ChemComm



ChemComm

H2O2-activated triplet-triplet annihilation upconversion via modulation of the fluorescence quantum yields of the triplet acceptor and the triplet-triplet-energy-transfer efficiency

Journal:	ChemComm
Manuscript ID:	CC-COM-05-2015-004325.R1
Article Type:	Communication
Date Submitted by the Author:	21-Jun-2015
Complete List of Authors:	Tao, Renjie; State Key Laboratory of Fine Chemicals, Zhao, Jianzhang; Dalian University of Technology, State Key Laboratory of Fine Chemicals Zhong, Fangfang; Dalian University of Technology, Zhang, Caishun; State Key Laboratory of Fine Chemicals, Yang, Wenbo; State Key Laboratory of Fine Chemicals, Xu, Kejing; State Key Laboratory of Fine Chemicals,

SCHOLARONE[™] Manuscripts Cite this: DOI: 10.1039/coxx00000x

www.rsc.org/chemcomm

COMMUNICATION

H₂O₂-activated triplet-triplet annihilation upconversion via modulation of the fluorescence quantum yields of the triplet acceptor and the triplettriplet-energy-transfer efficiency

Renjie Tao, Jianzhang Zhao,* Fangfang Zhong, Caishun Zhang, Wenbo Yang and Kejing Xu

s Received (in XXX, XXX) Xth XXXXXXXX 2015, Accepted Xth XXXXXXXX 2015 DOI: 10.1039/b000000x

Oxidation-activatable triplet-triplet annihilation (TTA) upconversion was achieved with 9,10-bis(diphenylphosphino)anthracene (BDPPA, nonfluorescent) as activatable triplet 10 acceptor/emitter, which can be oxidized to BDPPA-O (highly fluorescent) by H₂O₂ under mild condition, thus TTA upconversion was switched on by H₂O₂.

Triplet-triplet annihilation (TTA) upconversion have attracted much attention.¹⁻¹⁴ TTA upconversion is unique because of the requirement of only low power and incoherent photoexcitation (solar light is sufficient), strong absorption of visible light (ε = 80000 – 100000 M⁻¹ cm⁻¹), and high upconversion quantum yields (up to 20–30%).^{15,16} Moreover, the system is in supramolecular feature and it is readily tunable by using different ²⁰ triplet photosensitizers and triplet acceptors.² Recently TTA upconversion has been used in photovoltaics,^{17,18} in vivo luminescent bioimaging,^{19,20} and light-responsive actuators.²¹ However, in order to add more modulability to the application of TTA upconversion, optically or chemically-switchable TTA

²⁵ upconversion system is highly desired. But such TTA upconversion systems are rarely reported.^{22,23}

Concerning this aspect, reactive oxygen species (ROS)activatable TTA upconversion is of in great interest. ROS are endogenous chemical species and they play important roles in the ³⁰ metabolism of cells. For instance, hydrogen peroxide (H_2O_2) is involved in oxidative stress, and it is a second messenger in normal cellular signal transduction.^{24,25} Accordingly, a great

amount of fluorescent probes have been developed for detection of H_2O_2 in living cells.^{26–31} However, no H_2O_2 -activated TTA ³⁵ upconversion system was reported.^{1,2} Such a TTA upconversion system will be tremendously important for in vivo luminescence bioimaging of the endogenous H_2O_2 produced in living cells.

Herein we developed a H_2O_2 -activatavle TTA upconversion system, performed under mild conditions (room temperature).

⁴⁰ 9,10-bis(diphenylphosphino)anthracene (**BDPPA**. Scheme 1) was used as H_2O_2 -responsive triplet acceptor/emitter.²¹ **BDPPA** is non-fluorescent. Upon oxidation to 9,10-bis(diphenylphosphoryl)anthracene (**BDPPA-O**. highly fluorescent. Scheme 1) by H_2O_2 under room temperature, the TTA upconversion is ⁴⁵ switched on. **PdTPTBP** (TPTBP = tetraphenyltetrabenzoporphyrin, Scheme 1) was used as red lightexcitable triplet photosensitizer.^{32,33} Upon addition of H_2O_2 , redto-blue upconversion was achieved, which is distinctive to unaided eyes. This system may be useful for H_2O_2 -responsive ⁵⁰ luminescence bioimaging with TTA upconversion.

We noted that the **BDPPA-O** was prepared under mild condition by oxidation of **BDPPA** with H_2O_2 .^{21,34,35} Thus we envision that a H_2O_2 -activatable TTA upconversion can be developed with **BDPPA**. In the absence of H_2O_2 , ⁵⁵ **PdTPTBP/BDPPA** gives no upconversion due to the low fluorescence quantum yield of **BDPPA**. In the presence of H_2O_2 , **BDPPA** will be oxidized to **BDPPA-O**, which is highly fluorescent, thus the TTA upconversion will be switched on.



Scheme 1. Triplet photosensitizer PdTPTBP, and the triplet acceptors BDPPA and BDPPA-O used in this study.

Firstly the UV–vis absorption of **BDPPA** and **BDPPA-O** were studied (Fig. 1a). The two compounds give similar ⁷⁵ absorption bands in the range of 325 - 450 nm. **BDPPA-O** gives slightly red-shifted absorption band than **BDPPA.** In the presence of H₂O₂, **BDPPA** was oxidized to **BDPPA-O** under room temperature, which is demonstrated by the change of the UV–vis absorption spectra (Fig. 1b).

Significant fluorescence enhancement was observed upon oxidation of BDPPA (Fig. 2a). BDPPA shows negligible fluorescence, whereas BDPPA-O is strongly fluorescent (54-fold stronger than BDPPA. Optically matched solutions were used).^{21,34} We also carried out the *in situ* oxidation of BDPPA in ⁸⁵ the solution, and the fluorescence variation was monitored (Fig. 2b and ESI †, see Fig. S7 for the time course curve of the oxidation). Fast fluorescence enhancement was observed for BDPPA in the presence of H₂O₂ (*k* = 360 s⁻¹). The fluorescence

switching of **BDPPA** upon oxidation indicates that H_2O_2 activatable TTA upconversion can be achieved with **BDPPA** as the triplet energy acceptor/fluorescence emitter. The photophysical properties of the compounds were listed in Table 1.



¹⁵ Fig. 1 (a) UV–Vis spectra of **BDPPA** and **BDPPA-O**. (b) UV–Vis spectra of **BDPPA** after addition of 10 μ L 30% H₂O₂, $c = 1.0 \times 10^{-5}$ M in MeOH, 20 °C.



Fig. 2 (a) Fluorescence spectra of **BDPPA** and **BDPPA-O** (excited at 391 nm, optically matched solutions were used, A = 0.079 in MeOH); (b) Emission spectra of **BDPPA** after adding 30% H₂O₂ (10 µL). $c = 1.0 \times 30 \ 10^{-5}$ M in MeOH, 20 °C.

PdTPTBP was used as the triplet photosensitizer for the TTA upconversion (Fig. 3), because the absorption matches with the 635 nm solid state laser.^{32,33} **PdTPTBP/BDPPA** gives no TTA upconversion ($\Phi_{UC} = 0\%$). This result is tentatively attributed to ³⁵ the low fluorescence quantum yield of **BDPPA** ($\Phi_F = 1.4\%$). Interestingly, we noted the phosphorescence of **PdTPTBP** was quenched by half, which indicates TTET.^{1,2} With **BDPPA-O**, however, strong emission at 470 nm was observed. The emission band is the same to the prompt fluorescence of **BDPPA-O** itself.

- ⁴⁰ Excitation of **BDPPA-O** alone at 635 nm under the same condition does not produce any fluorescence emission at 470 nm, (see ESI †, Fig. S8) thus the TTA upconversion with **BDPPA-O** was verified (quantum yield $\Phi_{UC} = 1.8\%$). Interestingly, we found the quenching effect of **BDPPA-O** on the phosphorescence
- ⁴⁵ of **PdTPTBP** is much significant than that of **BDPPA**, which indicates more efficient TTET between **PdTPTBP** and **BDPPA-O** than that of **BDPPA** (confirmed in later section).³⁶

The TTA upconversion with in situ oxidation of **BDPPA** by H_2O_2 was also studied (Fig. 3b). No TTA upconversion was 50 observed for **BDPPA** (with **PdTPTBP** as the triplet





Fig. 3 (a) Upconversion spectra of BDPPA and BDPPA-O ($c = 7.0 \times 10^{-5}$ M in deaerated MeOH) after addition of PdTPTBP ($c = 1.0 \times 10^{-5}$ M) and PdTPTBP only ($c = 1.0 \times 10^{-5}$ M in deaerated MeOH), ⁷⁰ respectively. (b) Upconvertion spectra of mixture BDPPA (7.0×10^{-5} M in deaerated MeOH) and PdTPTBP ($c = 1.0 \times 10^{-5}$ M) after adding 20 μ L 30% H₂O₂. The asterisk in (a) and (b) indicated the scattered laser (635 nm, 10.0 mW, 27.8 mW cm⁻²). 20 °C.

The red-to-blue upconversion is visible to un-aided eyes (Fig. 75 4). No upconversion was observed with **BDPPA**, but the phosphorescence of the **PdTPTBP** was quenched (Fig. 4c). In the presence of **BDPPA-O**, strong blue emission was observed (Fig. 4a). In situ oxidation of **BDPPA** with H₂O₂ gives similar results (Fig. 4d–4f). Moreover, we found that ¹O₂ exerts negligible ⁸⁰ fluorescence enhancement on BDPPA as compared with that of H₂O₂ (see ESI[†], Fig. S11). These results infer that the H₂O₂-actiavated TTA upconversion may be used for in vivo imaging of H₂O₂, probably implemented in the form of polymer film,³⁷ microcapsule or microbeads.^{19,38–41} In this case the H₂O₂ may ⁸⁵ easily diffuse into the matrix to arouse the oxidation reaction, thus to switch ON the TTA upconversion.





Table 1.	Photophysica	l properties of the	compounds

	λ_{abs}^{a} / nm	e ^b	λ em / nm	Φ _F ^c / %	$\tau_{\rm F}^{d}$ / ns	$\tau_{\rm T}$ $^{e}/\mu{ m s}$
BDPPA	373/394/414	0.76/1.04/1.05	465	1.4	14.2	f
BDPPA-O	396/418/441	0.96/1.08/1.04	470	75.3	14.4	_ <i>f</i>
PdTPTBP	628	10.8	796	0	<i>f</i>	61.3

^{*a*} In MeOH ($c = 1.0 \times 10^{-5}$ M). ^{*b*} Molar absorption coefficient in toluene. $\varepsilon : 10^4$ M⁻¹ cm⁻¹. ^{*c*} Fluorescence quantum yield. ^{*d*} Fluorescence lifetime. ^{*e*} Triplet 105 excited state lifetime. ^{*f*} Not applicable.



Fig. 5 Nanosecond time-resolved transient absorption spectra of (a) PdTPTBP (10 μ M), (c) mixture of PdTPTBP (10 μ M) with BDPPA (30 30 μ M) and (e) mixture of PdTPTBP (10 μ M) with BDPPA-O (30 μ M); (b), (d) and (f) are the respective decay traces of (a), (c), and (e), with 445 nm nanosecond laser excitation. $c = 1.0 \times 10^{-5}$ M, in deaerated MeOH, 20 °C.

The TTA upconversion indicates that **BDPPA** and **BDPPA-O** exerts different quenching effect on the ³⁵ phosphorescence of **PdTPTBP** (Fig. 3). Thus we studied the TTET in detail with nanosecond transient absorption (TA) spectroscopy (Fig. 5).^{42,43} Bleaching bands at 435 nm and 625 nm were observed upon laser excitation for PdTPTBP (Fig. 5a), which is due to depletion of the ground state. Excited ⁴⁰ state absorption (ESA) in the range of 450 nm – 550 nm were observed, which is attributed to the $T_1 \rightarrow T_n$ transitions. In the presence of **BDPPA**, the optical density (O. D.) values of the bleaching band significantly reduced (Fig. 5c), the triplet state lifetime of **PdTPTBP** was also slightly quenched (Fig. 5d). ⁴⁵ These changes comfirm the TTET with **BDPPA** as the triplet energy acceptor. In the presence **BDPPA-O**, the O.D. values

energy acceptor. In the presence **BDPPA-O**, the O.D. values of the TA spectra of **PdTPTBP** were quenched to larger extent, and the triplet state lifetime of **PdTPTBP** was reduced more significantly (see ESI †, Table S1). These results so indicated that **BDPPA-O** is a more efficient quencher than **BDPPA**, which is in agreement with the results observed in the TTA upconversion (Fig. 3).⁴⁴

The quantitative comparison of the TTET was performed by using the Stern-Volmer quenching curves (Fig. 6). The ⁵⁵ figure clearly show that the **BDPPA-O** is more efficient then **BDPPA** as a triplet quencher, for which the Stern-Volmer constants are 6.3×10^4 M⁻¹ and 1.5×10^4 M⁻¹, respectively (Table 2).



⁷⁰ Fig. 6 Stern-Volmer plots of PdTPTBP with BDPPA and BDPPA-O as the triplet acceptor (quencher). c[PdTPTBP] = 1.0×10^{-5} M in deaerated MeOH, 20 °C.

Since the triplet photosensitizer is the same (PdTPTBP), the different TTET efficiency of the triplet acceptors may be 75 due to either the molecular size (diffusion-controlled bimolecular rate constants),⁴⁵ or the energy gap of the T₁ state of the photosensitizer and the triple acceptor.¹ Since **BDPPA** and **BDPPA-O** are with similar molecular size thus diffusioncontrolled bimolecular collision rate constants are similar (k_0 . ⁸⁰ Table 2).⁴⁵ In order to further reveal the different quenching efficiency of BDPPA and BDPPA-O on the triplet state of **PdTPTBP**, the T_1 state energy levels of the two acceptors were calculated with TD-DFT method, as 1.65 eV and 1.52 eV, respectively. The T₁ state energy level of PdTPTBP is 85 approximated as 1.56 eV with the phosphorescence. Thus, with larger energy gap between the T₁ states of the photosensitizer and acceptor, BDPPA-O is a better triplet acceptor (quencher) than BDPPA. This different TTET efficiency contributes to the different TTA upconversion 90 efficiency of two triplet acceptors.

In summary, we developed a H_2O_2 -activatable triplettriplet annihilation upconversion system, with **PdTPTBP** as triplet photosensitizer, and 9,10bis(diphenylphosphino)anthracene (**BDPPA**) as H_2O_2 -⁹⁵ activatable triplet acceptor/emitter. **BDPPA** is non-fluorescent ($\Phi_F = 1.4\%$), but it is able to be feasibly oxidized to 9,10bis(diphenylphosphoryl)anthracene (**BDPPA-O**) by H_2O_2 ,

	$K_{\rm sv}/10^4{ m M}^{-1}$ b	$k_0 / 10^{10} \mathrm{M}^{-1} \mathrm{s}^{-1} ^{c}$	$k_{\rm q}/ 10^8 { m M}^{-1}{ m s}^{-1} {}^d$	$f_{\mathrm{Q}}(\%)^{e}$	${\it P}_{ m UC} \left(\% ight)^{ m f}$
BDPPA	6.3	3.8	2.5	0.65	0
BDPPA-O	1.5	3.9	10.3	2.63	1.8

^{*a*} In deaerated MeOH. ^{*b*} Stern-Volmer quenching constants. ^{*c*} Diffusion-controlled bimolecular quenching rate constants. ^{*d*} Bimolecular quenching constants. ^{*e*} Quenching efficiency. ^{*f*} Upconversion yields measured with **M-1** (see ESI \dagger , Scheme. S1) as the standard ($\Phi_L = 10.1\%$ in DCM).

which is strongly fluorescent ($\Phi_F = 75.3\%$). Thus TTA upconversion was switched on with H₂O₂. Un–aided eye visible red-to-blue upconversion was observed. Moreover, nanosecond

- $_{\rm 5}$ transient absorption spectroscopy demonstrated that **BDPPA-O** is a more efficient triplet quencher than **BDPPA**, for which the Stern-Volmer constant is 6.3 \times 10⁴ M⁻¹ and 1.5 \times 10⁴ M⁻¹, respectively. The different energy transfer may also contribute to the different upconversion efficiencies. TD-DFT calculation
- ¹⁰ indicated that the T_1 state energy level of **BDPPA-O** (1.52 eV) is lower than that of **BDPPA** (1.65 eV). These results are useful for development of H_2O_2 -activitable TTA upconversion for in vivo monitoring endogenous H_2O_2 level in living cells or tissues, which can be implemented with microcapsules or microbeads.
- ¹⁵ We thank the NSFC (21273028, 21473020 and 21421005), Program for Changjiang Scholars and Innovative Research Team in University [IRT_13R06] and Ministry of Education (SRFDP-20120041130005) for financial support.

Notes and references

20 State Key Laboratory of Fine Chemicals, School of Chemical Engineering, E-208 West Campus, Dalian University of Technology, Dalian 116024, P. R. China. E-mail: zhaojzh@dlut.edu.cn

† Electronic Supplementary Information (ESI) available: molecular structural characterization and spectral studies. See 25 DOI: 10.1039/b000000x/

- Tanya N. Singh-Rachford and Felix N. Castellano, Coord. Chem. Rev., 2010, 254, 2560.
- 2. J. Zhao, S. Ji and H. Guo, RSC Adv., 2011, 1, 937.
- 3. P. Buchwalter, J. Rosé and P. Braunstein, Chem. Rev., 2015, 115, 395.
- 30 4. P. Ceroni, Chem. Eur., 2011, 17, 9560.
- 5. Y. C. Simon and C. Weder, J. Mater. Chem., 2012, 22, 20817.
- H. Chen, C. Hung, K. Wang, H. Chen, W. Fann, F. Chien, P. Chen, T. Chow, C. Hsu and S. Sun, *Chem. Commun.*, 2009, 27, 4064.
- 7. Y. Cheng, T. Khoury, R. G. C. R. Clady, M. J. Y. Taybebjee, N. J. Ekins-Daukes, M. J. Crossley and T. W. Schmidt, *Phys. Chem.*
- Chem. Phys., 2010, 12, 66.
 8. G. Bergamini, P. Ceroni, P. Fabbrizi and S. Cicchi. Chem. Commun., 2011, 47, 12780.
- 9. J. Peng, X. Jiang, X. Guo, D. Zhao and Y. Ma, *Chem. Commun.*, 2014, **50**, 7828.
- 10. P. Duan, N. Yanai and N. Kimizuka, Chem. Commun., 2014, 50, 13111.
- S. H. C. Askes, N. L. Mora, R. Harkes, R. I. Koning, B. Koster, T. Schmidt, A. Kros and S. Bonnet, *Chem. Commun.*, 2015, DOI: 10.1039/C5CC02197A.
- 12. P. Du, R. Eisenberg, Chem. Sci., 2010, 1, 502.
- A. Turshatov, D. Busko, Y. Aviasevich, T. Miteva, K. Landfester and S. Baluschev, *Chem. Phys. Chem.*, 2012, 13, 3112.
- T. N. Singh-Rachford, A. Haefele, R. Ziessel and F. N. Castellano, J. Am. Chem. Soc., 2008, 130, 16164.
- S. Baluschev, V. Yakutkin, T. Miteva, Y. Avlasevich, S. Chernov, S. Aleshchennov, A. Yasuda, K. Müllen and G. Wegner, *Angew. Chem. Int. Ed.*, 2007 46, 7693.
- A. Monguzzi, R. Tubino, S. Hoseinkhani, M. Campione and F. Meinardi, *Phys. Chem. Chem. Phys.*, 2012, 14, 4322.
- T. F. Schulze, J. Czolk, Y. Cheng, B. Fückel, R. W. MacQueen, T. Khoury, M. J. Crossley, B. Stannowski, K. Lips, U. Lemmer, A. Colsmann and T. W. Schmidt, *J. Phys. Chem. C*, 2012, **116**, 22794.
- A. Nattestad, Y. Y. Cheng, R. W. MacQueen, T. F. Schulze, F. W. Thompson, A. J. Mozer, B. Fückel, T. Khoury, M. J. Crossley, K. Lips, et al, *J. Phys. Chem. Lett.*, 2013, 4, 2073.
- 19. Q. Liu, T. Yang, W. Feng and F. Li, J. Am. Chem. Soc., 2012, 134, 5390.

- 20. Q. Liu, B. Yin, T. Yang, Y. Yang, Z. Shen, P. Yao and F. Li, J. Am. 5 Chem. Soc., 2013, **135**, 5029.
- 21. Z. Jiang, M. Xu, F. Li and Y. Yu, J. Am. Chem. Soc., 2013, 135, 16446.
- (a) X. Cui, J. Zhao, Y. Zhou, J. Ma and Y. Zhao, J. Am. Chem. Soc., 2014, **136**, 9256. (b) J. Ma, X. Cui, F. Wang, X. Wu, J. Zhao, X. Li. J. Org. Chem., 2014, **79**, 10855. (c) K. Xu, J. Zhao, X. Cui, J. Ma. J.
- *Phys. Chem. A*, 2015, **119**, 468.
 23. C. Zhang, J. Zhao, X. cui and X Wu, *J. Org. Chem.*, 2015, **80**, 5674.
- 24. Moto-oka, Nishi-ku, Fukuoka, *Anal. Bioanal. Chem.*, 2015, **30**, 5074.
- 25. E. W. Miller, C. J. Chang, *Curr. Opin. Chem. Biol.*, 2007, **11**, 620.
- ⁷⁵ 26. M. C. Y. Chang, A. Pralle, E. Y. Isacoff and C. J. Chang, *J. Am. Chem. Soc.*, 2004, **126**, 15392.
 - 27. D. Srikun, E. W. Miller, D. W. Domaille and C. J. Chang, J. Am. Chem. Soc., 2008, 130, 4596.
 - 28. N. Soh, O. Sakawaki, K. Makihara, Y. Odo, T. Fukaminato, T. Kawai, M. Irie, T. Imato, *Bioorg. Med. Chem.*, 2005, **13**, 1131.
- 29. K. Xu, B. Tang, H. Huang, G. Yang, Z. Chen, P. Li and L. An, *Chem. Commun.*, 2005, 5974.
- 30. L. Lo and C. Chu, Chem. Commun., 2003, 2728.
- M. Onoda, S. Uchiyama, A. Endo, H. Tokuyama, T. Santa and K.
 Imai, Org. Lett., 2005, 5, 1459.
- 32. X. Cui, J. Zhao, P. Yang and J. Sun, Chem. Commun., 2013, 49, 10221.
- 33. S. H. C. Askes, A. Bahreman and S. Bonnet, Angew. Chem. Int. Ed., 2014, 54, 1029.
- 90 34. Y. Zhao, L. Duan, X. Zhang, D. Zhang, J. Qiao, G. Dong, L. Wang and Y. Qiu, *RSC. Adv.*, 2013, **3**, 21453.
 - 35. Z. Fei, N. Kocher, C. J. M. Dipl.-Chem., H. I. Priv-Doz. And D. Stalke, *Angew. Chem. Int. Ed.*, 2003, 42, 783.
- 36. S. Ji, W. Wu, W. Wu, H. Guo, J. Zhao, *Angew. Chem. Int. Ed.*, 2011, 50, 1626.
- 37. R. R. Islangulov, J. Lott, C. Weder and F. N. Castellano, J. Am. Chem. Soc., 2007, **129**, 12652.
- 38. J. Kang, E. Reichmanis, Angew. Chem. Int. Ed., 2012, 51, 11841.
- 39. J. Kim, J. Kim, J. Am. Chem. Soc., 2012, 134, 17478.
- 100 40. C. Wohnhaas, V. Mailänder, M. Dröge, M. A. Filatov, D. Busko, Y. Avlasevich, S. Baluschev, T. Miteva, K. Landfester and A. Turshatov, *Macromol. Biosci.*, 2013, **13**, 1422.
 - 41. J. Kim, F. Deng, F. N. Castellano, J. Kim, ACS Photonics, 2014, 1, 382.
- 105 42. W. Wu, H. Guo, W. Wu, S. Ji and J. Zhao, J. Org. Chem., 2011, 76, 7056.
 - 43. L. Ma, H. Guo, Q. Li, S. Guo and J. Zhao, *Dalton Trans.*, 2012, **41**, 10680.
- 44. W. Wu, J. Sun, S. Ji, W. Wu, J. Zhao and H. Guo, *Dalton Trans.*, 2011, **40**, 11550.
 - 45. J. R. Lakowicz, *Principles of Fluorescence Spectroscopy*, Kluwer Academic/Plenum Publishers, New York, 2nd edn, 1999.