a novel luminogenic bioorthogonal probe. The probe is weakly emissive due to luminescence resonance energy transfer (LRET) to the tetrazine group. However, it showed dramatic emission enhancement upon cycloaddition reaction with a strained cyclooctene by altering the tetrazine structure and regaining the antenna effect. Interestingly, this tetrazine complex was also identified as a luminogenic bioorthogonal label and imaging reagent for cyclooctene-modified glycomes. We hope that these results will pave the way for future site-specific protein labelling and imaging in live cells.

Experimental section

Materials and instruments

Materials. TbCl₃·6H₂O (99%), diethylenetriaminepentaacetic acid (DTPA, ≥98%), NaOH (≥98%), *N*-hydroxysuccinimide (NHS, ≥98%), LaCl₃·7H₂O (99.99%), *N,N*-dimethylformamide (DMF, 99%), dimethyl sulfoxide (DMSO, 99%) and 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride (EDC, ≥98%) were purchased from Aladdin Reagent, Ltd (Shanghai, China); and *trans*-3-hydroxyl-cyclooctene (TCO, ≥98%) and TCO-NHS (≥98%) were purchased from Ruixi Biological Technology Co., Ltd (Xi'an, China). CCK-8 was obtained from Dojindo (Japan). Dulbecco's modified Eagle's medium (DMEM) cell culture medium, fetal bovine serum, streptomycin, and penicillin were purchased from Thermo Fisher

Scientific Co., Ltd (China). All chemicals used in this study were of analytical grade and reagents were used without further purification. The compound Ac₄ManNTCO was prepared (Fig. S28a†) according to our previous literature.⁵⁶ La-H and La-1 were introduced *via* coordinating between the central atom lanthanum ion (La³⁺) and the ligand of **Tb-M** and **Tb-1**, respectively (Fig. S28b†). All aqueous solutions were prepared using ultrapure deionized water (DI water), which was obtained through a Millipore Milli-Q water purification system (Billerica, USA) and had an electric resistance >18.2 MΩ.

Instruments and characterization. The ¹H NMR spectra were recorded on a Bruker Ultrashield 400 Plus NMR spectrometer. All chemical shifts are reported in the standard δ notation of parts per million. The UV-vis absorption spectra were recorded on a UV-vis spectrometer (Lambda 35 UV-vis spectrometer, PerkinElmer, USA) at room temperature. The emission lifetime measurements of **Tb-1** and **Tb-M** were recorded with a FLS-1000 fluorescence spectrophotometer (Edinburgh Instruments, Britain). High-performance liquid chromatography (HPLC) with absorbance detection was carried out using an Agilent7890B column and acetonitrile as the eluent. FT-IR spectra were recorded with KBr pellets on a Bio-Rad FTS 6000 spectrometer (Bio-Rad Company, Hercules, California, USA) at room temperature. The confocal fluorescence images of cell and zebrafish were collected on an Olympus FluoView FV10 microscope (Olympus Imaging America Inc., Japan). Cell viability was measured using a microplate reader (Infinite 200, Tecan, Switzerland).

Synthesis of compound 1. 4-[(1,2,4,5-Tetrazin-3-yl)phenyl]-methanamine (tetrazine-benzylamine) and 7-amino-1,2-dihydro-2-oxo-4-quinoline acetic acid were produced according to the literature, respectively.^{57,58} Quinolinone acid (21.8 mg, 0.1 mmol) was reactivated with *N*-hydroxysuccinimide by adding 1.2 equiv. of NHS (13.8 mg, 0.12 mmol) and 1.2 equiv. of DCC (24.7 mg, 0.12 mmol) in dry DMF and reacted for 2 hours at room temperature. 1.1 equiv. of tetrazine-benzylamine (22.1 mg, 0.11 mmol) was added to the above solution and stirred at room temperature until the reaction reached completion (monitored by TLC). DMF was removed by rotary evaporation, and the crude mixture was purified by silica gel column chromatography with CH₂Cl₂/CH₃CN (3 : 1–1 : 1, v/v) as the eluent yielding compound **1** (27.5 mg, 68.5%). ¹H NMR (400 MHz, d₆-DMSO): δ 11.23 (s, 1H), 8.74 (t, *J* = 4.0 Hz, 2H), 8.38 (d, *J* = 8.0 Hz, 2H), 7.49 (d, *J* = 8.0 Hz, 2H), 7.42 (d, *J* = 8.0 Hz, 1H), 6.44 (d, *J* = 8.0 Hz, 1H), 6.38 (s, 1H), 6.05 (s, 1H), 5.74 (d, *J* = 4.0 Hz, 2H), 4.41 (d, *J* = 4.0 Hz, 2H), 2.99 (s, 3H). ESI-MS (*m/z*): calcd for C₂₁H₁₉N₇O₂, 401.42 [M]; found, 402.2000.

Synthesis of compound 2. 1 equiv. of DTPA dianhydride (19.7 mg, 0.05 mmol) was dissolved in a mixture of DMSO, and 5 equiv. of Et₃N (34 mL) under N₂. 0.7 equiv. of compound **1** (14.1 mg, 0.035 mmol) dissolved in DMSO was added slowly to the above solution under vigorous stirring. The reaction mixture was stirred at room temperature for 2 h. After completion, the above solution was quenched by addition of 0.1 M HCl and pH was adjusted to 6.5. Then water was removed by rotary evaporation and the crude mixture was purified by ether precipitation yielding compound **2**. ¹H NMR (400 MHz, d₆-

DMSO): δ 11.23 (s, 1H), 7.95–8.54 (m, 7H), 7.51 (d, *J* = 4.0 Hz, 1H), 7.35 (d, *J* = 4.0 Hz, 1H), 6.42 (d, *J* = 8.0 Hz, 2H), 6.35 (s, 1H), 5.98 (d, *J* = 8.0 Hz, 1H), 4.38 (d, *J* = 4.0 Hz, 1H), 3.12 (d, *J* = 8.0 Hz, 8H), 2.43 (d, *J* = 4.0 Hz, 2H), 2.36 (s, 2H), 1.79 (s, 4H), 1.23 (s, 2H), 2.36 (t, *J* = 8.0 Hz, 1H). ESI-MS (*m/z*): calcd for C₃₅H₄₀N₁₀O₁₁, 776.75 [M]; found, 776.4000.

Synthesis of Tb-1. Addition of metals: TbCl₃·6H₂O (5.0 mg, 0.01 mmol) was added to compound **2** (15.4 mg, 0.02 mmol) in a DMSO/H₂O (2 : 1, v/v) solution, and pH was adjusted to 6.5 by addition of 0.1 M HCl, pH 6.5 buffer, and stirred for 3 h at room temperature. Then the water was removed by rotary evaporation and the crude mixture purified by ether precipitation affording **Tb-1**. ESI-MS (*m/z*): calcd for C₃₅H₃₆N₁₀O₁₁Tb³⁺, 931.64 [M]; found, 931.3000.

Calculation of the limit of detection (LOD). LOD was calculated using the formula,⁴⁸ LOD = 3δ/*k*. Here, δ represents the standard deviation of the fluorescence intensity of 3 blank solutions, and *k* represents the slope between fluorescence intensity and concentration.

Stability of Tb-1. The stability of **Tb-1** in the simulated physiological environment was detected as below: **Tb-1** solution containing 0 to 50% serum (pH = 7.4) in PBS (pH 7.4) were kept for different times (from 1 to 24 h). The luminescence emission spectra were then recorded.

Cell culture. Human lung adenocarcinoma cells (A549) and experimental mammary tumour-6 cells (EMT 6) were cultured in DMEM or normal human hepatic cell (LO2) were cultured in RIM1640 containing 10% fetal bovine serum and 1% penicillin/streptomycin at 37 °C in a humidified 5% CO₂ atmosphere. The cell density was determined using a hemocytometer before experimentation.

Cytotoxicity evaluation *in vitro*. Relative cell viabilities were determined by the standard CCK-8 assay. A549 cells and EMT6 or LO2 cells were seeded into 96-well plates (1 × 10⁴ cells per well). And the cells were cultured for 12 h at 37 °C in a humidified incubator, 100 μL different concentrations of **Tb-1** PBS (pH 7.4) solution were added into the wells. After this, the cells were incubated for another 12 h at 37 °C. Then the cells were washed using PBS buffer to remove the unbound compound, and fresh culture medium was added. After incubation at 37 °C for 12 h, solution containing 10% CCK-8 DMEM (100 μL) was added to these cells. After incubation for 2 h at 37 °C, OD450, the absorbance value at 450 nm, were measured with a microplate reader to determine cell viability.

Imaging glycans with cells. A549 cells were first pretreated with Ac₄ManNH₂ (50 μM) or Ac₄ManNTCO (50 μM) for 3 days and washed with PBS (pH 7.4) twice. This was followed by incubation with **Tb-1** (50 μM) for 3 h at 37 °C in a humidified incubator and images were obtained using an Olympus FluoView FV10 microscope (Olympus Imaging America Inc., Japan) without washing. The two-photon excited luminescence of Tb³⁺ by a 730 nm laser was recorded at 500–560 nm.

Western blot analysis of cells. A549 cells were pretreated with Ac₄ManNH₂ (50 μM) or Ac₄ManNTCO (50 μM) for 3 days and washed with PBS (pH 7.4) twice, then harvested from the plates and centrifuged at 1800 rpm for 3 min. The cells were

resuspended and incubated in 100 μL of lysis buffer (1% SDS, 100 mM Tris-HCl, pH 7.4) containing a protease inhibitor cocktail (Complete, EDTA-free) at 4 °C for 30 min and centrifuged for 20 min at 11 000 rpm to get supernatant solution. The solution lysates (5 mg mL^{-1} , 50 μL) were incubated with tetrazine-biotin (5 μL , 5 mM in DPBS) for 6 h at 37 °C. Then the samples were resolved by 12% SDS/PAGE and transferred to pure nitrocellulose blotting membranes (BioTrace NT; Pall Life Science, Pensacola, FL, USA). The membranes were blocked using 10 mM Tris/HCl (pH 7.4), 150 mM NaCl, and 0.1% Tween-20 containing 5% nonfat milk for 1.5 h. Then, the membrane was incubated with streptavidin-HRP (diluted 1:2000 in TBST) overnight at 4 °C. The membrane was developed using a HPR-AEC western blotting substrate.

Flow cytometry analysis. A549 cells were pretreated with $\text{Ac}_4\text{ManNTCO}$ (50 μM) or $\text{Ac}_4\text{ManNH}_2$ (50 μM) followed by tetrazine-FITC. Then, the cells were removed and washed with FACS buffer. 10 000 cells per sample were analysed on a FACS canto II flow cytometer (Becton Dickinson, Mountain View, CA, USA) with excitation at 488 nm.

Imaging glycans with zebrafish. All animal experiments were performed in compliance with the National Institutes of Health (NIH) guidelines for the Care and Use of Laboratory Animals of South China Normal University, and the experiments were approved by the Animal Ethics Committee of South China Normal University. Zebrafish Husbandry: adult wild-type zebrafish were maintained at 28.5 °C on a 14 hour light and 10 hour dark cycle. Embryos were obtained from natural spawning and were maintained in embryo medium (EM; 150 mM NaCl, 0.5 mM KCl, 1.0 mM CaCl_2 , 0.37 mM KH_2PO_4 , 0.05 mM Na_2HPO_4 , 2.0 mM MgSO_4 , and 0.71 mM NaHCO_3 in deionized water, pH 7.4).⁵⁹

Zebrafish embryos at the one-cell stage were dechorionated by 1 mg mL^{-1} protease (1 mg mL^{-1} in embryo medium) at 4 hours post-fertilization (hpf). And then these embryos were microinjected into the yolk with 5 nL of 5 mM $\text{Ac}_4\text{ManNTCO}$. Meanwhile, the embryos microinjected with 5 nL of 5 mM $\text{Ac}_4\text{ManNH}_2$ were used as control groups. The embryos were then cultured at 28 °C using the standard embryo culture protocol. PTU (131 μM) was included in the medium beginning at 12 hpf to inhibit melanin production. After 72 hpf, they were labeled with the TCO group on zebrafish using a similar strategy to that of cellular glycome labeling described above. The embryos were then removed from the $\text{Ac}_4\text{ManNTCO}$ or $\text{Ac}_4\text{ManNH}_2$ containing solution, rinsed, and incubated in embryo medium containing **Tb-1** (100 μM) for 3 h at 28 °C. The embryos were anesthetized with tricaine (2.6 μM in embryo medium) and mounted between two cover slips in embryo medium containing 0.6% low melting point agarose, 2.6 μM tricaine, and 131 μM PTU. After this, the embryos were detected using microscopy with excitation at 730 nm.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

This research was supported by the National Natural Science Foundation of China (21771065), the Guangdong Special Support Program (2017TQ04R138), the Science and Technology Program of Guangzhou (2019050001), the Natural Science Foundation of Guangdong (2019A1515012021), the Pearl River Nova Program of Guangzhou, Guangdong Province, China (201806010189), the Hong Kong Research Grants Council (HKBU1230019) and the Hong Kong Baptist University (RC-ICRS/18-19/01), and CAS-Croucher Funding Scheme for Joint Laboratories (CAS 18204).

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