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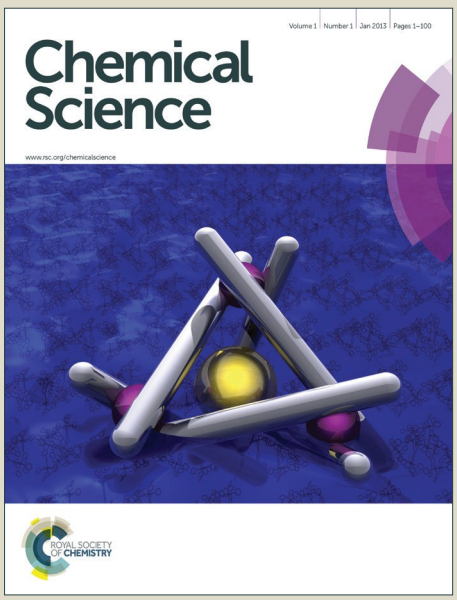

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ARTICLE

A Polyion Complex Sensor Array for Markerless and Noninvasive Identification of Differentiated Mesenchymal Stem Cells from Human Adipose Tissue

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Currently available methods for stem cell evaluation require both prior knowledge of specific markers and invasive cell lysis or staining, hampering development of stem cell product with assured safety and quality. Here, we present a strategy using optical cross-reactive sensor arrays for markerless and noninvasive identification of differentiated stem cell lineages with common laboratory equipment. A sensor array consists of a library of polyion complexes (PICs) between anionic enzymes and synthetic poly(ethylene glycol)-modified polyamines, which can recognize "secretomic signatures" in cell culture supernatants. Due to the reversible nature of PIC formation, incubation of diluted culture supernatants with PICs caused enzyme release through competitive interactions between secreted molecules and PICs, generating unique patterns of recovery in enzyme activity for individual cell types or lineages. Linear discriminant analysis of the patterns allowed not only normal/cancer cell discrimination but also lineage identification of osteogenic and adipogenic differentiation of human mesenchymal stem cells, and therefore providing an effective way to characterize cultured cells in the fields of regenerative medicine, tissue engineering and cell biology.

Introduction

There have been many recent attempts to exploit human stem cells in regenerative medicine, drug discovery, and disease modeling.[1] Every step in the process of stem cell product development must be continuously evaluated for potential safety and quality concerns, from the origin of the cells used through expansion, manipulation, and in some cases preclinical evaluation to eventual engraftment in the host.[2]

For evaluation of cultured stem cells, genetic and phenotypic analyses based on detecting markers for the specific cell state of interest were currently available, such as histochemistry, quantitative polymerase chain reaction assay (qPCR), and flow cytometry, used singly or in combination.[3] However, these methods require (i) prior knowledge of pairs of specific markers and corresponding antibodies, which is not identified in many situations; (ii) invasive cell lysis or staining, hampering continuous evaluation of the cells during the manufacturing process. It will therefore be critical for progress in stem cell research and application to develop a general solution for constructing systems that can identify cell states by a noninvasive and simple assay at desired timing without any information on markers.

Optical cross-reactive sensor array technology has been employed as an alternative to analytical methods using specific binding pairs, such as antibody/antigen.[4] This approach is based on pattern recognition of unique optical responses, i.e., sample signatures, for individual analytes obtained through cross-reactive, rather than specific, interactions between a library of cross-reactive receptors and analytes. Examples of their use for biological molecules include sensor arrays that can discriminate phosphates,[5] saccharides,[6] peptides,[7] and proteins.[8] We have also recently developed sensor arrays consisting of cross-reactive polyion complexes (PICs) for the discrimination of human plasma proteins[9] and structurally similar homologous albumins.[10] To date, optical cross-reactive sensor arrays have been applied to markerless but invasive discrimination of normal/cancer cells[6c,11] and stimulated cell lines[8d] by recognition of cell surfaces,[11a-c,e] lysates[8d,11d] and membrane extracts,[6c] while noninvasive stem cell identification has yet to be demonstrated.

Various molecules are secreted into the external medium from cultured cells. The entire set of secreted proteins is referred to as "the secretome", which reflects the functionality and state of a cell in a given environment and at a given time,[12] and is regarded as a rich source for discovery of cancer biomarkers in the biomedical field.[13] Secretome analyses have recently been used for mesenchymal stem cells (MSCs) to identify the autocrine/paracrine factors applicable in regenerative medicine.[14] MSCs are natural multipotent cells

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present in bone marrow, adipose, placental, and umbilical tissues, and have important roles in immunomodulation and tissue regeneration. Secretome analysis of MSCs using mass spectrometry and gel electrophoresis enabled detection of 100 – 300 proteins or more in the culture supernatants of cells,[15] and the components of secreted proteins were modulated by a variety of stimuli,[14b] including differentiation induction.[15,16] Inspired by recent progress in both optical cross-reactive sensor array and secretome analysis, we have applied a PIC sensor array to recognize "secretomic signatures" of culture supernatants for markerless and noninvasive identification of differentiated MSCs using only a standard microplate reader.

Results and discussion

In this study, an optical sensor array-based system was proposed as shown in Figure 1. A library of PICs between anionic enzymes and poly(ethylene glycol)-modified (PEGylated) polyamines was used as a source of cross-reactive receptors with the ability to translate the interactions between secreted molecules and PICs into a readable signal. The sensing strategy was based on our recent findings,[17] where reversible electrostatic-driven PIC formation between enzymes and PEGylated polyamines was accompanied by decreases in enzyme activity (Figure 1A). PICs may possess different affinities for secreted molecules in culture supernatants, and therefore incubation of culture supernatants with PICs would cause enzyme release through competitive interactions (Figure 1A). Consequently, unique patterns of recovery in enzyme activity for individual cell types or lineages would be generated. A PIC-based library is suited for tuning cross-reactivity to recognize signatures of complex biofluids, as reversible enzyme inhibition generally occurs through PIC formation with counter-charged polymers.[17] Enzyme activity is determined from the rate of increase in the concentration of fluorogenic 4-methylumbelliferone, which is catalytically cleaved from the substrates (see Figure S1). Therefore, background intensity from biofluids can be neglected. For sample preparation, total protein concentration of collected culture supernatants was first determined by the Bradford assay. Supernatants were then diluted for normalization to be able to recognize unique secretomic signatures of each cell type or lineage regardless of the density of cultured cells (Figure 1B).

To achieve recognition of secretomic signatures, we considered that potential candidate PICs possessing a variety of cross-reactivities toward secretomic molecules were required, and therefore we combined our previous strategies for construction of PICs—naturally occurring structural diversity of enzymes[9] and artificial structural diversity of PEGylated polyamines[10]—; six PICs between three anionic enzymes (GAO, GEC, and LAN; Figure 2A) and two synthesized structurally different PEGylated polyamines, quaternized poly(ethylene glycol)-*block*-poly(*N*,*N*-dimethylaminoethyl methacrylate) (PEG-*b*-QPAMA) (**P1**: hydrophilic; **P2**: hydrophobic and aromatic) (Figure 2B). As previously constructed PIC libraries discriminated proteins based

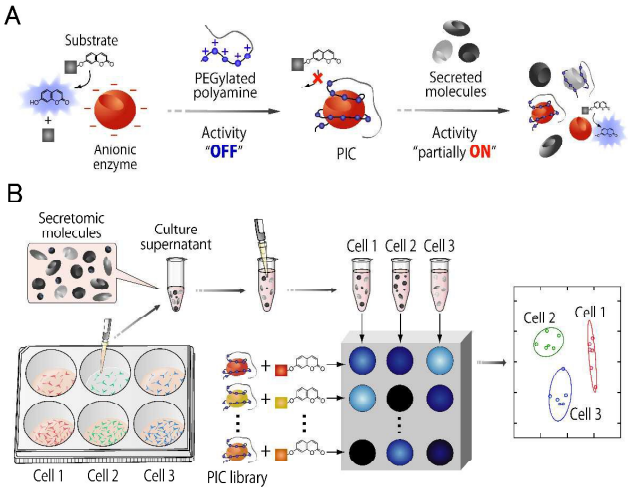
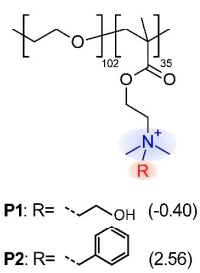


Figure 1. (A) Decrease in catalytic activity of anionic enzymes through reversible PIC formation with PEGylated polyamines, and subsequent partial recovery of activity through competitive interaction with secreted molecules. (B) A PIC sensor array for markerless and noninvasive identification of cell types and lineages. Culture supernatants collected from cells with varying seeding densities were diluted for normalization, followed by addition of PICs to generate activity patterns reflecting secretomic signatures for given cell types or lineages, and then the patterns were interpreted using a chemometric method.

A

Enzyme	Source	Abbr.	MW (kDa)	*p*I
β-Galactosidase	*Aspergillus oryzae*	GAO	110	5.2
β-Galactosidase	*Escherichia coli*	GEC	465	5.1
Lipase	*Aspergillus niger*	LAN	32	4.0

B



C

Cellular category	Source	Name
Cancer cell line	lung	A549
	bone	MG63
	liver	HuH7
Fibroblast	lung	NHLF
	dermal	NHDF
	fetal lung	WI-38
Mesenchymal stem cell	adipose	ADSC

Figure 2. Properties of (A) anionic enzymes, (B) PEGylated polyamines, and (C) human cells used in this study. Log P values of R groups in PEGylated polyamines are shown in parentheses, and these values were calculated by the program ALOPGs.[18]

predominantly on electrostatic signatures of proteins,[9,10] LAN was selected to provide binding affinity toward hydrophobic molecules for PIC libraries, which is expected from its peculiar capacity to adsorb on any hydrophobic interface.[19] From the titration of PEGylated polyamines to enzymes (Figures S1 and 3), a PIC library for analysis of culture supernatants was newly prepared. Note that higher hydrophobicity of R groups in PEGylated polyamines provided a greater effect on the decrease in activity of all enzymes, but differences in the inhibitory effect between **P1** and **P2** differed depending on enzymes, indicating the use of both enzymes and PEGylated

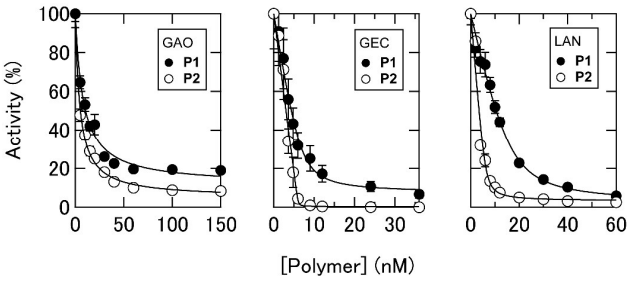


Figure 3. Changes in enzyme activities. Titration of PEGylated polyamines to 0.5 nM GAO, 0.2 nM GEC, and 10 nM LAN in 10 mM MOPS (pH 7.0) with 5% chemically defined serum-free CDCHO medium.

polyamines with different hydrophobicity is effective to increase cross-reactivity for PIC libraries.

As a proof-of-concept study, we first chose three different human cancer cell lines (Figure 2C): A549 (lung), MG63 (bone), and HuH7 (liver). After 16-hour incubation of each cell line seeded at 2.25×10^4 cells/cm^2 in DMEM supplemented with 10% fetal bovine serum, the medium was changed to chemically defined serum-free CDCHO medium. Culture supernatants were collected after 48 hours of incubation. These supernatants contained the following concentrations of proteins: 13.0 ± 3.0 μg/mL for A549, 22.7 ± 1.5 μg/mL for MG63, and 24.8 ± 7.2 μg/mL for HuH7 (The values are the averages of six parallel measurements with ± 1 S.D). Diluted culture supernatants at 5.0 μg/mL proteins were then mixed with a PIC library in 10 mM MOPS (pH 7.0), providing increases in the enzyme activities (Figure 4A, raw data of all PICs are shown in Table S1). The enzyme activity patterns were found to be reproducible and were likely characteristic of each cell type.

Generated data points (6 PICs × 3 cancer cell lines × 6 replicates) were subjected to linear discriminant analysis (LDA), which is a routinely used chemometric method for dimensional reduction to construct a set of orthogonal dimensions for describing the data, providing information on classification ability and a graphical output useful to gain insight into the clustering of the response data.[4a] Classification accuracy was initially calculated with the Jackknife classification procedure[20] to evaluate the discriminant capability of each PIC set as shown in Table S2 and investigate the mechanisms for sample discrimination. Accuracies of 56% – 89% using only one PIC were observed, whereas 100% accuracy was achieved using a combination of three PICs (GAO/**P1**, LAN/**P1**, and LAN/**P2**) (Table S2). First two discriminant scores, Z_1 and Z_2, were plotted to visualize how LDA clustered the patterns generated by the three PICs (Figure 4B). Discriminant scores were calculated using the discriminant functions Z_k, which are linear combinations of the descriptor variables with the greatest discriminating ability.

$$Z_k = a_1x_1 + a_2x_2 + \ldots + a_nx_n + C$$

where x_i are discriminating variables (enzyme activities in our case), a_i are discriminant weights, and C is a constant. A discriminant score plot showed that the different types of cancer cells clearly clustered into three nonoverlapping groups.

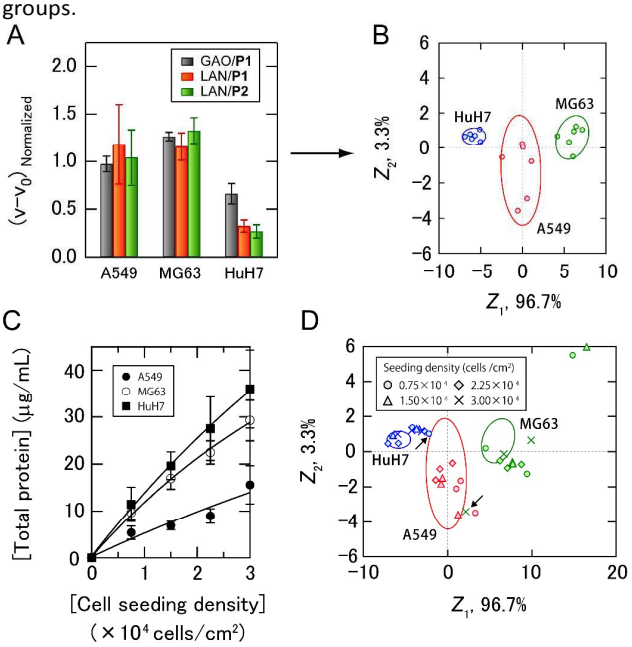


Figure 4. Sensing of human cancer cell lines. (A) Enzyme activity patterns for culture supernatants from three cancer cell lines seeded at 2.25×10^4 cells/cm^2. Each normalized value represents the average of six parallel measurements with 1 S.D. (details are shown in Table S1). (B) Discriminant score plot of the first two discriminant functions. The ellipses represent confidence intervals (± 1 S.D.) for the individual cancer cell lines. (C) Total protein concentrations of the culture supernatants with different seeding densities. The values are the averages of four parallel measurements with ± 1 S.D. (D) The effects of cell seeding density on pattern generation. Discriminant scores of enzyme activity patterns for the three kinds of cancer cell lines with various seeding densities were calculated using the first two discriminant functions obtained from training data. The ellipses are the same as those shown in (B), and the arrows indicate the misclassified samples.

The order of the first discriminant scores, accounting for 96.7% of the total variance, indicated that the value of activity recovery was critical for identification of cancer cell lines.

As secreted molecules are partly responsible for intercellular communication,[12,21] it can be assumed that the seeding density of cells affects pattern generation. Therefore, we examined whether our strategy could be used to identify cancer cell lines even when seeded at different densities. Total protein concentrations of the culture supernatants of cancer cells seeded at densities of $0.75 – 3.00\times10^4$ cells/cm^2 were fitted by nonlinear least-squares, as the linear approximations were slightly deviated with increasing seeding density of cells (Figure 4C), suggesting the influence of intercellular communications on the secretome. Newly obtained test data for diluted culture supernatants collected from the three kinds of cancer cells with different seeding densities were classified based on the shortest Mahalanobis distances to the aforementioned training data of three groups seeded at 2.25×10^4 cells/cm^2. Of the 33 diluted culture supernatants, only two samples were incorrectly identified (MG63 at 3.00×10^4 cells/cm^2 and HuH7 at 0.75×10^4 cells/cm^2), affording

an identification accuracy of 94% (Table S3). To visualize the differences between test and training data, the test data were transformed into discriminant scores

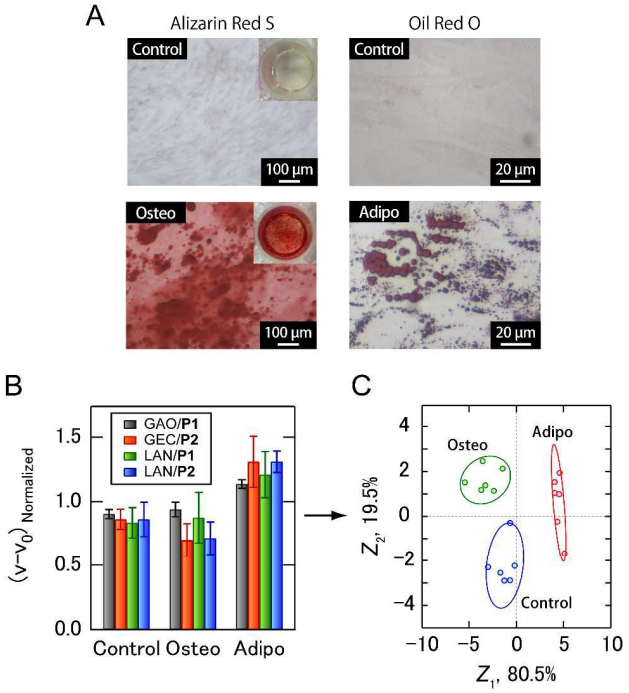


Figure 5. Sensing of ADSC differentiation. (A) Brightfield micrographs of ADSC cultured in control and differentiation media for 21 days. Cells were stained with Alizarin Red S (osteogenic differentiation) and Oil Red O (adipogenic differentiation). (B) Enzyme activity patterns for culture supernatants from three ADSC-derived lineages. Each normalized value represents the average of six parallel measurements with 1 S.D. (C) Discriminant score plot of the first two discriminant functions. The ellipses represent confidence intervals (± 1 S.D.) for the individual ADSC-derived lineages.

according to the first two discriminant functions, and plotted on the same 2D space as Figure 4B (Figure 4D). Interestingly, no systematic trend was found when changing seeding density of cancer cell lines. These results indicated that the ability of the PIC sensor array to discriminate cell types did not depend on the cell seeding density. Therefore, the analysis using PIC sensor array can be carried out at any arbitrary time even if the cells proliferate and change their density in a culture experiment. Taken together, our strategy allowed markerless and noninvasive classification of cancer cell lines with a broad range of density in culture by recognizing unique secretomic signatures of culture supernatants. In addition, by use of the standard curves shown in Figure 4C, the cell densities at the seeding time could be estimated (Table S3).

Neoplastic cell transformation and contamination by cancerous cells are important issues in cell culture. Our strategy also addressed this issue; only two PICs successfully discriminated culture supernatants from lung-derived normal (NHLF and WI38) and cancerous cells (A549) with different seeding densities (see Supplementary Section 3).

Building upon the noninvasive discrimination of normal/cancer cells using the PIC sensor array, we focused on differentiation of human MSCs, which have recently attracted attention for regenerative medicine due to their availability and potentially beneficial characteristics, including the ability

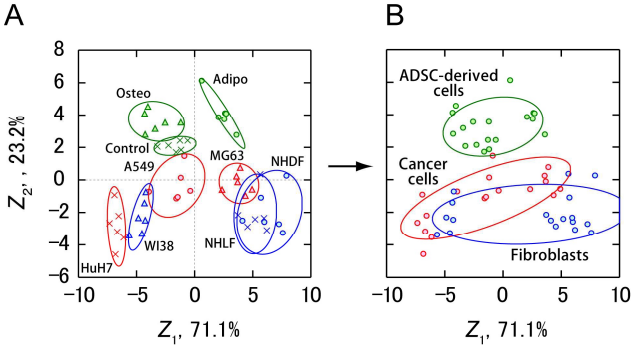


Figure 6. Sensing of cancer cells, fibroblasts, and ADSC-derived cells using four PICs (GAO/**P1**, GEC/**P1**, LAN/**P1**, LAN/**P2**). (A) Discriminant score plot of the first two discriminant functions. The enzyme activity patterns for nine types of cells obtained from four PICs were subjected to LDA, affording the best accuracy of 87%. (B) Clustering of all the data in (A) for each cellular category, as obtained from LDA analysis. The ellipses in (A) and (B) represent confidence intervals (± 1 S.D.) for the individual cells or cellular categories.

to differentiate into a variety of different cell types.[22] At present, the guided differentiation of MSCs is limited because the mechanisms governing the transition from uncommitted MSCs to differentiated cells have yet to be characterized in sufficient detail.[22a] In addition, traditional invasive assays can evaluate cultured cells only once, making it difficult to continue further differentiation induction even if stem cells are not yet differentiated. Therefore, the efficacy and cost of developing MSC-based products would be improved if differentiation of cells could be identified using culture supernatants. In this study, human adipose-derived stem cells (ADSCs) were selected as target MSCs. After osteogenic and adipogenic induction of ADSCs for 21 days (Figure 5A), culture supernatants prepared in CDCHO medium after 48 hours of incubation were collected. Culture supernatants of ADSC-derived cells were diluted to a concentration of 5.0 μg/mL protein, and then analyzed (Figure 5B). The combination of four PICs (GAO/**P1**, GEC/**P1**, LAN/**P1**, LAN/**P2**) showed 100% accuracy via Jackknife classification (Figure 5C and Table S4), and 83% accuracy was observed in a blind test (15 of 18) (Table S5).

It should be noted that PIC libraries consisting of only one type of enzyme (e.g., *GAO*/**P1** and *GAO*/**P2**) or only one type of PEG-*b*-QPAMA (e.g., GAO/***P1***, GEC/***P1***, and LAN/***P1***) were not capable of both discriminating cancer cell lines (Table S2) and identifying ADSC differentiation (Table S4), suggesting the effectiveness of the combined use of naturally occurring structural diversity of enzymes and artificial structural diversity of PEGylated polyamines. In addition, LAN-containing PICs played a significant role in discriminating secretomic signatures of culture supernatants in both cases (Figures 4 and 5). We have recently reported that electrostatic interaction was the main driving force for generating response patterns for plasma proteins when hydrophilic anionic enzymes were

used to construct PIC libraries.[9] To obtain more diverse response patterns, we have next used synthetic PEGylated polyamines with different hydrophobicities as PIC sources, enabling the discrimination of homologous albumins with very close resemblances in p*I*.[10] Taking the importance of hydrophobicity in account, in this study, we added highly hydrophobic LAN to PIC sources. Despite numerous secretome studies, very little has been reported on the differences in the abundances of secreted proteins and their physicochemical properties between cell types. However, LAN are expected to interact particularly with hydrophobic secreted proteins in culture supernatants, presumably resulted in the generation of unique responses that were different from those of GAO and GEC.

Finally, the activity patterns of all cells were combined and analyzed to evaluate whether they were generally indicative of cellular categories. Meta-analysis of this study showed that cancer cells, fibroblasts, and ADSC-derived cells likely clustered, respectively (Figures 6 and S2), indicating a potential correlation between cellular categories and the activity patterns reflecting secretomic molecules. Furthermore, ADSCs and fibroblasts were successfully discriminated with 100% accuracy using a combination of only two PICs (GAO/**P1** and LAN/**P2**) (Figure S4A and B), while discrimination of these cells based on morphological properties is not trivial due to the spindle-shaped fibroblast-like morphology of ADSCs. We also identified cancer cells and ADSC-derived cells with the accuracy of 100% using LAN/**P1** and LAN/**P2** combination (Figure S4C and D), an indication of the applicability of our system for identifying neoplastic transformation of stem cells.

Conclusions

We have applied a PIC sensor array for markerless and noninvasive discrimination of human cell types and lineage identification of differentiated stem cells. PIC sensor arrays for analysis of complex culture supernatants were newly constructed based on both naturally occurring structural diversity of enzymes and artificial structural diversity of PEGylated polyamines. PIC sensor arrays successfully recognized the phenotypic differences within the secretomic signatures in culture supernatants of the respective cells regardless of seeding density. The proposed array sensing system is the first secretomic-based approach that enables markerless and noninvasive identification of mesenchymal stem cell differentiation. Markerless identification with noninvasiveness is the most significant feature of our PIC sensor array. Traditional biomarker-based methods for endpoint cell evaluation require prior knowledge of specific markers and corresponding antibodies, which has not been identified in many situations. In addition, as our PIC sensor array does not need cell lysis or staining, evaluated cells can be used for other purposes or cell culture can be continued without damaging cells. The statistical processing can be automated by the use of analytical software, and therefore this approach will provide an effective way to characterize cultured cells with common laboratory equipment, such as the stepwise evaluation of the degree of differentiation of stem cells and prediction of lineage fates.

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Notes and references

1 (a) D. Rajamohan, E. Matsa, S. Kalra, J. Crutchley, A. Patel, V. George and C. Denning, *Bioessays*, 2013, **35**, 281; (b) M. B. Murphy, K. Moncivais and A. I. Caplan, *Exp. Mol. Med.*, 2013, **45**, e54; (c) V. Tabar and L. Studer, *Nat. Rev. Genet.*, 2014, **15**, 82.

2 D. G. Halme and D. A. Kessler, *N. Engl. J. Med.*, 2006, **355**, 1730.

3 O. Adewumi, B. Aflatoonian, L. Ahrlund-Richter, M. Amit, P. W. Andrews, G. Beighton, P. A. Bello, N. Benvenisty, L. S. Berry, S. Bevan, B. Blum, J. Brooking, K. G. Chen, A. B. Choo, G. A. Churchill, M. Corbel, I. Damjanov, J. S. Draper, P. Dvorak, K. Emanuelsson, R. A. Fleck, A. Ford, K. Gertow, M. Gertsenstein, P. J. Gokhale, R. S. Hamilton, A. Hampl, L. E. Healy, O. Hovatta, J. Hyllner, M. P. Imreh, J. Itskovitz-Eldor, J. Jackson, J. L. Johnson, M. Jones, K. Kee, B. L. King, B. B. Knowles, M. Lako, F. Lebrin, B. S. Mallon, D. Manning, Y. Mayshar, R. D. McKay, A. E. Michalska, M. Mikkola, M. Mileikovsky, S. L. Minger, H. D. Moore, C. L. Mummery, A. Nagy, N. Nakatsuji, C. M. O'Brien, S. K. Oh, C. Olsson, T. Otonkoski, K. Y. Park, R. Passier, H. Patel, M. Patel, R. Pedersen, M. F. Pera, M. S. Piekarczyk, R. A. Pera, B. E. Reubinoff, A. J. Robins, J. Rossant, P. Rugg-Gunn, T. C. Schulz, H. Semb, E. S. Sherrer, H. Siemen, G. N. Stacey, M. Stojkovic, H. Suemori, J. Szatkiewicz, T. Turetsky, T. Tuuri, S. van den Brink, K. Vintersten, S. Vuoristo, D. Ward, T. A. Weaver, L. A. Young and W. Zhang, *Nat. Biotechnol.*, 2007, **25**, 803.

4 (a) P. Anzenbacher Jr, P. Lubal, P. Bucek, M. A. Palacios and M. E. Kozelkova, *Chem. Soc. Rev.*, 2010, **39**, 3954; (b) J. R. Askim, M. Mahmoudi and K. S. Suslick, *Chem. Soc. Rev.*, 2013, **42**, 8649.

5 (a) G. V. Zyryanov, M. A. Palacios and P. Anzenbacher Jr, *Angew. Chem. Int. Ed.*, 2007, **46**, 7849; (b) Z. Yao, X. Feng, W. Hong, C. Li and G. Shi, *Chem. Commun.*, 2009, 4696; (c) A. M. Mallet, Y. Liu and M. Bonizzoni, *Chem. Commun.*, 2014, **50**, 5003.

6 (a) N. Y. Edwards, T. W. Sager, J. T. McDevitt and E. V. Anslyn, *J. Am. Chem. Soc.*, 2007, **129**, 13575; (b) S. H. Lim, C. J. Musto, E. Park, W. Zhong and K. S. Suslick, *Org. Lett.,* 2008, **10**, 4405; (c) K. L. Bicker, J. Sun, M. Harrell, Y. Zhang, M. M. Pena, P. R. Thompson and J. J. Lavigne, *Chem. Sci.*, 2012, **3**, 1147; (d) S. G. Elci, D. F. Moyano, S. Rana, G. Y. Tonga, R. L. Phillips, U. H. F. Bunz and V. M. Rottelo, *Chem. Sci.*, 2013, **4**, 2076.

7 (a) T. Zhang, N. Y. Edwards, M. Bonizzoni and E. V. Anslyn, *J. Am. Chem. Soc.*, 2009, **131**, 11976; (b) S. Rochat, J. Gao, X. Qian, F. Zaubitzer and K. Severin. *Chem. Eur. J.*, 2010, **16**, 104; (c) S. A. Minaker, K. D. Daze, M. C. Ma and F. Hof, *J. Am. Chem. Soc.*, 2012, **134**, 11674.

8 (a) M. De, S. Rana, H. Akpinar, O. R. Miranda, R. R. Arvizo, U. H. F. Bunz and V. M. Rotello, *Nat. Chem.*, 2009, **1**, 461; (b) O. R. Miranda, H. T. Chen, C. C. You, D. E. Mortenson, X. C. Yang,

U. H. F. Bunz and V. M. Rotello, *J. Am. Chem. Soc.*, 2010, **132**, 5285; (c) H. Pei, J. Li, M. Lv, J. Wang, J. Gao, J. Lu, Y. Li, Q. Huang, J. Hu and C. Fan, *J. Am. Chem. Soc.*, 2012, **134**, 13843; (d) D. Zamora-Olivares, T. S. Kaoud, K. N. Dalby and E. V. Anslyn, *J. Am. Chem. Soc.*, 2013, **135**, 14814; (e) D. Zamora-Olivares, T. S. Kaoud, J. Jose, A. Ellington, K. N. Dalby and E. V. Anslyn, *Angew. Chem. Int. Ed.*, 2014, **53**, 14064; (f) L. Motiei, Z. Pode, A. Koganitsky and D. Margulies, *Angew. Chem. Int. Ed.*, 2014, **53**, 9289.

9 S. Tomita and K. Yoshimoto, *Chem. Commun.*, 2013, **49**, 10430.

10 S. Tomita, T. Soejima, K. Shiraki and K. Yoshimoto, *Analyst*, 2014, **139**, 6100.

11 (a) A. Bajaj, O. R. Miranda, I. B. Kim, R. L. Phillips, D. J. Jerry, U. H. F. Bunz and V. M. Rotello, *Proc. Natl. Acad. Sci. U. S. A.*, 2009, **106**, 10912; (b) A. Bajaj, O. R. Miranda, R. Phillips, I. B. Kim, D. J. Jerry, U. H. F. Bunz and V. M. Rotello, *J. Am. Chem. Soc.*, 2010, **132**, 1018; (c) K. El-Boubbou, D. C. Zhu, C. Vasileiou, B. Borhan, D. Prosperi, W. Li and X. J. Huang, *J. Am. Chem. Soc.*, 2010, **132**, 4490; (d) S. Rana, A. K. Singla, A. Bajaj, S. G. Elci, O. R. Miranda, R. Mout, B. Yan, F. R. Jirik and V. M. Rotello, *ACS Nano*, 2012, **6**, 8233; (e) X. Yang, J. Li, H. Pei, Y. Zhao, X. Zuo, C. Fan and Q. Huang, *Anal. Chem.*, 2014, **86**, 3227.

12 K. J. Brown, C. A. Formolo, H. Seol, R. L. Marathi, S. Duguez, E. An, D. Pillai, J. Nazarian, B. R. Rood and Y. Hathout, *Expert Rev. Proteomics.*, 2012, **9**, 337.

13 (a) P. Dowling and M. Clynes, *Proteomics*, 2011, **11**, 794; (b) T. B. Schaaij-Visser, M. de Wit, S. W. Lam and C. R. Jimenez, *Biochim. Biophys. Acta.*, 2013, **1834**, 2242.

14 (a) A. J. Salgado, R. L. Reis, N. J. Sousa and J. M. Gimble, *Curr. Stem Cell Res. Ther.* 2010, **5**, 103; (b) H. K. Skalnikova, *Biochimie*, 2013, **95**, 2196.

15 (a) S. Zvonic, M. Lefevre, G. Kilroy, Z. E. Floyd, J. P. DeLany, I. Kheterpal, A. Gravois, R. Dow, A. White, X. Wu and J. M. Gimble, *Mol. Cell. Proteomics*, 2007, **6**, 18; (b) J. M. Kim, J. Kim, Y. H. Kim, K. T. Kim, S. H. Ryu, T. G. Lee, P. G. Suh, *J. Cell. Physiol.*, 2013, **228**, 216.

16 C. Chiellini, O. Cochet, L. Negroni, M. Samson, M. Poggi, G. Ailhaud, M. C. Alessi, C. Dani and E. Z. Amri, *BMC Mol. Biol.*, 2008, **9**, 26.

17 (a) S. Ganguli, K. Yoshimoto, S. Tomita, H. Sakuma, T. Matsuoka, K. Shiraki and Y. Nagasaki, *J. Am. Chem. Soc.* 2009, **131**, 6549; (b) S. Tomita, L. Ito, H. Yamaguchi, G. Konishi, Y. Nagasaki and K. Shiraki, *Soft Matter*, 2010, **6**, 5320; (c) S. Tomita and K. Shiraki, *J. Polym. Sci. Part A: Polym. Chem.* 2011, **49**, 3835; (d) T. Kurinomaru, S. Tomita, S. Kudo, S. Ganguli, Y. Nagasaki and K. Shiraki, *Langmuir*, 2012, **28**, 4334.

18 I. V. Tetko, J. Gasteiger, R. Todeschini, A. Mauri, D. Livingstone, P. Ertl, V. A. Palyulin, E. V. Radchenko, N. S. Zefirov, A. S. Makarenko, V. Y. Tanchuk and V. V. Prokopenko, *J. Comput.-Aided Mol. Des.*, 2005, **19** , 453.

19 G. Fernandez-Lorente, C. Ortiz, R. L. Segura, R. Fernandez-Lafuente, J. M. Guisan, J. M. Palomo, *Biotechnol. Bioeng.*, 2005, **92**, 773.

20 B. G. Tabachnick, and L. S. Fidell, *Using Multivariate Statistics*, Allyn & Bacon, Needham Heights, MA, 4th edn, 2001.

21 X. Zeng, P. Yang, B. Chen, X. Jin, Y. Liu, X. Zhao and S. J. Liang, *Proteomics*, 2013, **89**, 51.

22 (a) H. Mizuno, M. Tobita, A. C. Uysal, *Stem Cells*, 2012, **30**, 804; (b) S. Wang, X. Qu and R. C. Zhao, *J. Hematol. Oncol.* 2012, **5**, 19.