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ARTICLE TYPE

Fluorescent Magnesium Nanocomplex in Protein Scaffold for Cell Nuclei Imaging Application

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Here in, we report a facile strategy for the synthesis of water-soluble ultra-fine blue-green emitting fluorescent Magnesium nanoparticles-protein complex (MgNC). This MgNC is demonstrated to exhibit excellent photo stability and biocompatibility. It was also observed that MgNC stain cell nuclei with high specificity.

Ultrasensitive fluorescence based imaging techniques have received considerable attention in visualization of cell organelles or study of intracellular processes in cell biology and molecular spectroscopy¹. These techniques rely on bright fluorescent probes attached to DNA, proteins, or other biological components. Particularly, in cellular processes, the specific staining of cellular compartments is highly desirable for imaging subcellular structures to distinguish dissimilar cellular compartments and also to understand localization of the cell organelles. Therefore, researchers continue to utilize best available options in fluorescence techniques for *in-vivo* imaging of cellular processes. Among many cellular interactions, the imaging of the nucleus is particularly important because it contains deoxyribonucleic acid (DNA), which dictates most intracellular process and also it is the site for DNA replication and drug-DNA interactions^{1,2}. Many sensitive probes such as organic dyes and transition metal complexes have been used for imaging of the cells³⁻⁷. However, their cost and limited pH range restricts their use for imaging in cells.

QDs (such as CdSe, CdTe, and PbTe) have received broad attention due to their unique optical properties and biocompatibility. However, the applications of QDs are limited due to the release of Cd²⁺, Pb²⁺, or other heavy metal ions that exert cytotoxicity in *in-vivo* systems and also potential environmental hazard^{6,7}. In past few years, extensive studies have been performed on luminescent Au (gold) and Ag (silver) nanoclusters or nanocomplex (NC) using various templates such as peptides, dendrimers, polymers, and proteins⁸⁻¹³. The nanocomplex are attractive because of their high fluorescence, good photostability, less toxicity, good biocompatibility and solubility^{13e}. Most recently, biomolecules such as protein, enzyme, and peptide capped copper NC and carbon dots have been used in the biomedical imaging owing to their multicolor emission

capabilities and non-toxic nature¹⁴.

It is a well known fact that nucleic acids have affinity to bind divalent ions such as magnesium and stabilise DNA and RNA structure¹⁵. Magnesium ions have a critical role in maintaining the positional integrity of closely clustered phosphate groups. These clusters appear in numerous and distinct parts of the cell nucleus and cytoplasm, which coordinates with magnesium ion to form aggregates¹⁶. This prompted us to explore development of bioinspired magnesium nanoparticles for imaging of cell nucleus, which is the most important organelle controlling in the growth, proliferation, and apoptosis of a cell. Nevertheless, only a few experimental studies have given direct insight into magnesium nanoparticles¹⁷ primarily because of the difficulty in preparing highly stable and extremely tiny Mg nanoparticles. Additionally, subnanometer sized Mg intrinsically suffers from unstable colloidal dispersion of its particles. So, it is of great interest to develop stable, highly biocompatible and fluorescent Magnesium nanoparticles complexed with protein scaffold with visible emission. This MgNC was also observed to stain cell nuclei with high specificity.

Exclusively, we report for the first time synthesis of biocompatible, ultrafine MgNC by a simple one-pot chemical reduction method using bovine serum albumin (BSA) as protein

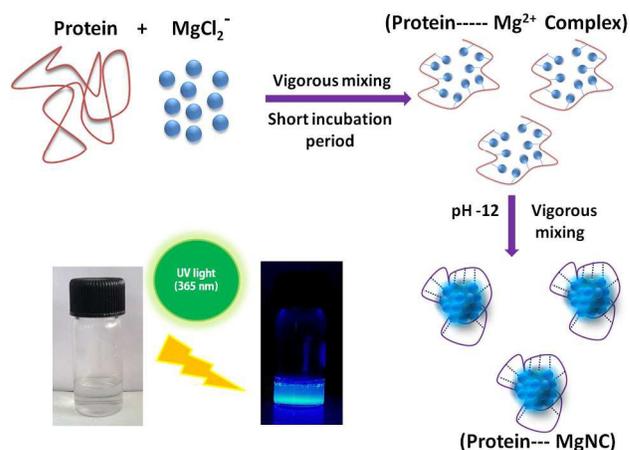


Fig.1 The facile synthetic route for producing BSA mediated synthesis of MgNC. (Inset: capped MgNC emitting blue-green fluorescence when excited with a handheld UV lamp (365 nm)).

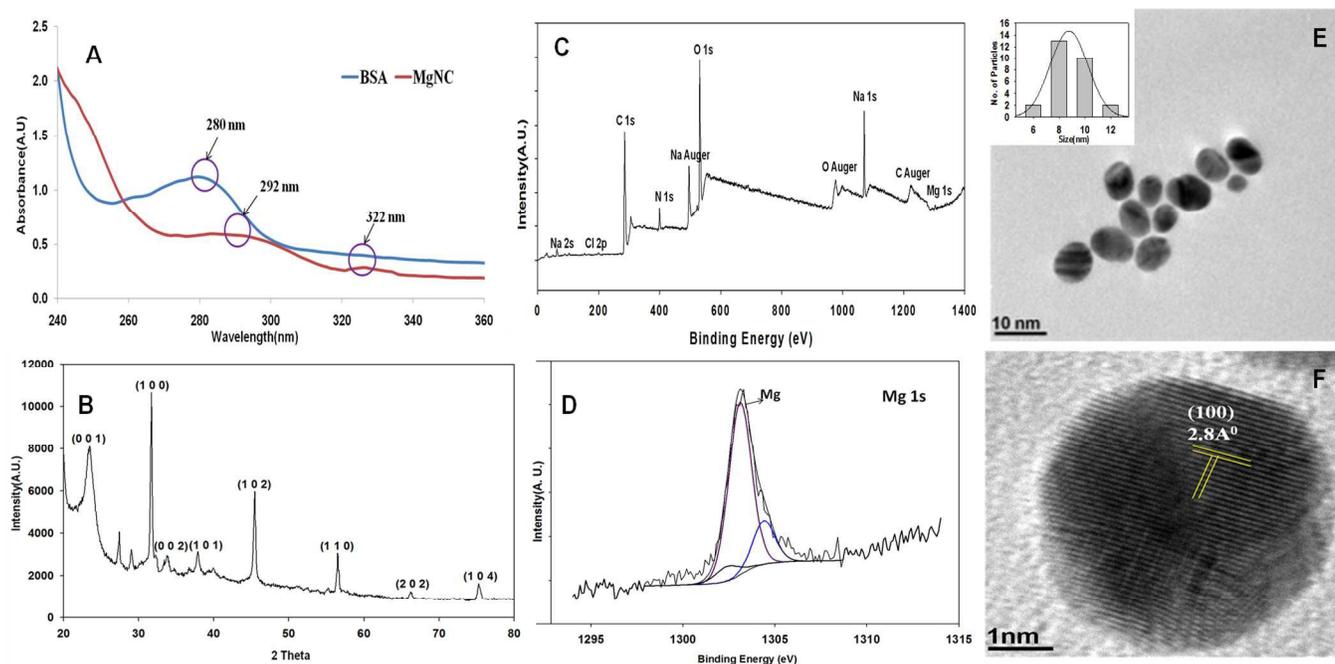


Fig. 2 (A) UV-Vis absorption spectra of BSA (blue line) and MgNC suspension (red line). (B) XRD pattern of MgNC indexed to the hexagonal phase of Mg (JCPDS 04-0770), (C) XPS survey spectrum of MgNC, (D) XPS spectrum in the Mg 1s region of MgNC, (E)(F) Low and high resolution transmission electron micrograph of MgNC (inset E: Mean particle size distribution)

matrix (**Fig.1**). It has been demonstrated that BSA can be used as the model protein for the synthesis and stabilization of metal nanostructures^{14b}. The as-synthesized bioconjugated MgNC exhibit emission and has been characterized using various spectroscopic and microscopic techniques. Further, using fluorescence microscopy, we studied the cellular uptake of MgNC, including uptake kinetics and intracellular localization.

A typical UV-Vis absorption spectrum of an aqueous solution containing free BSA and MgNC is presented in **Fig. 2A**. The absorption peak at 280 nm for bare BSA is attributed to the π - π^* transition of aromatic amino acid residue characteristic of polypeptide backbone structure C=O, whereas the absorption peaks at 292 and 322 nm indicate the shift in π - π^* transition and is attributed to the formation of MgNC (**Fig.2A**). The formation of any kind of oxidation product of the amino acids in protein is ruled out as also discussed in studies from other research groups^{14b}. The observed 322 nm absorption peak as well as the emission maxima revealed in the present study is purely due to the formation of MgNC. The low and high resolution transmission electron microscopy (HRTEM) image (**Fig. 2E,F**) shows average size of these ultrafine monodispersed MgNC of size ~ 8.5 nm (**Inset 2E**) which was calculated using the software (Image-J). Crystal lattice fringes with 2.8 Å spacing indicates the

(100) planes of the metallic Mg (**Inset 2F**). Further the average hydrodynamic size of MgNCs analyzed by DLS measurements was found to be 12 ± 0.7 nm. The observed change in size compared to TEM is attributed to protein swelling and fragmentation to accommodate MgNC. Protein fragmentation with multimodal distribution for the coexistence of MgNC and free protein as well as enhancement of aggregation is also observed (**Fig. S1**). It is well known that fragmentation of protein can occur when the pH of the environment changes drastically^{14b}. All together, these confirm the formation of protein- magnesium nanocomplex.

Powder X-ray diffraction (PXRD) of MgNC shows the crystalline nature of the MgNC and matched with the face-centered cubic structure of metallic magnesium (**Fig.2B**). The PXRD spectrum contains multiple peaks which are clearly distinguishable. X-ray diffraction patterns of as-synthesized MgNC match with diffraction patterns of hexagonal Mg (solid black line, JCPDS 04-0770). Peaks located at $2\theta = 23.3^\circ, 31.6^\circ, 33.6^\circ, 37.8^\circ, 45.3^\circ, 56.4^\circ, 66.1^\circ$ and 75.2° are assigned to the (001), (100), (002), (101), (102), (110), (202) and (104) planes of hexagonal magnesium, respectively. No significant diffraction peak is observed for magnesium oxide phase (JCPDS 89-7746) or any other magnesium phase except hexagonal Mg indicating that the product prepared via this route is single phase metallic magnesium.

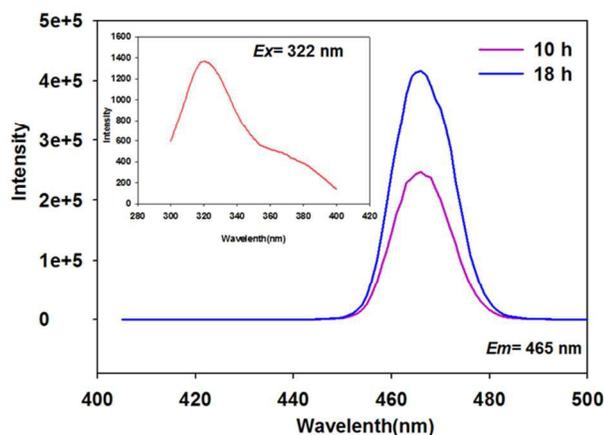


Fig. 3 Fluorescence emission spectra taken at different time intervals (10 h and 18 h), during the course of reduction.

Elemental identification and oxidation state of magnesium in MgNC were determined by X-ray photoelectron spectroscopy (XPS). The deconvolution of XPS spectra was carried out by CasaXPS software using Gaussian/Lorentzian (GL(30)) line shape and Shirley background correction. The XPS survey spectrum (Fig. 2C) shows the existence of C, N, O, Na, Cl and Mg in the magnesium nanocomplex. Fig. 2D shows the deconvoluted high resolution spectrum of Mg 1s with one intense peak centered at a binding energy of 1303.1 eV which is assigned to Mg(0). These results are consistent with XPS measurements performed by Gawande et al¹⁹. In addition, two minor peaks are also observed at binding energies of 1304.4 eV and 1302.4 eV, which can be associated to the formation of Mg-S and Mg-COO type species upon reaction with BSA located on the surface of the MgNC cluster. It is further in agreement with our observations of presence of thiol and carboxyl group features in FT-IR. The XPS spectrum (Fig. S2 A,B,C) shows the presence of C, N and O in the magnesium nanocomplex. The peaks with binding energy of C 1s (284.8 eV, 288.5 eV), N 1s (400 eV, 402.5 eV), and O 1s (533 eV) elements are probably due to the presence of BSA protein in MgNC and taken together, these data validate the successful synthesis of protein- magnesium nanocomplex.

To further investigate the interaction between BSA and magnesium during nanocomplex formation, fluorescence spectra was recorded. As shown in Fig. 3, the luminescence of MgNC revealed distinct excitation and emission maxima at $\lambda_{ex} \approx 322$ and $\lambda_{em} \approx 465$ nm, respectively. The peak position at 322 nm of the excitation spectrum was almost alike to the MgNC absorption band in Fig. 3. As the reaction progressed, a gradual increase in the luminescence ($\lambda_{em} \approx 465$ nm) from the solution was observed. After around 18 h of reaction, there was no further enhancement in the luminescence and the reaction was considered to be terminated. Quantum yield (QY) of MgNC was determined by using quinine sulfate (QY = 0.54) in 0.5 M sulfuric acid as a reference¹⁸. QY of MgNC was found to be 0.17 in ethanol. This luminescence of MgNC enabled their observation by *in-vitro* imaging with minimal interference from cellular autofluorescence. Further, in order to probe the role of NaOH in

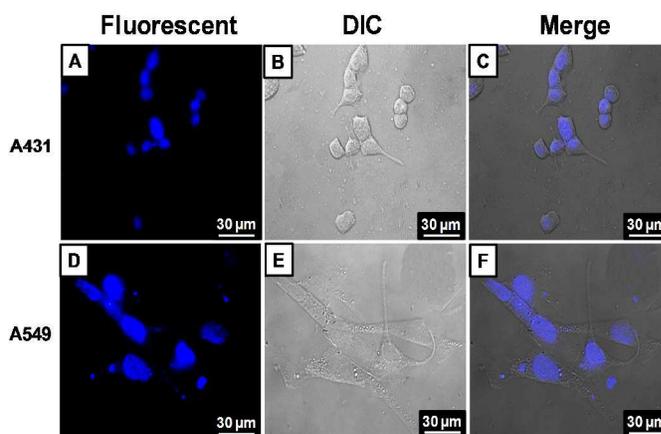


Fig. 4 Fluorescence microscope images of A549 and A431 cell lines after 24 h exposure with MgNC ($\lambda_{em} = 465$ nm); to A431 (A-C) and A549 cells (D-F). The images are pseudo coloured as blue which corresponds to the presence of MgNC

the synthesis of MgNC, we recorded the fluorescence of complex (Mg + BSA), without NaOH, but as expected we did not find any peak at $\lambda_{em} \approx 465$ nm (Fig.S3A). In addition, we also recorded UV-Vis spectra of complex without NaOH and we found the absorbance peak at the wavelength of 280 nm only, which is due to presence of BSA (Fig.S3B) and no absorbance signature corresponding to MgNC was found. Hence it is confirmed that fluorescence, post addition of NaOH, at $\lambda_{em} \approx 465$ nm and $\lambda_{ex} \approx 322$ nm is due to formation of MgNC and not from either BSA or pure salt alone or physical mixture of both.

The chemical stability of MgNC was determined at different salt concentrations such as 100 mM NaCl, and PBS buffer at pH 7.5. After 24h incubation, the UV-visible absorbance and emission spectrum of MgNC did not show any significant change suggesting high chemical stability of MgNC at varied salt concentrations (Fig.S4). FT-IR spectrum of the MgNC comparison with BSA, is shown in Fig.S5. The peaks in the spectra were assigned as follows: 1635 cm^{-1} corresponds to a protein with a high proportion of α -helix, which is the characteristic amide band of BSA; 1454 cm^{-1} is attributed to strong primary amine or scissoring while 3429 cm^{-1} is attributed to primary amines; 2937 cm^{-1} is ascribed to C-H vibration^{20a}. The disappearance of S-H stretching band (2499 cm^{-1}) of MgNC is also observed, suggesting the formation of metal-thiol bonds. The MgNC formed in the BSA solution could have been stabilized by a combination of Mg-S and Mg-COO⁻ bonding with the protein (via the amino acid residues in BSA), and the steric protection due to the bulkiness of the protein¹⁶. The relationship between the fluorescence intensity and pH was also studied. The pH value did not affect the fluorescence intensity of MgNC. We further investigated the photostability of MgNC over a period of 4 months and observed persistent emission (Fig.S6).

The high luminescence properties and superior photo and chemical stability of these MgNC encouraged us to investigate their possible bioimaging application. The A549 cell line was chosen to assess bio-compatibility using MTT assay at two different concentrations (50 and 100 $\mu\text{g}/\text{mL}$) of MgNC. No

significant cell death of A549 cells was observed at both the concentrations even after exposure for 24 h (Fig. S7). Moreover, to substantiate the cytotoxicity, nuclear morphological analysis was also done with MgNC (Fig.S8). Cells were treated with MgNC at 100 µg/mL for 24 h and cell nucleus were stained with DAPI as a control to elicit any apoptotic nuclei. Both MgNC treated and untreated cell's nuclei exhibited normal morphology. No change in nuclear morphology confirmed the non-toxic nature of MgNC even upto 100 µg/mL. This suggested that this nanocomplex is highly biocompatible and does not affect the proper functioning of the nucleus. Based on the aforesaid observations the concentration of 50 µg/mL of MgNC was selected for further experiments.

The A549 and A431 cells (6×10^4 cells/well) were exposed to 50 µg/mL of MgNC for 24 h to investigate their fluorescence imaging capability. Fluorescence microscopy of A549 and A431 cells treated with MgNC shows significant emitting fluorescence from the cells indicating efficient internalization. Interestingly, it is evident from the image that the emission arises from the cell nucleus (Fig. 4A,D). Further, nuclear localization was also confirmed by imaging the cells under DIC mode after treated with MgNC (Fig. 4B,E) as other cell components including the plasma membrane were unstained, establishing the potential of the MgNC for nuclear staining. The merged image (Fig. 4C,F) clearly supports that the fluorescent MgNC overlaps with dense nucleus region of DIC image. The A549 and A431 cells retains their normal morphology after MgNC exposure, supporting our observation with MTT. The high specificity of MgNC to cell nucleus may be due presence of free functional groups of denatured BSA over the surface of nanocomplex¹.

To see the distribution of particles in cells, we performed uptake kinetics study with 50 µg/mL of MgNC in A549 cells exposed for 1–24 hours. A549 cells were incubated with MgNC for different time (1 h, 3 h, 6 h, 12 h and 24 h) and the nuclei are also co-stained with DAPI. The nuclear and cytoplasmic regions are clearly evident under DIC mode of imaging. The merged image (Fig.S9) clearly supports that fluorescent MgNC and DAPI stained (blue) region overlaps with the dense nuclear region of DIC image. This observation suggest that the internalized MgNC eventually accumulate in cell nucleus. It is also interesting to note that within 1 h of incubation of MgNC, cells start to internalize the MgNC. It is also obvious from Fig. S9 that some MgNC are dispersed in cytoplasm (at 3 h and 6 h) while most of them have accumulated in nucleus. However, after 12 h of exposure, MgNC are mostly present in nuclear region and very limited green fluorescence can be seen in cytoplasmic region. This could be due to the fact that after 6 h cells do not internalize more MgNC, rather accumulate them in nucleus. Similar observation has been reported by Singh et al^{20b}, where carboxyfluorescein modified cerium oxide nanoparticles (< 10 nm) widely accumulate in cell nucleus. In order to investigate that the fluorescence is produced by MgNC only and not due to tyrosine and tryptophan residues present in BSA or any other species, we also performed imaging with unreduced MgNC (without NaOH). Exposure to cells showed no green fluorescence, as evident by Fig.S10, this

suggests that the fluorescence is due to formation of MgNC only and not from BSA or other residue.

In addition, to confirm the internalization of MgNC, flow cytometric analysis was carried out. A concentration dependent (10, 20, 30, 40, 50 µg/mL) increase in both forward scatter (FSC) as well as in side scatter (SSC) was observed (Fig S11). Interestingly, the increase in the FSC at concentration (10, 20, 30 µg/mL) was substantial and at the same time, SSC augmented at the concentrations of 40 and 50 µg/mL at 24 h. It clearly suggests that MgNC are internalized via plasma membrane without bilayer disruption, followed by their entrapment in nucleus as increase in forward scatter signifies the internalisation^{21a}. It can be concluded that the structural organization of surface chemical groups plays a key role in regulating cell-membrane diffusion. Early endosomes are known to gradually mature into late endosomes and eventually transform in lysosomes^{21b}. It may possibly be due to the “evasion” of MgNC from the lysosomes and ability to move towards the cell nucleus, an advantage compared to other nanoparticles. The aforesaid observations clearly suggest that these ultrasmall particles mainly follow an endosomal pathway for internalization and are ultimately transferred to nucleus.

In conclusion, we have demonstrated a simple method for preparing water-soluble blue green emitting fluorescent MgNC that exhibit superior fluorescence, high chemical and photo stability. The application of MgNC could be as fluorescent nanoprobe to stain nuclei. Further, these non-toxic MgNC could be used a potential drug carrier and DNA transfecting agents. It is expected that more characterisation will unravel the other possible bio-applications of these photo-stable MgNC.

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References:

1. V. Venkatesh, A. Shukla, S. Sivakumar, and S. Verma, *ACS Appl. Mater. Interfaces*, 2014, **6**, 2185–2191
2. D. H. M. Dam, J. H. Lee, P. N. Sisco, D. T. Co, M. Zhang, M. R. Wasielewski, T. W. Odom, *ACS Nano* 2012, **4**, 3318–3326
3. (a) L. Shang, S. Dong, G. U. Nienhaus, *Nano Today*, 2011, **6**, 401 (b) L. Shang, R. M. Dorlich, S. Brandholt, R. Schneider, V. Trouillet, M. Bruns, D. Gerthsen, G. U. Nienhaus, *Nanoscale*, 2011, **3**, 2009–2014; (c) P. Bian, J. Zhou, Y. Liu, Z. Ma, *Nanoscale* 2013, **5**, 6161–6166.
4. W. C. Chan, D. J. Maxwell, X. Gao, R. E. Bailey, M. Han, S. Nie, *Curr. Opin. Biotechnol.* 2002, **13**, 40–46.
5. R. Hardman, *Environ. Health Perspect.* 2006, **114**, 165–172.
6. (a) J. R. Roberts, J. M. Antonini, D. W. Porter, R. S. Chapman, J. F. Scabilloni, S. H. Young, *Part. Fibre Toxicol.* 2013, **10**, 1–17; (b) A. S. Karakoti, R. Shukla, R. Shanker, S. Singh, *Adv Colloid Interface Sci.*, 2015, **215**, 28–45
7. J. Zheng, C. A. Nicovich and R. M. Dickson, *Annu. Rev. Phys. Chem.*, 2007, **58**, 409.

8. H. Kawasaki, K. Hamaguchi, I. Osaka and R. Arakawa, *Adv. Funct. Mater.*, 2011, **21**, 3508.
9. H.W.Duan and S.M.Nie, *J. Am. Chem. Soc.*, 2007, **129**, 2412.
10. H. X. Xu and K. S. Suslick, *Adv. Mater.*, 2010, **22**, 1078.
11. (a)T. Vosch, Y. Antoku, J. C. Hsiang, C. I. Richards, J. I. Gonzalez and R. M. Dickson, *Proc. Natl. Acad. Sci. U. S. A.*, 2007, **104**, 12616,(b) N.Zhao, H. G Bagaria, M. S Wong, Y. Zu, *J. of Nanobiotech.* 2011, **9**,1-12
12. (a)Y. Pei, Y. Cao and X. C. Zeng, *J. Am. Chem. Soc.*, 2008, **130**,7830,(b) T. Chen, Y. Hu, Y. Cen, X. Chu, and Y. Lu, *J. Am. Chem. Soc.*, **2013**, **135**,31,11595–11602
13. (a)D.M.Chevrier,A.Chatt,P.J.Zhang,*Nanophotonics*,2012,**6**,0 64504-1-064504-1,(b)Z.Wang,R.Zhang,Z.Wang,H.F.Wang, Y.Wang,J. Zhao,F.Wang,W.Li,G.Niu,D.O.Kiesewetter, and X.Chen,*ACS Nano*,**2014**, **8**,12,12386–12396,(c)J-H Choi,H-J Hwang,S.W Shin,J-W.Choi,S. H. Um and B-K Oh, *Nanoscale*, **2015**,**7**, 9229-9237,(d) E. Ju, Z. Liu,Y.Du, Y.Tao,J.Ren,and X. Qu,*ACS Nano*, **2014**, **8**, 6, 6014–6023,(e)S.Singh,V.D’Britto,A.A.Prabhune,C.V.Ramana,A. Dhawan and B. L. V. Prasad,*New J.Chem.*,2010,**34**, 294-301
14. (a)H. Kawasaki, K. Hamaguchi, I. Osaka, R. Arakawa, *Adv Funct. Mater.* 2011, **21**, 3508-3515(b)N. Goswami,A. Giri, M. S. Bootharaju, P. L.Xavier,T.Pradeep and S.Kumar Pal, *Anal. Chem.* 2011, **83**, 9676–9680
15. C. Sissi and M. Palumbo,*Nucl.Acids Res.*,2009,**37**,3,702-711
16. G.J.He,W.F.Liu and Y.B.Yan,*Int.J.Mol.Sci.*2011,**12**,2901-2916
17. (a)S. B. Kalidindi and B. R.Jagirdar,*Inorg.Chem.*,2009,**48**, 10,4524–4529,(b)L.Haasa and A. Gedanken,Chem. Commun., 2008, 1795-1797
18. A. M. Brouwer, *Pure Appl. Chem.* 2011, **83**, 2213–2228
19. M.B Gawande,P. S Branco, K. Parghi,J.J Shrikhande,R. K. Pandey,C.A.A.Ghumman, N. Bundaleski,O.M.N.D. Teodoro, R. V Jayaram, *Catal. Sci. Technol.*,2011,**1**, 1653-1664
20. (a) S. Singh,U. M. Bhatta, P. V. Satyam, A. Dhawan, M.Sastrya and B. L. V. Prasad, *J. of Mater. Chem.*,2008,**18** 22,2601-2606;(b)S. Singh, A. Kumar, A. Karakoti, S. Seal, and W. T. Self, *Mol. Biosyst.* 2010,6,10,1813–1820
21. (a)H.Suzuki,T. Toyooka,Y.Ibuki,*EnvironSci.Technol.*, 2007, **41**,3018–3024,(b) J. P. Luzio, P. R. Pryor and N. A. Bright, *Nature Reviews Molecular Cell Biology* ,2007,**8**, 622-632

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