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COMMUNICATION

Renal-specific delivery of Prednisolone-folate conjugates for renal ischemia/reperfusion injury†

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Prednisolone-folate conjugate (PFC) was synthesized to achieve renal-targeted delivery and specific intracellular release of prednisolone. Our results highlight the significance of folate-mediated targeted delivery to kidney and the consequent *in vivo* therapeutic efficacy of PFC against renal ischemia/reperfusion injury.

Renal ischemia/reperfusion injury (IRI) commonly occurs in kidney transplantation, cardiopulmonary and aortic bypass surgery, trauma, hemorrhage, hypotension and burns which affects a large population of patients worldwide¹⁻³. As a leading cause for acute renal failure, renal IRI is associated with very high morbidity and mortality⁴. Glucocorticoids (GCs) are a class of steroid hormones that have been successfully used in clinic for their anti-inflammatory and immunosuppressive effects⁵. GCs have shown reno-protective effects against renal IRI by a receptor-dependent, non-genomic mechanism in the rat renal IRI model⁵⁻⁷. GCs exert effects by entering target cells and binding to the inactive cytoplasmic glucocorticoid receptors. The activated ligand-receptor complex then translocates into the nucleus to regulate gene transcription either directly or indirectly^{8,9}. However, extensive and severe adverse effects were reported for GCs when administered systemically mainly due to a non-specific distribution of GCs *in vivo*, including osteoporosis, peptic ulcer, hyperglycemia and weight gain^{10,11}. Thus, a renal specific delivery of GCs would be beneficial to improve their therapeutic efficacy against renal IRI while limiting systemic adverse effects.

Our lab has focused on the development of renal-specific delivery systems for years using prednisolone as the selected therapeutic molecule. Previously, several strategies have been explored to deliver prednisolone selectively to the kidney, *e.g.*, N-acetylated low molecular weight chitosan-prednisolone conjugates, prednisolone-glucose, and prednisolone-glucosamine conjugates which showed significantly higher intrarenal drug concentration compared to the free drug and the specificity to proximal tubule epithelial cells (PTECs)¹²⁻¹⁶. These glycoconjugates were designed to achieve renal targeting attributed to the specific interaction between sugar moieties and glucose transporters or megalin/cublin receptors that are extensively distributed in the renal proximal tubules. However, extensive distribution of glucose and other related transporters

throughout the human body present potential challenges on the targeted delivery of prednisolone to kidney via glycoconjugates. Therefore, more specific delivery strategies need to be developed to overcome existing challenges. In general, a successful renal-specific delivery system requires: i). an efficient renal-targeting moiety, *e.g.*, ligands that can interact with specific receptors in the kidney; ii). good aqueous solubility that favors rapid distribution to the kidney via systemic administration; iii). a suitable linker that is stable in the blood circulation and can further be cleaved to achieve drug release at the target site.

Herein, we report a new prednisolone conjugate candidate derived from prednisolone and folate to selectively deliver prednisolone to PTECs in the kidney. High folate receptor- α expression was well demonstrated in the PTECs thus making it an excellent target for specific drug delivery to the kidney^{17,18}. Previously, folate-conjugated agents for tumor imaging were shown to distribute specifically in the kidney in addition to the tumor site^{19,20}. This is mostly likely due to a relatively high level of folate receptor- α expression in PTECs. Moreover, the re-absorption of filtered solutes by PTECs may also contribute to the accumulation of folate conjugates in the kidney. However, tumor-targeted folate conjugates did not display nephrotoxicity which implies that folate conjugates might undergo rapid exocytosis and might be further transported across the basolateral membrane and into the circulation^{19,21}. Folate conjugates are suggested to enter cells via receptor-mediated endocytosis and undergo stages of endosomal and lysosomal metabolism and degradation in the cytoplasm. Folate receptor-containing endosomal pH was proven around 6.5 which is critical to the intracellular release of parent drug²². Based on these findings, an acid-labile acetal linker was introduced in the design of prednisolone-folate conjugate (PFC) to allow specific cleavage and activation of the prodrug in the endosomes of PTECs. The structure of acetals offers a reasonable stability under extracellular conditions while undergoing fast hydrolysis in an acidic milieu²³. Derived from previous studies^{20,24,25}, the proposed PFC structure consists of a water soluble pentapeptide spacer rendering the conjugate more water-soluble, a folic acid moiety with targetability to PTECs in the kidney, an acid-labile linker that allows the intracellular release of drug and the therapeutic moiety of prednisolone (Fig. 1). A straight forward synthetic strategy was thus developed in the study to synthesize PFC (Scheme S1, ESI†).

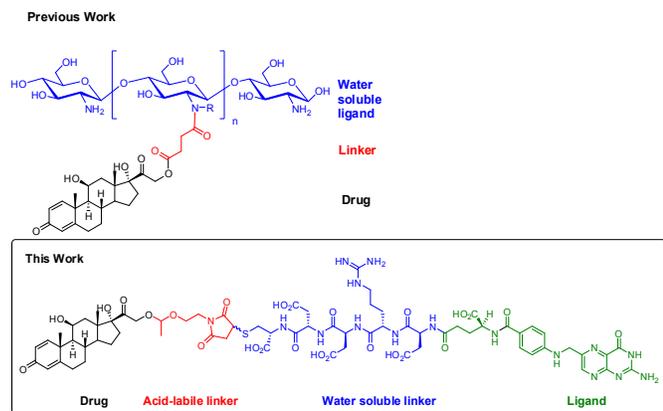


Fig. 1 Renal-specific delivery of prednisolone. (R=H or Ac)

To confirm cleavage and release of prednisolone under mildly acidic conditions, the *in vitro* stability of PFC was evaluated at varying pH conditions. PFC showed an aqueous solubility of about 3.27 mg/mL in the neutral buffer saline while prednisolone was reported to be nearly insoluble in water. At 37°C, 99% of PFC remained unchanged for 29h. Also, PFC remained stable in the rat plasma with 90% of the conjugates remaining unchanged for 6h and 85% of the conjugates remaining unchanged for 12h (Fig. S1, ESI†). Maintaining plasma stability is critical to the successful delivery of the conjugate to the target organ through systemic administration without encountering quick hydrolysis or degradation in the plasma. At pH 5, less than 50% of the conjugate was hydrolyzed by 12h, while at pH 2, over 60% of the conjugate was hydrolyzed by 4h (Fig. S2, ESI†). Thus, the acid-labile property of the conjugate would trigger the specific release of prednisolone in the mildly acidic endosomes via folate-receptor mediated endocytosis while remaining stable in the circulation.

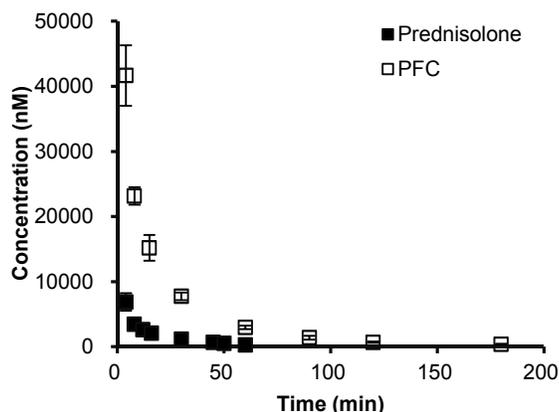


Fig. 2. The time-concentration profile of prednisolone in plasma after injection of prednisolone and prednisolone-folate conjugate (PFC). Data represent mean \pm S.D. (n=3).

To gain insight into the *in vivo* profiles of the PFC conjugate, the pharmacokinetic profiles of PFC and prednisolone were evaluated in rats. Both PFC and prednisolone displayed a profile with decreasing plasma concentration over time *in vivo* (Fig. 2). Interestingly, PFC displayed significantly higher plasma concentrations at all time points than prednisolone ($p < 0.05$). In contrast to prednisolone, a 7.7-fold higher AUC_{0-t} value and a 1.9-fold higher mean-residence-

time (MRT_{0-t}) value were observed for PFC. Pharmacokinetic parameters, including maximum plasma concentration (C_{max}), relative uptake efficiency ($R_{ekidney}$) and concentration efficiency ($C_{ekidney}$) of PFC and prednisolone in kidney, are presented in Table S2 (ESI†).

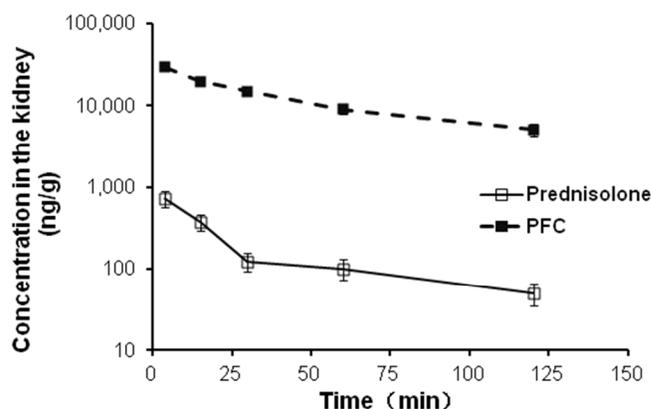


Fig. 3 The concentration-time profile of prednisolone and PFC in rat kidney after intravenous injection (equivalent to 3 mg·kg⁻¹ prednisolone). Data represent mean \pm S.D., (n=5).

Regarding kidney bioavailability (Fig. 3), PFC displayed significantly higher levels of distribution in the rat kidney compared to free prednisolone over the course of investigation ($p < 0.05$). Compared to previously reported prednisolone conjugate systems, a remarkably higher $R_{ekidney}$ of 116.83 was observed for PFC (Table S3, ESI†), while a $R_{ekidney}$ of 5.64 was reported for prednisolone-glucosamine system²⁶, which provided strong evidence for selecting folate as the targeting ligand for specific renal drug delivery.

To further evaluate the kidney targetability of PFC, the tissue distribution profile of PFC and prednisolone was measured at 4 and 30 min after i.v. injection (Fig. 4). At 4 min, PFC displayed significantly higher distributions in both kidney and plasma than in other organs, while concurrently prednisolone showed higher distributions in intestine and pancreas. At 30 min, PFC maintained the specific distribution in the kidney with a much higher concentration compared to all other organs. However, PFC also displayed increased distribution in the intestine at 30 min after single i.v. injection, a phenomenon that has been previously reported.²⁵ The increase in the intestinal distribution may be due to the folate receptor expression in the intestine. Additionally, promiscuous organic anion transporters (OATP) may also contribute to the transport of folate conjugates in the intestine. Studies showed that folate conjugates avoided being recognized and transported by OATP through increasing the steric hindrance of the linker linking folate and therapeutic agent.²⁷

To investigate the reno-protective effect of PFC against renal IRI in rats, a rat renal IRI model was established with some modifications to the previous study.²⁸ PFC, which was consecutively given at 3mg/kg for three days, effectively prevented and alleviated acute injury to renal tubules by maintaining serum creatine (CRER) and blood urea nitrogen (BUN) at normal levels compared to the sham group (Table S4, ESI†). Furthermore, the renal morphology of

ischemia/reperfusion (I/R) group displayed common renal damages such as dilatation of the tubular lumen, degeneration of the renal tubular epithelial cells, infiltration of leukocytes, and formation of protein casts (Fig. 5B). Compared with I/R group, renal damages were relieved to a certain extent in the I/R group treated with prednisolone, though dilatation of the tubular lumen, degeneration of tubular epithelia cells and protein casts were observed (Fig. 5D). Only mild degeneration of the renal tubular cells, dilatation of the tubular lumen and few casts were observed in the I/R group treated with PFC compared to the sham-operated group (Fig. 5A and C) which demonstrated a better therapeutic performance for PFC *in vivo* and thus a great potential in renal IRI treatment.

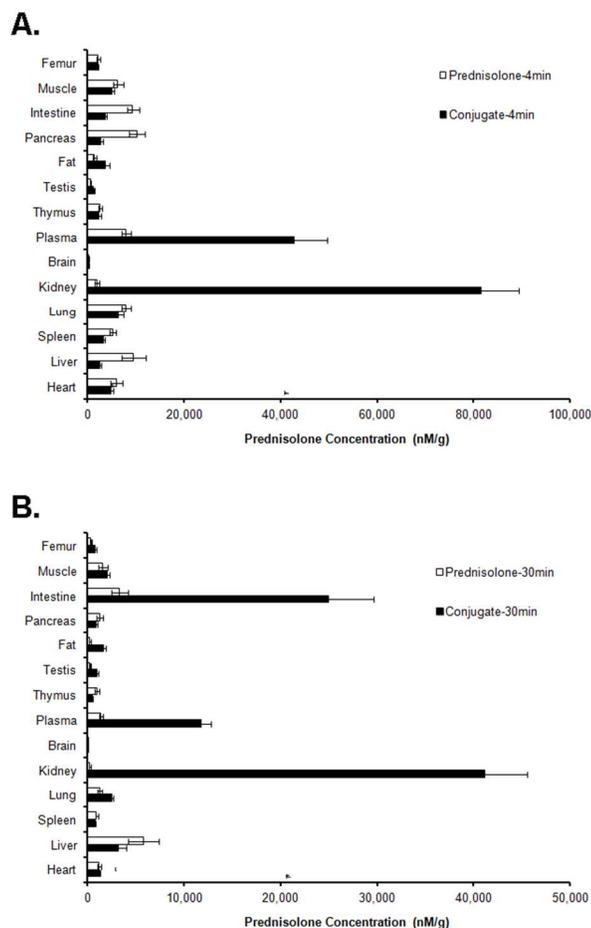


Fig 4. Tissue distribution in rats 4 min (A) and 30 min (B) after i.v. injection of prednisolone (white columns) and PFC (black columns). Data represent mean \pm SD, (n=5).

In summary, we utilized folate as the targeting ligand to achieve renal-specific delivery of prednisolone. PFC was demonstrated to have improved water solubility, plasma stability, and acid-sensitive properties that are crucial to achieving targeted kidney delivery via systemic administration. Moreover, PFC displayed excellent renal targetability *in vivo*, which was also proven to successfully reverse the disease progression in the rat renal I/R model indicating that PFC can serve as a potential prodrug candidate for renal IRI therapy.

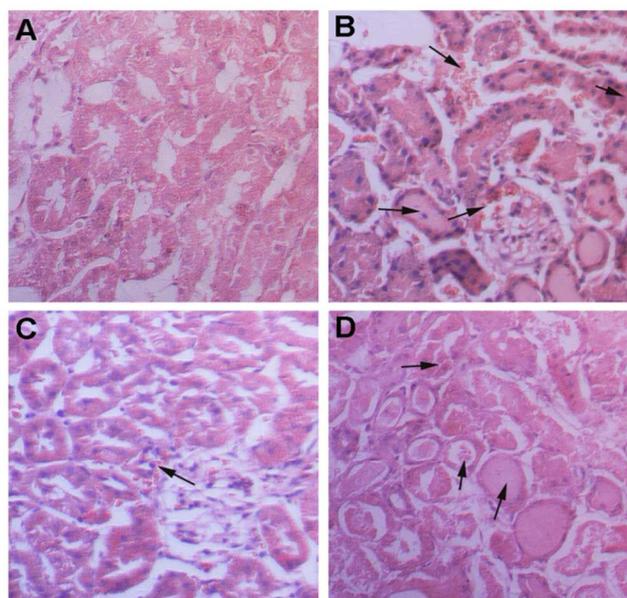


Fig 5. Renal morphology of sham-operated (A), I/R (B), I/R + PFC (C) and I/R + prednisolone (D) treated kidneys by hematoxylin-eosin staining. Prednisolone treated group showed significantly more severe damages than the PFC treated group. Magnification for all panels is 200 \times . Black arrows in panels B, C and D indicate the areas of damage. These are representative sections from five animals analyzed for each condition.

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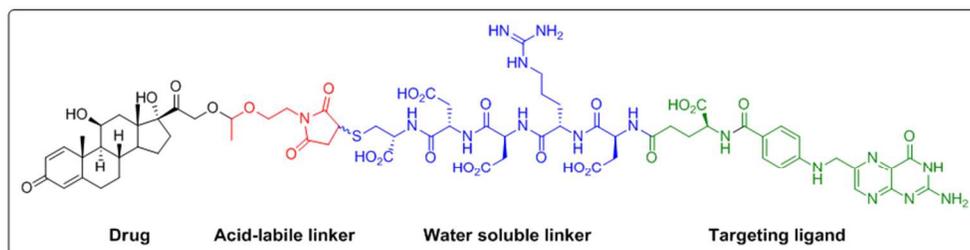
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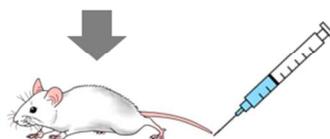
[†] Electronic Supplementary Information (ESI) available: Experimental methods, characterization data. See DOI: 10.1039/c000000x/

1. Z. Aydin, A. J. van Zonneveld, J. W. de Fijter and T. J. Rabelink, *Nephrology Dialysis Transplantation*, 2007, **22**, 342-346.
2. C. M. Mangano, L. S. Diamondstone, J. G. Ramsay, A. Aggarwal, A. Herskowitz, D. T. Mangano and G. Multictr Study Perioperative Ischemia Res, *Annals of Internal Medicine*, 1998, **128**, 194-203.
3. A. Kazmers, L. Jacobs and A. Perkins, *Journal of Surgical Research*, 1997, **67**, 62-66.
4. G. M. Chertow, E. Burdick, M. Honour, J. V. Bonventre and D. W. Bates, *Journal of the American Society of Nephrology*, 2005, **16**, 3365-3370.
5. S. Kumar, D. A. Allen, J. E. Kieswich, N. S. A. Patel, S. Harwood, E. Mazzon, S. Cuzzocrea, M. J. Raftery, C. Thiernemann and M. M. Yaqoob, *Journal of the American Society of Nephrology*, 2009, **20**, 2412-2425.

6. K. Salmela, L. Wramner, F. Ekberg, I. Hauser, O. Bentdal, L. E. Lins, H. Isoniemi, L. Backman, N. Persson, H. H. Neumayer, P. F. Jorgensen, C. Spieker, B. Hendry, A. Nicholls, G. Kirste and G. Hasche, *Transplantation*, 1999, **67**, 729-736.
7. A. Reutzel-Selke, T. Zschockelt, C. Denecke, U. Bachmann, A. Jurisch, J. Pratschke, G. Schmidbauer, H. D. Volk, P. Neuhaus and S. G. Tullius, *Transplantation*, 2003, **75**, 1786-1792.
8. C. Stahn, M. Loewenberg, D. W. Hommes and F. Buttgerit, *Molecular and Cellular Endocrinology*, 2007, **275**, 71-78.
9. H. Schacke, A. Schottelius, W. D. Docke, P. Strehlke, S. Jaroch, N. Schmees, H. Rehwinkel, H. Hennekes and K. Asadullah, *Proceedings of the National Academy of Sciences of the United States of America*, 2004, **101**, 227-232.
10. H. Schacke, W. D. Docke and K. Asadullah, *Pharmacology & Therapeutics*, 2002, **96**, 23-43.
11. E. Bakina, Z. Wu, M. Rosenblum and D. Farquhar, *Journal of Medicinal Chemistry*, 1997, **40**, 4013-4018.
12. Z.-X. Yuan, X. Sun, T. Gong, H. Ding, Y. Fu and Z.-R. Zhang, *Journal of Drug Targeting*, 2007, **15**, 269-278.
13. Z.-x. Yuan, Z.-r. Zhang, D. Zhu, X. Sun, T. Gong, J. Liu and C.-t. Luan, *Molecular Pharmaceutics*, 2009, **6**, 305-314.
14. Z.-x. Yuan, J.-j. Li, D. Zhu, X. Sun, T. Gong and Z.-r. Zhang, *Journal of Drug Targeting*, 2011, **19**, 540-551.
15. Y. Lin, Y. P. Li, X. H. Wang, T. Gong, L. Zhang and X. Sun, *Journal of Controlled Release*, 2013, **167**, 148-156.
16. X. Liu, W. Li, Z. Liang, X. Zhang, Y. Guo, T. Gong and Z. Zhang, *Archiv Der Pharmazie*, 2012, **345**, 925-933.
17. H. Birn, O. Spiegelstein, E. I. Christensen and R. H. Finnell, *Journal of the American Society of Nephrology*, 2005, **16**, 608-615.
18. B. A. Kamen and A. K. Smith, *Advanced Drug Delivery Reviews*, 2004, **56**, 1085-1097.
19. C. P. Leamon, J. A. Reddy, I. R. Vlahov, P. J. Kleindl, M. Vetzal and E. Westrick, *Bioconjugate Chemistry*, 2006, **17**, 1226-1232.
20. C. P. Leamon, M. A. Parker, I. R. Vlahov, L. C. Xu, J. A. Reddy, M. Vetzal and N. Douglas, *Bioconjugate Chemistry*, 2002, **13**, 1200-1210.
21. C. P. Leamon, J. A. Reddy, I. R. Vlahov, E. Westrick, A. Dawson, R. Dorton, M. Vetzal, H. K. Santhapuram and V. Wang, *Molecular Pharmaceutics*, 2007, **4**, 659-667.
22. J. Yang, H. Chen, I. R. Vlahov, J.-X. Cheng and P. S. Low, *Journal of Pharmacology and Experimental Therapeutics*, 2007, **321**, 462-468.
23. A. Schlossbauer, C. Dohmen, D. Schaffert, E. Wagner and T. Bein, *Angewandte Chemie-International Edition*, 2011, **50**, 6828-6830.
24. J. M. Shillingford, C. P. Leamon, I. R. Vlahov and T. Weimbs, *Journal of the American Society of Nephrology*, 2012, **23**, 1674-1681.
25. S. F. Knight, K. Kundu, G. Joseph, S. Dikalov, D. Weiss, N. Murthy and W. R. Taylor, *Journal of the American Society of Nephrology*, 2012, **23**, 793-800.
26. Y. Lin, Y. Li, X. Wang, T. Gong, L. Zhang and X. Sun, *Journal of Controlled Release*, 2013, **167**, 148-156.
27. C. P. Leamon, J. A. Reddy, P. J. Klein, I. R. Vlahov, R. Dorton, A. Bloomfield, M. Nelson, E. Westrick, N. Parker, K. Bruna, M. Vetzal, M. Gehrke, J. S. Nicoson, R. A. Messmann, P. M. LoRusso and E. A. Sausville, *Journal of Pharmacology and Experimental Therapeutics*, 2011, **336**, 336-343.
28. Z. Zhang, Q. Zheng, J. Han, G. Gao, J. Liu, T. Gong, Z. Gu, Y. Huang, X. Sun and Q. He, *Biomaterials*, 2009, **30**, 1372-1381.



Prednisolone-Folate conjugate (PFC)



Kidney targeting

Systemically administered PFC achieved renal-specific distribution and largely ameliorated renal ischemia/reperfusion injuries.

78x59mm (300 x 300 DPI)