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Observe without disturbing: Tracer particles sense local stresses in cell collectives without affecting the cancer cell dynamics

Sumit Sinha,†*a*Himadri Samanta,†*^b* and D. Thirumalai∗*a*,*^b*

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Measurements of local stresses on the cancer cells (CCs), inferred by embedding inert compressible tracer particles (TPs) in a growing multicellular spheroid (MCS), show that pressure decreases monotonically as the distance from the core of the MCS increases. How faithfully do the TPs report the local stresses in the CCs is an important question because pressure buildup in the MCS is dynamically generated due to CC division, which implies that the CC dynamics should be minimally altered by the TPs. Here using theory and simulations, we show that although the TP dynamics is unusual, exhibiting sub-diffusive behavior on times less than the CC division times and hyper-diffusive dynamics in the long-time limit, they do not affect the long-time CC dynamics. The CC pressure profile within the MCS, which decays from a high value at the core to the periphery, is almost identical with and without the TPs. That the TPs have a small effect on the local stresses in the MCS implies that they are reasonale reporters of the CC microenvironment.

The interplay between short-range forces and non-equilibrium processes arising from cell division and apoptosis gives rise to unexpected dynamics in the collective migration of cancer cells $1-9$. An example is the dynamics of cancer cells (CCs) in a growing multicellular spheroid (MCS), which is relevant in cancer metastasis^{7,8}. Imaging experiments show that collective migration of a group of cells that maintain contact for a long period of time exhibits far from equilibrium behavior $8,10-13$. The experimental studies have served as an impetus to develop simulations and theoretical models $14-19$, which have established that proliferation as well as CC dynamics are governed by mechanical feedback and the microenvironment associated with cells in the MCS. Dynamics in a growing MCS is reminiscent of the influence of active forces in abiotic systems $20-22$. In a growing MCS, the analogue of active forces are self-generated²³, arising from biological events characterized by cell growth, division and apoptosis.

Although it has long been recognized that the stresses in the tumor interior are enhanced $24-26$, it is only recently that direct nonuniform pressure variations in the MCS has been measured. Following the early pioneering experiments $24-26$, several experimental techniques have been introduced in order to quantify the local stresses or pressure on the CCs^{27-32} . The stresses within MCSs were measured 32 by embedding micron-sized inert deformable polyacrylamide beads, which we refer to as tracer particles (TPs). The reduction in the volume of the TPs in the presence of the colon carcinoma cancer cells, was used to estimate the strain on the TPs. From the calibrated stress-strain plot for the TPs, measured in the absence of the CCs, they obtained the stress value at the location of the TPs. By assuming that the stresses associated with the fluorescently labeled TPs faithfully report the pressure of the CCs, it was argued that the pressure propagates nonuniformly across the tumor. It was concluded that the TPs could be used as local stress sensors.

The experiments raise an important question: *What should be the characteristics of the TPs for them to be faithful sensors of stresses in an evolving MCS?* This question emerges naturally from the experimental results $32,33$. We believe that the following criteria must be satisfied : (a) Because in a growing MCS, pressure is dynamically generated, predominantly by cell division, the TPs should have negligible effect on the CC dynamics. (b) The radial pressure profile in the MCS with and without the TPs must be qualitatively similar. (c) Finally, the TPs should have negligible effect on the distribution of pressure on the cells across the MCS. In essence, to the extent possible, the TPs should "observe" but not "disturb" the CC microenvironment.

In order to assess the influence of TPs on the fate of cancer cells, we used theory and simulations by embedding the TPs in an evolving MCS. We first show that the CCs, whose dynamics is sub-diffusive for $t \lesssim \tau$ (τ is the cell division time), exhibits superdiffusive behavior at long times with the MSD, $\Delta_{CC}(t) \sim t^{\alpha_{CC}}$. Exponent α_{CC} does not change appreciably from the value in the

^a Department of Physics, University of Texas at Austin, TX 78712.

^b Department of Chemistry, University of Texas at Austin, TX 78712.

[†] S.S. and H.S.S contributed equally to this work.

To whom correspondence should be addressed. E-mail: dave.thirumalai@gmail.com

Fig. 1 Difference between the CCs (pink sphere) and the TPs (magenta sphere). The cells grow if the local pressure, *p*, is less than a critical pressure *pc*. Cell division, also regulated by local pressure *p*, creates two daughter CCs (blue and green sphere), each with $R_d = \frac{R_m}{2^{1/3}}$. Their relative position is displaced by $2R_m(1-\frac{1}{2^{1/3}})$ with the orientation being random with respect to the sphere center. The TP is inert but deformable.

absence of the TPs. It could be argued that because the tracer density in the long time limit is small compared to cell density, the CC dynamics and in particular α_{CC} should not change. However, there is a rich literature in glass forming materials in which by pinning a small fraction (roughly 2-3%) of particles (analogous to TPs) the dynamics of unpinned particles (CCs in our case) are significantly slowed down, roughly by an order of magnitude, (see the curve corresponding to $T=0.46$ in figure 3 of reference 34). Thus, it is not *a priori* clear why the CC dynamics would not be impacted by the TPs even in the long time limit. Thus, criterion (a) described above is satisfied. We find that the pressure profiles vary radially across the MCS. The pressure is larger near the core and decreases as the boundary of the tumor is approached. Most importantly, the radial pressure profiles, not their magnitudes, of the CCs with and without the TPs are identical. It is important to realize that it is difficult to experimentally measure profiles in the absence of the TPs. Thus, it is only through simulations could we can assess if criterion (b) is qualitatively satisfied. We note parenthetically that the motility of the TPs is unusual. On short times ($t\lesssim \tau$) $\Delta_{TP}\sim t^{\beta_{TP}}$ with β_{TP} < 1. In the long-time limit, $(t > \tau)$, the TPs undergo hyper-diffusive motion, $\Delta_{TP}(t) \sim t^{\alpha_{TP}}$ with $\alpha_{TP} > 2$. Despite the unusual TP dynamics, it has only a small effect on the CCs, thus obeying criterion (c). Because TP is a passive reporter of the CC microenvironment, we conclude that deformable gel-like TPs may be used as sensors of local stresses in the tumor.

Results:

In order to investigate the effects of the inert but soft TPs on the properties of the CCs, we developed a theory supplemented by computer simulations of an agent based model. The complementary approaches allow us to draw sharp conclusions, which show that despite the unusual non-equilibrium dynamics the TPs, driven by the self-generated random forces created by the CC division, they faithfully report, at least qualitatively, on the local stresses within the solid MCS.

Theoretical Considerations:We consider the dynamics of deformable inert TPs in a growing MCS in a dissipative environment by neglecting inertial effects. In contrast to the TPs, the CCs grow and divide at a given cell division rate, and also undergo apoptosis (see Fig.(1)). The CCs and TPs experience systematic shortranged attractive and repulsive forces arising from the other CCs and $TPs¹⁴$. To model a growing MCS, we modify the density equation for the CCs by adding a non-linear source term, $\propto \phi(\phi_0 - \phi)$ that accounts for cell birth and apoptosis, and a non-equilibrium noise term that breaks the CC number conservation 35 . The noise, f_{ϕ} , satisfies $=\delta(\mathbf{r}-\mathbf{r}')\delta(t-t')$. The source term, $\propto \phi(\phi_0 - \phi)$, represents cell division and apoptosis, with $\phi_0 = \frac{2k_b}{k_a}$ ^{35,36}. The coefficient, $\sqrt{k_b\phi + \frac{k_a}{2}\phi^2}$ of f_ϕ , is the noise strength, corresponding to number fluctuations.

In order to obtain analytic insights, we first derive equations that describe the dynamics of the TPs and CCs. We introduce the density fields, $\phi(\mathbf{r},t) = \sum_i \delta[\mathbf{r} - \mathbf{r}_i(t)]$ for the CCs, and $\psi(\mathbf{r},t) =$ $\sum_i \delta[\mathbf{r}-\mathbf{r}_i(t)]$ for the TPs. A formally exact Langevin equation for $\phi(\mathbf{r},t)$ and $\psi(\mathbf{r},t)$ may be derived using the Dean's method ³⁷ that accounts for diffusion and systematic non-linear interactions. The equations for $\phi(\mathbf{r},t)$ and $\psi(\mathbf{r},t)$ are,

$$
\frac{\partial \psi(\mathbf{r},t)}{\partial t} = D_{\psi} \nabla^2 \psi(\mathbf{r},t) + \nabla \cdot (\psi(\mathbf{r},t) \mathbf{J}) + \tilde{\eta}_{\psi},
$$
(1)

$$
\frac{\partial \phi(\mathbf{r},t)}{\partial t} = D_{\phi} \nabla^2 \phi(\mathbf{r},t) + \nabla \cdot (\phi(\mathbf{r},t) \mathbf{J}) + \frac{k_a}{2} \phi(\frac{2k_b}{k_a} - \phi)
$$

$$
+ \sqrt{k_b \phi + \frac{k_a}{2} \phi^2} f_{\phi} + \tilde{\eta}_{\phi}
$$
(2)

where $J = \int_{\mathbf{r}'} [\psi(\mathbf{r}',t) + \phi(\mathbf{r}',t)] \nabla U(\mathbf{r}-\mathbf{r}'), \quad \tilde{\eta}_{\psi}(\mathbf{r},t) =$ $\nabla \cdot \left(\eta_\psi(\mathbf{r},t) \psi^{1/2}(\mathbf{r},t)\right), \quad \tilde{\eta}_\phi(\mathbf{r},t) = \nabla \cdot \left(\eta_\phi(\mathbf{r},t) \phi^{1/2}(\mathbf{r},t)\right), \quad \text{and}$ $\eta_{\phi,\psi}$ satisfies $\langle \eta_{\phi,\psi}(\mathbf{r},t)\eta_{\phi,\psi}(\mathbf{r}',t')\rangle = \delta(\mathbf{r}-\mathbf{r}')\delta(t-t')$. The second term in Eq.(1) accounts for the TP-TP interactions $(\nabla \cdot (\psi(\mathbf{r},t))_{\mathbf{r}'} \psi(\mathbf{r}',t) \nabla U(\mathbf{r}-\mathbf{r}'))$ and TP-CC interactions, $(\nabla \cdot (\psi({\bf r},t) \int_{{\bf r}'} \phi({\bf r}',t) \nabla U({\bf r}-{\bf r}'))).$ The influence of the CCs on the TP dynamics is reflected in the TP-CC coupling. The third term in Eq.(2) results from cell division and apoptosis, and the fourth term in Eq.(2) is the non-equilibrium noise. It should be noted that Eq.(1) does not satisfy the fluctuation-dissipation theorem in a growing tumor.

The coupled stochastic integro-differential (SID) equations are difficult to solve analytically $38,39$. Even numerical solutions in the absence of TPs, are usually obtained by replacing $\tilde{\eta}_{\phi}(\mathbf{r},t)$, the last term in Eq.(2), by $\tilde{\eta}_{\phi}(\mathbf{r},t) = \nabla \cdot (\tilde{\eta}_{\phi}(\mathbf{r},t)\phi_0^{1/2})$, where ϕ_0 is a constant. Instead of using uncontrolled approximations, we solved the exact coupled SIDs $(Eq.(1)$ and $Eq.(2))$ numerically (see the appendix for details) in order to calculate the needed correlation functions from which the exponents characterizing the mean square displacement may be derived. For instance, from the time-dependent decay of the correlation function $C_{\psi\psi}(t) = \int d^3r C_{\psi\psi}(\mathbf{r},t)$, we can extract the dynamical exponent, *z*. Previously¹⁴, we had argued that $C_{\psi\psi}(t) \sim t^{1-(2+d)/z}$ (*d* is the spatial dimension). The correlation function $C_{\psi\psi}(t)/C_{\psi\psi}(0)$ decays in two stages, depending on t/τ , where τ is the cell division time (Fig.(2a)). For $t/\tau < 1$, we find that $C_{\psi\psi}(t) \sim t^{-3/7}$ from which we obtain $z = 7/2$. The MSD ($\Delta_{TP}(t)$) exponent for the TPs is related to *z* as $\Delta_{TP}(t) \sim t^{\beta_{TP}} \sim t^{2/z}$, which implies $\beta_{TP} = 4/7$.

Fig. 2 (a) The normalized time-dependent density correlation function plot for the TPs. The cell cycle time is $\tau = 54,000$ s. The integrated time $(t \approx 4\tau)$ is long enough to obtain accurate exponents from the power law decay for both short $(t < \tau)$ and long times $(t > \tau)$. The plots for three box sizes confirm that the decay of the correlation function is independent of the system size. (b) MSD of the TPs (Δ_{TP}) using the Gaussian potential. The plots are for 3 cell cycle times (blue (0.5τ) , red (τ) , and green (2τ) . Time to reach the hyper-diffusive behavior, which is preceded by a jamming regime $(\Delta_{TP} \sim t^{\beta_{TP}}, \beta_{TP} = 0.11$, shown in black dashed line), increases with τ. The inset highlights the hyper-diffusive regime ($\frac{t}{\tilde{\tau}} > 1$). τ̃ is the cell cycle time for the respective curves. Time is divided by $\frac{1}{\tilde{\tau}}$. $\Delta_{TP} \sim t^{\alpha_{TP}}, \alpha_{TP} = 2.3$ (dashed black line).

Similarly, at long ($t/\tau > 1$) times $C_{\psi\psi}(t) \sim t^{-33/7}$ from which we deduce that $\Delta_{TP}(t) \sim t^{\alpha_{TP}}$ with $\alpha_{TP} = 16/7$. Thus, the numerical solution of the exact equations together with the scaling ansatz predict that, in the long time, the TPs undergo hyper-diffusive motion in the presence of CCs. It should be noted that we have extracted the exponents even though the data spans less than a decade in time. Our goal is to make connections with simulations and more importantly experiments. In the latter case, the observations are made for time spanning about $(2-3)\tau$, which is not that dissimilar from the range probed in numerical simulations of the SIDS.

Simulations: In order to validate the theoretical predictions and decipher the mechanisms for the origin of the CC driven hyperdiffusive motion, we simulated a 3D MCS with embedded TPs using an agent-based model^{14,16,23,40–43} (see Figure S1 in the SI). The cancer cells, treated as interacting soft deformable spheres, grow with time, and divide into two identical daughter cells (Fig. (1)) upon reaching the mitotic radius (R_m) . We used, $\tau = \frac{1}{k_b}$ 54,000*s* as the cell division time. The CCs also undergo apoptosis at the rate $k_a \ll k_b$. The sizes and the number of TPs are held constant, which means that the density of the TPs decreases, as in the experiments, as the tumor grows.

We used a pressure inhibition mechanism to model the observed growth dynamics in solid tumors¹⁴. Dormancy or the growth phase of the CCs depends on the local microenvironment, which is determined by the pressure on the *i th* cell (Fig.(1)). Cell division and the placement of the daughter cells stochastically alter the dynamics of the CCs. More importantly, cell division generates active forces, referred to self-generated active force

 $(SGAF)$ ^{23,42}, that persist for a period of time, thus affecting the dynamics of the neighboring cells. To determine the TP dynamics, we model the CC-CC, CC-TP, and TP-TP interactions using two potentials (Gaussian and Hertz) to ensure that the qualitative results are robust (simulation details are described in the Appendix).

Mean Square Displacement of the TPs $(\Delta_{TP}(t))$ **: The simula**tions show that, in the limit $t < \frac{1}{k_b}$, the TPs exhibit sub-diffusive behavior, $\Delta_{TP}(t) = \langle [\mathbf{r}(t) - \mathbf{r}(0)]^2 \rangle \sim t^{\beta_{TP}}$, with $\beta_{TP} < 1$ (Fig.(2b)). Note, $\Delta_{TP}(t)$ was calculated by averaging over $\approx 2,000$ trajectories using the Gaussian potential. The simulations confirm (qualitatively) the theoretical predictions that the dynamics on times less than than the cell division time is sub-diffusive. We also calculated $\Delta_{TP}(t)$, for the Hertz potential (Figure S3a in the SI). Similar behavior is found in both the cases, which suggests that the conclusions are robust.

The modest increase in the MSD at short times is reflected as a near plateau in time dependence of ∆*T P*(*t*) (Fig.(2b)). The duration of the plateau (Fig.(2b)) increases as the cell cycle time increases. Although jamming behavior predicted theoretically is consistent with the simulations, the numerical values of β_{TP} differ (compare Fig.(2a) and (2b)), implying that the dynamics in the t/τ < 1 limit depends on the details of the model.

In the $t > \frac{1}{k_b}$ limit, theory and simulations predict hyperdiffusive dynamics for the TPs, $\Delta_{TP} \sim t^{\alpha_{TP}}$ with $\alpha_{TP} \approx 2.3$. Variations in $k_b = \tau^{-1}$ do not change the value of α_{TP} (Fig.(2b)). It merely changes the coefficients of the linear term and the value of $\frac{2k_b}{k_a}$. Therefore, α_{TP} is independent of the cell division rate in the long time limit. Simulation results are in excellent agreement with the theoretical predictions. For the Gaussian potential, we

Fig. 3 CC dynamics in the presence of the TPs. ∆*CC* using the Hertz potential. The curves are for different TP radius (magenta - $r_{TP} = 2$ *r*_{*CC*}, green - *r*_{*TP*} = *r*_{*CC*}, and red - *r*_{*TP*} = 0.75*r*_{*CC*}, and blue - *r*_{*TP*} = 0.5*r*_{*CC*}, where $r_{CC} = 4.5 \mu m$ is the average cell radius. Inset shows Δ_{CC} for the four curves, focusing on the super-diffusive regime. The black dashed line is drawn with $\alpha_{CC} = 1.47$.

obtain $\alpha_{TP} \approx 2.3$ (see the inset in Fig.(2b)) for 0.5τ, τ and 2τ. Thus, α_{TP} is independent of cell cycle time, as argued above.

Note that TP-TP interactions play an insignificant role in the dynamics of TPs or the CCs (Figure S4 in the SI). They merely alter the amplitude of Δ_{TP} in the intermediate time without affecting the long time dynamics. We conclude that the long-time behavior is universal, in the sense that it is impervious to the details of the interaction but is determined by cell division and apoptosis rates.

Impact of TP on CC dynamics: Interestingly, the time-dependent changes in $\Delta_{CC}(t)$ are not significantly affected by the TPs. Fig.(3) shows that changing the tracer size affects only the amplitude of the Δ_{CC} in the intermediate time without altering the α_{CC} values (see also figure S5 in the SI). This is because the TP size only changes the nature of the short-range interactions without introducing any new length scale. Since, α_{CC} is a consequence of the long-range spatial and temporal correlations that emerge because of cell division and apoptosis, it is independent of the tracer size (see the details in the SI).

In the absence of the TPs, birth and apoptosis determine the CC dynamics in the long time regime, yielding $\alpha_{CC} = 1.33^{14,15}$. When the TPs are present, the CCs continue to exhibit superdiffusive motion with a modest increase in α_{CC} (Table II in the SI). The origin of the predicted super-diffusive behavior in the CCs is related to the long range spatial correlation that arises due the SGAF (self-generated active force) that is related to cell division. This is reflected in the CC pair-correlation, $g(r)$ = $\frac{V}{4\pi r^2 N^2} \sum_{i=1}^N \sum_{j \neq i}^N \delta(r - |\mathbf{r}_i - \mathbf{r}_j|) \sim r^{-0.5}$, in the presence and absence of TPs at $t \approx 8\tau$ (Fig.(4a)). The dynamically-induced longrange CC correlations is independent of the TPs, thus explaining the insignificant effect of the TPs on the CC dynamics.

Mechanistic Origin: To explain the finding that, $\alpha_{TP} > \alpha_{CC}$, we calculated the force-force autocorrelation function, $FFA = \langle \mathbf{F}(t + \mathbf{F}) \rangle$ $(t_d) \cdot \mathbf{F}(t)$ _{*t*}. Here, $\langle ... \rangle_t$ is the time average and t_d is the duration of the delay time. Since, the TPs (CCs) exhibit hyper-diffusion (super-diffusion), we expect that the FFA of the TPs should decay slower relative to the CCs. This is confirmed in Fig.(4b), which shows that the FFA for the TPs (CCs) decays as $t_d^{-0.7}$ ($t_d^{-1.2}$). Thus, the motion of the inactive TP is significantly more persistent than the CCs, which explains the hyper-diffusive nature of the TPs.

Physically, the mobilities of the TPs are mainly driven by CC cell division. At each cell division, the CCs impart an impulsive force to the TPs. The active force propels them in a directed manner for a period of time. As the CC density grows the number of cell divisions increases, resulting in enhanced motility of the TPs, which is also reflected in the slow decay of the FFA of the TPs. The active forces generated by the CCs produces the observed hyper diffusive behavior in the TPs.

To illustrate the difference between the dynamics of the TPs and CCs, we calculated the angle (θ) between two consecutive time steps in a trajectory. We define $\cos(\theta(t, \delta t)) = \frac{\delta \mathbf{r}(t+\delta t) \cdot \delta \mathbf{r}(t)}{|\delta \mathbf{r}(t+\delta t)| |\delta \mathbf{r}(t)|)}$, where $\delta \mathbf{r}(t) = \mathbf{r}(t + \delta t) - \mathbf{r}(t)$. Fig.(5a) shows the ensemble and time averaged $\cos\theta$ distribution for $\frac{\delta t}{\tau} = 1$. If the motion of the CCs and TPs were diffusive, the distribution of $\cos\theta$ would be uniformly distributed from -1 to 1. However, Fig.(5a) shows that $P(\cos\theta)$ is skewed towards unity implying that the motion of both the TPs and CCs are persistent. Interestingly, the skewness is more pronounced for the TPs compared to the CCs (Fig.(5a)). The enhanced persistence of TP motion due to SGAF is also reflected in the FFA(t) displayed in Fig.(4b). The slower decay of FFA(t) (Fig.(4b)) in the TPs compared to the CCs is neglected in the distribution of $P[cos(\theta)]$, shown in Fig.(5a). During each cell division, the motion of the CCs is randomized, and hence the persistence is small compared to TPs. This explains the hyperdiffusive (super-diffusive) for the TPs (CCs) and thus $\alpha_{TP} > \alpha_{CC}$ in the long time limit (see Fig.(5a), which shows TPs movement is much more persistent than CCs).

Monotonic decrease of pressure from the core to periphery of the MCS: To assess the effectiveness of the TPs as sensors of the local microenvironment, we calculated the radial pressure (Eq.(10) in the Appendix) profile for both the CCs and TPs at $t = 7.5\tau$. In order to distinguish between the local pressure due to the TPs and CCs, we calculated pressure experienced just by the TPs. Finally, we computed pressure in a system consisting of only the CCs. By comparing the results from different simulations, we establish that TPs are good reporters of the local stresses within a single tumor.

From the results shown in Fig.(5b), we draw the following conclusions. (i) All three curves qualitatively capture the pressure profile found in the experiment 32, even though the protocol used in the experiment and simulation is different. From the results in Fig.(5b) several conclusions that are relevant to experiments may be drawn. Pressure decreases roughly by a factor of four, as the distance, *r*, from the center of the tumor increases. The pressure is almost constant in the core, with a decrease that can be fit using the logistic function, as the boundary of the tumor is reached

b) two dashed lines (black and orange) are power law fits to *g*(*r*) in the large *r* limit. **(b)** Force force autocorrelation (FFA), as a function of delay time exponent of -0.7 (-1.2). Fig. 4 (a) CC correlation function, $(g(r))$, as a function of inter-cellular distance. Red (blue) curve shows $g(r)$ in presence (absence) of the TPs. The (t_d) . The red (blue) curve shows the FFA for TPs (CCs). Inset shows FFA on log-log scale. The black (yellow) dashed line is a power law fit with

data1

solid tumor. important point being that the stress is non-uniform across the solid tumor was fit using a power law ($\sim r^{-\beta}$ with $\beta \approx 0.2$) (see Figure(4b) .(5b)). The experimental data, shown in the inset the experiment³²). Both the fits account for the da occ t d (Fig.(5b)). The experimental data, shown in the inset in Fig.(5b), in the experiment 32). Both the fits account for the data with the

reason is that in experiments high stress is uniformly applied by compressing the spherical tumor. In contrast, in our simulations \mathbf{u} lations qualitatively reproduce the experimental pressure profile, 0.3 10-3 pressure sensors in colon carcinoma cells. Although our simu-(ii) The experiments used reduction in volume of the polyacrythe magnitude of the stress in the simulations is about three orlamide microbeads upon external compression $(= 5KPa)$ as local ders of magnitude less compared to experiments. The most likely pressure arises explicitly due to SGAF arising from an interplay of systematic forces, cell division, as well as mechanical feedback. The simulated MCS is not intended to mimic the characteristics of the colon carcinoma cells studied in the experiment 32 . The simulations illustrate that the TPs are good pressure sensors. It should also be noted that the magnitude of the pressure is also dependent of the tissue type. For example, a recent experiment 33 , using fluorescently labeled water-soluble peptide-based microtubes, showed that the measured pressure $\approx 0.4KPa$. The qualitative agreement between experiments and simulations (Fig.(5b)) suggests that the dynamical rearrangement due to cell division and apoptosis and mechanical feedback controls the observed radial variations in the pressure profiles.

(iii) The higher value of pressure in the core of the MCS as the MCS grows is a consequence of jamming, which increase the local density. Because of the rapid cell division rate, the pressure experienced by cells in the interior is high (Figs. 5b, 5c and 5d). As a result, the local stresses in the core of the MCS do not have enough time to relax. The higher core density implies that pressure in the interior increases and exceeds *pc*, which further inhibits proliferation, thus deceasing the number of cell divisions. The reduction in the number of cell divisions, which is a mechanism of stress relaxation, leads to an increase in pressure at $r < R_0$ (Figs. (5b, 5c and 5d)). As *r* increases, the jamming effects diminish, and local stress relaxation is faster resulting in the pressure on the cells being less than *pc*. Consequently, the CCs proliferate, resulting in a decrease in self-generated stresses, and consequently, a decreases in the magnitude of the pressure (Figs. (5b, 5c and 5d)).

(iv) Most importantly, the CC pressure profiles are unaltered even in the presence of the TPs (compare blue circles and greenish-yellow squares in Fig. 5b, which shows that the latter does faithfully report the microenvironment of the CCs. In addition to the radial profile, we also show that the CC stress distribution is unaltered with and without TPs in Fig.5e. The plot in Fig. 5f establishes that the stress distribution associated with the TPs roughly approximates the CC stress distribution, implying that TPs can serve as reasonable reporters of not only the radial profile (Fig. 5b) but also the CC stress distribution. It should be pointed out that the magnitude of the pressure calculated with only the TPs is less than the values obtained using the stresses using only CCs. In this sense, TPs only qualitatively report the stresses experienced by the CCs, which imply that the TPs are not perfect reporters. This situation likely exists in experiments as well. Using the current experiments, the contribution to the stresses arising from TPs and CCs cannot be estimated. Based on our simulations, we would assert that the pressure reported using the characteristics of the TPs provide only a lower bound. We should add that the differences between the pressure reported using TPs does not differ significantly from the CC pressure.

The simulations also provide plausible reasons for the decrease in the pressure from the center to the boundary of the tumor. Pressure on the *i th* cell located at a distance *r* from the tumor cen-

Fig. 5 **(a)** Distribution of cos(θ) for the TPs and CCs. θ is the angle between two consecutive steps that a given cell takes in a trajectory. The red (blue) plot is for the TPs (CCs). The distribution is skewed to $cos(\theta) > 0$, indicative of persistent motion. Extent of skewness is greater for the TPs than CCs. The inset shows the *P*[cos(θ)] of CCs with and without TPs. **(b)**Pressure as a function of the radial distance (*r* scaled by *R*0: tumor radius ≈ 100 $μm$) from tumor center. The blue circles correspond to local pressure measured using both CCs and TPs. The red data points is the local pressure measured using just the TPs in simulations containing both the CCs and TPs. The greenish yellow squares give the local pressure obtained in simulations with only the CCs. The black and green dashed lines is logistic fit. The inset shows the radial pressure profile in experiments (green data points). The red dashed lines are logistic fits. Note that the magnitude of pressure measured in simulations and experiments are in qualitative agreement. (c) Pressure probability distribution, $\mathscr{P}(\frac{p_i}{p_c})$ for CCs. In the main figure, orange (blue) distributions correspond to stress distribution in the CCs whose distance is greater (less) than 80 μ m (20 μ m). Similarly in the inset, orange (blue) distributions correspond to stress distribution on CCs whose distance is greater (less) than 60 µ*m* (30 µ*m*). (d) Same as (c), except the distributions correspond to TPs. (e) The blue (orange) distribution corresponds to the stress distributions on the CCs in the presence (absence) of the TPs. (f) The blue (orange) distribution are the stress distributions on all the CCs (TPs) in the spheroid. 6 | Journal Name, [year], **[vol.]**,1–10

ter depends not only on the total force exerted by the neighboring cells but also is controlled by cell division, which is a dynamical process. As the tumor evolves through cell division, the local density $\phi(\mathbf{r})$ of the cells at small **r** increases. More importantly, cell division induces local stresses⁴⁴, which would relax on time scales that depend on $\phi(\mathbf{r})$. Thus, cell division induced jamming leading to an increase in $\phi(\mathbf{r})$, and stress relaxation times are intimately related. If the local stress relaxation time decreases as the local density $\phi(\mathbf{r})$ decreases, the pressure would be small. It is clear that near the periphery of the tumor $\phi(\mathbf{r})$ is smaller than at the core, which explains the origin of the pressure profile in Fig. 5b.

Conclusions: We used simulations and theory to elucidate the dynamics of inert tracer particles that are embedded in an evolving multicellular spheroid (MCS). Surprisingly, theory and simulations show that for $k_b t > 1$ the TPs exhibit hyper-diffusive behavior, which does not depend on the details of interaction between the cells. In contrast, the exponent, α_{CC} , characterizing the long-time behavior of $\Delta_{CC}(t)$ is nearly the same with or without the TP, and is also independent of the probe radius. This suggests that TPs may be used to probe the behavior of cancer cells within a solid tumor, which cannot be established using experiments alone.

Direct connections to experiments were made by showing that the simulated and measured stress profiles are qualitatively similar. This is unexpected because the protocol used in experiments (response to an isotropically applied stress to a spheroid) is completely different in the simulations in which stresses within the MCS are self-generated. The higher stress values in the interior of the spheroid imply that proliferation is suppressed in the core. The most important finding is that the CC pressure profiles are virtually identical, and is independent of the presence or absence of TPs (Fig.(5(b)). Thus, we conclude that the TPs are reasonable reporters of the CC microenvironment in a growing solid tumor.

The radial dependence of local stresses in Fig.(5(b) is related to the non-uniform proliferation of cells in the MCS. In the models used in the simulations, mechanical feedback controls the growth of the MCS. This implies that if the local stresses exceed a critical value then the cells become dormant, and cannot grow or divide until the neighboring cells rearrange to decrease the pressure on a labeled cell. However, as the MCS evolves, the pressure on cells that are in the interior could exceed a critical value (p_c) due to jamming. In contrast, the pressure on cells near the MCS boundary is typically below *pc*, which results in enhanced proliferation. It is the non-uniformity in the cell division rates due to mechanical feedback that not only is the cause of the radial variation in pressure in the MCS but also gives rise to the complicated radial dependent dynamics (glass-like slow dynamics in the interior to super diffusion near the periphery)¹⁶. The implication is that the effective diffusion constants of CCs as well as the TPs also vary with distance from the center of the MCS.

It is not uncommon to measure properties of equilibrium systems (such as simple and molecular fluids) by embedding tracer particles. In these systems, the measurements are reasonable because fluctuation-dissipation theorem (FDT) holds. In contrast, a growing tumor, driven by cell division, is out of equilibrium, thus violating the FDT. Nevertheless, the response of tracer particles could be used to determine accurately the local stresses, as shown here explicitly. More generally, it appears that the theoretical and simulation approaches developed here are applicable in a range of active systems that are always out of equilibrium.

In the current model, we have neglected active forces arising from self-propulsion because the matrix viscosity is very large. However, in cases where the matrix viscosity is low, it will be important to incorporate the role of self-propulsion. In this scenario, physics due to self-propulsion, cell growth and division will be coupled, which provides a new direction for active matter research.

Appendix:

Numerical solution of the SIDs (Eq.(1) and Eq.(2)): We numerically integrated Eq.(1) and Eq.(2) by discretizing space in a cubic box with periodic boundary conditions. The size of the 3D box with $L_x = L_y = L_z$ is varied from 300 to 500 μ *m* to ensure that the results are not affected by finite size. The 3D box was divided into grids, each with volume $\delta x \times \delta y \times \delta z$ with $\delta x = \delta y = \delta z = 10 \mu m$. The parameters used in the numerical integration scheme are shown in Table I in the SI. Note that the precise shape of the box does not affect the numerical solutions nor does the value of *dx* as long the solutions converge without numerical instability.

Initial Conditions: The initial conditions for both Ψ and ϕ fields are Gaussian, where the tracer density is roughly ten times smaller than the cancer cell denisty.

Simulations: In order to test the theoretical predictions, and provide mechanistic insights underlying the unusual dynamics of the TPs in the MCS, we simulated a 3D tumor spheroid with embedded TPs. Following previous studies, we used an agent based model 14,40 for the tumor spheroid. The cells are treated as deformable spheres. The size of the CCs increase with time, and divide into two identical cells upon reaching a critical mitotic radius (R_m) , as showin in Fig. 1. The mean cell cycle time, $\tau = 54,000$ *s*, which is a realistic value for fibroblast cells. The CCs can also undergo apoptosis with rate k_a . The birth rate, $k_b = \frac{1}{\tau}$ is large compared to the death rate ($\frac{k_b}{k_a} = 20$), which mimics the growth of the MCS. As in the theory, the TPs are inert, and their sizes and the number are constant in the simulations. We use two different potentials for the CC-CC, CC-TP and TP-TP interactions.

Hertz potential: The form of the Hertz forces between the CCs is the same as in the previous studies $14,16,40,45,46$. The physical properties of the CC, such as the radius, elastic modulus, membrane receptor, and E-cadherin concentration characterize the strength of the inter-cellular interactions. The elastic forces between two spheres with radii R_i and R_j , is given by,

$$
F_{ij}^{el} = \frac{h_{ij}^{\frac{3}{2}}}{\frac{3}{4}(\frac{1 - v_i^2}{E_i} + \frac{1 - v_j^2}{E_j})(\sqrt{\frac{1}{R_i} + \frac{1}{R_j}})},
$$
(3)

where *Ei* and ν*i* are, respectively, the elastic modulus and Poisson ratio of the i^{th} cell. We assume that E_i and v_i are independent of i . Since the CCs or the TPs are deformable, the elastic force depends on the overlap, h_{ij} , between two cells. The adhesive force, F_{ij}^{ad} , between the CCs is proportional to the area of contact $(A_{ij})^{47}$, and is calculated using, ⁴⁵,

$$
F_{ij}^{ad} = A_{ij} f^{ad} \Lambda_0,\tag{4}
$$

where Λ_0 is unity in the present study.

Repulsive and adhesive forces in Eqs.(3) and (4) act along the unit vector \vec{n}_{ij} connecting the centers of cells *j* and *i*. Therefore, the net force on cell *i* (\vec{F}_i^H) is given by the sum over the nearest neighbors [NN(i)],

$$
\vec{F}_i^H = \Sigma_{j\epsilon NN(i)} (F_{ij}^{el} - F_{ij}^{ad}) \vec{n}_{ij}.
$$
 (5)

To model the TP-TP and TP-CC interactions, we assume that the TPs are CC-like objects 32. Therefore, CC-TP and TP-TP interactions are the same as CC-CC interactions. The parameters used in the simulations using the Hertz potential (Eqs.(3) and (4)) are given in Table I in the SI. We initiated the simulations with 100 TPs and 100 CCs. Because the number of the TPs is a constant, it follows that the TP density decreases as the tumor size grows with time. The coordinates of the CCs and TPs at $t = 0$ were sampled using a normal distribution with zero mean, and standard deviation 50 μ m. The initial radii of the CCs and TPs were sampled from a normal distribution with mean 4.5 μ *m*, and a dispersion of 0.5 µ*m*.

Gaussian potential: In the theoretical treatment, we assumed that the CC-CC interaction is given by a sum of Gaussian terms (Eq. (S2) in the SI). For this potential, the force \mathbf{F}_{ij}^G on cell *i*, exerted by cell *j*, is,

$$
\mathbf{F}_{ij}^G = \frac{1}{(2\pi)^{3/2}} \left[\frac{v e^{\frac{-r^2}{2\lambda^2}}}{\lambda^5} - \frac{\kappa e^{\frac{-r^2}{2\sigma^2}}}{\sigma^5} \right] \mathbf{r}
$$
 (6)

where **r** is **r**(*i*) – **r**(*j*). We write λ and σ as $\lambda = \tilde{\lambda}(R_i + R_i)$ and $\sigma = \tilde{\sigma}(R_i + R_j)$, as the ranges of interactions corresponding to the repulsive and attractive interactions, respectively. In our simulations, the CCs grow and divide, their radii change in time, and therefore λ and σ also change with time. However, since these interactions are short-ranged, we assume that they are constant in order to be consistent with the assumption in the theory. We fixed $\lambda = \lambda(2R_d)$ and $\sigma = \tilde{\sigma}(2R_d)$, where $R_d \approx 4\mu m$ is the size of a daughter cell (introduced in the next section). For simplicity, we write the force as $\mathbf{F}_{ij}^G = [\frac{\widetilde{\gamma}e^{\frac{-r^2}{2\lambda^2}}}{\lambda^2}]$ $\frac{\partial^2 F^2}{\partial \lambda^2}$ – $\frac{\tilde{\kappa}e^{\frac{-r^2}{2\sigma^2}}}{\sigma^2}$ **]r**, where $\tilde{v} = \frac{1}{(2\pi)^{3/2}} \frac{v}{\lambda^3}$ and $\widetilde{\kappa} = \frac{1}{(2\pi)^{3/2}} \frac{\kappa}{\sigma^3}$. In the simulations, we fixed $\widetilde{\nu} = 0.03$, $\widetilde{\lambda} = 0.28$, $\widetilde{\kappa} = 0.003$ and $\widetilde{\sigma} = 0.4$.

Cell division, Dormancy and Apoptosis: The CCs are either dormant or in the growth phase depending on the microenvironment, which is assessed using the value of the local pressure. The pressure on cell *i* (*pi*) due to *NN*(*i*) neighboring cells is calculated using the Irving-Kirkwood equation,

$$
p_i = \frac{1}{3V_i} \Sigma_{j\epsilon NN(i)} \mathbf{F}_{ij} \cdot d\mathbf{r}_{ij},\tag{7}
$$

where \mathbf{F}_{ij} is the force on the *i*th cell due to *j*th cell and $d\mathbf{r}_{ij} =$ $\mathbf{r}_i - \mathbf{r}_j$. The volume of the *i*th cell (*V*_{*i*}) is $\frac{4}{3}\pi R_i^3$, where R_i is the cell radius. If p_i exceeds a pre-assigned critical limit p_c (= 1.7 Pa) the CC enters a dormant phase. The dormancy criterion serves as a source of mechanical feedback, which limits the growth of the tumor spheroid^{48–52}. The volume of a growing cell increases at a constant rate, r_V . The cell radius is updated from a Gaussian distribution with the mean rate $\dot{R} = (4\pi R^2)^{-1} r_V$. Over the cell cycle time τ , the cell radius increases as,

$$
r_V = \frac{2\pi (R_m)^3}{3\tau},\tag{8}
$$

where R_m is the mitotic radius. A cell divides once it grows to the fixed mitotic radius. To ensure volume conservation, upon cell division, we use $R_d = R_m 2^{-1/3}$ as the radius of the daughter cells. The resulting daughter cells are placed at a center-to-center distance $d = 2R_m(1 - 2^{-1/3})$ (Fig.(1)). The direction of the new cell location is chosen randomly from a uniform distribution on a unit sphere. At times $\sim k_a^{-1}$, a randomly chosen cell undergoes apoptosis.

Although the model does not have detailed description of the important proliferation gradients that are present in tumors, it is included in an approximate manner. In our model, the spatial gradients in cell division arise due to the indirect bio-mechanical feedback. In the agent-based model, the cell division predominantly occurs on the periphery as the self-generated stress is higher in the core than in the periphery. Therefore, we have considered a minimal model which takes into account spatial heterogeneity of cell-division.

Equation of Motion: The equation of motion governing the dynamics of the TP and CCs is taken to be,

$$
\dot{\vec{r}}_i = \frac{\vec{F}_i}{\gamma_i},\tag{9}
$$

where $\dot{\vec{r}}_i$ is the velocity of *ith* CC or TP, \vec{F}_i is the force on *ith* CC/TP (see equation 5 and 6), and γ is the damping term. The details including the rationale for neglecting the random forces and inertial forces, are given elsewhere 14 . The role of internal friction generated by cell-cell adhesion is neglected. We had accounted for internal friction in our previous work 14 , and found that the value was was small compared to the drag from the matrix. One could include such terms by following a procedure used recently in⁵³ if the viscosity of the ECM is small.

Radial dependence of pressure: We calculated the dependence

of pressure $P(r)$ at a distance *r* from the center of the tumor using,

$$
P(r) = \frac{1}{n(r)} \sum_{i=1}^{n(r)} p_i,
$$
 (10)

where p_i is the pressure (Eq.(7)) on the i^{th} cell located between *r* and $r + dr$, and $n(r)$ is the number of cells in the same annulus. It is worth remembering that if the MCS is at equilibrium, $P(r)$ would be independent of *r*. Because in the simulations, cell proliferation is enhanced compared to the core of the MCS, there is more rapid relaxation of the stress at the MCS boundary, which result in reduced pressure.

In order to show that cell division is more likely to occur away from the core than in the interior of the MCS, we also calculated distribution $(P(p_i(r)))$ of pressure experienced by cells in the annulus between *r* and $r + dr$. We calculated $p_i(r)$ using the equation similar to Eq.(10) except in the $n(r)$ is the number of cells in the spherical volume between r and $r + dr$.

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