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Binary, ternary and quaternary mixture toxicity of benzo[a]pyrene, arsenic, cadmium and lead in HepG2 cells

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24 **ABSTRACT**

25 Polycyclic aromatic hydrocarbons (PAHs) and heavy metal/lloid(s) are common
26 environmental pollutants. Toxicological interaction data on benzo[a]pyrene (B[a]P) and
27 heavy metal/lloid(s) are lacking. In this study, we have determined the combined toxicity of
28 B[a]P, arsenic (As), cadmium (Cd) and lead (Pb) in HepG2 cells. Binary, ternary and
29 quaternary mixture toxicity of B[a]P and heavy metal/lloid(s) was predicted by using the
30 combination index (CI)- isobologram method. This method is useful to predict quantitative
31 nature of interaction between chemicals at different effect (inhibitory concentration) levels
32 from 0.1 to 99% using computerised quantitation. A total of 11 mixtures including six binary
33 mixtures, four ternary and one quaternary mixtures of B[a]P and heavy metal/lloid(s) were
34 evaluated for their interactions. Cytotoxicity of individual and multi-component mixtures was
35 evaluated by MTS assay. The selected concentrations for individual dose response study was
36 0-100 μ M - B[a]P; 0-40 μ M - Cd; 0-400 μ M - As and Pb. The individual dose response
37 results showed that all four chemicals were toxic to liver cells with Cd being the most potent
38 toxicant. Mixtures of B[a]P and heavy metal/lloid(s) were prepared based on their individual
39 D_m concentration using 1:1 ratio and exposed to HepG2 cells. By using the CI-isobologram
40 method, the predicted interactions between these chemicals were synergism, additivity or
41 antagonism at different effect levels. All the mixtures except ternary mixture of B[a]P + As +
42 Pb displayed synergism at lower effect level (IC_{10} - IC_{30}), additivity, synergism or antagonism
43 at 50-90% effect levels. Among these mixtures, mixtures of heavy metal/lloid(s) (both binary
44 and ternary combinations) and quaternary mixture of B[a]P + As + Cd+ Pb showed strong
45 synergistic response at lower effect levels compared to other mixtures. The predicted
46 interaction response by the CI method was compared with classical models of concentration
47 addition and independent action. The CI method displayed improved prediction power
48 compared to classical models. The predicted synergistic interaction between B[a]P and heavy

49 metal/loid(s) may have important implication on human health risk assessment of these
50 mixed chemicals mixtures at contaminated sites.

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68 **ABBREVIATIONS**

69 As – Arsenic

70 ATCC- American type culture collection

71 ATSDR- Agency for toxic substances and disease registry

72 B[a]P- Benzo[a]pyrene

73 CA- Concentration addition

74 Cd- Cadmium

75 CI- Combination index

76 Cr- Chromium

77 D_m- Concentration for 50% effect (e.g., 50% inhibition of cell viability or inhibitory
78 concentration)

79 DMEM- Dulbecco's Modified Eagle Medium

80 DMSO- Dimethyl sulfoxide

81 EDTA- Ethylene diamine tetra acetic acid

82 H- hours

83 HepG2- Human hepatocellular carcinoma cell

84 IA- Independent action

85 IARC- International agency for research on cancer

86 IPQ- Index on prediction quality

- 87 MT- Metallothionein
- 88 MTS – tetrazolium compound [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-
- 89 (4-sulfophenyl)-2H-tetrazolium]
- 90 OD- Optical density
- 91 PAHs- Polycyclic aromatic hydrocarbons
- 92 Pb- Lead
- 93 PBS- Phosphate buffered saline
- 94 r- Linear correlation coefficient of the median-effect plot
- 95 μ M- Micro molar
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INTRODUCTION

Benzo[a]pyrene (B[a]P), a polycyclic aromatic hydrocarbon (PAH) is formed during the process of incomplete combustion of organic material (eg. gasoline and wood) and found widely in tobacco smoke and grilled foods. Anthropogenic and industrial activities results in elevated concentration of B[a]P in environment.¹ B[a]P has been classified as class I carcinogen by IARC and it also causes teratogenicity, neurotoxicity and immunotoxicity.²

Arsenic (As), cadmium (Cd) and lead (Pb) are naturally occurring substances in the environment. Besides their natural occurrence, manufacturing and industrial activities such as mining, smelting operations, storage and disposal of waste from these process results in elevated concentrations of these contaminants in the environment.³ Human exposure to heavy metal/lloid(s) has been associated with a wide range of toxic effects.⁴ Exposure to arsenic causes skin toxicity, liver injury, peripheral neuropathy, cardiovascular diseases and immunotoxicity.⁵ Cd exposure results in testicular damage, pulmonary edema, osteomalacia, renal and hepatic dysfunction in humans.⁶ Pb causes reduction in haemoglobin synthesis, neurological, neurobehavioral and developmental effects in children.⁷ Furthermore, As and Cd are classified as class I carcinogen and Pb is classified as Class II carcinogen.

As, Pb, Cd and B[a]P have been ranked as 1, 2, 7 and 8th priority pollutants as per the Agency for Toxic Substances and Disease Registry (ATSDR) substance priority list.⁸ Mixed contamination of PAHs and heavy metal/lloid(s) are ubiquitous in environment and has received significant attention due to their known adverse health effects to humans and other organisms.⁹⁻¹² Soil is one of the major sink for these mixed contaminants and their co-occurrence at various contaminated sites around the world has been reported.¹³⁻¹⁷

Humans are very rarely exposed to individual contaminants of B[a]P or heavy metal/lloid(s), exposure is more likely to mixture of chemicals from the contaminated sites. The available

130 toxicity data for B[a]P and heavy metal/lloid(s) mixtures is very limited for their risk
131 assessment. Human health assessment of B[a]P and heavy metal/lloid(s) is carried out based
132 on their individual toxicity. The reported interaction toxicity data for mixtures of As, Cd and
133 Pb is limited to their binary combinations and the data are inconsistent for same endpoints
134 from study to study and less relevant in terms of end points compared to their individual
135 toxicity data.¹⁸ In the literature, only two reports were available for metal mixtures toxicity
136 beyond their binary combinations.¹⁹⁻²⁰ In case of B[a]P and metal mixtures, few studies were
137 reported on As and Cd effect on B[a]P's genotoxicity and metabolism.²¹⁻²⁴ To the best of our
138 knowledge there is no detailed study available with respect to assessment of multi-component
139 mixture toxicity of B[a]P and heavy metal/lloid(s).

140 One possible route for human exposure for these chemicals from soil is by oral ingestion.
141 Liver, the organ for biotransformation is one of the main target organs for exposed chemicals.
142 B[a]P and heavy metal/lloid(s) accumulate in liver followed by their absorption and cause
143 toxicity to liver cells.²⁵⁻³³ In this study, we have used HepG2 (hepatocellular carcinoma) cell
144 line to evaluate the interaction toxicity. This cell line is well established for toxicological
145 research and possesses a liver-like enzyme pattern including the enzymes for
146 biotransformation and the level of CYP enzymes expression although it is lower compared to
147 primary hepatocytes.³⁴⁻³⁵

148 Concentration addition (CA) and independent action (IA) models are extensively used to
149 predict the mixture toxicity of chemicals.³⁶⁻³⁸ Combination Index (CI) method developed by
150 Chou and Talalay³⁹ is commonly used for drug combination studies and in recent years this
151 method has been used by various researchers to determine the interaction between
152 environmental chemicals for ecotoxicity risk assessment.⁴⁰⁻⁴² One distinct advantage of this
153 model is prior knowledge about chemicals mode of action is not required in order to
154 determine their interaction.

The objective of this study is to determine the interaction cytotoxicity of B[a]P, As, Cd and Pb in HepG2 cells. Binary, ternary and quaternary mixture toxicities of B[a]P and heavy metal/lloid(s) were predicted by using the combination index (CI)- isobologram method.

MATERIALS & METHODS

Chemicals

Cell culture medium DMEM (Dulbecco's Modified Eagle Medium), Trypsin-EDTA (0.25%), penicillin-streptomycin solution and fetal bovine serum (FBS) were purchased from Gibco[®] (Life Technologies, VIC, Australia). CellTiter 96[®] Aqueous One Solution Cell Proliferation Assay (G3581) was purchased from Promega Corporation, Madison, USA. Benzo[a]pyrene (B[a]P) (CAS ID-50-32-8), cadmium chloride (CAS ID-10108-64-2), lead acetate (CAS ID-6080-56-4) and sodium arsenite (CAS ID-7784-46-5) were purchased from Sigma-Aldrich, St. Louis, USA.

Cell culture

HepG2 cells were obtained from American type culture collection (ATCC), USA (ATCC No. HB-8065). This cell line is derived from a liver hepatocellular carcinoma of a 15 year old Caucasian male. After thawing, cells were maintained as sub confluent monolayer in a 75 cm² culture flask using the modified DMEM medium with 10% (v/v) FBS + penicillin-streptomycin (50 Units/ml). Cells were used for experiment after two week of thawing from cryopreserved stock. On the day of harvest, the cell monolayer was rinsed with 1 x PBS and pre-warmed 0.25% trypsin-EDTA solution was added. Cells (1×10^4 cells/well) were seeded into 96 well plates and allowed to attach for 24 h before chemical treatment. During the experiment period, cells were maintained in a CO₂ incubator at 37 °C under 5% CO₂.

178 Stock solution of B[a]P was prepared in DMSO, whereas solutions of heavy metal/lloid(s)
179 (As, Cd and Pb) were prepared in MilliQ water respectively. Working solutions were
180 prepared in DMEM medium and added to the plates reaching a final concentration of 1% v/v
181 of vehicle control (DMSO or MilliQ water).

182 **Chemical Treatment**

183 Individual dose response was evaluated for B[a]P and heavy metal/lloid(s) by exposing them
184 to HepG2 cells for 24 h. Selected concentrations were 0 to 100 μ M- B[a]P, 0 to 40 μ M -Cd
185 and 0 to 400 μ M for both As and Pb. Treatment was carried out in triplicate for vehicle
186 (DMSO or MQ water) and chemicals. The details of selected individual concentrations were
187 provided in Table S1 of supplementary information.

188 **Cytotoxicity assay**

189 Cell viability was determined using CellTiter 96® aqueous one solution, which is a
190 colorimetric method for determining the number of viable cells. This solution contains a
191 tetrazolium compound, MTS [3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-
192 (4-sulfophenyl)-2H-tetrazolium, inner salt] and an electron coupling reagent PES (phenazine
193 ethosulfate). The principle of this assay is based on metabolic reduction of tetrazolium salts.
194 Viable cells with active metabolism convert MTS salt into purple colored formazan product.
195 The exact mechanism of metabolic reduction is not well understood, enzymes from
196 endoplasmic reticulum, mitochondria and from cell surface is attributed to the observed
197 metabolic reduction activity.

198 After a chemical treatment period of 24 h, the treatment medium was carefully aspirated
199 using multi-channel micropipette and 20 μ L of MTS reagent + 100 μ L of DMEM was added

200 to each well and incubated for another 2-3 h. The plates were placed in a microplate reader
201 and the absorbance was measured at 490 nm.

202 Cell viability was calculated as shown in equation (1) after blanking.

203
$$\text{Viability (\%)} = \frac{100 \times \text{mean optical density (OD) in single chemical or mixtures treated samples}}{\text{mean OD in vehicle control treated samples}} \text{ ---- (1)}$$

204 Mixture experiments

205 Mixture studies were conducted for a total of 11 mixtures including six binary mixtures (As +
206 Cd, As + Pb, Cd + Pb, B[a]P + As, B[a]P + Cd and B[a]P + Pb), four ternary mixtures (As +
207 Cd + Pb, B[a]P + As + Cd, B[a]P + As + Pb and B[a]P + Cd + Pb) and one quaternary
208 mixture (B[a]P + As + Cd + Pb). Each mixture was prepared based on their individual Dm
209 concentration and mixed at 1:1 ratio. The mixtures were further diluted serially at two fold
210 for 8 times. Three independent experiments were conducted for chemical mixtures and
211 treatment was carried out in triplicate for each concentration and vehicle control (DMSO/or
212 MQ water). The details of individual chemical concentrations in mixtures were provided in
213 Table S1 of supplementary information.

214 Individual and mixtures toxicity determination

215 Individual toxicity of B[a]P, As, Cd and Pb and their binary, ternary and quaternary
216 interaction was determined by using the median effect and combination index (CI)
217 isobologram respectively.³⁹ This method is based on median-effect principle (mass-action
218 law) which defines the relationship between concentration of individual/mixtures and
219 cytotoxic effect, independent of number of chemicals and also mechanism of action. In this
220 method, response for individual and their combination is determined by using the median-
221 effect equation (2).

$$f_a = (D/D_m)^m \quad (2)$$

D is the concentration of chemicals,

D_m is the concentration for 50% effect (e.g., 50% inhibition of cell viability or IC_{50}),

f_a is the fraction affected by dose D (e.g., 0.50 if cell viability is inhibited by 50%),

f_u is the unaffected fraction (therefore, $f_a = 1 - f_u$), and

m is the coefficient of the sigmoidicity of the dose-effect curve: $m = 1$, $m > 1$, and $m < 1$

indicate hyperbolic, sigmoidal, and negative sigmoidal dose-effect curve, respectively.

D_m and m values for each chemical were calculated by using median-effect plot by taking

account of both potency (D_m) and shape (m) parameters. These parameters (D_m and m) were

used to calculate concentrations of individual chemical and their combination required to

produce various effect levels (e.g. 10, 30, 50 and 90% effect) by using the median-effect

equation.

Combination index (CI) values were calculated for n number of chemical combination at x%

effect by using equation (3).

$${}^n(CI)_x = \sum_{j=1}^n \frac{(D_j)}{(D_x)} = \sum_{j=1}^n \frac{(D_x)^{1-n} \{ [D_j] / \sum_1^n [D] \}}{(D_m)_j \{ (f_{ax})_j / [1 - (f_{ax})_j] \}^{1/m_j}} \quad (3)$$

${}^n(CI)_x$ is the combination index for n chemicals at x% inhibition (e.g., cytotoxicity)

$(D_x)_{1-n}$ is the sum of the concentration of n chemicals that exerts x% inhibition in combination

$\{ [D_j] / \sum_1^n [D] \}$ is the proportionality of the concentration of each of n chemicals that exerts x% inhibition in combination

$(D_m)_j \{ (f_{ax})_j / [1 - (f_{ax})_j] \}^{1/m_j}$ is the concentration of each chemicals alone that exerts x% inhibition

f_a is the fraction affected by individual chemical/mixture concentration.

Mixture toxicity prediction by concentration addition, independent action and CI-isobologram method

The predicted dose response by CI method was compared with classical models of CA and IA.

CA model is used for chemicals which are having similar mode of action.

$$\text{Concentration addition (CA)} = EC_{\text{mix}} = \sum_{i=1}^n \left(\frac{p_i}{EC_{xi}} \right)^{-1} \quad (4)$$

EC_{mix} is the effect concentration of the mixture provoking $x\%$ effect

EC_{xi} is the concentration of component i provoking the same effect ($x\%$) as the mixture when applied individually

p_i is the molar concentration ratio of the i^{th} component in the mixture.

For chemicals with different mode of action, interaction is calculated by using equation (5)

$$\text{Independent action (IA)} = E(C_{\text{mix}}) = 1 - \prod_{i=1}^n (1 - E(C_i)) \quad (5)$$

C_{mix} and $E(C_{\text{mix}})$ are the total concentration and total effect of the mixture respectively

$E(C_i)$ is the effect of the i^{th} component with the concentration of ci in the mixture.

The predictive dose response for CI method is computed by using equation (6).

$$CI = \left(\sum_{i=1}^n \frac{p_i}{EC_{xi} \times CI_{\text{comp}}^i} \right)^{-1} \quad (6)$$

CI_{comp} is the computed CI value for each mixtures at different effect level (0.1-97%) from independent experimental toxicity data of the mixtures.³⁵ Dose response for each mixture was predicted using computed CI values from individual mixture toxicity experiment and different f_a levels.

Quantitative relationship between predicted and observed effect was determined by index on prediction quality (IPQ).⁴³ Predicted dose response from three different models CA, IA and

CI was compared with experimental dose response of multi-component mixtures of B[a]P and heavy metal/lloid(s).

If the predicted effect is greater than observed value, the prediction quality is determined by using equation (7)

For $EC_{\text{predicted}} > EC_{\text{observed}}$, $IPQ = \left(\frac{EC_{\text{predicted}}}{EC_{\text{observed}}} \right) - 1$ ----- (7)

If the predicted value is less than observed value, the prediction quality is determined by using equation (8)

For $EC_{\text{predicted}} < EC_{\text{observed}}$, $IPQ = - \left(\frac{EC_{\text{observed}}}{EC_{\text{predicted}}} \right) + 1$ ----- (8)

IPQ value of zero indicates exact prediction by reference models, IPQ value <0 indicates overestimation; values>0 indicates underestimation of mixture effects.

Data analysis

Computer program CompuSyn developed by Chou and Martin⁴⁴ was used for the calculation of dose-effect curve parameters, CI values and f_a -CI plot. The details of CI values and corresponding descriptions are presented in Table 1.

RESULTS

Individual toxicity of B[a]P and heavy metal/lloid(s)

The individual dose response cytotoxicity results showed that both B[a]P and heavy metal/lloid(s) were toxic to HepG2 cells and Cd was found to be the most toxic compared to other chemicals with D_m of 2.70 μ M followed by B[a]P (D_m -37 μ M). The D_m concentration for As and Pb is 159 and 217 μ M respectively. Dose response curve parameters D_m (concentration for 50% effect on cell viability or IC_{50}), m (coefficient of the sigmoidicity of the dose-effect curve), r (linear correlation coefficient of the median-effect plot) for individual chemicals are presented in Table 2.

Toxicological interactions of multi-component mixtures of B[a]P and heavy metal/lloid(s)

Dose effect curve parameters (D_m , m and r) for binary mixtures of heavy metal/lloid(s) (As + Cd, Cd+ Pb and As+ Pb), binary mixtures of B[a]P with heavy metal/lloid(s) (B[a]P + As, B[a]P + Cd and B[a]P + Pb), ternary mixtures (Cd + As + Pb, B[a]P + As + Cd, B[a]P + AS + Pb and B[a]P + Cd + Pb) and quaternary mixtures (B[a]P + As + Cd+ Pb) and their cytotoxicity to HepG2 cells are summarised in Table 2. Different type of interaction (synergism, antagonism or additivity) between these mixtures and combination index plot (f_a -CI plot) are presented in Figure 1. For each mixture, average CI values for four effect level (IC_{10} , IC_{30} , IC_{50} and IC_{90}) are summarised in Table 2.

Binary mixture toxicity of heavy metal/lloid(s)

All the three heavy metal/lloid(s) mixtures (As + Cd, Cd + Pb and As + Pb) displayed synergistic response at 10% level with strong synergism was observed for mixture of As + Pb with CI value of 0.25. The observed synergism was continued up to 30 and 50% level for Cd+ Pb and As + Pb mixtures respectively. Binary mixture of As + Cd showed additive response at 30-50% level. All the mixtures showed antagonistic effect at IC_{90} level and mixture of As + Pb displayed strong antagonism (CI value 3.3).

Binary mixture toxicity of B[a]P and heavy metal/lloid(s)

Binary mixture of B[a]P + Cd showed synergistic effect up to 50% level and antagonism at 90% level. Mixtures of B[a]P + As, B[a]P + Pb displayed synergism at 10-30% level, additivity at 50% level and antagonism at 90% level. All three mixtures displayed same levels of synergistic response at lower effect levels (10-30% level).

Ternary and quaternary mixtures toxicity of B[a]P and heavy metal/lloid(s)

313 Ternary mixtures of B[a]P + As + Cd and B[a]P + Cd + Pb displayed synergism at 10-50%
 314 level and antagonism at 90% level. Mixture of B[a]P + As + Pb displayed antagonistic
 315 response for entire effect level (10-90%). Ternary mixture of heavy metal/lloid(s) showed
 316 synergism at 10-50% level and antagonism at 90% level. Among all ternary mixtures, heavy
 317 metal/lloid(s) mixtures showed strong synergism at lower effect level with CI value of 0.34.

318 In case of quaternary mixture, synergism was observed up to 50% effect level (strong
 319 synergism at 10% level, CI-0.29) and antagonism at 90% level respectively.

320 **Comparison of mixture effect predicted by CA, IA and CI methods**

321 The dose response parameters and predicted dose response effect for all three models CA, IA
 322 and CI along with experimental data are presented in Table 3 and Figures 2 to 4 respectively.

323 The IPQ value for all three models were less than one for all the mixtures except ternary
 324 mixture of B[a]P + As +Pb. This indicates good agreement between observed and predicted
 325 effects. An individual comparison of IPQ values for three reference models (CA, IA and CI)
 326 showed that prediction power of CI model is superior to classical models of CA and IA
 327 (comparatively IPQ value for CI method is less for six out of 11 mixtures compared to CA
 328 and IA models). In addition, CI method accurately predicts the synergistic interaction
 329 between these mixtures.

330 **DISCUSSION**

331 In this study, we have determined the multi-component interaction between B[a]P and heavy
 332 metal/lloid(s) in HepG2 cells by exposing the chemicals mixtures for 24 h and the CI method
 333 was used to predict the mixture toxicity. The treatment period of 24 h was selected based on
 334 the doubling time of HepG2 cells (24 h) and also as reported by International Workshop on *In*
 335 *Vitro* Methods for Assessing Acute Systemic Toxicity.⁴⁵ Cytotoxicity of individual chemical

and mixtures was determined by MTS assay. Various methods including viability assays based on metabolism reductase activity (MTS, XTS, resazurin dye reduction assays), bioluminescent ATP assays and enzyme linked cytotoxicity assays (LDH leakage assays) are used to determine cytotoxicity or cell viability. Each assay is having its own advantages and disadvantages. Eg. Neutral red viability assay is a classic example of a scientifically useful and validated method which is impractical and rarely used. MTS assay has been widely used with great success in toxicity testing. This assay has been well characterized and it is stable, cost-effective, simple to use and suitable for high-throughput testing.⁴⁶

The results showed different kind of interactions between B[a]P and heavy metal/lloid(s). Synergism was the predominant response at lower effect level (IC₁₀₋₃₀) for all mixtures with the exception of ternary mixture of B[a]P + As+ Pb. In case of binary mixtures, all six mixtures showed various degree of synergism (strong to moderate synergism) at these effect levels. The percentage of binary mixtures displaying synergism at IC₁₀, IC₃₀, and IC₅₀ was 100, 83 and 33% respectively. For ternary and quaternary mixtures, the observed synergism at IC₁₀ was extended up to IC₅₀ level. A total of 75% of ternary mixtures showed synergistic response at IC₁₀₋₅₀ level.

The observed effect for heavy metal/lloid(s) mixtures toxicity i.e. synergism and antagonism at low and higher effect level, respectively is supported by findings of Bae *et al.*¹⁹ and Klutse *et al.*²⁰ Cytotoxic interactions among As, Cd, Cr and Pb was determined by Bae *et al.*¹⁹ using four immortal human keratinocytes and observed similar findings of synergism and antagonism at lower and higher dose of mixtures respectively. Klutse *et al.*²⁰ studied the interaction between As, Cd, Pb and Hg in MCF cell line and observed that combined toxicity changed from synergistic to additive to antagonistic with increasing concentration of the metal mixtures.

One possible explanation for observed antagonistic response at higher effect level is due to presence of strong defense mechanism in the liver. A low molecular weight protein metallothionein (MT) synthesis was upregulated followed by Cd exposure which protects the liver against Cd toxicity.⁴⁷⁻⁴⁸ Arsenic also induces MT genes and heat shock protein which may reduce the toxicity of As and of Cd by detoxification.⁴⁹⁻⁵⁰ Cellular glutathione level plays an important role in resistance to metal toxicity.⁵¹⁻⁵⁴ Heavy metal/loid(s) (As, Cd and Pb) are also reported to diminish the toxicity (carcinogenic potential) of B[a]P by decreasing B[a]P mediated induction of its bioactivation by CYP1A1.²³ In addition, Nrf2 (nuclear factor erythroid 2 -related factor 2)-antioxidant response element (ARE) pathway is activated followed by these chemicals exposure. Nrf2-ARE pathway plays a major role in protection of cells from heavy metal/loid(s) and B[a]P induced oxidative-stress.⁵⁵⁻⁵⁹ In our laboratory, we have used HepG2-Nrf2 reporter system to confirm the activation of Nrf2-ARE pathway by B[a]P and heavy metal/loid(s) (data not shown). Co-exposure of Cd and B[a]P resulted in significant increase in expression of Nrf2-responsive genes and cellular glutathione levels.⁶⁰ Combined defense activities like Nrf2-ARE pathway activation, induction of MT and heat shock protein, antioxidant protective effect of glutathione and metabolic inhibition of B[a]P by metal/loid(s) may contribute to protection of cells from mixture toxicity and the observed antagonistic response at higher effect level of mixtures.

The observed interaction between B[a]P and heavy metal/loids especially synergism at different effect level cannot be explained in terms of the known mechanism of action. In few cases these metal/loid(s) were reported to enhance the toxicity of B[a]P. Arsenic was found to have synergistic effect on cytotoxicity of B[a]P in human lung carcinoma cells⁶¹ and also potentiate the genotoxicity of B[a]P.^{21,61-62} In case of Cd and B[a]P combination, the available report for their genotoxicity is inconsistent as both synergistic^{22,63} and antagonistic effect was reported by various authors.^{20,57} There is no detailed report available for these

mixture toxicity especially for higher order mixtures (ternary and quaternary). We have reported the mixture toxicity of multi- component mixtures of B[a]P and heavy metal/loids(s) and showed different nature of interactions between these mixtures.

CA and IA models are traditionally used to predict the mixture toxicity effects of chemicals. Cedergreen et al.,⁶⁴ reported that only half of the 158 mixture data was accurately predicted by either CA or IA models. CI method was successfully used to study the interaction between drugs/chemicals for ecotoxicity risk assessment and reported to have improved predictive power compared to classical models.⁴¹⁻⁴³ We have used CI method to predict the interaction between environmental chemicals using human cell lines and observed that this method is useful to predict the synergistic interaction between B[a]P and heavy metal/loid(s) at lower effect level.

CONCLUSIONS

Binary, ternary and quaternary mixture toxicity of B[a]P, As, Cd and Pb was determined in HepG2 cells and the CI-isobologram method was used to predict the mixture toxicity. The results showed that the nature of interaction varies according to the effect levels and number of components in the mixtures. This multi-component mixture of B[a]P and heavy metal/loid(s) displayed synergistic and antagonistic interaction at low and higher effect levels respectively. The observed synergistic effect increases with complexity of mixture. Given the complexity of the chemical-biological interaction in the cells, further interaction studies on other endpoints such as oxidative stress, genotoxicity and toxicokinetic interaction can provide a more complete data set for health risk assessment of these mixed contaminants. CI method was found to have improved predictive power of CA and IA models. The predicted synergism among these common environmental contaminants may have important implications on human risk assessment.

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CONFLICT OF INTEREST

There is no conflict of interest.

SUPPLEMENTARY INFORMATION

Supplementary information to article is attached.

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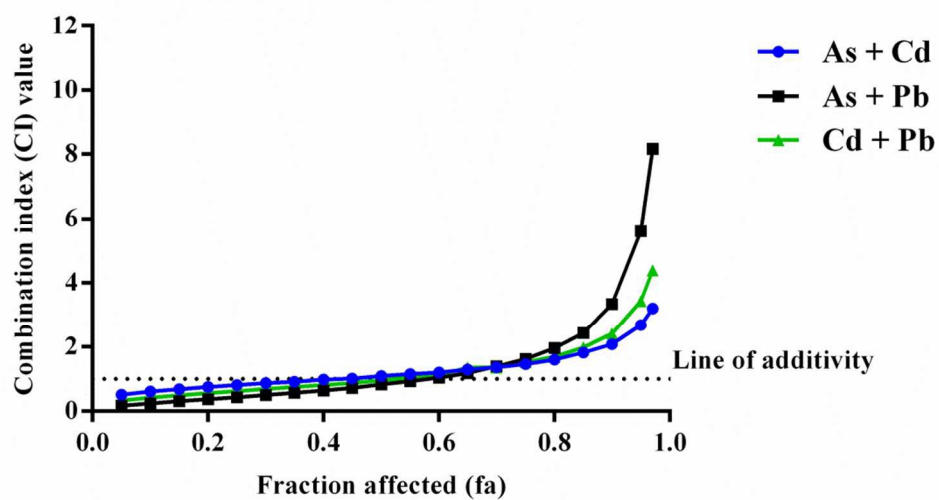
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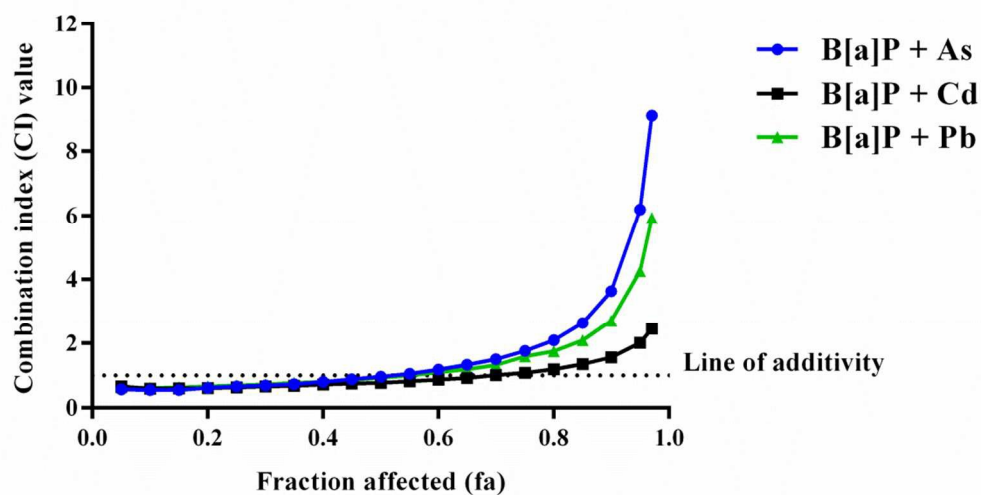
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Binary mixtures of As, Cd and Pb



Binary mixtures of B[a]P, As, Cd and Pb



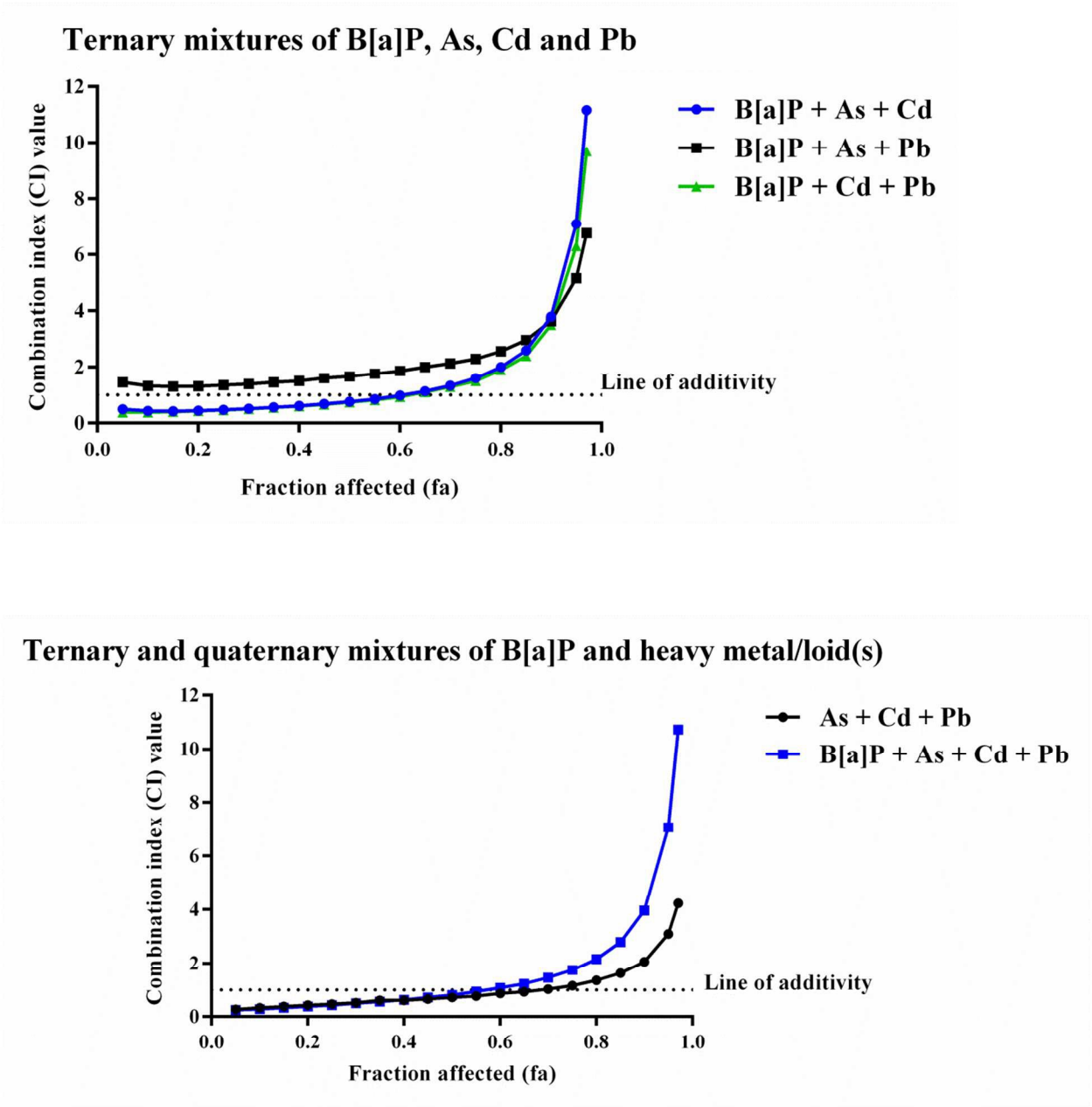
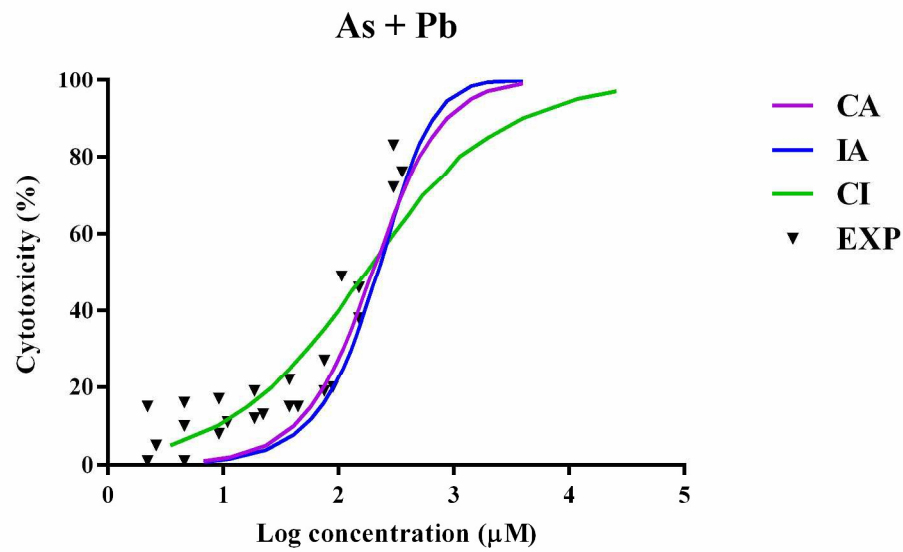
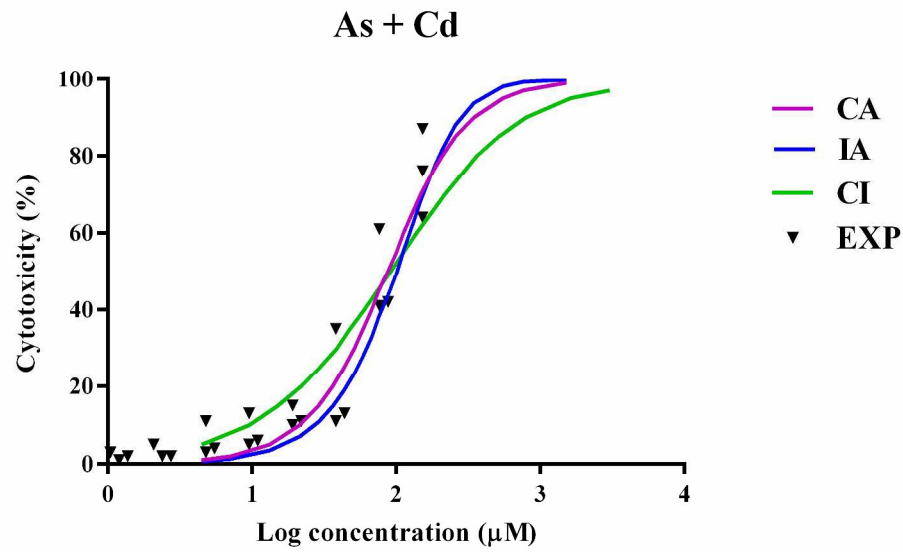


Figure 1. Combination index plot (f_a -CI plot) for multi-component mixtures of benzo[a]pyrene (B[a]P) and heavy metal/loid(s) (arsenic (As), cadmium (Cd) and lead (Pb) in HePG2 cells. CI values are plotted as a function of cell viability (f_a). CI < 1, = 1 and > 1 indicates synergism, additive and antagonism respectively.



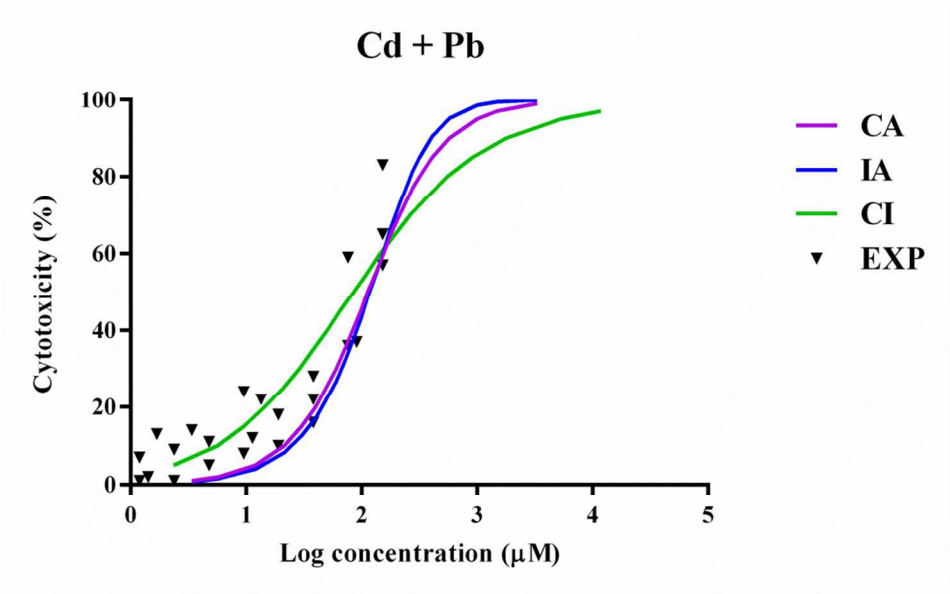
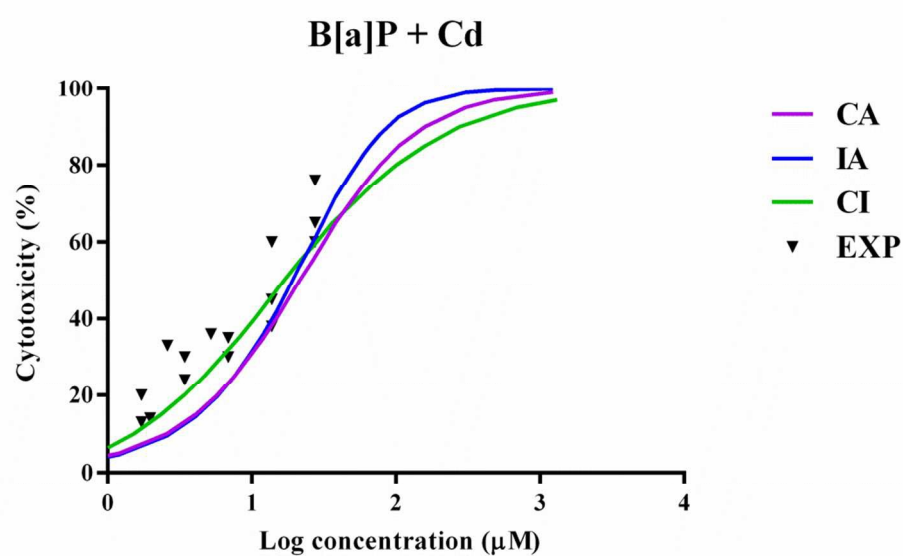
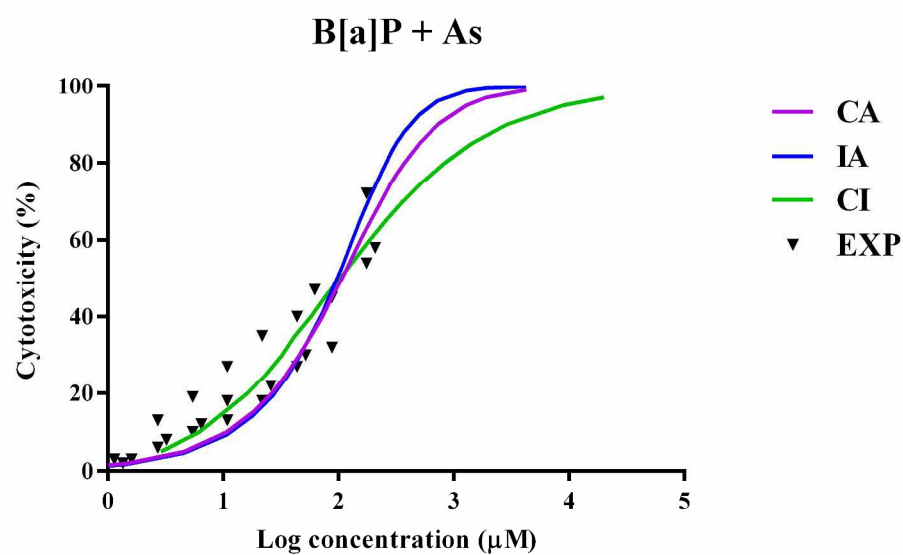


Figure 2: Comparison of dose-response effect of binary mixtures of heavy metal/loid(s) (As + Cd, As + Pb and Cd + Pb) predicted by concentration addition (CA), independent action (IA), combination-index (CI) methods and observed experimental toxicity effect (EXP) in HepG2 cells. Experimental values are expressed as mean values from three independent experiments. As –arsenic; Cd –cadmium and Pb-lead.



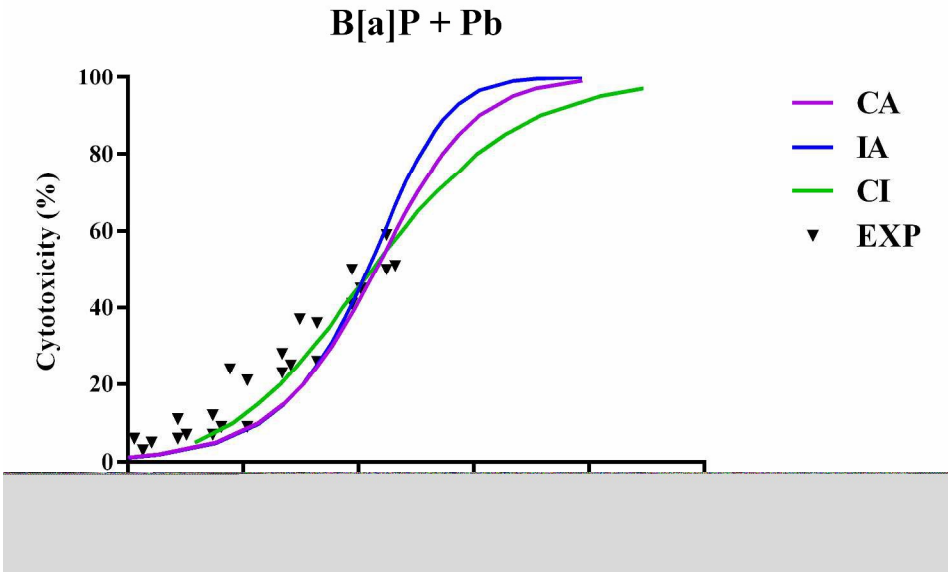
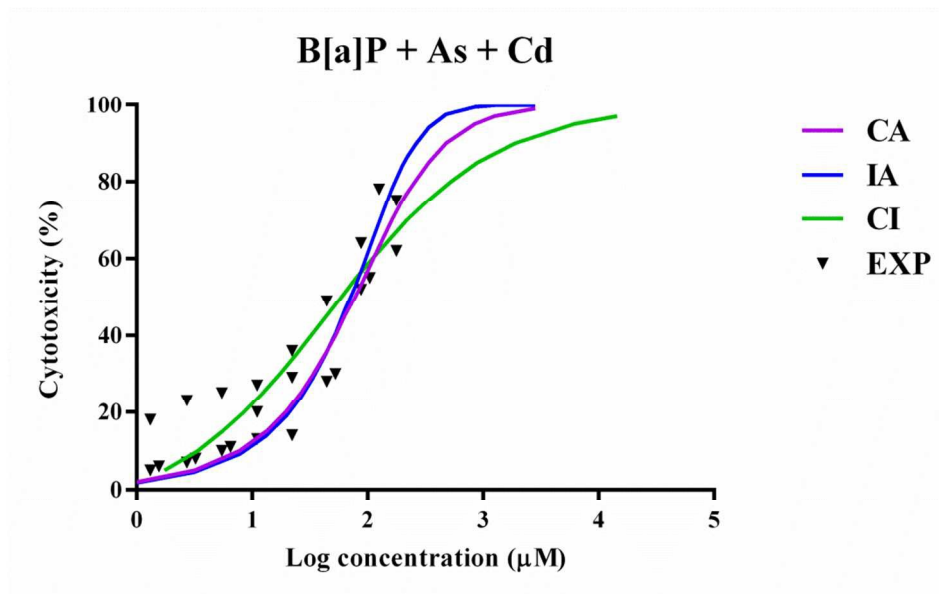
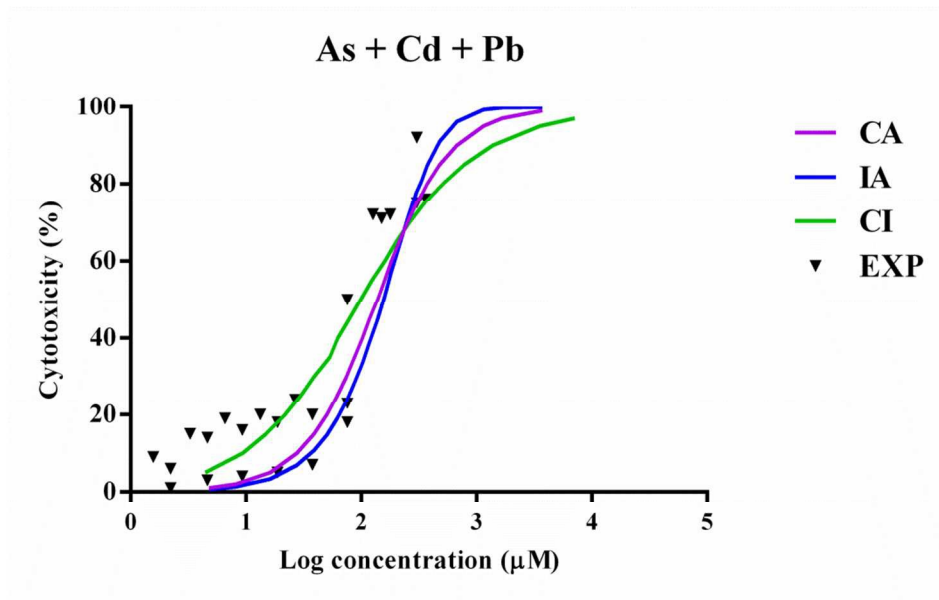
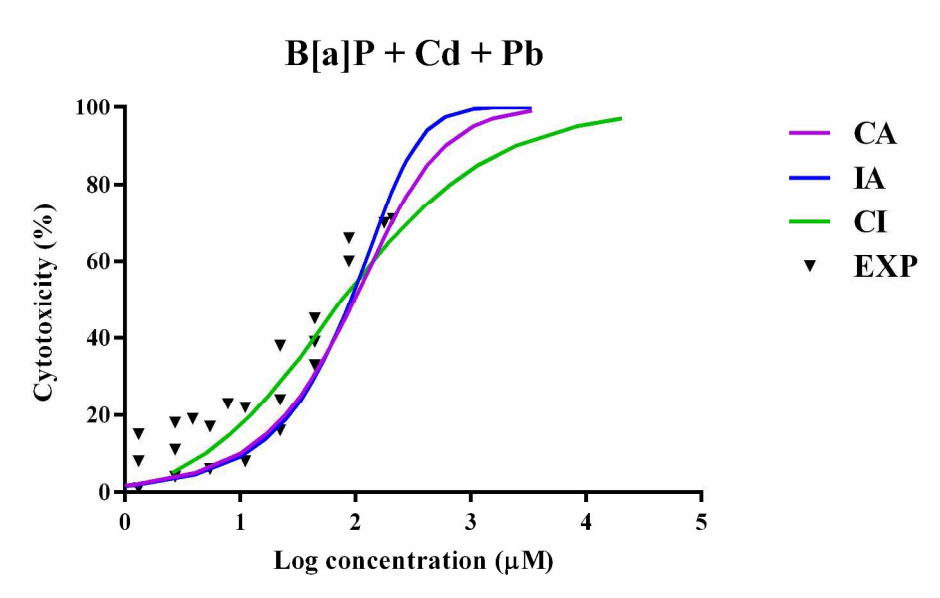
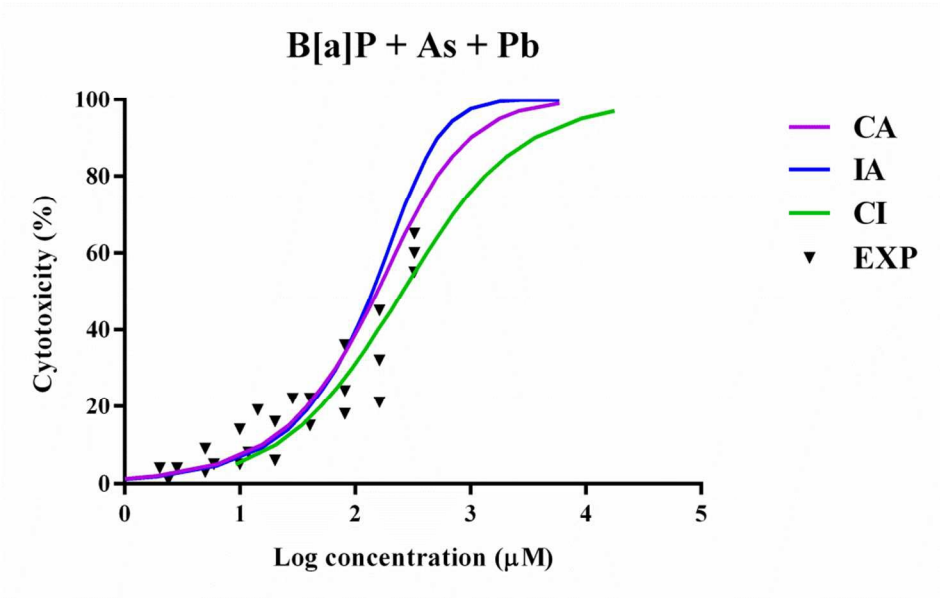


Figure 3: Comparison of dose-response effect of binary mixtures of B[a]P and heavy metal/lloid(s) (B[a]P + As, B[a]P + Cd and B[a]P + Pb) predicted by concentration addition (CA), independent action (IA), combination-index (CI) methods and observed experimental toxicity effect (EXP) in HepG2 cells. Experimental values are expressed as mean values from three independent experiments. B[a]P – benzo[a]pyrene; As –arsenic; Cd -cadmium and Pb-lead.





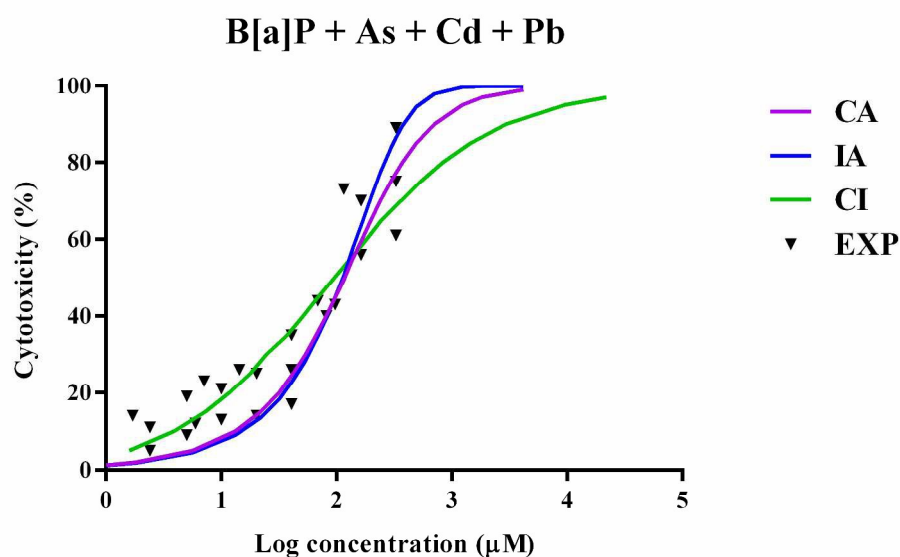


Figure 4: Comparison of dose-response effect of multi-component mixtures of B[a]P and heavy metal/lloid(s) predicted by concentration addition (CA), independent action (IA), combination-index (CI) methods and observed experimental toxicity effect (EXP) in HepG2 cells. Experimental values are expressed as mean values from three independent experiments. B[a]P – benzo[a]pyrene; As –arsenic; Cd -cadmium and Pb-lead.

Table 1: Description for combination index values

Combination index (CI) range	Description
<0.1	Very strong synergism
0.1-0.3	Strong synergism
0.3-0.7	synergism
0.7-0.85	Moderate synergism
0.85-0.90	Slight synergism
0.90-1.10	Nearly additive
1.10-1.20	Slight antagonism
1.20-1.45	Moderate antagonism
1.45-3.3	antagonism
3.3-10	Strong antagonism
>10	Very strong antagonism

Table 2: Dose response parameters and combination index (CI) values for multi-component mixtures of B[a]P and heavy metal/lloid(s) in HepG2 cells

CI values for different effect levels and description											
Chemical combination	D _m (μM)	m	r	IC ₁₀		IC ₃₀		IC ₅₀		IC ₉₀	
As	159	1.5 ± 0.25	0.94	---		---		---		---	
Cd	2.70	1.4 ± 0.10	0.97	---		---		---		---	
Pb	217	1.2 ± 0.32	0.96	---		---		---		---	
B[a]P	37.0	0.9 ± 0.11	0.97	---		---		---		---	
Binary mixtures- heavy metal/lloid(s) mixtures											
As + Cd	87.7	1.0 ± 0.17	0.97	0.62 ± 0.27	SYN	0.88 ± 0.36	ADD	1.1 ± 0.36	ADD	2.1 ± 0.43	ANT
As + Pb	170.0	0.78 ± 0.30	0.92	0.25 ± 0.19	SYN	0.51 ± 0.22	SYN	0.83 ± 0.23	SYN	3.3 ± 0.92	ANT
Cd + Pb	93.2	0.91 ± 0.07	0.94	0.43 ± 0.17	SYN	0.69 ± 0.35	SYN	0.97 ± 0.55	ADD	2.40 ± 1.5	ANT
Binary mixtures- B[a]P + heavy metal/lloid(s) mixtures											
B[a]P + As	109.5	0.73 ± 0.07	0.97	0.55 ± 0.48	SYN	0.70 ± 0.36	SYN	0.97 ± 0.38	ADD	3.6 ± 1.8	ANT
B[a]P + Cd	13.7	0.88 ± 0.02	0.95	0.60 ± 0.30	SYN	0.66 ± 0.25	SYN	0.78 ± 0.37	SYN	1.6 ± 0.90	ANT
B[a]P + Pb	132.7	0.68 ± 0.05	0.97	0.64 ± 0.30	SYN	0.73 ± 0.13	SYN	0.93 ± 0.26	ADD	2.7 ± 0.57	ANT
Ternary mixtures of B[a]P and heavy metal/lloid(s)											
As+ Cd + Pb	101.7	0.93 ± 0.24	0.92	0.34 ± 0.29	SYN	0.52 ± 0.28	SYN	0.72 ± 0.22	SYN	2.1 ± 0.81	ANT
B[a]P + As + Cd	73.9	0.67 ± 0.12	0.98	0.43 ± 0.39	SYN	0.52 ± 0.26	SYN	0.77 ± 0.26	SYN	3.8 ± 1.5	ANT
B[a]P + As + Pb	305	0.74 ± 0.14	0.97	1.33 ± 0.76	ANT	1.40 ± 0.62	ANT	1.67 ± 0.64	ANT	3.6 ± 1.0	ANT
B[a]P + Cd + Pb	67.0	0.74 ± 0.23	0.97	0.53 ± 0.52	SYN	0.59 ± 0.36	SYN	0.78 ± 0.20	SYN	3.3 ± 1.5	ANT
Quaternary mixtures of B[a]P and heavy metal/lloid(s)											
B[a]P + As + Cd + As	68.0	0.7 ± 0.13	0.95	0.29 ± 0.19	SYN	0.50 ± 0.25	SYN	0.81 ± 0.35	SYN	4.0 ± 1.4	ANT

Values are mean or mean ± SD for D_m, m and CI values; Values are from three independent experiments for each mixture. D_m- concentration for 50% effect on cell viability; m- coefficient of the sigmoidicity of the dose-effect curve; r-linear correlation coefficient of the median-effect plot; CI- Combination index; IC- Inhibitory concentration; ADD-additivity; ANT-antagonism; SYN-synergism.

Table 3: Comparison of experimental data and dose response effect predicted by concentration addition (CA), independent action (IA) models and combination index- isobologram (CI) method.

Prediction by CA, IA and CI method													IPQ		
Experimental data			CA			IA			CI			CA	IA	CI	
D _m	m	r	D _m	m	r	D _m	m	r	D _m	m	r				
Binary mixtures- heavy metal/lloid(s) mixtures															
As + Cd	87.7	1.0 ± 0.17	0.97	84.25	1.4	0.99	73.3	1.8	0.99	102.4	0.91	0.99	-0.04	-0.20	0.17
As + Pb	170.0	0.78 ± 0.30	0.92	212	1.4	0.99	174.0	1.6	0.99	182	0.74	0.91	0.24	0.02	0.07
Cd+ Pb	93.2	0.91 ± 0.07	0.94	119.3	1.2	0.99	96	1.5	0.99	110	0.86	0.99	0.28	0.03	0.18
Binary mixtures- B[a]P + heavy metal/lloid(s) mixtures															
B[a]P + As	109.5	0.73 ± 0.07	0.97	86.0	1.0	0.98	57.5	1.2	0.95	102.2	0.70	0.99	-0.27	-0.9	-0.07
B[a]P + Cd	13.7	0.88 ± 0.02	0.95	18.4	1.0	0.99	13.6	1.3	0.97	16.1	0.85	0.99	0.34	-0.007	0.18
B[a]P+ Pb	132.7	0.68 ± 0.05	0.97	116	0.89	0.99	76.3	1.1	0.97	135.3	0.68	0.99	-0.14	-0.74	0.02
Ternary mixtures of B[a]P and heavy metal/lloid(s)															
Cd+ Pb+ As	101.7	0.93 ± 0.24	0.92	135.3	1.4	0.99	117	1.8	0.97	100.3	0.85	0.99	0.33	0.15	-0.01
B[a]P +As + Cd	73.9	0.67 ± 0.12	0.98	62.3	0.92	0.98	40.1	1.2	0.95	86.8	0.63	0.99	-0.19	-0.84	0.13
B[a]P + As + Pb	305	0.74 ± 0.14	0.97	119	0.96	0.98	77.3	1.3	0.96	254	0.84	0.99	-1.6	-2.9	-0.20
B[a]P + Cd + Pb	67.0	0.74 ± 0.23	0.97	75.0	0.91	0.98	47.3	1.3	0.95	63.3	0.7	0.99	0.12	-0.42	0.06
Quaternary mixtures of B[a]P and heavy metal/lloid(s)															
B[a]P + As + Cd + Pb	68.0	0.7 ± 0.13	0.95	86.5	1.0	0.98	53.0	1.4	0.96	46	0.78	0.99	0.27	-0.28	-0.48

Values are mean or mean ± SD for experimental data; For CA, IA and CI- values are mean; D_m- concentration for 50% effect on cell viability; m- coefficient of the sigmoidicity of the dose-effect curve; r- linear correlation coefficient of the median-effect plot; CA- concentration addition; IA- Independent action; CI- Combination index- isobologram; IPQ- index on prediction quality. IPQ= 0 indicates perfect agreement between observed and predicted effect; IPQ < 0-overestimation; IPQ >0-underestimation.

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Highlights:

Multi-component mixtures of B[a]P and heavy metal/lloid(s) toxicity was determined in HepG2 cells. Concentration dependent synergism, additive or antagonism was predicted by combination-index isobologram method.

