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Solid lipid nanoparticles (SLNs): Delivery vehicles for food bioactives

N. P. Aditya, Sanghoon Ko*

Department of Food Science and Technology, Sejong University, Seoul, Korea

* Corresponding author: Sanghoon Ko

Tel.: +82 (2) 3408 3260; fax: +82 (2) 3408 4319

E-mail address: sanghoonko@sejong.ac.kr (Sanghoon Ko)

Institutes:

Department of Food Science and Technology

Sejong University

98 Gunja-dong, Gwangjin-gu

Seoul 143-747, Korea

Abstract

Bioactives which are isolated from different sources like plant, animal etc. are known to be ideal candidates to treat and prevent chronic health problems such as obesity, hypertension, cardiovascular diseases, cancer etc. Unfortunately, due to change in life style and food habit, humans are consuming healthy bioactives less than recommended. Hence there is an increasing demand from consumers for food products which are fortified with these bioactives. However, addition of these healthy bioactives into food products for fortification is a challenging job. The main cause is their susceptibility to a complex matrix present in the food product and gastro intestinal tract (GIT) such as pH, temperature, enzymes, etc. Though, colossal effort has been put by researchers to successfully translate drug delivery technology for bioactives delivery to protect these susceptible bioactives during production, storage and consumption, successes is limited. Due to their near impeccable suitability for the delivery of bioactives in terms of toxicity, scalability, acceptability, solid lipid nanoparticles (SLNs) are drawing attention from food technologists. In this review, effort has been made present an overview about expectations and suitability of SLNs for delivery of bioactives, selection of the ingredients and their importance in achieving those expectations and industrially feasible methodology for production of SLNs.

Keywords: bioactive, solid lipid nanoparticle, drug delivery, stability, surfactant

1. Introduction

As quoted by Hippocrates, the father of modern medicine “Let food be thy medicine and medicine be thy food”, in the last few years there has been a growing realization of the pivotal link between diet, health and well-being.¹ Thus consumer prefers food products which are tasty and healthy. Further, consumer expects food to provide some additional physiological functions for promoting general body development and disease prevention. These attributes required to the food are provided by bioactive molecules which possess biological activity in addition to their nutritional value.²

Bioactive molecules occur naturally in plant and animal products, normally at very low concentrations. Hence fortification is highly desired to increase the concentration of specific and desired bioactives in the food products.³ Unfortunately, direct addition of these healthy bioactives to food products results in the unwanted change in the organoleptic properties of food products (e.g. bad odor and taste, undesired mouthfeel) and also destabilizes the product by forming aggregates and sediments. Even then, due to their pharmacokinetic mismatch (lessened stability in gastro intestinal tract (GIT) and absorption variability) a consumer doesn't get full benefits after consumption of food products without their desired organoleptic attributes.^{1,4}

The use of encapsulation technology which is originally used in pharmaceutical industries to increase the stability and bioavailability of drug molecules is also has been in use in food sector for several years for similar purposes with required modification.^{1,5,6} Encapsulation can be defined as “Processes of entrapping sensitive materials into the matrix of the carrier material which forms the protecting wall and are generally resistant to the sensitive environment for which entrapped materials are not”.^{7,8} Encapsulation processes may result in the formation of carriers from micro- to nanometer size depending on various factors which are used for their

fabrication (e.g. energy input, presence of surface acting materials, core and wall material physicochemical properties etc.).⁹ In this regard, since last two decades, food scientists and industries are thriving hard to develop novel delivery systems to carry food bioactives which can be used to fortify food products with desired bioactives or combination of bioactive molecules. Problems which are associated with food bioactives are shown in Figure 1.

The main objectives of this review is to provide insight into the expectations from bioactive delivery systems and suitability of solid lipid nanoparticles (SLNs) to fulfill those expectations and to describe how lipid nanostructures can be engineered to meet those expectations.

2. Expectations from bioactives encapsulation technology

Ideally the developed encapsulation system to deliver bioactives is expected to possess the following properties:

- A. It is suitable to entrap bioactives with different physicochemical properties (e.g. melting point, solubility, stability, origin etc.) in maximum quantity.
- B. It has capacity to deliver the entrapped cargos to the right place in right time and right concentration.
- C. It can protect the entrapped bioactives both from adverse environment (temperature, oxygen, light etc.) in the complex matrix of the food products and during consumption until entrapped bioactives reaches the desired site of action (pH, ionic strength, enzymes and microbes etc.).
- D. It should prove to be affordable when cost to benefit ratio is considered.

- E. It should be versatile enough to allow its use in different types of food products (liquids, powders, gels etc.).
- F. It should not interfere with desired qualities of the final food products (taste, odor, appearance, viscosity etc.).
- G. It should be easy to prepare but stable in complex processing and storage conditions to use in processed food products.
- H. It should be easily scalable
- I. It should be stable upon sterilization using methods which are generally used to sterilize food products (thermal treatments, UV sterilization etc.).
- J. Raw materials for fabrication should be easily available in abundance.
- K. It should be stable in various food formats (e.g. liquid, solid, gel etc.). Thus it needs to be stable in aqueous dispersions and also upon lyophilization and spray drying.

In order to obtain above mentioned characteristics from the designed delivery systems, the major determining factors are

- A. Selection of carrier materials and
- B. Production method.

In this review, each of these aspects will be discussed in detail.

3. Selection of carrier materials

Variety of materials is available to use as carrier materials to obtain the desired characteristics as mentioned above. However limitations among them are certified as “generally recognized as safe” (GRAS) which can be added to food products to obtain the desired functional attributes

like controlled and targeted delivery, taste and odor masking and to protect from degradation etc.^{1, 10} Further it is noteworthy that due to preference for fully natural materials as carrier materials among the consumers and producers, several GRAS approved materials which are used in drug delivery are not preferred for use in food sector to deliver bioactives. In addition, presence of non food particles in food products in large quantity also compromises nutritional value of products.¹¹ Hence, in food sector, delivery systems which are comprised of natural food additives are preferred.

Important factors which need to be considered before selecting the carrier material are,

- A. Interaction between functional ingredient and carrier material (carrier and bioactive interaction)
- B. Intended use (products in which it needs to be added and processing and storage condition of that particular products etc.),
- C. Effect of the carrier material on the consistency and the structure of the product,
- D. Presence of other co-excipients presence in the system and their interaction behavior with the carrier materials,
- E. Biological barriers which should be overcome in order to obtain desired biological activity,
- F. Toxicity upon chronic ingestion.

3.1. Lipids and lipid based delivery systems

Lipids are the group of natural molecules which include mono-, di- and triglycerides, fats, waxes, sterols, phospholipids etc.¹² These lipids are important constituents for existence of life. Starting from the fetal growth they are required in all stages of life to perform specific activity or to

maintain general health of the organism. In living organism they provide energy, act as reserve source of energy and aid the absorption of fat soluble vitamins, nutraceuticals and drug molecules.¹³ Lipids play important role as the components which give desired qualities like texture, aroma, color, flavor, satiety, mouthfeel and rheological properties to food products.¹⁴

Thus due to their high acceptability, non toxicity and suitability to use in food products, lipid based delivery systems are rapidly gaining interest among the food scientists for application in food systems to deliver bioactives.^{9, 10} When referring to lipid based delivery systems, it includes liposomes, solid lipid nanoparticles (SLN), nanostructured lipid carriers (NLC), emulsions, micelles etc.¹⁵ In this article, we will provide an overview about SLN which is the most rapidly developing carrier systems among lipid based delivery systems for food application due to their inherent characteristic i.e. stability in complex systems and ability to withhold the bioactives within the core of the nanoparticles which suit food application. These characteristics are less seen in other lipid based delivery systems like liposomes and emulsions (Figure 2). Due to high cost and reports of toxicity, synthetic polymeric nanoparticles are less preferred in food delivery compared to drug delivery systems.¹⁶

3.1.1. Solid lipid nanoparticles (SLNs)

These SLNs which are first introduced in the year 1991 as drug carriers can be fabricated in submicron size range from 50 to 500 nm using high melting point lipids (i.e. lipids solid at room temperature and also at body temperature) and stabilized by surfactant(s) which are biodegradable, biocompatible and non toxic.^{17, 18} SLN based on para-acyl-calix(4)arenes have been prepared and studied for the purpose of drug delivery.¹⁹ Though these nanostructures can be used to entrap and deliver both hydrophilic and hydrophobic molecules, they are preferred for

the delivery of hydrophobic molecules. The main reason is the high affinity of hydrophobic molecules to the hydrophobic matrix formed in the SLN compared to the low affinity of the hydrophilic molecules to the hydrophobic matrix.¹¹ Still in the past, there are reports where these SLNs were used to deliver nutraceuticals, proteins, and drugs.^{11, 20}

3.1.2. The main features of SLNs which promote their food application

3.1.2.1. Production

- A. SLNs can be fabricated using methods like high pressure homogenization (HPH) and Microfluidization which can be easily scalable for industrial production. Several lipid based nanoparticle formulations are in the market for drug and cosmeceutical delivery.
- B. SLNs can be fabricated without using organic solvents which otherwise are not acceptable to use in food products.
- C. SLNs can be sterilized using techniques which are used to sterilize food products (heat sterilization, UV sterilization etc.).
- D. SLNs can be obtained both in the liquid and solid forms. Hence they are easy to use both in solid and liquid food products.

3.1.2.2. Acceptability

1. Lipids, which are used to fabricate SLNs are non toxic and biocompatible. Several food grade materials are available (e.g. tristearin is FDA approved food additive [21CFR172.811]).
2. Raw materials used in the production of SLNs such as lipids, surfactants and co-surfactants are easily available in large scale with preferred quality.

3. Food grade surfactants like polyoxyethylene sorbitan monooleate (e.g. Tween 80), proteins etc., can be used to fabricate SLNs.
4. In order to be best suited for oral delivery, this is the preferred route of administration for nutraceuticals.

3.1.2.3. Biological activity

1. In order to be best suited for oral delivery, this is the preferred route of administration for nutraceuticals.
2. SLN provide chemical protection to the encapsulated nutraceuticals against chemical degradation in the product (pH, salt, temperature etc.) during storage.
3. SLN protects susceptible nutraceuticals against degradation in GIT.
4. SLN provide chemical protection to the encapsulated nutraceuticals against chemical degradation in the product (pH, salt, temperature etc.) during storage.
5. SLN enhances bioaccessibility of nutraceuticals in the intestine by forming micelles.
6. Rate and site of nutraceutical release can be controlled by varying SLN size and composition (surfactant, co-surfactant etc.).

In the next part of this article, an important aspect which needs to be considered before fabricating SLNs to obtain optimal benefit such as various industrially feasible methods for the production, advantages and disadvantages with regarding their application in food products are discussed in detail.

3.2. Thing to be considered in the formulation of SLNs for bioactives delivery

Excipients such as lipids, surfactants, co-surfactants, and cryoprotectants etc. which are used to fabricate SLNs determine their fate with regarding to their ability for encapsulation and loading efficiency, targetability, stability etc. Hence selections of proper and compatible excipients are highly desired. The common excipients which are generally used to fabricate SLNs are listed in Table 1. Important factors which need to be reviewed carefully before designing delivery system are as follows.

3.2.1. Selection of lipids

Various lipids with different physicochemical properties (melting point, composition, chain length etc.) with GRAS status which can be readily incorporated into food products are available for fabrication of SLNs. Solid lipids which are used to fabricate SLNs including triglycerides, fatty acids, waxes, steroids, partial glycerides, hard fats etc.²¹ Various categories of lipids have their own advantages and disadvantages. Hence it is important to know the characteristics such as miscibility, solubility of bioactive in the lipid, physical and chemical structure of the lipid and bioactive, polymorphic state of lipid etc., before selecting lipid for formulation development.

Important factors needs to be considered before selection of lipids are as follows.

3.2.1.1. Loading and entrapment efficiency

To reduce the concentration of carrier material required to fortify the product with required amount of bioactives which otherwise may interfere with original product quality, it is highly desired to select the carrier material with highest loading efficiency for the desired bioactives. In this regard, it is highly warranted to study the solubility of chosen bioactives in the various lipids with different physicochemical characteristics.²² Although this methodology is quite often used

before designing drug delivery systems, mention of this kind of study in food bioactives delivery is rare.²³

In general lipophilic nature of the glycerides (e.g. trimyristin, C=14; tripalmitin, C=16; tristearin, C=18) increases with the increase in the hydrocarbon chain length. This increases solubility and loading efficiency of hydrophobic bioactives within the SLNs. Further, lipids with different hydrocarbon chain lengths which are mixtures of mono-, di-, and triglycerides (e.g. glycerylbehenate and glycerylmonostearate, GMS) form nanoparticles with many lattice defects. These defects provide the accommodation for the guest molecules (bioactives) to stay inside the nanoparticle matrix. In one of our studies, maximum curcumin loading and entrapment efficiency was observed in GMS which is composed of mono-, di-, and triglycerides compared to trimyristin (TM) and tristearin (TS) which are composed of >95 % triglycerides.²⁴ Recently, we have shown that, in addition to hydrophobicity of lipid and bioactive molecules, molecular weight of the bioactives also plays important role in determining the loading efficiency. Loading efficiency decreases with increase in molecular weight of the bioactives. In case of curcumin and genistein co-loading, though the curcumin ($\log P = 3.1$) is more hydrophobic than genistein ($\log P = 2.9$), genistein loading efficiency was significantly more than curcumin.¹³ Another important factor which determines the loading efficiency of lipid nanoparticles is recrystallization. If complete recrystallization is not imposed by cooling below lipid critical recrystallization temperature, soon after the formation of SLN during fabrication, instead of solid matrix liquid droplet resembling the emulsion is formed. In this metastable super cooled condition, these nanoparticles lose the advantage of having solid matrix and decrease their loading capacity and capacity to retain the bioactives within the lipid matrix for longer period (Figure 2).^{5, 24, 25}

Further, for delivery of hydrophilic molecules, instead of SLNs, lipid drug conjugates (complexation of bioactives with lipid molecules) are fabricated first and later they were converted into nanoparticles. This method decreases the requirement of carrier materials.²⁶ Though this technology is used in pharmaceutical industry to deliver various hydrophilic drug molecules, its application in food industry is still not explored. In future it will be worthy to study the lipid bioactive conjugate using bioactives like catechin and rutin which are less soluble in lipid.

3.2.1.2. Stability

Stability is important for successful incorporation of fabricated SLNs into food products. Due to complex nature of food matrix (temperature, pH, ionic strength etc.) and also sterilization methods used to store foods make these nanoparticles vulnerable for degradation by aggregation, coalescence, creaming, and sedimentation. Hence extra care needs to be taken to fabricate SLNs for food application in comparison to pharmaceutical application. On the other hand, shelf life of food products is shorter compared to pharmaceutical products. Hence stability of the SLNs for a few months is sufficient in most of the cases whereas it is expected to be at least two years in SLNs fabricated for drug delivery.

Crystallinity of the lipid plays an important role in determining the stability of the nanoparticles. Fabrication of SLNs using lipids without lattice defects (e.g. monoacid triglycerides like tripalmitate, bee wax, cetylpalmitate) results in the bioactive expulsion from the nanoparticle matrix due to polymorphic transition of unstable α subcells into stable β subcells.²⁷

²⁸ Conversion of α subcell into β subcells abrogates the space located in the matrix of the SLNs which provide accommodation for bioactives. Said so, it is very important to balance the

composition of mono-, di-, and triglycerides. Earlier studies have shown that, among the glycerides, tribehenin which composes 15% of monoglycerides shows better stability compared to glycerylmonostearate (~50% monoglycerides) and tripalmitate (<5% monoglycerides). Monoglycerides possess the surfactant properties and hence stabilizes the SLNs up to certain extent. However above certain concentration, they destabilize the nanostructures.²⁸ Even though lattice defects could be slightly increased by mixing two lipids, trivial increase in loading efficiency and stability was observed.²⁹

Another important factor which determines the stability of SLN is their crystallization rate and temperature. If proper care is not taken during fabrication to induce controlled crystallization in time, then formed nanoparticles become vulnerable for degradation. The uncontrolled crystallization leads to conversion of spherical structures into platelet like structures which lacks the controlled release property and stability.¹⁶ Jenning and co-authors reported the variation in the formation of α and β crystals upon changing the cooling temperature. Fast cooling (10 °C/ min) resulted in the formation of predominant α crystals whereas decreasing the cooling rate to 2 °C/ min resulted in the formation of β crystals.³⁰ Similar results were also published by other research groups.³¹

In this direction, recently we fabricated SLN stabilized with non ionic surfactant Tween 80 and stability of these SLNs (aqueous stability, dispersability, flow behavior, size etc.) was studied under various environmental conditions which may exist in food products. Presence of electrolytes (NaCl) resulted in increased viscosity of the SLN suspension. Further increasing NaCl concentration resulted in conversion of Newtonian flow behavior to non-Newtonian flow behavior. Since most of the food products like beverages contain electrolytes in the range of 20 mM, proper selection of surfactants are warranted to avoid the aggregation and gelation of SLNs

in such conditions.³² Several other studies also reported similar effect of ionic strength and pH on stability of SLNs.³³

With regarding to chemical stability of lipid, lipids with longer hydrocarbon chain length (tristearin) are known to have highest stability (~ 4% degradation upon storage). Further, in most cases degradation of lipid was within ~10%.³⁴ Though, in food sector stability up to 2 years is not expected, these lipid degradation studies were conducted only with regarding to their stability in pharmaceutical formulations. Presence of complex food matrix may alter the stability profile of lipids. Unfortunately, as per our knowledge, no study has been conducted regarding stability of various solid lipids in the complex food structures (various temperature, pH, ionic strength etc.). Hence, there is a need to conduct these studies in the near future to successfully incorporate these SLNs into industrially processed foods.

3.2.1.3. Particle size

Particle size of SLN is an important criterion in designing delivery systems for bioactives for the reason that it affects organoleptic properties like appearance, stability, texture, and mouthfeel etc. and biological performance (dissolution, absorption etc.) of food products.^{17, 35} In this regard, chemical nature (type of fatty acid chain, degree of unsaturation, melting point, shape and surface area of lipid crystals, hydrophilicity etc.) plays an important role.³⁶ From earlier studies it is evident that, lipid with higher melting point forms larger nanoparticles due to increased viscosity.³⁷ Attesting these observations, in one of our studies, curcumin loaded SLNs fabricated using higher melting point lipid resulted in the formation of bigger SLNs compared to SLNs fabricated using the lipids with lower melting point. SLN fabricated using tristearin (MP: 70°C) had the size of 314 ± 23 nm compared to SLNs fabricated using glycerylmonostearate (MP: 56°C)

) which had the size of 111 ± 3 nm.³⁸ Further, with regarding to its application in food products, recent studies from our laboratory and others have shown that gradual increase in electrolyte concentration resulted in particle growth in Tween 80 stabilized SLN. This is evident by size increase of the SLNs. However pH of the solution has little effect on the stability of these SLNs.³²

3.2.1.4. Bioavailability

In general, bioavailability refers to the amount of bioactives available to act at the site of physiological activity in the living system. Lipid nanoparticles are used to increase oral bioavailability of poorly water soluble bioactives such as curcumin, quercetin etc. and bioactives which has low cell permeability like catechin.³⁹

Though, SLN digestion is a complex process, it can be categorized into three main phases, (1) digestive phase; (2) absorption phase and (3) entering the blood circulation.⁴⁰ In the beginning digestion is initiated by the hydrolysis of triglycerides (TGs) in to diglyceride (DGs) and fatty acids (FFAs) by gastric and lingual lipases in the stomach (Figure 3). Further, shear force generated during antral contraction, retropulsion and gastric emptying results in the formation of emulsions from the hydrolyzed lipids. In the duodenum, presence of lipids stimulates the release of bile salts, pancreatic fluids and binary lipids from gall bladder which further assists in the formation emulsions. In addition, bile salts which are amphiphilic in nature (one side is hydrophilic and another side of the same molecule is hydrophobic) acts on this lipids and align themselves to face their hydrophobic part towards the lipid droplets and forms the layer around droplets. This increases lipid droplets surface area and hydrophilicity and also avoids their re-aggregation. Here the Lipase which is a hydrophilic enzyme acts on the remaining triglycerides

and diglycerides to break apart glycerol back bone and fatty acid chains by breaking the bond which link them. As a result from each triglyceride 2 fatty acids and one monoglycerides (MGs) are released⁴¹. This released FFAs and MGs will form different structures like micelles, mixed micelles, vesicles etc. Here the bioactive compounds will be trapped inside these structures. The presence of bile salts increase their solubility in lumen of the gut (>1000 fold) due to which required concentration gradient for the diffusion of these structures into enterocytes is generated.⁴² While taking up these structures, bioactives which were trapped inside these structures will also get absorbed. In the cytosol (cell compartment) absorbed bioactive compounds will directly go to portal circulation if they have the sufficient solubility ($p < 5$). In case of bioactives which are highly hydrophobic ($p > 5$), add on to the intestinal lipoproteins (chylomicrons) in the enterocytes which were formed by the re-esterification of MGs and FFAs into TGs either by monoacyl glycerol pathway or phosphatidic pathway. Here these chylomicrons fuses with basolateral cell membrane of the enterocytes and transported to the circulation through thoracic lymph duct.

As in other factors, lipid composition plays an important role in determining the bioavailability pattern of entrapped nutraceutical molecules. In one of the recent studies from our laboratory it was found that lipid nanocarriers like NLCs which are composed of lipids with medium hydrocarbon chain length ($C = < 12$) has increased the bioavailability of hydrophobic bioactives such as quercetin compared to SLNs which are fabricated using long hydrocarbon chain length ($C = 21$). This could be attributed to the formation of medium chain free fatty acids after digestion which would migrate to the surrounding aqueous phase without interfering in the lipase activity resulting in the increased bioavailability. Whereas in case of long chain triglycerides ($C = 21$),

formation of long chain free fatty acids after digestion accumulates in the oil and water interface resulting in lipase activity inhibition and reduced bioavailability.⁴³

3.2.2. Surfactants

Surfactant plays a vital role in determining the physicochemical properties of SLNs such as size, surface charge, stability, polymorphic transition etc. Till date, various types of surfactants either alone or combination which are acceptable for direct addition into food products have been used to stabilize lipid nanoparticles including SLNs. Various surfactants which have been used or which are suitable to use in food products are listed in Table 1. During production, melted lipid resembles the emulsions. In these conditions surfactants align themselves in the water in oil (W/O) or oil in water (O/W) interface and provide stability to the nanoparticles after solidification by avoiding direct contact between them in the solution which otherwise leads to flocculation and coalescence.^{10, 31} With regarding to surfactants, factors which determine the stability of nanoparticles are surfactant type and concentration.

3.2.2.1. Surfactant type

One among the key challenges for the successful stabilization of SLNs in complex food matrix is selection of suitable food grade surfactant. Combinations of ionic and non ionic surfactants are used to obtain matrix stability and suspension stability of SLNs in drug formulations. However for food application, ionic surfactants are not preferred due to toxicity issues.³² Though biopolymers are non toxic their use in food is restricted due to their high emulsifying ability which induces gelling during storage.¹⁰ Hence proteins and polyoxyethylene sorbitan monooleate (Tweens) are preferred surfactants.

The selection of surfactant for the particular formulation depends on lipid matrix type and intended use. The surfactant property which determines the ability to act as surface active agents are molecular weight, chemical structure, hydrophile-lipophile balance (HLB), surface charge etc. Surfactants contain both hydrophilic and a lipophilic elements which assist them to align them in W/O, O/W or water-in-oil-in-water (W/O/W) interface. Selections of surfactants are usually done by calculating their HLB value. HLB is given by the balance between the size and strength of the hydrophilic and the lipophilic groups in a surfactant molecule.³⁶ According to Griffin classification, surfactants which has the HLB value < 9 are hydrophobic, < 11 are hydrophilic and in between them are in between.¹⁷ Usually HLBs are preferred by deciding type of interface which needs to be fabricated. In general HLB 3.5-6 supports formation of W/O emulsion and HLB 8-18 results in O/W emulsion. However, different types of lipids require different HLBs in order to ensure formation of stable nanoparticles.⁴⁴ Severino et al. reported superior stability of SLNs with the combination of polysorbate 80 and sorbitan trioleate due to their structure compatibility.⁴⁵ Another important aspect about the use of surfactants in SLN fabrication is controlling crystallization processes. An amphiphilic molecule with high hydrophilicity covers the new surfaces which are emerged due to conversion of α crystal to β crystals during storage (polymeric transition). In pharmaceutical formulation, recrystallization is inhibited by using bile salts like sodium glycocholate, sodium taurodeoxycholate etc. However in food products these bile salts are not preferred due to cost constraints and their bitter taste.^{46, 47}

In summary, ideal surfactant needs to stabilize the SLN against physical instability as well as to modulate crystallization (to prevent recrystallization). To achieve this in the formulations, combinations of surfactants with synergistic activity are used. Generally non ionic surfactants which provide stabilization against recrystallization are used in combination with

ionic surfactants which stabilize SLN by avoiding aggregation and flocculation. Further, combination of surfactants efficiently covers the surface area as well as produces sufficient viscosity to promote the nanoparticle stability.⁴⁸ Since, ionic surfactants are not preferred in food product; highly biocompatible molecules which can provide both suspension stability and matrix stability are warranted.

Recently, Salminen and co-workers reported significant increase in stability of aromatic amino acids stabilized SLNs with regarding to the retention of α crystals compared to SLN stabilized with other two synthetic surfactants such as pluronic F68, Tween 60 and 80. The main reason for their superior stability is due to the presence of both hydrophilic and hydrophobic areas with high water solubility. These characteristics allow the surfactant molecules to cover the new area formed during conversion of α crystal to β crystal. Recently in our laboratory we successfully stabilized SLN using β -lactoglobulin; the fabricated SLNs showed particle stabilization and matrix stabilization for more than 30 days in room temperature (22 ± 3 °C). This is due to the presence of essential characteristics in β -lactoglobulin to become an ideal surfactant (amphiphilic with pronounced hydrophobic areas, but highly water soluble). In the near future, stability of these β -lactoglobulin stabilized SLN will be studied in complex food matrix (unpublished data). Recent study from our laboratory revealed that Tweens successfully stabilizes the SLN in various pH conditions. However presence of electrolytes destabilized the SLN stabilized by Tweens.

3.2.2.2. Surfactant concentration

Presence of number of surfactant molecules on the surface area of the nanoparticles also affects the stability of SLNs. Presence of sufficiently high number of surfactants during the formation of

new surface area after conversion of bulk lipids into SLNs ensures stabilization by decreasing surface tension. Further availability of a few excess molecules of surfactant also effectively decreases the conversion of unstable α crystals of lipids into more thermodynamically stable β form during the storage.^{27, 32} Said so, its concentration should be optimum. Excess use of surfactant results in decreased loading efficiency and induces burst release.⁴⁸

4. Production of SLNs

There are several industrially feasible methods available to fabricate the SLNs. Generally SLNs are fabricated using either top down or bottom up methods as shown in Figure 4.

Some of the methods which are used in fabricating the bioactives loaded SLNs for food application are listed below.

A. Top down technology

- (a) High pressure homogenization (HPH)⁴⁹
- (b) Microfluidization⁵⁰
- (c) Membrane contactor method⁵¹

B. Bottom up technology

- (a) Emulsification–solvent evaporation⁵²
- (b) Emulsification–solvent diffusion⁵³

In principle, selection of method depends on the physicochemical characteristics of the excipients and bioactives, quantity of production etc. Among the above mentioned methods, HPH and microfluidization are the most preferred ones due to their ease in scalability and non toxicity. Hence HPH and microfluidization which are industrially most feasible methods are explained below in detail.

4.1. High pressure homogenization (HPH)

Initially this method has been used to fabricate nanoemulsion for parenteral nutrition production.⁹ In food industry, this technique is quite often used to improve the stability, shelf life, taste and quality of food products. Thus HPH is highly reliable and powerful technique to produce SLNs for food application. In HPH, lipid and bioactive melt is passed through the valve gap where required operating pressure (100- 2000 bar) can be created by piston pump. Here it breaks up the oil and water phases and form small W/O or O/W emulsion due to high shear stress and cavitation forces. After repeatedly exposing the oil and water phases to high shear stress and cavitation forces, emulsion with required size can be obtained. The factors which mainly determine the quality of SLN produced by HPH are homogenization pressure, temperature, number of cycles etc.^{9, 16} Homogenization can be done in both hot (melted lipid) and cold (powdered lipid) condition which helps to avoid the degradation of heat sensitive bioactives like lycopene. The disadvantages are its high energy dependence and distribution of bioactives into aqueous phase during homogenization. Figure 5 illustrates the schematic procedure for fabrication of SLN using HPH.

4.2. Microfluidization

Like HPH, microfluidization is a high energy emulsification method used to fabricate both W/O or O/W emulsion. This methodology can be used to fabricate SLNs at both laboratory and industrial scale and it is easily scalable. In food industry, it is used to fabricate emulsions and to homogenize proteins (whey protein, trypsin, milk etc.). In microfluidizer, oil and water phases breakup occurs from high turbulence and shear created by the collision of two impinging jets

oriented at 180° to each other. Here also operating pressure up to 150 MPa can be created by using piston pump. Size of the emulsion formed depends on operating pressure and the number of passes. Increasing operating pressure and the number of passes decreases size of the SLNs.

The specific advantages of these high energy emulsification methods (HPH and microfluidization) regarding bioactives loaded SLN production are as following

- A. SLN can be produced in both hot and cold conditions which help to avoid the degradation of heat sensitive bioactives like curcumin, quercetin etc., (Figure 5).
- B. Lipids are food ingredients or additives. These helps to maintain the required organoleptic properties (transparency, mouth feel) of food products even after the addition of SLNs.
- C. SLNs can be produced without using organic solvents.
- D. No contamination from instruments as in case of high shear homogenization (ultra sonication) occurs.
- E. HPH and microfluidization are easily scalable.

Conclusion

Though, SLNs do not fulfill all the criteria to be used as a sole carrier system for bioactives, they have several advantages which include the non toxic composition (physiological compounds), avoidance of organic compounds for production, high entrapment efficiency, well established industrially feasible method for production etc.

The main disadvantages are low loading efficiency for hydrophilic bioactives, polymorphic transition during storage and lack of proper evidence for their behavior in complex food matrix etc. However these disadvantages can be successfully avoided by selecting suitable

excipients for production, compatible bioactives for incorporation and finally choosing suitable production and storage conditions.

Presence of SLNs in the food products will help to materialize the need of bioactives enriched products which can deliver their health benefits to consumers without losing their nutritional properties as well as increase the profitability of food industries. In the future, bioactives entrapped in SLNs can be expected to take the central stage in the development of fortified food products.

In conclusion, SLN are very multifaceted delivery structure with several advantages and drawbacks to use them as bioactives delivery vehicles. In future, lot of efforts needs to be to put for fabrication of SLN which are particularly tuned for the delivery of bioactives to successfully use them in industrially processed food products.

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Reference

1. A. R. Patel and K. P. Velikov, *LWT-Food Sci Technol*, 2011, 44, 1958-1964.
2. J. E. Norton, G. A. Wallis, F. Spyropoulos, P. J. Lillford and I. T. Norton, *Annu Rev Food Sci T*, 2014, 5, 177-195.
3. A. Das and C. K. Sen, in *Nutraceutical and Functional Food Regulations in the United States and Around the World*, ed. D. Bagchi, Academic Press, San Diego, 2nd edn., 2014, DOI: <http://dx.doi.org/10.1016/B978-0-12-405870-5.00002-5>, pp. 13-39.
4. M. F. Yao, H. Xiao and D. J. McClements, *Annu Rev Food Sci T*, 2014, 5, 53-81.
5. M. A. Neves, J. Hashemi and C. Prentice, *Curr. Opin. Food Sci.*, 2015, 1, 7-12.
6. E. Acosta, *Curr Opin Colloid Interface Sci*, 2009, 14, 3-15.
7. V. Nedovic, A. Kalusevic, V. Manojlovic, S. Levic and B. Bugarski, *Procedia Food Sci*, 2011, 1, 1806-1815.
8. F. Tamjidi, M. Shahedi, J. Varshosaz and A. Nasirpour, *Innov Food Sci Emerg Technol.*, 2013, 19, 29-43.
9. W. Mehnert and K. Mäder, *Adv Drug Deliver Rev*, 2012, 64, Supplement, 83-101.
10. F. Tamjidi, M. Shahedi, J. Varshosaz and A. Nasirpour, *Innov Food Sci Emerg Technol.*, 2013, 19, 29-43.
11. S. Singh, A. K. Dobhal, A. Jain, J. K. Pandit and S. Chakraborty, *Chem Pharm Bull*, 2010, 58, 650-655.
12. S. Phan, S. Salentinig, E. Gilbert, T. A. Darwish, A. Hawley, R. Nixon-Luke, G. Bryant and B. J. Boyd, *J. Colloid Interface Sci.*, in press, DOI: <http://dx.doi.org/10.1016/j.jcis.2014.11.026>.

13. N. P. Aditya, M. Shim, I. Lee, Y. Lee, M. H. Im and S. Ko, *J Agr Food Chem*, 2013, 61, 1878-1883.
14. J. B. German, in *Impact of Processing on Food Safety*, eds. L. Jackson, M. Knize and J. Morgan, Springer US, 1999, vol. 459, ch. 3, pp. 23-50.
15. M. Fathi, M. R. Mozafari and M. Mohebbi, *Trends Food Sci Tech*, 2012, 23, 13-27.
16. H. Harde, M. Das and S. Jain, *Expert Opin Drug Del*, 2011, 8, 1407-1424.
17. P. Severino, T. Andreani, A. S. Macedo, J. F. Fangueiro, M. H. A. Santana, A. M. Silva and E. B. Souto, *Drug Deliv*, 2012, 2012, 10.
18. M. Trotta, M. E. Carlotti, M. Gallarate, G. P. Zara, E. Muntoni and L. Battaglia, *J Disper Sci Technol*, 2011, 32, 1041-1045.
19. P. Shahgaldian, M. Cesario, P. Goreloff and A. W. Coleman, *Chem Commun*, 2002, DOI: Doi 10.1039/B111367b, 326-327.
20. J. F. Fangueiro, E. Gonzalez-Mira, P. Martins-Lopes, M. A. Egea, M. L. Garcia, S. B. Souto and E. B. Souto, *Pharm Dev Technol*, 2013, 18, 545-549.
21. K. Ahmed, Y. Li, D. J. McClements and H. Xiao, *Food Chem*, 2012, 132, 799-807.
22. K. Vivek, H. Reddy and R. S. R. Murthy, *AAPS PharmSciTech*, 2007, 8.
23. N. P. Aditya, S. Aditya, H.-J. Yang, H. W. Kim, S. O. Park, J. Lee and S. Ko, *J Funct Foods*, 2015, 15, 35-43.
24. A. P. Nayak, W. Tiyaboonchai, S. Patankar, B. Madhusudhan and E. B. Souto, *Colloid Surface B*, 2010, 81, 263-273.
25. K. Westesen and H. Bunjes, *Int. J. Pharm.*, 1995, 115, 129-131.
26. C. Olbrich, A. Gessner, O. Kayser and R. H. Muller, *J Drug Target*, 2002, 10, 387-396.

27. H. Salminen, T. Helgason, S. Aulbach, B. Kristinsson, K. Kristbergsson and J. Weiss, *J. Colloid Interface Sci.*, 2014, 426, 256-263.
28. V. Jenning and S. H. Gohla, *J Microencapsul*, 2001, 18, 149-158.
29. K. Westesen, B. Siekmann and M. H. J. Koch, *Int. J. Pharm.*, 1993, 93, 189-199.
30. V. Jenning, A. F. Thünemann and S. H. Gohla, *Int. J. Pharm.*, 2000, 199, 167-177.
31. J. Weiss, E. A. Decker, D. J. McClements, K. Kristbergsson, T. Helgason and T. Awad, *Food Biophys*, 2008, 3, 146-154.
32. K. O. Choi, N. P. Aditya and S. Ko, *Food Chem*, 2014, 147, 239-244.
33. Z. G. Cui, K. Z. Shi, Y. Z. Cui and B. P. Binks, *Colloids Surf A Physicochem Eng Asp*, 2008, 329, 67-74.
34. A. Radomska-Soukharev, *Adv Drug Deliver Rev*, 2007, 59, 411-418.
35. V. Kakkar, S. Singh, D. Singla and I. P. Kaur, *Mol Nutr Food Res*, 2011, 55, 495-503.
36. D. J. Hauss, *Adv Drug Deliver Rev*, 2007, 59, 667-676.
37. M. Uner and G. Yener, *Int J Nanomed*, 2007, 2, 289-300.
38. A. P. Nayak, W. Tiyaboonchai, S. Patankar, B. Madhusudhan and E. B. Souto, *Colloids Surf., B*, 2010, 81, 263-273.
39. N. P. Aditya, S. Aditya, H. Yang, H. W. Kim, S. O. Park and S. Ko, *Food Chem*, 2015, 173, 7-13.
40. L. H. Reddy and R. S. Murthy, *Indian J Exp Biol*, 2002, 40, 1097-1109.
41. Y. Sun, Z. Xia, J. Zheng, P. Qiu, L. Zhang, D. J. McClements and H. Xiao, *J Funct Foods*, 2015, 13, 61-70.
42. H. Westergaard and J. M. Dietschy, *J. Clin. Invest.*, 1976, 58, 97-108.

43. L. Salvia-Trujillo, C. Qian, O. Martín-Belloso and D. J. McClements, *Food Chem*, 2013, 139, 878-884.
44. T. Schmidts, D. Dobler, C. Nissing and F. Runkel, *J. Colloid Interface Sci.*, 2009, 338, 184-192.
45. P. Severino, S. C. Pinho, E. B. Souto and M. H. A. Santana, *Colloids Surf., B*, 2011, 86, 125-130.
46. K. Westesen, H. Bunjes and M. H. J. Koch, *J Control Release*, 1997, 48, 223-236.
47. K. Westesen and B. Siekmann, *Int. J. Pharm.*, 1997, 151, 35-45.
48. R. Cavalli, O. Caputo, E. Marengo, F. Pattarino and M. R. Gasco, *Pharmazie*, 1998, 53, 392-396.
49. N. P. Aditya, A. S. Macedo, S. Doktorovova, E. B. Souto, S. Kim, P.-S. Chang and S. Ko, *LWT-Food Sci Technol*, 2014, 59, 115-121.
50. L. Lee and I. T. Norton, *J. Food Eng.*, 2013, 114, 158-163.
51. D. M. Lloyd, I. T. Norton and F. Spyropoulos, *J. Membr. Sci.*, 2014, 466, 8-17.
52. M. Fathi, Á. Martín and D. J. McClements, *Trends Food Sci Tech*, 2014, 39, 18-39.
53. A. Imbrogno, E. Piacentini, E. Drioli and L. Giorno, *J. Membr. Sci.*, 2014, 467, 262-268.
54. J. Sun, C. Bi, H. M. Chan, S. Sun, Q. Zhang and Y. Zheng, *Colloids Surf., B*, 2013, 111, 367-375.
55. R. Cortesi, E. Esposito, G. Luca and C. Nastruzzi, *Biomaterials*, 2002, 23, 2283-2294.
56. L. H. Reddy, K. Vivek, N. Bakshi and R. S. R. Murthy, *Pharm Dev Technol*, 2006, 11, 167-177.
57. H. Li, X. Zhao, Y. Ma, G. Zhai, L. Li and H. Lou, *J Control Release*, 2009, 133, 238-244.

58. K. Teskač and J. Kristl, *Int. J. Pharm.*, 2010, 390, 61-69.
59. S. Jose, S. S. Anju, T. A. Cinu, N. A. Aleykutty, S. Thomas and E. B. Souto, *Int. J. Pharm.*, 2014, 474, 6-13.
60. J. Zhang and E. Smith, *J Pharm Sci-US*, 2011, 100, 896-903.
61. D. T. Baviskar, A. S. Amritkar, H. S. Chaudhari and D. K. Jain, *Pharmazie*, 2012, 67, 701-705.
62. S. A. Wissing and R. H. Muller, *Pharmazie*, 2001, 56, 783-786.
63. L. Montenegro, A. Campisi, M. G. Sarpietro, C. Carbone, R. Acquaviva, G. Raciti and G. Puglisi, *Drug Dev Ind Pharm*, 2011, 37, 737-746.
64. H. Yuan, J. Chen, Y.-Z. Du, F.-Q. Hu, S. Zeng and H.-L. Zhao, *Colloids Surf., B*, 2007, 58, 157-164.
65. J. O. Woo, M. Misran, P. F. Lee and L. P. Tan, *Sci World J*, 2014, DOI: Artn 205703
Doi 10.1155/2014/205703.
66. S. Y. Xie, L. Y. Zhu, Z. O. Dong, Y. Wang, X. F. Wang and W. Z. Zhou, *Int J Nanomed*, 2011, 6, 547-555.
67. D. Chirio, M. Gallarate, E. Peira, L. Battaglia, L. Serpe and M. Trotta, *J Microencapsul*, 2011, 28, 537-548.
68. L. Battaglia, M. Gallarate, R. Cavalli and M. Trotta, *J Microencapsul*, 2010, 27, 78-85.
69. N. Scholer, H. Hahn, R. H. Muller and O. Liesenfeld, *Int. J. Pharm.*, 2002, 231, 167-176.
70. S. Jebors, A. Leydier, Q. Z. Wu, B. B. Ghera, M. Malbouyre and A. W. Coleman, *J Microencapsul*, 2010, 27, 561-571.
71. I. Montasser, P. Shahgaldian, F. Perret and A. W. Coleman, *Int J Mol Sci*, 2013, 14, 21899-21942.

72. J. Varshosaz, M. Tabbakhian and M. Y. Mohammadi, *J Liposome Res*, 2010, 20, 286-296.
73. T. Helgason, T. S. Awad, K. Kristbergsson, E. A. Decker, D. J. McClements and J. Weiss, *J Agr Food Chem*, 2009, 57, 8033-8040.
74. M. Sznitowska, M. Gajewska, S. Janicki, A. Radwanska and G. Lukowski, *Eur J Pharm Biopharm*, 2001, 52, 159-163.
75. J. Yi, T. I. Lam, W. Yokoyama, L. W. Cheng and F. Zhong, *J Agr Food Chem*, 2014, 62, 1096-1104.
76. C. M. Keck, A. Kovačević, R. H. Müller, S. Savić, G. Vuleta and J. Milić, *Int. J. Pharm.*, 2014, 474, 33-41.

Figure Legends

Figure 1. Problems associated with food bioactives for attaining optimal product stability and in body performance.

Figure 2. Comparison between lipid based delivery systems with regarding to their stability in complex food matrix and gastro intestinal track (GIT).

Figure 3. Schematic representation of digestion and absorption of bioactive compounds during oral intake using solid lipid nanoparticles (SLN).

Figure 4. Top down and bottom up technology for the production of solid lipid nanoparticles (SLNs).

Figure 5. Illustration of solid lipid nanoparticles (SLNs) production by hot and cold homogenization method.

Table1. Excipients and surfactants used in the fabrication of SLNs

Items	Food-grade materials	References
	Triacylglycerols	
	Trimyristin (Dynasan 114)	38, 54
	Tripalmitin (Dynasan 116)	17, 38
	Tristearin (Dynasan 118)	55, 56
	Mono-, di- and triglycerides mixtures	
	Glyceryl monostearate (Imwitor 900)	29, 49, 57
	Glyceryl behenate (Compritol 888 ATO)	35, 58, 59
	Waxes	
Excipients	Bee wax	60, 61
	Cetyl palmitate	62, 63
	Hard fats	
	Stearic acid	45, 64, 65
	Palmitic acid	66, 67
	Behenic acid	68, 69
	Other lipid	
	Para-acyl-calix(4)arenes	70, 71

	Polyoxyethylene sorbitan monooleate	
	Polysorbate 20 (16.7)	72
	Polysorbate 60 (14.9)	73
	Polysorbate 80 (15)	73
	Lecithin	
Surfactants	Soy and egg lecithin (4.0)	57, 73, 74
(HLB)	Protein	
	Whey protein	75
	Others	
	Alkyl polyglucosides	76
	Amino acids	27

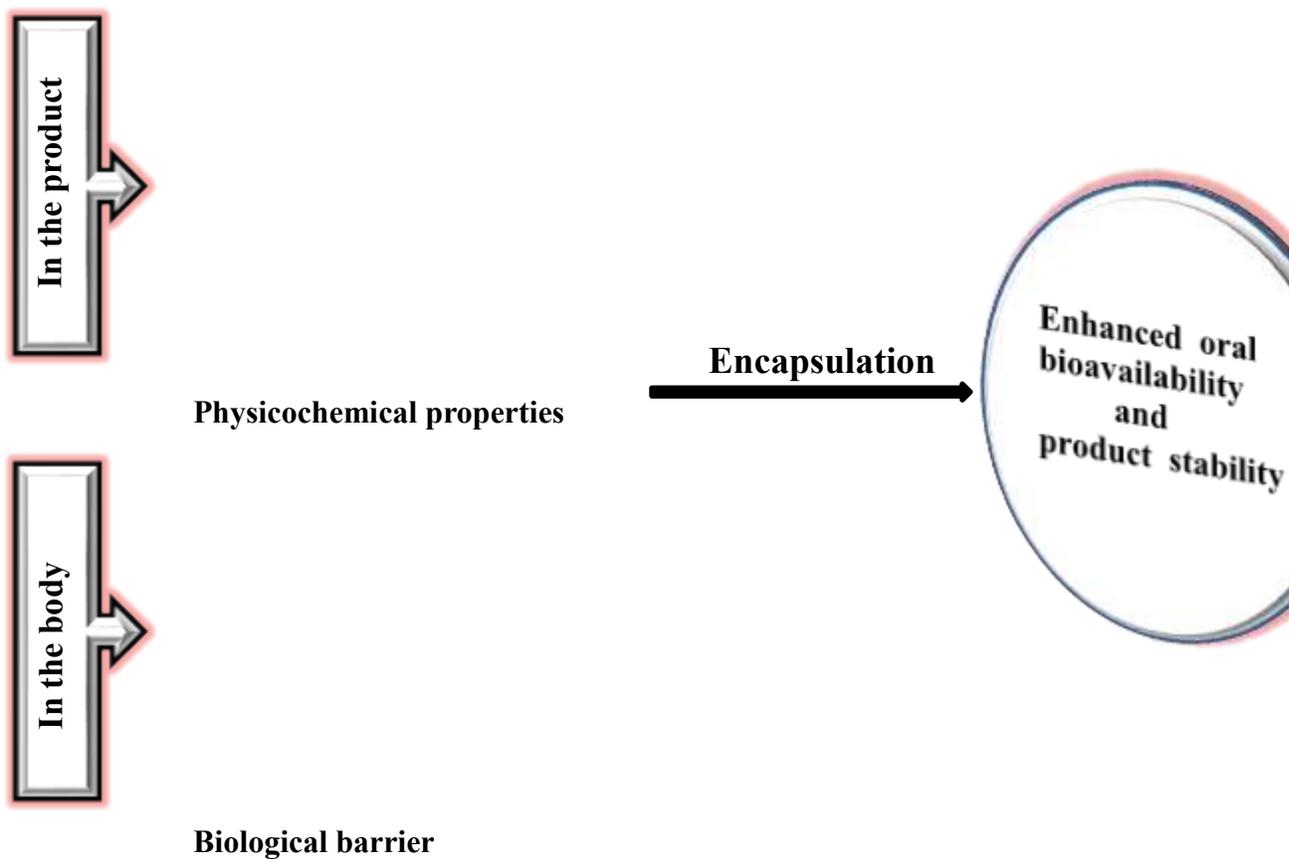


Figure 1

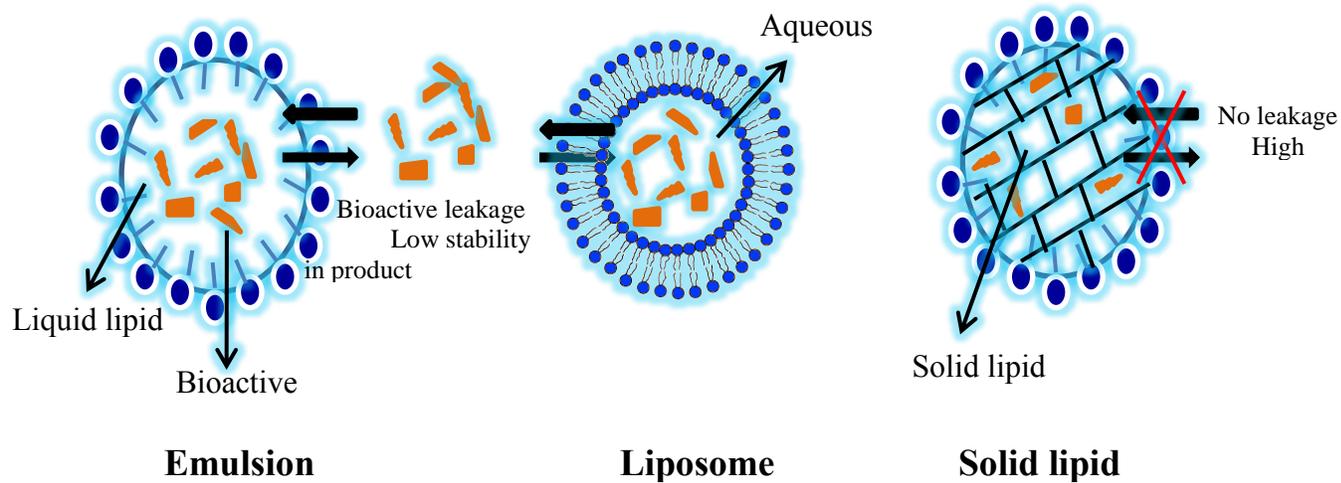


Figure 2

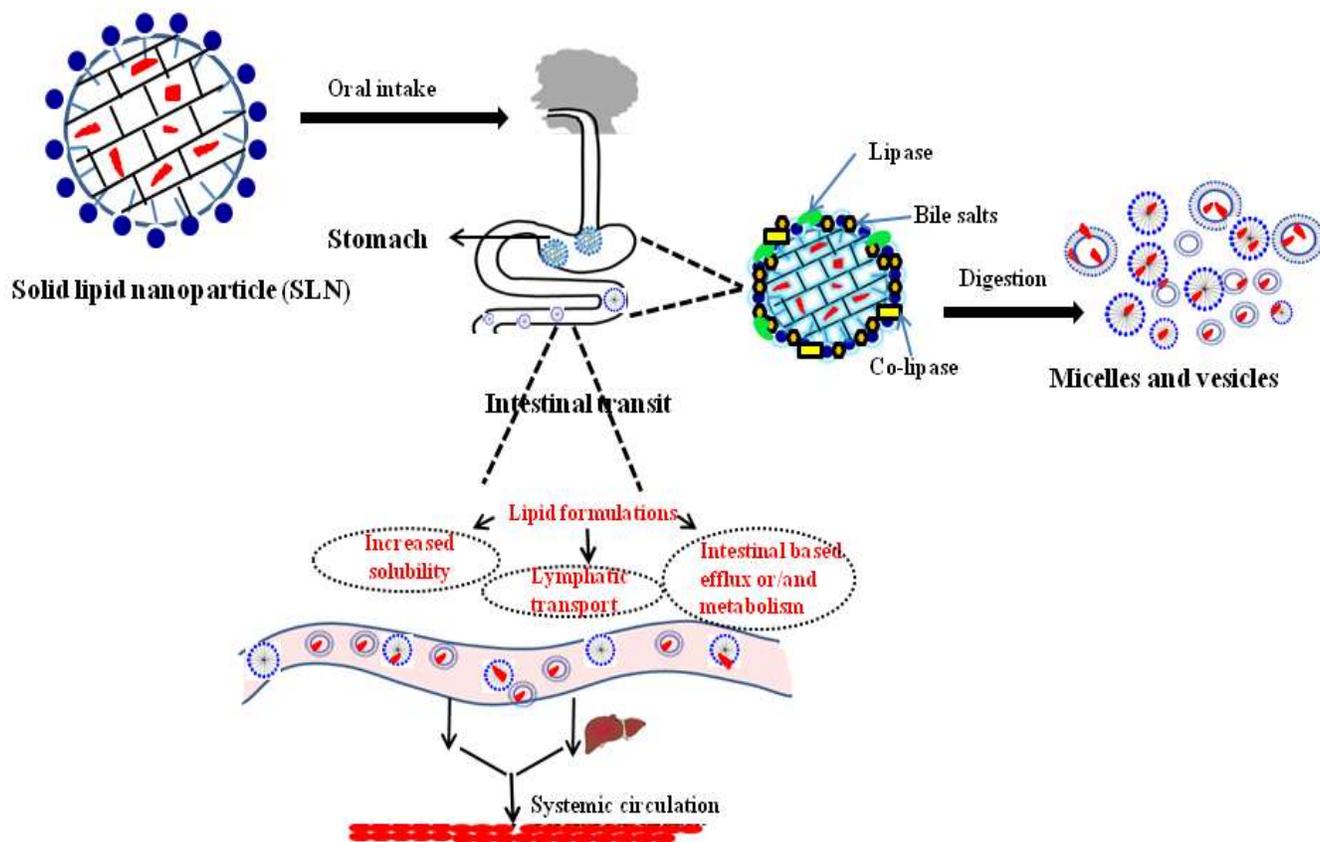


Figure 3

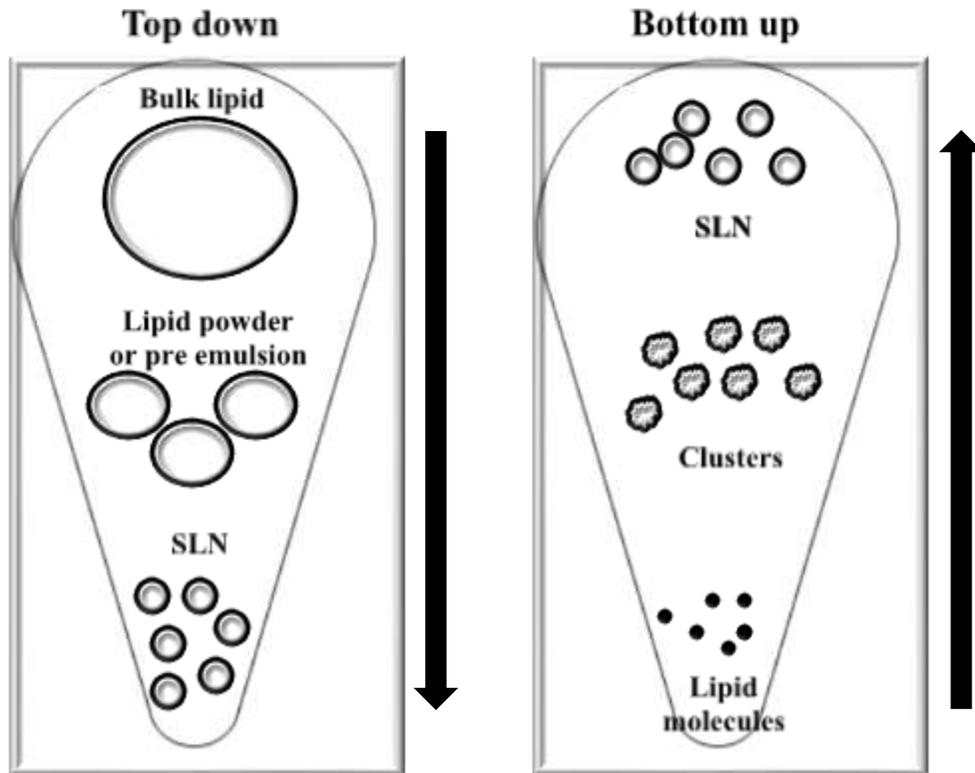


Figure 4

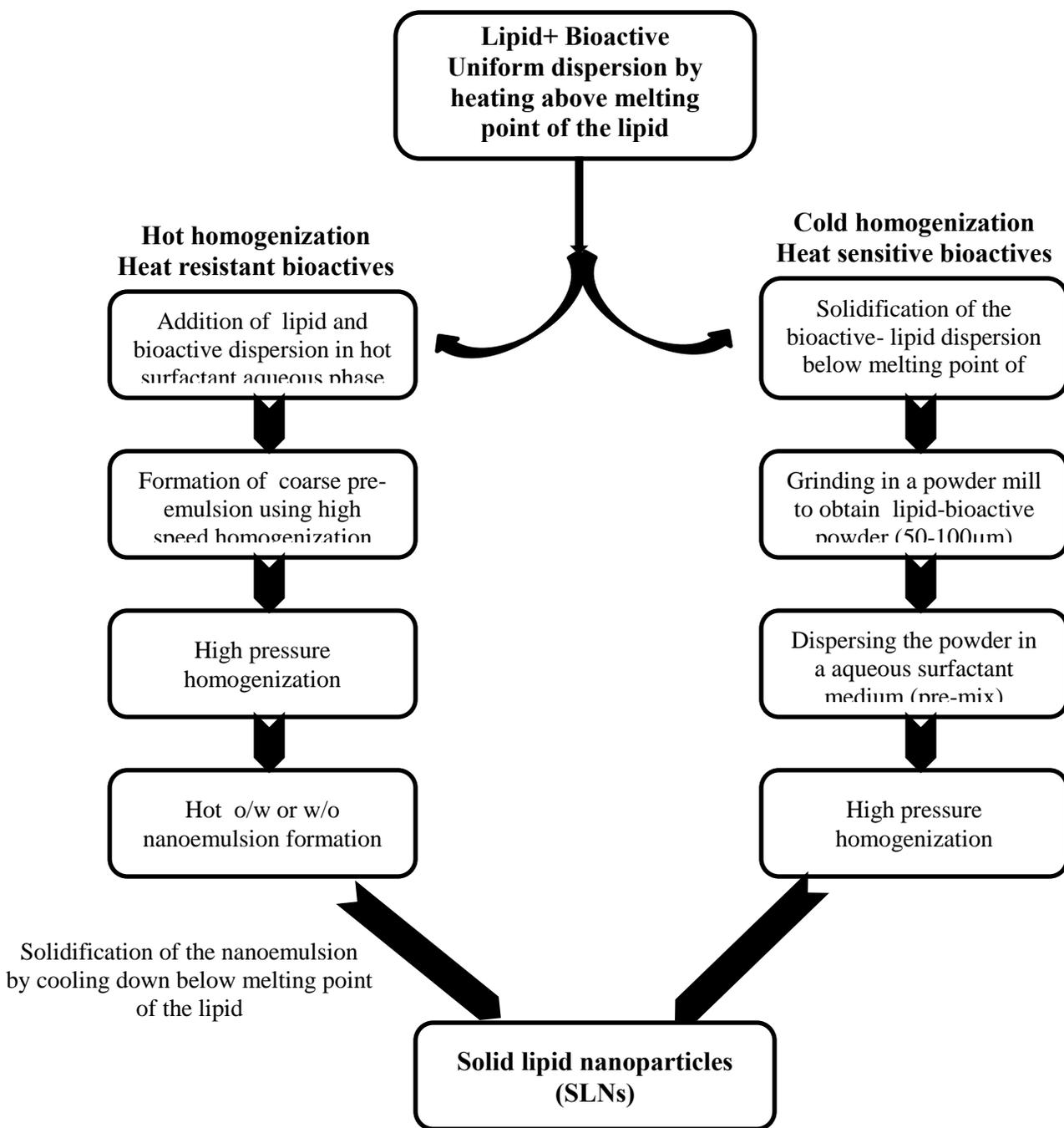


Figure 5