

# Analytical Methods

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4 1 **Post Derivative TLC Densitometric Stability Indicating Assay for Mesterolone**  
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6 2 **and Quantum Chemical Calculations**  
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3 **ABSTRACT**  
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5 Mesterolone is a synthetic androgenic-anabolic steroid (AAS) used for the cure of infertility and  
6 hypogonadism. Mesterolone cannot be analyzed by a UV detector because of the lack of a  
7 chromophore and this puts limitations on its direct UV analysis. This paper reports development  
8 and validation of a simple and cost-effective post-derivatized TLC-densitometric method using  
9 ceric sulfate as staining reagent for the analysis of mesterolone and its degraded products under  
10 various stress conditions, as recommended in the ICH guidelines. The limit of detection and limit  
11 of quantification for mesterolone in this method were found to be 1.64 and 4.97 ng/spot  
12 respectively. The current study suggests that mesterolone is very prone to photochemical  
13 degradation while quantum chemical calculations carried out as an integral part of this study  
14 supplement the assessed reactivity of mesterolone under applied stress conditions. The developed  
15 stability-indicating TLC-densitometric method can be employed for analysis of mesterolone in  
16 the presence of its degradation products.

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37 **Keywords:** mesterolone, stress degradation, stability-indicating, TLC-densitometry, quantum  
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## 1 Introduction

2 Androgenic-anabolic steroids (AAS) are testosterone derived drugs that stimulate protein  
3 synthesis resulting in accelerated rate of food consumption, increased muscle growth, body mass  
4 and enhanced performance by binding to androgen receptors that maintain male characteristics <sup>1</sup>.  
5 The pharmaceutical market has many formulations available that either contains individual  
6 steroids or their combinations in various dosage forms. In spite of the fact that World Anti-  
7 Doping Agency (WADA), International Olympic Committee (IOC) and other sports authorities  
8 have restricted the use of AASs, their self-administration is continuously practiced by many  
9 individuals, particularly sportsmen, is a desire to enhance athletic performance, aggressiveness,  
10 body strength and shape <sup>2-4</sup>. Mesterolone (1 $\alpha$ -methyl-5 $\alpha$ -androstan-17 $\beta$ -ol-3-one), first  
11 synthesized by Wiechert and patented in 1968 <sup>5</sup>, is generally used for the cure of infertility and  
12 hypogonadism <sup>6,7</sup> and is one of the most utilized androgenic-anabolic steroids (AAS) by athletes  
13 due to its antiestrogenic activity. The analysis of mesterolone, therefore requires much attention  
14 because of its presence on the WADA prohibited substances list.

15 The International Conference on Harmonization (ICH) and Food and Drug Administration  
16 (FDA) require stress degradation analysis of drugs to explain the inherent stability characteristics  
17 of the active component. These guidelines emphasize to explore features possibly affected by  
18 various environmental factors such as light, humidity, oxidation, pH variation and temperature.  
19 Validated stability-indicating analysis methods therefore must be employed for quality  
20 assurance, safety and efficacy of pharmaceutical formulations. During stress degradation studies,  
21 Quantum-chemical calculations can serve as useful tool for prediction of degradation pathways  
22 and identification of degradation prone sites in a molecule. These calculations serve their  
23 purpose equally well when applied to stability-indicating studies of either drug formulations or

1 lead molecules in drug discovery. Furthermore, the determination of HOMO (Highest occupied  
2 molecular orbital) and LUMO (Lowest unoccupied molecular orbital) size can lead to an  
3 estimation of overall reactivity of the molecule using the density of electrons dispersed over  
4 these orbitals<sup>8-15</sup>.

5 Various analytical methods based on gas chromatography-mass spectrometry (GC-MS),  
6 spectrophotometry and matrix-assisted laser desorption/ionization time-of-flight spectrometry  
7 have been previously published for the analysis of mesterolone in different matrices<sup>16-20</sup>  
8 however, to the best of our knowledge no method has so far been reported for the photochemical  
9 degradation studies and stability indicating TLC-densitometric analysis of mesterolone.

10 A number of benefits make TLC-densitometry a method of choice for routine chemical analysis.

11 In comparison to HPLC, TLC-densitometry is cost effective and environment friendly as it  
12 consumes minimum amount of solvents. This technique offers reduced analysis time by being  
13 able to process several samples simultaneously.

14 In addition to our previous contributions to the development of various chromatographic  
15 methods for the analysis of biologically and pharmaceutically important compounds<sup>21-27</sup>, this  
16 study describes a simple stability-indicating TLC-densitometric method for the analysis of  
17 mesterolone in pharmaceutical products as per ICH guidelines<sup>28</sup> and also establishes  
18 mesterolone's reactivity under various stress conditions.

## 19 **Experimental**

### 20 **Standards and chemicals**

21 Mesterolone was purchased from Tokyo Chemical Industry Co., Ltd. Proviron tablets  
22 (containing 25 mg mesterolone / tablet manufactured by Bayer Schering Pharma) were procured  
23 from local pharmaceutical market in Karachi, Pakistan. Sodium hydroxide (NaOH) was acquired

1 from BioM Laboratories (Cerritos, USA). Hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>, 35% v/v) and hydrochloric  
2 acid (HCl) were purchased from Fisher Scientific (UK). Precoated silica gel aluminum sheets  
3 (60F<sub>254</sub>, 20 cm × 20 cm) were ordered from Merck (Germany). Deionized water used throughout  
4 this study was obtained by passing distilled water from Millipore Milli-Q Integral Water  
5 Purification System (Bedford, USA). Others solvent required for sample preparation and analysis  
6 were purchased from Merck, (Germany).

### 7 **Instrumentation and computational analysis**

8 Samples were sprayed as bands of 6mm width on precoated silica gel aluminum plate 60 (F<sub>254</sub>,  
9 (20 cm × 10 cm with 0.20 mm layer thickness) by CAMAG automatic TLC sample applicator  
10 (Linomat 5) using CAMAG 100 μL glass syringe. A constant application rate of 0.1 μL/s was  
11 maintained and two sample bands were separated by a distance of 9.1 mm. Monochromatic  
12 bandwidth was set at 20 nm while each track was scanned thrice with baseline correction. Mobile  
13 phase composition was hexane:acetone (6.5:3.5 v/v) while a total of 10 mL mobile phase was  
14 used per chromatographic analysis. Linear ascending development was carried out in CAMAG  
15 twin trough glass chamber (20 cm×10 cm) in an unsaturated condition. The chromatographic  
16 process was carried out at room temperature (25 ± 2 °C) with relative humidity 43 ± 5% while  
17 the length of run chromatogram was 8 cm. Developed TLC plate was dried for 3-5 minutes using  
18 hot air from a hair dryer and subjected to staining process using CAMAG Chromatogram  
19 Immersion Device. TLC plate immersed in the staining reagent (Ceric sulfate) was heated for 4-5  
20 minutes at 110-120 °C on CAMAG TLC Plate Heater 3. Developed TLC plate was scanned at  
21 λ<sub>max</sub> 466 nm on CAMAG TLC Scanner 3 operating in reflection absorbance mode and controlled  
22 by winCATS software. Slit dimensions were set at 5 mm × 0.45 mm and scanning speed was 10

1 mm s<sup>-1</sup>. Radiation source was a deuterium lamp operating in 190 nm to 400 nm wavelength range and evaluation was carried out *via* peak areas using linear regression analysis.

Quantum chemical calculations were also performed to determine spatial electron distribution on HOMO and LUMO and for initial geometry interpretation of mesterolone. Initial geometry was optimized using Spartan 08 v 1.2.0 (Wavefunction, CA, USA) by means of density functional theory (DFT) method with Becke-3-Le-Yang-Parr (B3LYP) and 6-31G\*\* basis sets on a PC equipped with an Intel® Core™ i3 processor running Microsoft Windows 7 (64-bit edition).

### 8 **Calibration curve of standard mesterolone**

Calibration curve was constructed using six standard solutions prepared by independently weighing mesterolone and making up the individual volumes with methanol. The solutions were stored at 4 °C prior to analysis. Five µL from each standard solution was spotted three times on the TLC plate at various concentrations (200, 400, 600, 800, 1000, and 1200 ng per spot) and subjected to chromatographic analysis. The spotted plate was developed as described in experimental section. The procedure was repeated six times to obtain an average standard calibration curve in the range 200–1200 ng per spot. Residual linearity test was used to verify the linearity of standard calibration curve.

### 17 **Method validation**

ICH guidelines were followed throughout method validation process. Sensitivity of the method was determined with respect to LOD, LOQ and correlation coefficient. Linearity was evaluated by spotting 5 µL from each dilution three times on the TLC plate to give concentration range of 200-1200 ng per spot and subjected to chromatographic analysis. The limit of detection (LOD) and limit of quantification (LOQ) were estimated at 3 and 10 times of the noise level. Moreover, LOD and LOQ were also experimentally confirmed by diluting the known concentrations of

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3 1 mesterolone till the average instrumental responses were approximately 3 and 10 times of the  
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5 2 standard deviation of the responses for six replicates. Method precision was determined by intra-  
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7 3 and inter-day analyses of mesterolone for repeatability and reproducibility. This was achieved by  
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9 4 spotting three different concentrations (300, 500 and 700 ng/spot) on the TLC plate. Repeated  
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11 5 analyses were performed on the same day for the intra-day analysis and on the next day for inter-  
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13 6 day analyses. Results were listed as percent relative standard deviation (% R.S.D.). Robustness  
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15 7 was assessed by deliberately varying different parameters within the range of  $\pm 5\%$  at three  
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17 8 different concentration levels such as using different types of TLC plates e.g., silica-gel glass  
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19 9 plates and aluminum sheets procured from other supplier (Macherey-Nagel, Germany), varying  
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21 10 mobile phase composition, varying time from spotting to chromatography, varying time from  
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23 11 chromatography to derivatization, varying time for derivatization and varying time from  
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25 12 derivatization to scanning. Recovery studies were carried out on pre-analyzed samples for the  
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27 13 assessment of method accuracy by spiking with extra 50, 100 and 150% standard mesterolone.  
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29 14 This procedure was also followed in six replicates.  
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### 36 15 **Forced degradation study of mesterolone**

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38 16 50 mg of mesterolone was dissolved in 50 mL of methanol to achieve  $1 \text{ mg.mL}^{-1}$  solution which  
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40 17 was used for each experiment. Stress degradation studies were performed using parallel sixteen  
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42 18 reaction vessels synthesizer (Smart Start Synthesizer, Chem Speed Ltd., Switzerland).  
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44 19 3 mL of stock solution of mesterolone ( $1 \text{ mg.mL}^{-1}$ ) was mixed with 3 mL of each 0.1N, 1N and  
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46 20 5N NaOH independently for alkaline hydrolysis and the consequential blends were refluxed at  
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48 21  $80^\circ\text{C}$  for two hours in the dark to avoid any possible photodegradation of the compound. For  
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50 22 neutral hydrolysis, 3 mL of methanolic stock solution was mixed with 3 mL of milli Q water and  
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52 23 refluxed for two hours in the dark. For the experiment of acidic hydrolysis, 3 mL of methanolic  
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1 stock solution of mesterolone was mixed with 3 mL of each 0.1 N, 1N and 5N HCl  
2 independently for acidic hydrolysis and the consequential blends were refluxed at 80 °C for two  
3 hours in the dark. For thermal degradation, 3 mL of methanolic stock solution of mesterolone  
4 was refluxed at 80 °C for two hours in the dark. Experiment on the oxidation degradation was  
5 carried out by mixing 3 mL of the stock solution with 3 mL of H<sub>2</sub>O<sub>2</sub> (35% v/v) and refluxed at 80  
6 °C for two hours in dark.

7 Photochemical stability of mesterolone was studied by exposure of powdered mesterolone to  
8 direct sunlight for seven days at 30 ± 2 °C from 09:00 am to 05:00 pm. 1 mg of sunlight exposed  
9 mesterolone was dissolved in 2 mL of methanol to give stock solution for the treated standard.  
10 Powdered mesterolone was stored in oven for 4 hrs at 90 °C for dry heat-induced degradation.  
11 Stock solution for dry heat exposed mesterolone was prepared by dissolving 1 mg of treated  
12 standard in 2 mL of methanol. 3 mL methanolic stock solution of mesterolone was mixed with 3  
13 mL of H<sub>2</sub>O<sub>2</sub> (35% v/v) kept for 24 hours for hydrogen peroxide induced oxidation at room  
14 temperature. 2 µL (1000 ng/spot) each of alkali treated solutions, neutral hydrolysis solutions,  
15 acid treated solutions, oxidation degraded solution, dry heat-treated solution, light exposed  
16 solution, hydrogen peroxide induced oxidized mesterolone solution and wet heated solutions  
17 were applied on TLC plates in triplicate and the densitograms were developed as described in  
18 experimental section.

#### 19 **Analysis of marketed drug formulation**

20 A total of 20 mesterolone tablets (label claim: 25 mg mesterolone per tablet) were accurately  
21 weighed and powdered with the help of mortar and pestle for 3-5 minutes. 10 mg of the  
22 homogenized powder was accurately weight and transferred to a volumetric flask containing 10  
23 mL methanol. The mixture was sonicated for 10 minutes to ensure complete extraction of

1 mesterolone from the tablet matrix. The resulting solution was filtered and 2 mL of it were  
diluted to 10 mL with methanol. 5  $\mu\text{L}$  (1000 ng spot<sup>-1</sup>) of the diluted solution was applied on  
TLC sheet followed by development, staining and scanning.

#### **ESI-QqTOF-MS/MS analysis of photo-degraded products**

The standard mesterolone and its photo-degraded products were collected from the spots  
separated from the TLC plate and dissolved in methanol. Working dilution was prepared in a  
mixture of 1:1 acetonitrile and water, containing 0.1% formic acid. Analysis was performed by  
electrospray ionization (ESI) and collision-induced dissociation (CID), positive ion mode, on a  
tandem mass spectrometer (QSTAR XL mass spectrometer Applied Biosystems/MDS Sciex,  
Darmstadt, Germany) coupled with Agilent 1100 HPLC system. High purity nitrogen gas was  
used as the curtain gas and collision gas (generated using Peak Scientific nitrogen generator).  
The ESI interface conditions were as follows: ion spray capillary voltage of 5500 V, curtain gas  
flow rate 20 L min<sup>-1</sup>, nebulizer gas flow rate 30 L min<sup>-1</sup>, DP1 60 V, DP2 10 V, and focusing  
potential of 265 V. The collision energy was swept from 20 to 45 eV for MS/MS experiment.  
Samples were introduced into the mass spectrometer using a Harvard syringe pump (Holliston,  
MA) at a flow rate of 5  $\mu\text{L min}^{-1}$ .

## **Results and discussion**

### **Optimization of method**

The purpose of chromatography was to develop a stability-indicating method for mesterolone.  
Solutions containing standard mesterolone and its degraded products were spotted on TLC plates  
and developed in various mobile phases. Different solvents system were tried to separate  
mesterolone from its degraded products in good resolution. Peak widths and  $R_f$  values of  
mesterolone in different solvent systems are listed in Table 1. Best results were obtained when

1 the mobile phase was a 6.5:3.5 v/v mixture of hexane and acetone in unsaturated condition  
2 (Figure. 1A). Peak for mesterolone appeared at  $R_f = 0.53 \pm 0.03$  while peaks for its  
3 photodegradation products (D1 and D2) appeared at  $R_f = 0.46$  and  $R_f = 0.67$ , respectively.  
4 Staining reagents like phosphomolybdic acid (PMA), ceric sulfate, ceric ammonium sulfate and  
5 vanillin were tried for derivatization while best results were obtained with ceric sulfate staining.  
6 Spots stained with ceric sulfate were brighter and sharper than with other derivatization agents.  
7 Six different concentrations of mesterolone (200, 400, 600, 800, 1000 and 1200 ng/spot),  
8 prepared in methanol, were simultaneously spotted in triplicate on aluminum TLC plates and  
9 developed as described in experimental section. Peak area and concentration were subjected to  
10 least squares linear regression analysis to calculate the calibration equation and correlation  
11 coefficient. The linear regression data for the calibration curves ( $n = 6$ ) showed linearity ( $r =$   
12  $0.9967 \pm 0.002$ ) over the range of 200-1200 ng /spot. In the residual linearity test, a random  
13 pattern of residuals versus applied standard concentrations showed linear model for standard  
14 calibration curve.

### 15 **Method validation**

16 The repeatability of the sample analysis was assessed and tabulated as percentage relative  
17 standard deviation (R.S.D.) between two analyses and was found to be less than 1% when  
18 performed by two different analysts. Similarly, intra- and inter-day analysis of mesterolone was  
19 found to be reproducible as the R.S.D. was less than 1% (Supplementary file 1). For the  
20 assessment of robustness of our developed method, % R.S.D. was calculated for three different  
21 criteria as the concentration was varied (300, 500 and 700 ng). Average %R.S.D. was 0.62 for  
22 change in solvent composition, 0.48 for varying TLC plates, 0.62 for varying time from spotting  
23 to chromatography, 0.49 for varying time duration for chromatography to staining, and 0.48 for

1 duration for staining and 0.6 for varying time duration from staining to scanning (Supplementary  
2 file 2). RSD % was found to be less than 1.5% for every criterion which shows that the  
3 developed method is reasonable robust. This method was found to be sensitive as LOD for  
4 standard mesterolone at signal/noise ratio of 3:1 was found to be 1.64 ng per spot while LOQ at  
5 signal/noise ratio 10:1 was found to be 4.97 ng/spot. To check the peak purity of mesterolone the  
6 spectra were compared at peak start, peak apex and peak end positions of the spot. Good  
7 correlation,  $r^2$  (start to middle) = 0.9997 and  $r^2$  (middle to end) = 0.9996 were obtained between  
8 standard and sample spectra of mesterolone. The recovery of the method was found to be 97.5–  
9 99.2% after spiking with 50, 100 and 150% of additional standard mesterolone (Supplementary  
10 file 3). All of the validation parameters have been summarized in Table 2.

### 11 **Analysis of pharmaceutical product**

12 The chromatogram of mesterolone extracted from tablets afforded a compact spot at  $R_f = 0.53 \pm$   
13  $0.03$  while no interference was observed from the excipients generally added in tablet  
14 formulations (Figure. 1B). Mesterolone content of the tablet was found to be  $24.847 \pm 0.304$  mg  
15 (99.39 %) with an R.S.D. of 1.22%. This low % R.S.D. value indicates the usefulness of this  
16 method for usual analysis of mesterolone in pharmaceutical products.

### 17 **Stability indicating property of mesterolone**

18 The chromatogram for light exposed sample of mesterolone showed additional peaks for two  
19 degradation products at  $R_f = 0.46$  (D1) and  $R_f = 0.67$  (D2), respectively. The spots for  
20 degradation products were well resolved from the standard mesterolone spot as shown in Figure.  
21 1C. On the other hand, samples degraded under acid, base, heat and oxidation conditions showed  
22 no additional peaks. Table 3 shows the  $R_f$  values of degradation products and the percent  
23 degradation of mesterolone under the mentioned stress conditions.

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3 1 Current stability indicating assay of mesterolone was also aided by computational calculations of  
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5 2 HOMO and LUMO. These calculations led to the explanation of observed degradation of  
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7 3 mesterolone under different conditions. Mesterolone was found to be quite stable under thermal  
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9 4 degradation conditions and this can be explained by its calculated values of enthalpy (-  
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11 5 2429219.48 kJ/mol) and free energy (-2429399.48 kJ/mol). These values suggest a high  
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13 6 thermodynamic stability for mesterolone and hence it does not degrade even under accelerated  
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15 7 thermal conditions. It was seen from the calculation results that the HOMO is spread from the  
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17 8 methyl group of C-1 to C-4 suggesting the presence atomic centers vulnerable to electrophilic  
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19 9 attack in this region. In principle, acidic conditions should result in protonation in this region  
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21 10 which should not lead to any substantial degradation of the molecule. This assumption was  
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23 11 supported by our experimental findings where only 1 – 4 % degradation under acidic conditions  
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25 12 was observed. The LUMO orbital was found to be spread in the region from C-2 to C-4 which is  
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27 13 the enolizable region. However, the mesterolone molecule owing to its high thermodynamic  
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29 14 stability does not undergo aldol type reactions. This was again supported by experimental data  
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31 15 where only 1 – 5 % degradation under alkaline conditions was observed. If the findings from the  
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33 16 HOMO and LUMO calculations are combined (Figure. 2), the observation of two  
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35 17 photodegradation products (D1 and D2) can be explained. The formation of D1 is probably due  
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37 18 to cleavage of the bond between C-2 and C-3. Photo-degraded product D2 was found to be a  
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39 19 dimer of mesterolone which is probably dimerized by an enolate formation at C-2 and the attack  
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41 20 of enolate on the C-3 of other molecule. The thermodynamic stability of the molecule may have  
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43 21 a role in these findings as UV light can provide energy required for the molecule to undergo such  
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3 1 The elemental compositions of both photo-degraded products were determined by precise mass  
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6 2 measurements from the ESI-QqTOF-MS. ESI-QqTOF-MS (positive mode) scan of mesterolone  
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8 3 and its degraded products showed the peaks of  $[M+H]^+$  at  $m/z$  305.2497 (MS), 337.2743 (D1)  
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10 4 and 591.4780 (D2) corresponding to the molecular formulae  $C_{20}H_{33}O_2^+$  (calc. 305.2475),  
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12 5  $C_{21}H_{37}O_3^+$  (calc. 337.2737) and  $C_{40}H_{63}O_3^+$  (calc. 591.4771), respectively. MS/MS spectra of  
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14 6 standard mesterolone and its photo-degraded products are provided in Figure. 3. Elemental  
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16 7 compositions of both photo-degraded products are listed in Table 4.

## 8 **Conclusion**

9 The current study establishes the intrinsic stability of mesterolone under various stress  
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11 10 conditions. Of all the stress conditions investigated, mesterolone was found to be most  
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13 11 susceptible to photochemical degradation. Two photo degraded products were observed with  
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15 12 significantly different  $R_f$  values. The developed stability-indicating TLC densitometric method  
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17 13 was found to be rapid, accurate and precise and can be efficiently used for the direct  
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19 14 identification and quantification of mesterolone in its pharmaceutical formulations. The  
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21 15 computational calculations also provided a clear understanding of mesterolone reactivity under  
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23 16 photochemical conditions. In addition, the characterization of photo-degraded products will be  
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25 17 useful for the better understanding of degradation pathways.  
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## 19 **Competing interests**

20 The authors declare that they have no competing interests.

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(Project Reference No. 5-9/PAS/54).

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3 **1 Supplementary Material**  
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6 2 The following supplementary files are available with the online version of this paper in pdf  
7  
8 3 format.

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10 4 Supplementary file 1. Precision and accuracy for quality control standard of mesterolone

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12 5 Supplementary file 2. Robustness testing (n = 6)

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14 6 Supplementary file 3. Recovery studies (n = 6)  
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1 **Figure and Table Legends**

2 **Figure. 1.** TLC chromatogram of standard mesterolone (1000 ng/ spot),  $R_f = 0.53 \pm 0.03$  (a),  
3 TLC Chromatogram of pharmaceutical product Proviron (b), chromatogram of photo degraded  
4 mesterolone (1000 ng/ spot), degradant (D1) has  $R_f = 0.46$  and degradant (D2) has  $R_f = 0.67$  (c):  
5 mobile phase; hexane: acetone (6.5:3.5, v/v)

6 **Figure. 2.** Optimized molecule geometry (a), LUMO (b) and HOMO (c) orbitals of mesterolone

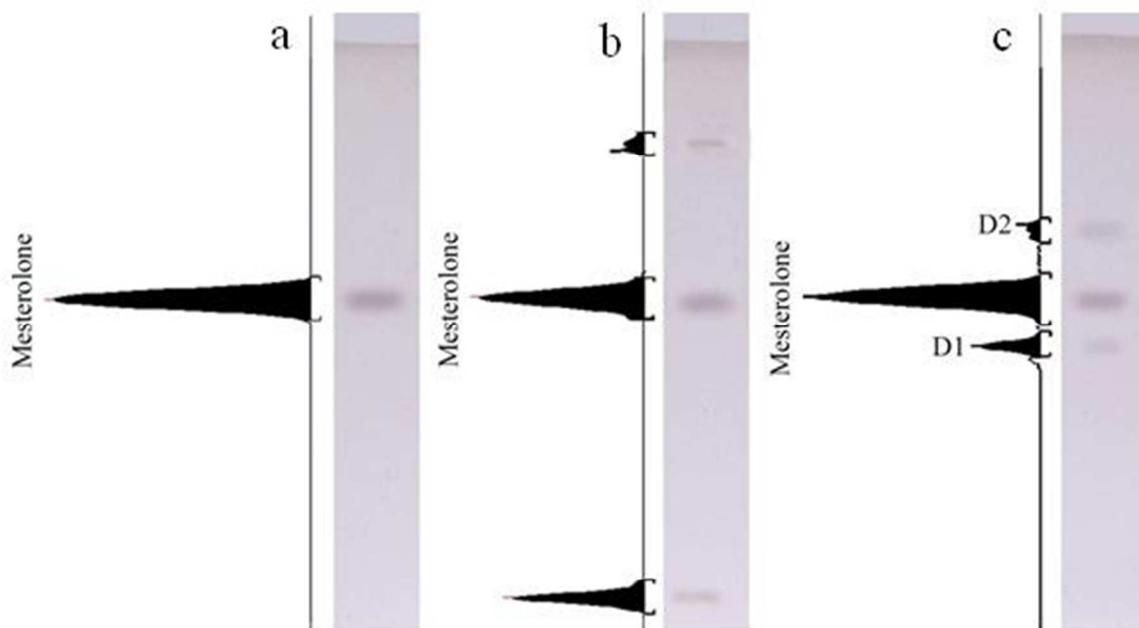
7 **Figure. 3.** Product ion spectra of mesterolone (A) and its photo-degraded products D1 (B), D2,  
8 at collision energy of 25 eV.

9 **Table 1.**  $R_f$  values and peak widths of mesterolone in different mobile phases

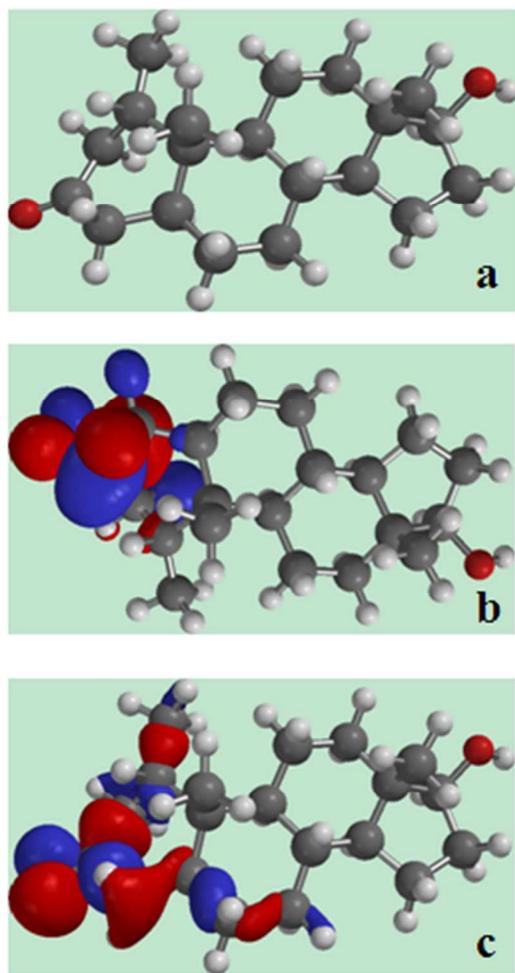
10 **Table 2.** Summary of validation parameters

11 **Table 3.** Summary of stress degradation studies of mesterolone

12 **Table 4.** Elemental composition of the Photo-degraded products of mesterolone.  
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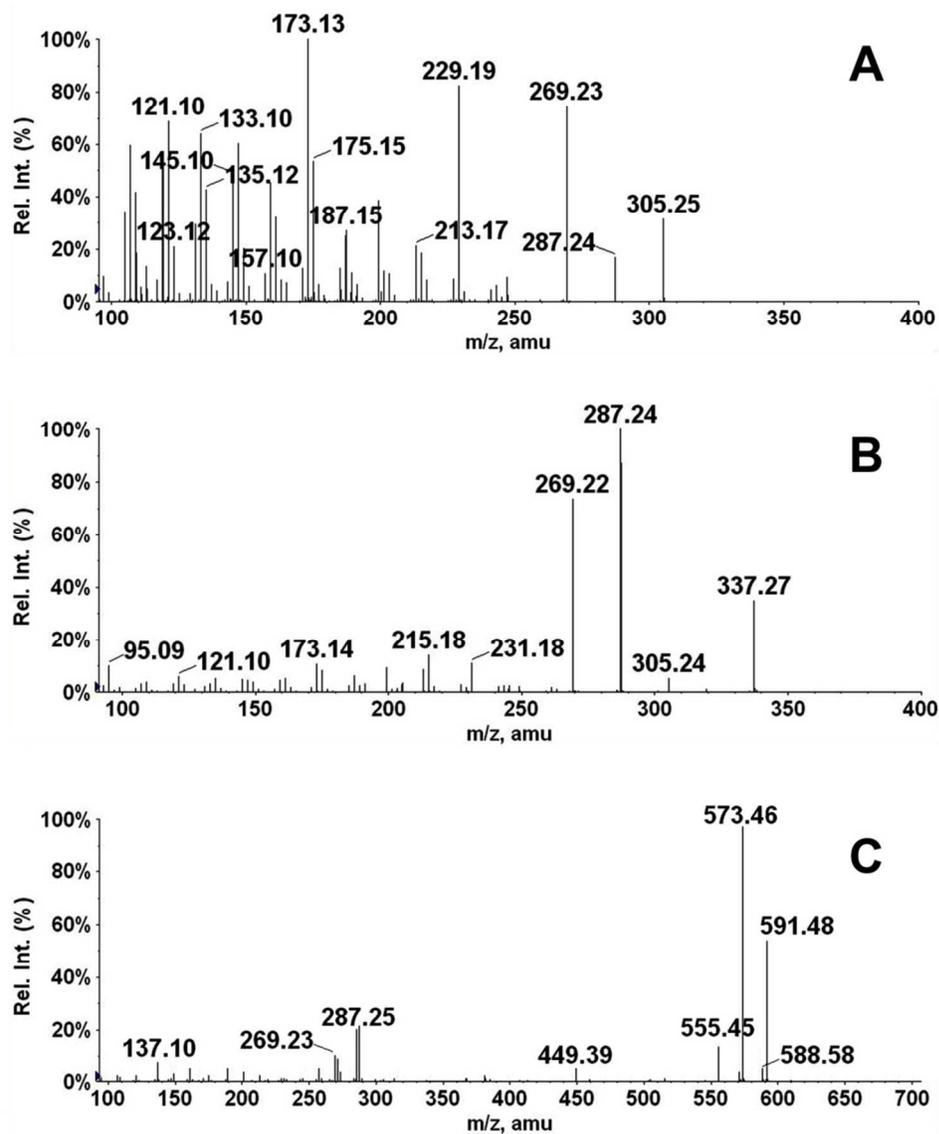
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27 **Figure 1.** TLC chromatogram of standard mesterolone (1000 ng/ spot),  $R_f = 0.53 \pm 0.03$  (a),  
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29 TLC Chromatogram of pharmaceutical product Proviron (b), chromatogram of photo degraded  
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2 **Figure. 2.** Optimized molecule geometry (a), LUMO (b) and HOMO (c) orbitals of mesterolone

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**Figure 3.** Product ion spectra of mesterolone (A) and its photo-degraded products D1 (B), D2, at collision energy of 25 eV.

**Table 1.**  $R_f$  values and peak widths of mesterolone in different mobile phases

S. No	Solvent Composition	Proportion(v/v)	Retention Factor $R_f$	Peak Width $\Delta R_f$ (cm)
1	Hexane-ethyl acetate	8:2	0.19	0.08
2	Chloroform-methanol	9.4:0.6	0.21	0.11
3	Dichloromethane-methanol	9.2:0.8	0.14	0.07
4	Hexane-acetone	6:4	0.57	0.07
5	Hexane-acetone	6.5:3.5	0.53	0.06
6	Hexane-acetone	7:3	0.45	0.07

**Table 2.** Summary of validation parameters

Parameter	Data of standard mesterolone (at $\lambda_{\max}$ 466 nm)
Linearity range	200-1200 ng/spot
Correlation coefficient	0.9967 $\pm$ 0.002
Limit of detection (LOD)	1.64 ng/spot
Limit of quantification (LOQ)	4.97 ng/spot
Y = mx + c	Y = 7.28X + 525.49
Slope $\pm$ SD	7.28 $\pm$ 0.32
Intercept $\pm$ SD	525.49 $\pm$ 3.528
Intra-day analysis (n=3), % R.S.D.	0.367
Inter-day analysis (n=3), % R.S.D.	0533
Robustness	Robust
Specificity	Specific

**Table 3.** Summary of stress degradation studies of mesterolone

Degradation conditions	% Recovery	% Degradation	$R_f$ of degraded products
<b>Acidic hydrolysis<sup>a</sup></b>			
0.1N HCl	98.27	1.73	Not detected
1N HCl	97.35	2.65	Not detected
5N HCl	95.47	4.53	Not detected
<b>Basic hydrolysis<sup>a</sup></b>			
0.1N NaOH	98.32	1.68	Not detected
1N NaOH	96.59	3.41	Not detected
5N NaOH	94.61	5.39	Not detected
<b>Neutral hydrolysis<sup>a</sup></b>			
Wet heating <sup>a</sup>	98.53	1.47	Not detected
Dry heating	98.46	1.54	Not detected
<b>Oxidation<sup>a</sup></b>			
5%v/v H <sub>2</sub> O <sub>2</sub>	97.62	2.38	Not detected
35%v/v H <sub>2</sub> O <sub>2</sub>	98.69	1.31	Not detected
Oxidation at room temp	95.43	4.57	Not detected
<b>Photostability-daylight</b>	98.83	1.17	Not detected
	82.57	17.43	0.46, 0.67

<sup>a</sup>Reflux in parallel synthesizer for two hours at 80 °C

**Table 4.** Elemental composition of the photo-degraded products of mesterolone.

Product ID	Proposed formula	Observed mass	Calculated mass	Error (ppm)
MS	$C_{20}H_{33}O_2^+$	305.2497	305.2475	7.1842
D1	$C_{21}H_{37}O_3^+$	337.2743	337.2737	1.7142
D2	$C_{40}H_{63}O_3^+$	591.4780	591.4771	1.3984

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