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ARTICLE

New *Securinega* alkaloids with anti-HIV activity from *Flueggea virosa*

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Chemical fractionation of the ethanolic extract of *Flueggea virosa* yielded a group of *Securinega* alkaloids including flueggenines E (1) and F (2) as novel hybrid structures, flueggenines G–I (3–5) as new dimers and fluevirosines E–H (6–9) as new trimers, along with six known biosynthetically related compounds. The diverse structures of these isolates were characterized via comprehensive spectroscopic analyses and comparison with literature data. Compounds 1 and 2 are rare *Securinega* alkaloid hybrids incorporating tryptamine and piperidine residues, respectively, while 3 represents the first example bearing a securinine-type monomeric unit among *Securinega* alkaloid dimers. An *in vitro* anti-HIV screening of all available alkaloids revealed weak to moderate activities for over half of these isolates with EC₅₀ values ranging from 7.8 to 122 μM. Among the tested compounds, the known dimer flueggenine D exhibited the best activity with an EC₅₀ of 7.8 ± 0.8 μM.

Introduction

The genus *Flueggea* Willd. (family Euphorbiaceae) comprises only ca. 12 species but occurs worldwide in both tropical and temperate regions.¹ The *Flueggea* plants are well known in the literature for their diverse secondary metabolites among which the *Securinega* alkaloids are the most investigated class of compounds.² *F. virosa* (Roxb. ex Willd.) Voigt, one of the four Chinese endemic species, is used as folk medicine by local residents for the treatment of eczema and rheumatoid arthritis.¹ Previous chemical investigations into the alkaloidal constituents of this herb have yielded all four types of *Securinega* alkaloids including securinine-type, norsecurinine-type, neosecurinine and neonorsecurinine structures.^{2–4} Of particular note, *F. virosa* has so far proven to be the only species to produce *Securinega* alkaloid oligomers that are all norsecurinine-derived.²

Our continuous study of the alkaloids from *F. virosa* collected at different sites since 2006 has led to the identification of 21 new structures from monomer to pentamer.^{4–8} It is worthwhile to note that the previous discovery of higher level (n > 2) oligomeric alkaloids was based on a biogenetic proposal⁶ and relied on a MS-guided separation strategy.^{7,8} In addition to the above-intended

search for routine oligomers, we also noticed some irregular MS peaks indicative of the likely presence of new types of alkaloid oligomers. Further fractionation of the remaining fractions returned two new alkaloid hybrids, flueggenines E (1) and F (2), three new dimers, flueggenines G–I (3–5), and four new trimers, fluevirosines E–H (6–9) (Fig. 1), as well as a known dimer and five known monomers. The new compounds were characterized by spectroscopic means and NMR comparison with known alkaloids. Both new and known compounds were tested *in vitro* for their anti-HIV effects with more than half showing weak to mild activities. A detailed account of the isolation, structure elucidation and anti-HIV evaluation of these *Securinega* alkaloids is presented below.

Results and discussion

Positive mode high resolution electrospray ionization mass spectrometry [(+)-HRESIMS] analysis of alkaloid 1 revealed a protonated molecular ion at *m/z* 378.2193 (calcd 378.2182), supportive of a molecular formula of C₂₃H₂₇N₃O₂. Inspection of the IR spectrum indicated the existence of an α,β-conjugated lactone moiety (1747 and 1631 cm⁻¹),⁹ which was further confirmed by the ¹³C resonances at δ_c 173.7, 173.5 and 110.3. The NMR data (Tables 1 and 2) for 1 displayed signals for a singlet methyl (δ_c 39.5; δ_h 2.56), seven sp³ methylenes (two *N*-bonded at δ_c 57.4 & 55.4), three sp³ *N*-connected methines (δ_c 66.8, 64.7 & 63.4), five sp² methines (δ_h 7.60, 7.37, 7.20, 7.12 & 7.03), four quaternary carbons (a sp³ oxygenated at δ_c 92.4 and three sp² ones), and an active proton (δ_h 8.01). The aforementioned NMR observations were in accord with a 14,15-dihydronorsecurinine fragment⁶ and a tryptamine residue,⁹ and this was supported by careful examination of 2D NMR data (Fig. 2). Finally, the connection between the two

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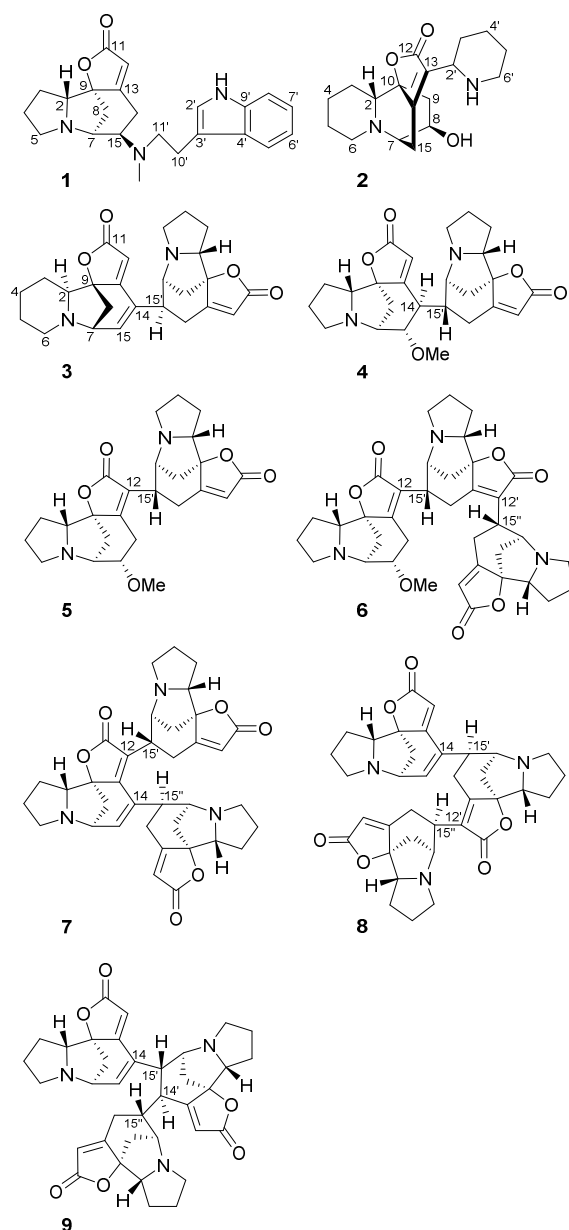


Fig. 1 New alkaloid oligomers from *F. virosa*.

monomeric units was secured by the diagnostic HMBC correlations from the *N*-methyl protons to C-15 (δ_c 64.7) and C-11' (δ_c 55.4) and from H₂-11' to C-15, thus defining the planar structure of **1** as shown. The relative configuration of **1** was assigned on the grounds of excellent NMR comparisons (especially proton couplings) with 15 β -substituted 14,15-dihydronorsecurinine derivatives,^{6,10} which was corroborated by subsequent acquisition of ROESY data (Fig. 2) with key correlations of H-2/H-14 β and H-15/H-8b. The absolute configuration of **1** was further established via CD data with Cotton effects at 260 (−7.1) and 220 (0.5) nm similar to those observed for a series of 14,15-dihydronorsecurinine

Table 1. ¹³C NMR data (125 MHz) for alkaloids 1–5.

No.	1 ^a	2 ^a	3 ^a	4 ^b	5 ^a
2	66.8	62.8	63.2	65.1	66.2
3	29.3	25.9	27.0	29.3	29.26 ^f
4	26.8	24.8	24.3	26.82 ^c	26.8 ^g
5	57.4	26.8	25.6	57.8 ^d	57.3
6		52.62	48.9		
7	63.4	57.8	58.5	64.1	63.4
8	33.8	66.7	42.2	31.1	30.4
9	92.4	36.5	89.7	92.6	90.6
10		84.8			
11	173.5		173.2	172.62 ^e	173.0
12	110.3	173.4	103.6	114.7	122.9
13	173.7	117.4	171.8	172.8	165.4
14	27.4	173.6	134.7	44.2	28.1
15	64.7	22.7	135.4	81.5	78.7
2'	121.6	52.60	67.0	65.8	66.9
3'	114.5	28.3	29.0	29.0	29.25 ^f
4'	127.6	23.2	26.6	26.76 ^c	26.9 ^g
5'	118.9	23.5	57.5	57.9 ^d	57.7
6'	119.5	46.0			
7'	122.3		65.0	64.8	64.6
8'	111.4		35.8	30.5	30.0
9'	136.4		91.9	91.6	92.5
10'	22.5				
11'	55.4		172.6	172.4	172.8
12'			110.2	111.7	110.7
13'			172.8	172.60 ^e	174.7
14'			29.1	27.2	26.3
15'			42.9	41.5	39.2
OMe				56.6	57.1
NMe	39.5				

^a Measured in CDCl₃; ^b Measured in CDCl₃ with 5% CD₃OD; ^{f–g} Interchangeable assignments.

derivatives isolated from *F. leucopyra*.¹⁰ Alkaloid **1** was thus identified unambiguously and was named flueggenine E after the known dimers flueggenines A–D^{5,7} from the same species. A literature search revealed only three examples of *Securinega*

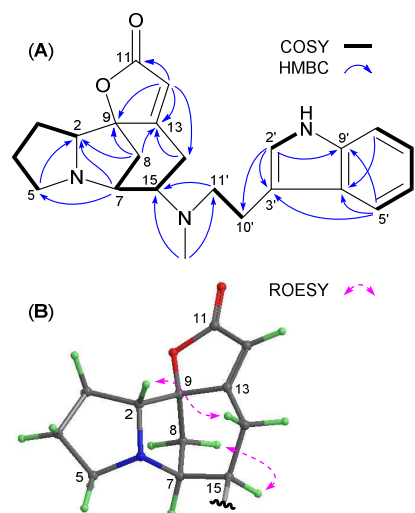


Fig. 2 Key 2D NMR correlations for alkaloid **1** (tryptamine moiety not shown in the 3D structure).

Table 2. ^1H NMR data (500 MHz) for alkaloids **1**–**5**.

No.	1 ^a	2 ^a	3 ^a	4 ^b	5 ^a
2	3.12 (dd, 8.9, 7.0)	2.18 (br d, 10.5)	2.04 (dd, 11.3, 2.6)	2.96 (dd, 8.8, 7.0)	3.05 (dd, 8.9, 6.9)
3	1.88 (m)	β 1.60 (m)	1.64 (m)	1.87 (m)	1.85–1.94 (m)
	1.73 (m)	α 1.38 (m)	1.54 (m)	1.77 (m)	1.71–1.80 (m)
4	1.93 (m)	1.86 (m)	1.89 (m)	1.95 (m)	1.90–1.98 (m)
	1.73 (m)	1.26 (m)	1.22 (m)	1.68 (m)	1.66–1.76 (m)
5	3.35 (m)	1.57 (2H, m)	1.62 (m)	3.25–3.32 (m)	3.31 (m)
	2.58 (m)		1.59 (m)	b 2.52–2.63 (m)	2.62 (m)
6		α 2.80 (m)	2.98 (m)		
		β 2.63 (m)	2.41 (m)		
7	3.27 (br d, 6.4)	2.94 (m)	3.85 (dd, 5.6, 4.2)	3.21 (dd, 5.6, 3.9)	3.13 (m)
8	a 2.50 (dd, 11.2, 6.4)	4.13 (dd, 8.7, 5.0)	2.50 (dd, 9.1, 4.2)	a 2.32 (dd, 11.1, 5.6)	a 2.27 (dd, 11.1, 5.6)
	b 1.29 (d, 11.2)		1.72 (d, 9.1)	b 1.75 (d, 11.1)	b 1.70 (d, 11.1)
9		a 2.82 (dd, 13.4, 8.7)			
		b 1.19 (d, 13.4)			
12	5.63 (d, 2.3)		5.56 (s)	5.72 (br s)	
14	α 3.03 (m)			2.61 (d, 12.8)	2.93 (d, 16.4)
	β 2.69 (ddd, 14.7, 11.1, 2.3)				2.66 (ddd, 16.4, 5.3, 1.7)
15	2.62 (ddd, 11.1, 5.3, 1.5)	α 3.22 (dd, 19.4, 2.4)	6.67 (d, 5.6)	3.48 (d, 3.9)	3.57 (dd, 5.3, 4.1)
		β 2.76 (dd, 19.4, 2.0)			
1'	8.01 (br s)				
2'	7.03 (d, 2.2)	3.86 (dd, 12.3, 3.1)	3.10 (dd, 8.8, 7.0)	3.15 (dd, 9.0, 7.0)	3.20 (dd, 8.8, 6.8)
3'		2.02 (m)	1.93 (m)	1.91 (m)	1.85–1.94 (m)
		1.87 (m)	1.79 (m)	1.75 (m)	1.71–1.80 (m)
4'		1.98 (m)	1.96 (m)	1.93 (m)	1.90–1.98 (m)
		1.55 (m)	1.60 (m)	1.66 (m)	1.66–1.76 (m)
5'	7.60 (br d, 7.9)	1.83 (m)	3.22 (m)	3.25–3.32 (m)	3.39 (m)
		1.71 (m)	2.59 (m)	2.52–2.63 (m)	2.64 (m)
6'	7.12 (ddd, 7.9, 7.2, 0.9)	3.39 (br d, 12.9)			
		2.92 (ddd, 12.9, 12.3, 3.2)			
7'	7.20 (ddd, 8.2, 7.2, 1.1)		3.01 (m)	2.80 (m)	3.17 (m)
8'	7.37 (br d, 8.2)		a 2.61 (m)	a 2.42 (dd, 11.3, 5.6)	2.29 (dd, 11.6, 5.8)
			b 1.49 (d, 11.0)	b 1.56 (d, 11.3)	1.54 (d, 11.6)
10'	2.95 (2H, m)				
11'	3.01 (m), 2.97 (m)				
12'			5.67 (d, 2.1)	5.72 (br s)	5.74 (d, 2.4)
14'			2.96 (m)	2.91 (m, 2H)	3.11 (ddd, 17.3, 9.6, 2.4)
			2.67 (m)		2.91 (d, 17.3)
15'			2.53 (m)	2.76 (m)	3.36 (br d, 9.6)
NMe	2.56 (s)				
OMe				3.30 (s)	3.20 (s)

^a Measured in CDCl_3 ; ^b Measured in CDCl_3 with 5% CD_3OD .

alkaloids oligomerized with other classes of alkaloids: margaritarine, bearing a securinine-type unit and a tryptamine tryptamine residue, reported in 1991;⁹ and secu'amamines F and G, incorporating a neosecurinine unit and a piperidine moiety, reported in 2009.¹¹ Flueggene E (**1**) represents the fourth *Securinea* alkaloid hybrid, but the first with a norsecurinine-type monomeric unit.

Flueggene F (**2**) was assigned a molecular formula of $\text{C}_{18}\text{H}_{26}\text{N}_2\text{O}_3$ based on the (+)-HRESIMS ion at m/z 319.2029 ($[\text{M} + \text{H}]^+$, calcd 319.2022) indicative of an isomer of secu'amamine F.¹¹ Analysis of the NMR data (Tables 1 and 2) for **2** confirmed this hypothesis with characteristic signals for a neosecurinine substructure and a 2-substituted piperidine moiety. Further examination of 2D NMR data (Fig. 3) established the planar

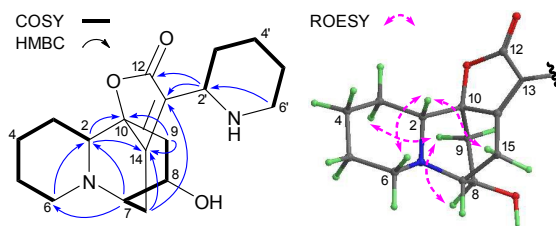


Fig. 3 Key 2D NMR correlations for alkaloid **2** (piperidine moiety not shown in the 3D structure).

structure of **2** to be identical with that of secu'amamine F, and the critical HMBC correlations from H-2' (δ_{H} 2.56) to C-12 (δ_{C} 173.4) and C-13 (δ_{C} 117.4) corroborated the linkage between C-2' and C-13. The relative configuration of the neosecurinine part in **2** was considered to be the same as that of virosine B¹² via their highly similar NMR data (particularly the comparable proton couplings) at the corresponding stereocenters (C-2, C-7 and C-8). The ROESY correlations (Fig. 3) of H-2/H-15 β , H-3 α /H-9 α and H-9 α /H-8 also supported this assignment. Although the relative configuration at C-2' remained unassigned due to its distance from the core structure, it was apparent from the coupling patterns (J = 12.3, 3.1 Hz) that H-2' was axially oriented. The absolute configuration of the neosecurinine part in **2** was determined as shown on the basis of its CD data with Cotton effects at 283 (0.73) and 250 (−1.9) nm.¹²

The molecular formula of C₂₅H₂₈N₂O₄ for flueggeine G (**3**) was established by HRESIMS analysis at m/z 420.2049 (Δ mmu 0). The NMR data (Tables 1 and 2) for **3** revealed diagnostic resonances for two γ -carbons (δ_{C} 91.9 and 89.7) of the lactones in securinine-/norsecurinine-type alkaloids, suggestive of its dimeric nature. Analysis of 2D NMR data (Fig. 4) established one securinine-type fragment and one dihydronorsecurinine unit that were connected from C-15' to C-14 via the key HMBC correlations of H-15'/C-13 & C-14. The relative configuration of **3** was assigned on the basis of ROESY data (†ESI Fig. S24) and NMR comparison (†ESI Table S1) with virosecurinine¹³ and flueggeine A.⁶ Excellent comparisons between the resonances of monomeric unit A and virosecurinine, and those of monomeric unit B and the dihydronorsecurinine moiety of flueggeine A, supported common configurations at all corresponding chiral centers. In addition, the configuration at the C-15' stereocenter in **3** was also confirmed by the ROESY correlation of H-15' (δ_{H} 2.53) with H-8'b (δ_{H} 1.49). As with the other alkaloid oligomers from the same species,⁶ alkaloid **3** is likely to be biosynthesized from virosecurinine and (−)-norsecurinine, and thus retains the absolute configurations from the two monomeric precursors. Meanwhile, the CD spectrum (Fig. 5) of **3** showed Cotton effects arising from the $\alpha,\beta,\gamma,\delta$ -conjugated lactone

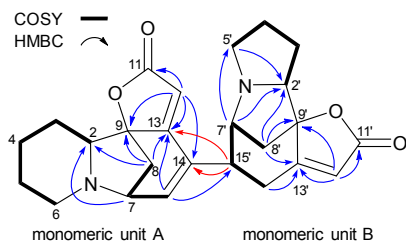


Fig. 4 Key 2D NMR correlations for alkaloid **3**.

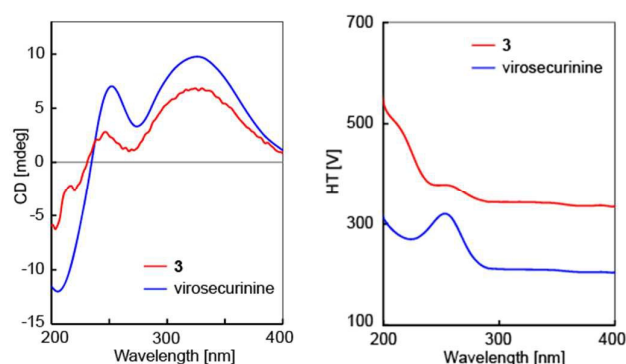


Fig. 5 CD and UV spectra for alkaloid **3** and virosecurinine.

chromophore comparable to that of virosecurinine, confirmative of the above-mentioned biogenetic correlation. Alkaloid **3** was thereby elucidated to be the first dimeric example derived from a securinine-type monomeric unit.

Flueggenines H (**4**) and I (**5**) gave the same molecular formula of C₂₅H₃₀N₂O₅ as determined from the (+)-HRESIMS ions at m/z 439.2230 and 439.2218 ($[M + H]^+$, calcd 439.2233), respectively. Analysis of the NMR data (Tables 1 and 2) for **4** indicated a MeOH adduct of flueggeine D⁷ with diagnostic signals for two sp³ methines (δ_{C} 44.2 & 81.5; δ_{H} 2.61 & 3.48) and a methoxyl group (δ_{C} 56.6; δ_{H} 3.30) replacing those for Δ^{14} in the latter. The acquisition 2D NMR data (Fig. 6) facilitated the aforementioned structural assignment revealing key ¹H–¹H COSY correlations of H-14/H-15' and HMBC correlation from the methoxyl protons to C-15 (δ_{C} 44.2). The relative configurations at the chiral centers of C-14 and C-15 as shown were assigned via the diagnostic coupling pattern of H-14/H-15 ($J_{14,15}$ = 0 Hz)⁸ and the ROESY crosspeaks of OCH₃/H-8b and H-14/H-8'b. The relative stereostructure of **4** was hence characterized. As with the case of **4** and flueggeine D, the NMR data (Tables 1 and 2) for **5** also suggested it to be a MeOH adduct at Δ^{14} of flueggeine C⁷ and this was supported by the absence of the olefinic resonances and the presence of those for two sp³ methines and one methoxyl group. Examination of 2D NMR data (†ESI Fig. Sa) further confirmed this structural assignment. The relative configuration at the new C-15 stereocenter was suggested by the ROESY correlation of OCH₃/H-8b, while the configurations at the other stereocenters were consistent with their counterparts in flueggeine C⁷ based on excellent NMR comparisons and ROESY data (†ESI Fig. S42).

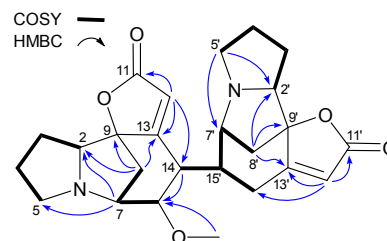


Fig. 6 Key 2D NMR correlations for alkaloid **4**.

The molecular formula of $C_{37}H_{44}N_3O_7$ for **6** was determined through ^{13}C NMR data and the (+)-HRESIMS ion at m/z 642.3173 ($[M + H]^+$, calcd 642.3179). Analysis of the NMR data (Tables 3 and 4) for **6** established that it was a trimer derived from **5** by the addition of a dihydronorsecurinine unit at C-12'. Indeed, the absence of the H-12' signal and the presence of those for a dihydronorsecurinine residue were supportive of this structural variation, which was further confirmed by 2D NMR data (1H -ESI Fig. S5) with corroborative HMBC correlations from H-15'' (δ_H 3.30) to C-11' (δ_C 172.4), C-12' (δ_C 121.2) and C-13' (δ_C 168.6). The new C-15'' chiral center was considered to possess identical relative configuration with C-15' based on the shielded C-8'' signal (δ_C 30.2),^{6,7} while those at all other stereocenters were assigned as drawn via NMR comparison with **5** and analysis of ROESY data (1H -ESI Fig. S51). The structure of **6** was hence elucidated and was named flueviroisine E after the known trimers flueviroisines A–D.^{6,7}

Table 3. ^{13}C NMR data (125 MHz) for alkaloids **6–9** in $CDCl_3$.

No.	6	7	8	9
2	66.5 ^a	65.1	65.1	65.5 ^j
3	29.3 ^b	29.3 ^e	29.5 ^g	29.5
4	26.9 ^c	26.88 ^f	27.0 ^h	27.0 ^k
5	57.55 ^d	54.8	55.2	55.3
7	63.3	58.8	59.5	59.4
8	30.8	36.4	36.3	35.7
9	90.4	91.4	92.3	92.2
11	173.4	172.1	172.34 ⁱ	172.1
12	122.6	120.6	107.4	107.0
13	166.8	162.9	169.3	169.4
14	27.9	132.8	133.2	134.9
15	78.6	139.9	139.2	141.1
2'	67.1	66.5	67.5	65.1
3'	29.21 ^b	29.22 ^e	29.4 ^g	29.2
4'	26.84 ^c	26.88 ^f	26.93 ^h	26.8 ^k
5'	57.52 ^d	57.8	57.8	58.1
7'	66.0	67.5	64.3	65.4 ^j
8'	30.4	30.4	35.8	31.5
9'	90.4	91.8	90.7	92.9
11'	172.4	172.8	173.9	171.8
12'	121.2	109.8	122.3	113.9
13'	168.6	173.5	166.1	173.2
14'	25.1	26.76 ^f	26.9 ^h	42.9
15'	37.9	38.8	43.1	47.7
2''	66.4 ^a	67.1	67.2	66.0
3''	29.16 ^b	29.18 ^e	29.2 ^g	29.1
4''	26.80 ^c	26.86 ^f	26.7 ^h	26.8 ^k
5''	57.46 ^d	58.1	57.5	57.9
7''	65.5	65.7	65.2	64.8
8''	30.2	36.0	35.7	30.2
9''	92.1	91.7	91.9	91.6
11''	172.6	173.4	173.0	172.5
12''	110.0	110.9	110.6	112.0
13''	174.0	172.3	172.31 ⁱ	171.9
14''	25.8	28.4	27.8	27.4
15''	38.5	42.9	39.3	44.3
OMe	57.0			

^{a–k} Interchangeable assignments.

Flueviroisines **F** (**7**) and **G** (**8**) exhibited *quasi*-molecular ion peaks in the (+)-HRESIMS spectra at m/z 610.2934 and 610.2910 ($[M + H]^+$), respectively, corresponding to the same molecular formula of $C_{36}H_{39}N_3O_6$ and indicative of a pair of isomers. The NMR data (Tables 3 and 4) for **7** were highly similar to those for flueviroisine D⁷ with the only difference being attributable to signals around C-15'', which suggested **7** to be a likely 15''-epimer of the latter. Analysis of 2D NMR data (1H -ESI Fig. S5) for **7** confirmed that they possess the same planar structure bearing one norsecurinine and two dihydronorsecurinine moieties. Compared to flueviroisine D, the coupling pattern of H₂-14'' with H-15'', the chemical shift (δ_C 36.0) for C-8'' and the ROESY correlation of H-8''b/H-15'' all supported an inverted configuration at C-15''. The relative configurations at all other stereocenters were determined to be the same as those in flueviroisine D by their excellent NMR comparisons and analysis of ROESY data (1H -ESI Fig. S60). Similar to the case of **7** and flueviroisine D, alkaloid **8** was determined to be the 15''-epimer of flueviroisine A⁶ by analysis of the NMR data (Tables 3 and 4). The relative configuration at C-15'' was established as described for **7**, and inspection of 2D NMR data (1H - 1H COSY, HMBC and ROESY, 1H -ESI Fig. S5 & S60) further corroborated the aforementioned assignment. The structures of **7** and **8** were thus characterized.

Flueviroisine H (**9**) gave a molecular formula ($C_{36}H_{39}N_3O_6$) same as **8** based on the (+)-HRESIMS ion at m/z 610.2924 ($[M + H]^+$, calcd 610.2917). Compared to **8**, the NMR data (Tables 3 and 4) for **9** also exhibited the presence of one norsecurinine and two dihydronorsecurinine fragments with extra signals for olefinic proton (δ_H 5.69, H-12') and methine group (δ_H 2.78 & δ_C 42.9, CH-14') but absence of those for a methylene and a quaternary carbon, which suggested a distinct oligomerization pattern. Further analysis of 2D NMR data (Fig. 7) confirmed the presence of the aforementioned monomeric substructures and the connections between them with a confirmatory 1H - 1H COSY correlation of H-14'/H-15'' and HMBC correlations from H-15' (δ_H 2.98) to C-13 (δ_C 169.4), C-14 (δ_C 134.9), C-15 (δ_C 141.4) and C-15'' (δ_C 44.3). The relative configurations at C-14', C-15' and C-15'' were established to be identical with those at the corresponding chiral centers in flueviroisine E⁸ via the same coupling patterns of H-14', H-15' and H-15'', while the

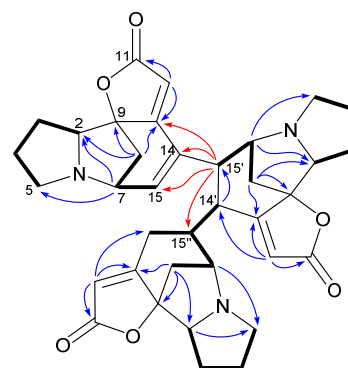


Fig. 7 Key 2D NMR correlations for alkaloid **9**.

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Table 4. ^1H NMR data (500 MHz) for alkaloids **6–9** in CDCl_3 .

No.	6	7	8	9
2	3.20 (dd, 8.9, 6.8)	3.06 (m)	3.15 (8.9, 7.1)	3.07 (m)
3	1.81–1.96 (m)	1.89–1.99 (m)	2.01 (m)	1.99 (m)
	1.66–1.80 (m)	1.72–1.82 (m)	1.73–1.83 (m)	1.70–1.83 (m)
4	1.90–2.02 (m)	1.92–2.01 (m)	1.92–2.03 (m)	1.92–2.02 (m)
	1.64–1.78 (m)	1.68–1.79 (m)	1.63–1.77 (m)	1.66–1.79 (m)
5	3.28–3.40 (m)	3.26 (m)	3.32 (m)	3.22 (m)
	2.55–2.69 (m)	2.52 (m)	2.54 (m)	2.49 (m)
7	3.17 (m)	3.66 (dd, 6.5, 4.5)	3.70 (dd, 6.5, 4.5)	3.61 (dd, 6.5, 4.6)
8	2.28 (m)	2.54 (dd, 10.6, 4.5)	2.60 (dd, 10.5, 4.5)	2.54 (dd, 10.6, 4.6)
	1.72 (d, 11.0)	1.64 (d, 10.6)	1.74 (d, 10.5)	1.57 (d, 10.6)
12			5.72 (s)	5.82 (s)
14	2.76 (d, 16.6)			
	2.70 (dd, 16.6, 5.1)			
15	3.61 (dd, 5.1, 4.2)	6.92 (d, 6.5)	6.73 (d, 6.5)	6.26 (d, 6.5)
2'	3.15 (m)	3.17 (dd, 8.9, 6.9)	3.12 (m)	3.11 (dd, 8.9, 6.7)
3'	1.81–1.96 (m)	1.89–1.99 (m)	1.87–1.95 (m)	1.88–1.97 (m)
	1.66–1.80 (m)	1.72–1.82 (m)	1.73–1.83 (m)	1.70–1.83 (m)
4'	1.90–2.02 (m)	1.92–2.01 (m)	1.92–2.03 (m)	1.92–2.02 (m)
	1.64–1.78 (m)	1.68–1.79 (m)	1.63–1.77 (m)	1.66–1.79 (m)
5'	3.28–3.40 (m)	3.30 (m)	3.24 (m)	3.37 (m)
	2.55–2.69 (m)	2.53 (m)	2.52 (m)	2.54 (m)
7'	2.94 (m)	2.85 (dd, 5.5, 2.5)	3.22 (brd, 5.9)	2.97 (d, 5.4)
8'	2.26 (dd, 11.5, 5.8)	2.32 (dd, 11.3, 5.5)	2.62 (dd, 11.1, 5.9)	2.32 (dd, 11.4, 5.4)
	1.63 (d, 11.5)	2.17 (d, 11.3)	1.47 (d, 11.1)	1.52 (d, 11.4)
12'		5.66 (br s)		5.69 (s)
14'	2.95 (m, 2H)	2.99 (m, 2H)	3.56 (dd, 16.0, 5.0)	2.78 (d, 10.1)
			2.75 (dd, 16.0, 12.4)	
15'	3.34 (m)	3.42 (m)	2.51 (m)	2.98 (s)
2''	3.01 (m)	3.14 (dd, 9.0, 7.0)	3.10 (dd, 9.2, 6.9)	3.15 (dd, 8.9, 6.9)
3''	1.81–1.96 (m)	1.89–1.99 (m)	1.87–1.95 (m)	1.88–1.97 (m)
	1.66–1.80 (m)	1.72–1.82 (m)	1.73–1.83 (m)	1.70–1.83 (m)
4''	1.90–2.02 (m)	1.92–2.01 (m)	1.92–2.03 (m)	1.92–2.02 (m)
	1.64–1.78 (m)	1.68–1.79 (m)	1.63–1.77 (m)	1.66–1.79 (m)
5''	3.28–3.40 (m)	3.27 (m)	3.13 (m)	3.30 (m)
	2.55–2.69 (m)	2.54 (m)	2.58 (m)	2.59 (m)
7''	2.96 (m)	3.00 (m)	3.01 (m)	2.82 (m)
8''	2.30 (dd, 11.3, 6.3)	2.64 (dd, 10.9, 5.9)	2.55 (dd, 11.1, 6.1)	2.40 (dd, 11.2, 5.7)
	1.88 (d, 11.3)	1.56 (d, 10.9)	1.53 (d, 11.1)	1.55 (d, 11.2)
12''	5.67 (d, 2.1)	5.72 (d, 2.0)	5.66 (d, 2.0)	5.59 (d, 1.5)
14''	3.11 (d, 16.0)	2.86 (m)	2.99 (dd, 14.7, 5.2)	2.94 (m)
	3.02 (m)	2.78 (ddd, 15.9, 12.9, 2.0)	2.81 (ddd, 14.7, 12.1, 2.0)	2.59 (m)
15''	3.30 (m)	2.90 (m)	2.88 (m)	2.94 (m)
OMe	3.26 (s)			

^{a-c} Assignments might be interchangeable.

other stereocenters were assigned on the basis of excellent NMR comparisons with the above-described alkaloids. The absolute configurations of **7–9** were determined to be as shown by analysis of their CD spectra, which displayed Cotton

effects at 270 (–10.5), 270 (–14.6) and 267 (–13.2) nm, respectively.⁸

In addition to the above-mentioned new alkaloids, six known ones, flueggeainol,¹⁴ bubbialine,¹⁵ (+)-14,15-dihydronorsecurine,¹⁶ 14,15-epoxynorsecurinine,¹⁷ 15 α -

methoxy-14,15-dihydronorsecurine,¹⁰ and flueggine B,¹⁸ were also obtained and identified by full spectroscopic analyses and comparison with reported data in the literature.

All available *Securinega* alkaloids from *F. virosa*, including those (virosecurinine,⁶ viroallosecurinine,⁶ (–)-norsecurinine,⁶ flueggenines C and D,⁷ and fluevirosine D⁷) reported previously, were tested for their *in vitro* anti-HIV activities on HIV-1 NL 4-3 infected MT4 cells and nevirapine was used as the positive control.¹⁹ The assay results (Table 5) revealed that more than half of these compounds showed mild protection on MT-4 cells against the HIV-induced cytopathic effect (EC₅₀ 10–100 μM) without displaying cytotoxicity (CC₅₀ >100 μM). The best anti-HIV activity was observed for a known dimer, flueggine D, with an EC₅₀ of 7.8 ± 0.8 μM (selective index = 12.6).

Experimental

General experimental procedures

Optical rotations were obtained on a Rudolph Autopol VI automatic polarimeter. UV and IR spectra were acquired on a Shimadzu UV-2550 UV/Visible spectrophotometer and a Perkin-Elmer 577 spectrometer, respectively. NMR experiments were performed on a Bruker AM-500 spectrometer with a cryoprobe referenced to deuterated solvent peaks (δ_{H} 7.26 and δ_{C} 77.23 for CDCl₃). LR- and HR-EIMS analyses (70 eV) were performed on a Finnigan MAT 95 mass spectrometer. LR- and HR-ESIMS experiments were conducted on Bruker Daltonics esquire3000plus and Waters LCT Premier XE spectrometers, respectively. Pre-coated silica gel GF254 plates (Yantai Huiyou Silica Gel Exploitation

Company, Ltd., China) were used for TLC analyses and separations. Silica gel H (300–400 mesh, Qingdao Haiyang Chemical Plant, Ltd., China), Sephadex LH-20 gel (Pharmacia Biotech, Sweden), and amino silica gel (20–45 μm, Fuji Silysia Chemical, Ltd., Japan) were used for column chromatography (CC). HPLC purifications were carried out on a Waters 1525 binary pump system equipped with a 2489 UV/Visible detector and a XBridge Prep C18 column (5 μm, 10 × 250 mm). All solvents used for CC were of at least analytical grade (Shanghai Chemical Reagents Company, Ltd., China), and solvents used for UV, $[\alpha]_{\text{D}}$, and NMR measurements were of suitable chromatographic grades from Merck or Sigma-Aldrich.

Plant materials

The plant materials (twigs and leaves) of *F. virosa* were collected in April 2007 from Xishuangbanna of Yunnan Province and in September 2007 from Gongchen of Guangxi Zhuang Autonomous Region, respectively. They were authenticated by Prof. Y. K. Xu from Xishuangbanna Tropical Botanical Garden and by Prof. S. Q. Tang from Guangxi Normal University, respectively. The voucher specimens have been deposited in the herbarium of Shanghai Institute of Materia Medica (Accession numbers: 2007-FV-1Y and 2007-FV-2Y, respectively).

Extraction and isolation

The crude alkaloids (11.1 g) of the Xishuangbanna species were obtained as previously reported.⁵ Fractionation of the crude alkaloids with a MCI gel column (MeOH-H₂O, 3:7 to 7:3) yielded four major fractions F1–F4. After crystallization of virosecurinine from F3, the mother solution was subjected to silica gel CC (CHCl₃-CH₃OH, 500:1 to 10:1) to furnish ten sub-fractions, the fourth of which was further purified by Sephadex LH-20 CC (in MeOH) to give flueggine G (**3**, 3.0 mg).

A total of 28.2 g crude alkaloids were prepared from the Gongchen species through the same procedures as described formerly.⁷ Fractionation of the crude alkaloids with a silica gel column (petroleum ether-EtOAc-HNEt₂, 5:1:0.1 to 1:2:0.2) returned six major fractions F1–F6, and F4 (7.76 g) was further subjected to silica gel CC (CHCl₃-MeOH, 100:1 to 5:1) to afford subfractions F4a–F4g. Subfraction F4b was sequentially separated by silica gel CC (petroleum ether-EtOAc-HNEt₂, 5:1:0.1 to 1.5:1:0.1), amino silica gel CC (petroleum ether-EtOAc, 1:1 to 1:2), and preparative TLC (petroleum ether-EtOAc-HNEt₂, 1.5:1:0.2) to give 14,15-epoxynorsecurinine (6.6 mg) and 15α-methoxy-14,15-dihydronorsecurine (8.5 mg). Subfraction F4d was processed with amino silica gel (CHCl₃-MeOH, 50:1) to furnish three elutions, and **4** (1.9 mg) and (+)-14,15-dihydronorsecurine (4.5 mg) were purified from the first and third elutions by HPLC with MeCN-H₂O system (20–80% and 20–40%, respectively, over 20 min). Subfraction F4e was fractionated by HPLC with MeCN-H₂O system (30–45% over 20 min) to yield **5** (5.6 mg) and a mixture which was further separated by HPLC with MeOH-H₂O as mobile phase (35–60% over 20 min) to give **9** (2.5 mg) and **7** (3.7 mg).

Fraction F5 (3.27 g) was first separated over silica gel (CHCl₃-MeOH, 50:1 to 5:1) to afford subfractions F5a–F5f, and six

Table 5. Anti-HIV activities of alkaloids from *F. virosa* (in μM)

Compds.	EC ₅₀	CC ₅₀
flueggine E (1)	42.6 ± 4.3	>100
flueggine F (2)	–	–
flueggine G (3)	–	–
flueggine H (4)	122 ± 12	–
flueggine I (5)	63.9 ± 6.5	–
fluevirosine E (6)	–	–
fluevirosine F (7)	–	–
fluevirosine G (8)	58.7 ± 5.6	–
fluevirosine H (9)	108 ± 10	–
flueggeinol	41.9 ± 4.2	>100
bubbialine	–	–
14,15-dihydronorsecurine	–	–
14,15-epoxynorsecurinine	85.5 ± 8.7	–
15α-methoxy-14,15-dihydronorsecurine	89.0 ± 9.2	–
flueggine B	79.6 ± 8.1	–
flueggine C	–	–
flueggine D	7.8 ± 0.8	97.9
fluevirosine D	–	–
virosecurinine	19.3 ± 2.0	>100
viroallosecurinine	56.4 ± 5.7	–
(–)-norsecurinine	43.0 ± 4.4	>100
nevirapine	0.119 ± 0.012	>100

more elutions (F5a1–F5a6) were further obtained by subsequent fractionation of F5a (2.06 g) on a silica gel column (petroleum ether–EtOAc–HNEt₂, 4:1:0.1 to 2:1:0.1). Flueggeainol (1.8 mg) was obtained from F5a1 by HPLC purification (30–60% MeCN–H₂O over 15 min), while bubbialine (9.6 mg) and **1** (2.3 mg) were acquired from F5a6 also by HPLC separation (MeCN–H₂O, 25–30% over 15 min, to 50% in 0.5 min then to 80% over 6 min). Subfraction F5c was further fractionated by HPLC (25–60% MeCN–H₂O over 20 min) to furnish **8** (1.9 mg), while subfractions F5g and F5i yielded flueggine B (1.9 mg) and **2** (2.7 mg) via HPLC purifications eluted with gradient (25–60% over 20 min) and isocratic (37%) MeCN–H₂O, respectively. All solvent systems used for HPLC separations were at a flow rate of 3.5 mL/min and modified with 0.02% HNEt₂ unless specified.

Characterization of new compounds

Flueggine E (1). Off-white solid; $[\alpha]_{\text{D}}^{23}$ 38 (c 0.23, MeOH); UV (MeOH) λ_{max} (log ϵ) 260 (3.93), 220 (4.46) nm; CD (MeOH) λ ($\Delta\epsilon$) 260 (–7.1), 220 (0.5), 210 (2.1) nm; IR (KBr) ν_{max} 2964, 1747, 1631, 1599, 1564, 1552, 1381, 1354, 1221, 1075, 911, 744 cm^{–1}; ¹H and ¹³C NMR data see Tables 1 and 2; ESIMS m/z 378.2 [M + H]⁺, 376.1 [M – H][–]; (+)-HRESIMS m/z 378.2193 [M + H]⁺ (calcd for C₂₃H₂₈N₃O₂, 378.2182).

Flueggine F (2). Off-white solid; $[\alpha]_{\text{D}}^{23}$ 15 (c 0.27, MeOH); UV (MeOH) λ_{max} (log ϵ) 219 (4.05) nm; CD (MeOH) λ ($\Delta\epsilon$) 283 (0.3), 250 (–0.7), 223 (8.5) nm; IR (KBr) ν_{max} 2941, 2850, 1745, 1670, 1442, 1414, 1344, 1201, 1174, 1124, 1080, 1014 cm^{–1}; ¹H and ¹³C NMR data see Tables 1 and 2; ESIMS m/z 319.2 [M + H]⁺, 637.2 [2M + H]⁺; (+)-HRESIMS m/z 319.2029 [M + H]⁺ (calcd for C₁₈H₂₇N₂O₃, 319.2022).

Flueggine G (3). Off-white solid; $[\alpha]_{\text{D}}^{23}$ 245 (c 0.10, MeOH); UV (MeOH) λ_{max} (log ϵ) 257 (3.74), 212 (4.10) nm; CD (MeOH) λ ($\Delta\epsilon$) 324 (3.5), 247 (1.5) nm; IR (KBr) ν_{max} 3431 (br, H₂O), 2926, 2852, 1755, 1649, 1618, 1259, 1076 cm^{–1}; ¹H and ¹³C NMR data see Tables 1 and 2; EIMS (70 eV) m/z 420 ([M]⁺, 12), 337 (19), 191 (36), 149 (14), 84 (100); HREIMS m/z 420.2049 [M]⁺ (calcd for C₂₅H₂₈N₂O₄, 420.2049).

Flueggine H (4). Off-white solid; $[\alpha]_{\text{D}}^{25}$ –10 (c 0.19, MeOH); UV (MeOH) λ_{max} (log ϵ) 215 (4.14) nm; IR (KBr) ν_{max} 2960, 2927, 1758, 1641, 1465, 1385, 1227, 1144, 1080, 975, 918 cm^{–1}; ¹H and ¹³C NMR data see Tables 1 and 2; ESIMS m/z 439.3 [M + H]⁺, 877.4 [2M + H]⁺; (+)-HRESIMS m/z 439.2230 [M + H]⁺ (calcd for C₂₅H₃₁N₂O₅, 439.2233).

Flueggine I (5). Off-white solid; $[\alpha]_{\text{D}}^{25}$ –0.7 (c 0.56, MeOH); UV (MeOH) λ_{max} (log ϵ) 216 (4.41) nm; IR (KBr) ν_{max} 2963, 2874, 1754, 1670, 1643, 1460, 1420, 1292, 1233, 1220, 1097, 1081, 1065, 919, 784 cm^{–1}; ¹H and ¹³C NMR data see Tables 1 and 2; ESIMS m/z 439.4 [M + H]⁺, 877.6 [2M + H]⁺; (+)-HRESIMS m/z 439.2218 [M + H]⁺ (calcd for C₂₅H₃₁N₂O₅, 439.2233).

Fluevirosine E (6). Off-white solid; $[\alpha]_{\text{D}}^{25}$ 4.5 (c 0.36, MeOH); UV (MeOH) λ_{max} (log ϵ) 224 (4.33) nm; IR (KBr) ν_{max} 2960, 2928, 1752, 1645, 1458, 1421, 1291, 1232, 1114, 1067, 998, 918 cm^{–1}; ¹H and ¹³C NMR data see Tables 3 and 4; ESIMS m/z 642.4 [M + H]⁺, 1283.8 [2M + H]⁺; (+)-HRESIMS m/z 642.3173 [M + H]⁺ (calcd for C₃₇H₄₄N₃O₇, 642.3179).

Fluevirosine F (7). Off-white solid; $[\alpha]_{\text{D}}^{25}$ –24 (c 0.37, MeOH); UV (MeOH) λ_{max} (log ϵ) 270 (3.99), 214 (4.26) nm; CD (MeOH) λ ($\Delta\epsilon$) 270 (–10.5) nm; IR (KBr) ν_{max} 2958, 2875, 1757, 1645, 1616, 1460, 1418, 1262, 1223, 1198, 1144, 1117, 1077, 978, 917 cm^{–1}; ¹H and ¹³C NMR data see Tables 3 and 4; ESIMS m/z 305.7 [M + 2H]²⁺, 610.4 [M + H]⁺, 1219.6 [2M + H]⁺; (+)-HRESIMS m/z 610.2934 [M + H]⁺ (calcd for C₃₆H₄₀N₃O₆, 610.2917).

Fluevirosine G (8). Off-white solid; $[\alpha]_{\text{D}}^{25}$ –1.8 (c 0.19, MeOH); UV (MeOH) λ_{max} (log ϵ) 259 (3.99), 217 (4.39) nm; CD (MeOH) λ ($\Delta\epsilon$) 270 (–14.6) nm; IR (KBr) ν_{max} 2961, 2872, 1755, 1646, 1622, 1458, 1284, 1243, 1222, 1144, 1111, 1079, 973, 915 cm^{–1}; ¹H and ¹³C NMR data see Tables 3 and 4; ESIMS m/z 305.8 [M + 2H]²⁺, 610.4 [M + H]⁺; (+)-HRESIMS m/z 610.2910 [M + H]⁺ (calcd for C₃₆H₄₀N₃O₆, 610.2917).

Fluevirosine H (9). Off-white solid; $[\alpha]_{\text{D}}^{25}$ –84 (c 0.25, MeOH); UV (MeOH) λ_{max} (log ϵ) 260 (4.04), 216 (4.31) nm; CD (MeOH) λ ($\Delta\epsilon$) 267 (–13.2) nm; IR (KBr) ν_{max} 2961, 2874, 1756, 1644, 1622, 1484, 1458, 1286, 1248, 1231, 1143, 1113, 1077, 966, 918, 854 cm^{–1}; ¹H and ¹³C NMR data see Tables 3 and 4; ESIMS m/z 305.8 [M + 2H]²⁺, 610.4 [M + H]⁺, 1219.6 [2M + H]⁺; (+)-HRESIMS m/z 610.2924 [M + H]⁺ (calcd for C₃₆H₄₀N₃O₆, 610.2917).

Bioassays

The anti-HIV and cytotoxic activities of the tested alkaloids on MT-4 cell cultures were measured as described previously¹⁹ with minor modifications. Briefly, MT-4 cells were added to 96-well plates containing serial dilutions of the tested compounds. HIV-1 NL 4-3 was used to infect the MT-4 cells at a final multiplicity of infection (MOI) of 0.03. The assay plates were incubated in a humidified incubator at 37 °C under 5% CO₂. After 3 or 4 days, 10 mL of MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] (5 mg/mL in PBS) was added into each well and the plates were incubated for another 3 h at 37 °C. A lysis buffer (10% (v/v) Triton X-100 in acidified isopropanol) was added to each well to lyse the cells and to solubilize the formazan crystal. The plates were read at a wavelength of 570 nm on a FLUOStar Optima plate reader (BMG Labtech). EC₅₀ (50% effective concentration) is defined as the concentration of each compound achieving 50% protection on MT-4 cells against the HIV-induced cytopathic effect, and CC₅₀ (50% cytotoxic concentration) in parallel is defined as the concentration of each compound killing 50% of the MT-4 cells. Both values were determined using Prism 5.0 software (GraphPad, San Diego, CA).

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Notes and references

1. B. Li, M. G. Gilbert, G. Fischer and C. A. Meyer, *Flora of China*, Science Press: Beijing, Missouri Botanical Garden Press: St. Louis, 2008, Vol 11, p 178.
2. E. Chirkin, W. Atkalian and F.-H. Porée, *The Alkaloids: Chemistry and Biology*, Academic Press: Pittsburgh, 2015, Vol 74, pp 1-120.
3. X. H. Li, M. M. Cao, Y. Zhang, S. L. Li, Y. T. Di and X. J. Hao, *Tetrahedron Lett.* 2014, **55**, 6101-6104.
4. L. S. Gan and J. M. Yue, *Nat. Prod. Commun.*, 2006, **1**, 819-823.
5. L. S. Gan, C. Q. Fan, S. P. Yang, Y. Wu, L. P. Lin, J. Ding and J. M. Yue, *Org. Lett.*, 2006, **8**, 2285-2288.
6. H. Zhang, C. R. Zhang, K. K. Zhu, A. H. Gao, C. Luo, J. Li and J. M. Yue, *Org. Lett.*, 2013, **15**, 120-123.
7. H. Zhang, W. Wei and J. M. Yue, *Tetrahedron*, 2013, **69**, 3942-3946.
8. H. Zhang, Y. S. Han, M. A. Wainberg and J. M. Yue, *Tetrahedron*, 2015, **71**, 3671-3679.
9. D. Arbain, A. A. Birkbeck, L. T. Byrne, M. V. Sargent, B. W. Skelton and A. H. White, *J. Chem. Soc.-Perkin Trans. 1*, 1991, 1863-1869.
10. G. Wang, Y. Wang, X. Zhang, Y. i, X. Yao and W. Ye, *Chem. Pharm. Bull.*, 2010, **58**, 390-393.
11. A. Ohsaki, T. Nagaoka, K. Yoneda and A. Kishida, *Tetrahedron Lett.*, 2009, **50**, 6965-6967.
12. G. C. Wang, Y. Wang, Q. Li, J. P. Liang, X. Q. Zhang, X. S. Yao and W. C. Ye, *Helv. Chim. Acta*, 2008, **91**, 1124-1129.
13. H. Wu and J. Zhou, *Zhongguo Zhongyao Zazhi*, 2004, **29**, 535-537.
14. M. Chen and L. Hou, *Zhiwu Xuebao*, 1985, **27**, 625-629.
15. A. Ahond, J. Guilhem, J. Hamon, J. Hurtado, C. Poupat, J. Pusset, M. Pusset, T. Sevenet and P. Potier, *J. Nat. Prod.*, 1990, **53**, 875-881.
16. G. Han, M. G. LaPorte, J. J. Folmer, K. M. Werner and S. M. Weinreb, *J. Org. Chem.*, 2000, **65**, 6293-6306.
17. E. V. Dehmlow, M. Guntenhoner and T. Van Ree, *Phytochemistry*, 1999, **52**, 1715-1716.
18. B. X. Zhao, Y. Wang, D. M. Zhang, R. W. Jiang, G. C. Wang, J. M. Shi, X. J. Huang, W. M. Chen, C. T. Che and W. C. Ye, *Org. Lett.*, 2011, **13**, 3888-3891.
19. C. Pannecouque, D. Daelemans and E. De Clercq, *Nat. Protoc.*, 2008, **3**, 427-434.

Graphical Abstract

New *Securinega* alkaloids with anti-HIV activity from *Flueggea virosa*

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A series of *Securinega* alkaloid hybrids and oligomers with anti HIV activity were isolated and identified from *Flueggea virosa*.

