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Design and synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones with C-3' side chains as potent Met kinase inhibitors†

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Abstract

Pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold **4** with various C-3' side chains were designed as potent Met kinase inhibitors. Structural optimization led to compounds **10**, **20**, and **22–24** that demonstrated subnanomolar IC₅₀ values in the biochemical assay. The potent compound **20** inhibited Met with IC₅₀ value of 0.37 nM and the proliferation of MKN45 cells with IC₅₀ of 0.22 μ M. It suppressed Met autophosphorylation with the downstream signaling through Gab-1, PLC- γ , FAK, Akt, STAT3, and ERK in cell. Complete inhibition of STAT3 and ERK phosphorylation was observed in MKN45 cells treated with **20** at the concentration of 100 nM. A computation simulation study was performed to reveal the interaction of **20** with Met.

Introduction

Met is a transmembrane receptor tyrosine kinase expressed in normal and malignant cells. Upon binding to its natural ligand hepatocyte growth factor/scatter factor (HGF/SF), Met induces several signaling pathways responsible for cell proliferation, motility, migration, and survival.^{1–3} The cellular events play an important role in normal embryonic development and wound healing⁴ nevertheless overexpression of HGF and/or Met or activating mutation of Met can lead to cancer.⁵ As a result, Met is considered as a potential target for cancer therapy.^{6–8}

C-5 substituted pyrrole–indolin-2-ones SU11274 (1)⁹ and PHA665752 (2)¹⁰ are the prototype Met kinase inhibitors from in silico design before the three dimensional structure of Met was characterized (Figure 1). They are obtained using the homology model of Met built from the closely related FGFR1 kinase.¹¹ The C-4' amino–amido side chains of **1** and **2** were designed to modulate the pharmacokinetic properties¹² and would provide additional interactions with the enzyme to enhance the activities. Though **1** and **2** are only used as the biology tool, subsequent scaffold hopping based on the structure of **2** led to PF-2341066 (**3**),¹³ the most advanced Met and ALK kinase inhibitor recently approved by FDA for the treatment of NSCLC (see Figure 1).

Figure 1

Based on the literatures reporting the discovery of pyrrole–indolin-2-ones as multikinase inhibitors for VEGFR-2, FGFR1, and PDGFR- β kinases, different inhibition profiles are observed with different positions of the side chains.^{14,15} For FGFR1 inhibition, moving the C-4' side chain in a pyrrole–indolin-2-one to the C-3' position provided compounds with 5- to 10-fold increased potency (see the structures of 1 and 2 in Figure 1). As compound 2 was designed from the Met homology model of FGFR1, we envisaged that pyrrole–indolin-2-ones with C-3' side chains would also be potent Met inhibitors. Currently, the role of C-3' side chain for Met inhibition has not been studied in detail.

In this study, we used pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-one of scaffold **4** as the template to study the role of C-3' substituents for the inhibition of Met kinase. Except for the C-3' substituents, scaffold **4** possesses a pyrrole-fused cyclohexanone moiety for patent purposes.¹⁶

Subsequent optimization led to compounds 10, 20, and 22–24 that demonstrated subnanomolar IC_{50} values for Met inhibition in the biochemical assay. A computation modeling was used to reveal the different interactions of the C-3' side chain of 4 with Met in comparison with the C-4' side chain of 2. We expected the structural information would contribute to the further design of new scaffolds against Met, as in the case of designing 3 from 1 and 2.

Results and discussion

Chemistry

Scheme 1 presents the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7–12 of scaffold **4** bearing C-3' carboxyl-, ethoxycarbonyl-, and amido-ethyl side chains ($X = CO_2H$, CO_2Et , and $CONR^2R^3$ in **4**, Figure 1). 2-Formylpyrrole **5**, prepared according to our published procedures,¹⁷ was condensed with 5-[(2,6-dichlorobenzyl)sulfonyl]indolin-2-one (**6**) to provide compound **7** bearing a C-3' carboxylethyl group in 89% yield. Esterification of **7** in EtOH in the presence of SOCl₂ provided **8** that possessed a C-3' ethoxycarbonylethyl group in 92% yield. Amidation of the carboxyl group in **7** by various amines using CDI as the coupling agent provided pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones **9–12** bearing a C-3' amido-ethyl group in 70–85% yields.

Scheme 1

For the synthesis of pyrrole–indolin-2-ones **20–24** bearing C-3' aminopropyl side chains (X = $CH_2NR^2R^3$ in **4**, Figure 1), pyrroles **13a–e** served as the starting material to react with phenylsulfonyl chloride (PhSO₂Cl) in the presence of NaOH to provide the N¹-protected pyrrole **14a–e** in 86–94% yields (Scheme 2). The carbonyl groups in **14a–e** could be reacted with ethane-1,2-dithiol to give the dithiane **15a–e** in 87–99% yields. Removal of phenylsulfonyl group in **15a–e** under basic condition provided **16a–e** in 96% to >99% yields, which was treated with LiAlH₄ in THF at room temperature to afford amine **17a–e** in 60–96% yields. Introduction of a formyl group to the C-2 position of **17a–e** was accomplished by use of Vilsmeier-Haack reaction to generate **18a–e** in 71–97% yields. Deprotection of the dithianyl group using HgCl₂ afforded **19a–e** in 66–93% yields, which was eventually condensed with 5-(2,6-dichlorobenzylsulfonyl)indolin-2-one (**6**) to

provide the corresponding 20–24 with aminopropyl groups at their C-3' positions in 60–78% yields.

Scheme 2

In vitro potency

The IC₅₀ values of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7–12 and 20–24 for the inhibition of Met kinase and the proliferation of MKN45 cells (gastric carcinoma) that carry an amplified Met proto-oncogene¹⁸ are presented in Table 1. We also tested the potencies of 7-12 and **20–24** for the inhibition of Aurora kinases for comparison due to pyrrole–indolin-2-ones possessing a C-5 bulky group and a C-3 carboxylethyl side chain were found to show Aurora kinase inhibitory study.¹⁷ activities in our previous As shown in Table 1, the prototype pyrrole-5-(2,6-dichlorobenzyl)sulfonylindolin-2-one 7 bearing a C-3' carboxyethyl group displayed an IC₅₀ value of 30.40 nM for Met inhibition. However, it did not show significant antiproliferative activity for MKN45 cells (IC₅₀ > 10 μ M). Compound 8, the ethyl ester analog to 7, showed a slightly improved potency (IC₅₀: 20.20 nM) as well as enhanced antiproliferative activity for MKN45 (IC₅₀: 3.25μ M). These two compounds did not significantly inhibit Aurora kinases expect 7 inhibited Aurora A with an IC_{50} value of 855 nM.

Table 1

For further improving the Met and MKN45 inhibitory activities, we evaluated compounds 9-12 with the C-3' amidoethyl substituents. These compounds showed remarkable improvements for Met inhibition with IC₅₀ values less than 10 nM (IC₅₀ for **2**: 9 nM).¹⁹ The most potent amido compound **10** with a *N*,*N*-diethylaminoethylamino (Et₂NCH₂CH₂NH–) substituent inhibited Met at subnanomolar concentration (IC₅₀: 0.15 nM) and showed antiproliferative activity for MKN45 cells at low micromolar concentration (IC₅₀: 2.34 μ M). The *N*-methylpiperazinyl analog **12** displayed the most potent antiproliferative activity (MKN45 IC₅₀: 0.99 μ M) among the amido compounds **9–12**. Respective comparison of kinase and cell activities of **9** with **11** and **10** with **12** suggested that the NH group in the amido moiety would profit the enzymatic but not the cell activity. The difference in the cell activities of these compounds might come from their different cell permeability or inhibition profiles against other kinases, e.g., VEGFR-2, PDGFR, and FGFR1.¹⁵

Pyrrole-5-(2,6-dichlorobenzyl)sulforylindolin-2-ones 20-24 bound with C-3' aminopropyl groups showed enhanced activities both for Met kinase and MKN45 cells (Table 1). Except for 21 with a diethylamino group, these compounds inhibited Met at subnanomolar concentrations (IC_{50} : 0.26–0.43 nM) and demonstrated better antiproliferative activities than their carboxyl, ester, and amido analogs (i.e. 7–12) on MKN45 cells with IC_{50} values less than 1.0 μ M. The promising results from the biochemical shown in Table 1 indicated assay that pyrrole-5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold 4 with C-3' side chains also showed promising Met kinase inhibitory activity as their C-4' analog 2. Furthermore, the use of the C-3' ethyl ester, amido, and amino side chains also diminished the activity of the corresponding compounds against Aurora A and Aurora B kinases ($IC_{50} > 1.0 \mu M$, Table 1) in comparison with the use of a C-3' carboxyl side chain (compound 7). A great selectivity (~100- to 1000-fold) was thus achieved for Met over Aurora kinases for the compounds in Table 1.

Western blotting

As shown in Table 1, the great enhancement for the Met inhibition by compounds **20** and **22–24** did not reflect on their antiproliferative activities (MKN45 IC₅₀: 0.22–0.82 μ M). We then characterized the effect of potent compound **20** (MKN45 IC₅₀: 0.22 μ M) on Met signal transduction in MKN45 cells by Western blot, and the results are presented in Figure 2. Strong inhibition of tyrosine phosphorylation in the Met kinase domain (pY1234/1235) was observed with **20** at a concentration of 1.1 μ M and the reduced level of phosphorylated Met (IC₅₀ ~ 0.1–1.1 μ M) correlated to the antiproliferative IC₅₀ value of **20** (0.22 μ M). The phosphorylation of Met docking protein Gab-1 was completely inhibited at the concentration of 0.4 μ M. Compound **20** also potently inhibited the evidence of Met inhibition by **20** in cell. Regarded as the downstream cellular effectors of Met inhibition, the phosphorylation of STAT3 and ERK in MKN45 cells was completely abolished by **20** at the concentration below 100 nM, showing better correlation to the results from biochemical assay than the antiproliferative activity.

Figure 2

Kinase profiling

Since compound **20** shared a similar scaffold with the multikinase inhibitor sunitinib,¹⁴ we explored its inhibition profile against a panel of 451 kinases at the concentration of 1.0 μ M on DiscoveRx Kinomescan scanMax platform.²⁰ The results revealed that CAMKK1, CDKL2, CSF1R, DCAMKL3, FLT3 (D835Y), FLT3 (ITD), GRK1, GRK7, IRAK4, JAK2 (JH1domain-catalytic), MAP4K2, MAP4K3, MAP4K4, MAP4K5, MET, MET(M1250T), MET(Y1235D), MST2, PDGFRB, RSK3 (Kin.Dom.1-N-terminal), TAOK1, TAOK2, TAOK3, TNIK, TRKB, and YSK4 kinases were inhibited significantly by **20** with a percentage of control of < 10% (see the table in the Supporting Information).

Molecular modeling

To explore the interaction of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold **4** with Met, we modeled the binding mode of the potent compound **20** as well as reference compound **2** in Met (PDB code 2RFS)¹² using docking simulations. After energy minimization, the binding poses of **20** and **2** were overlapped as illustrated in Figure 3. Obviously, **20** shared similar binding pose to **2** in Met. Both compounds formed two H-bonds from N¹–H and N¹⁷-H with Pro1158 and Met1160, respectively, and an H-bond from C²=O with Met1160 in the hinge region. The oxygen atom of the C-5 sulfonyl group formed an H-bond with Asp1222, and the 2,6-dichlorophenyl fitted the hydrophobic pocket formed by Val1092, Tyr1230, and Gly1085 from the analysis of Ligplot.²¹ However, the side chain at the C-3' position of **20** headed to the different region from the C-4' amino–amido group in **2**, and thus interacted with different residues in Met other than **2**: **20** interacted with Ile1084 and **2** interacted with Tyr1159 and His1094 (Figure 3). The hydrophilic interaction of the C-3' dimethylamino group might contribute the promising activity of **20**.

Figure 3

Conclusions

Pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones of scaffold **4** with a C-3' side chain was designed as potent Met kinase inhibitors. Structural optimization led to compounds **10**, **20**, and **22–24** that showed subnanomolar IC_{50} values in the biochemical assay. Computation modeling

revealed that the C-3' side chain of **4** interacted with Ile1084 and would contribute to its promising activity. The potent compound **20** possessed an IC₅₀ value of 0.37 nM also inhibited the proliferation of MKN45 cells (IC₅₀: 0.22 μ M). Cellular events including the inhibition of Met autophosphorylation and the downstream signaling through Gab-1, PLC- γ , FAK, Akt, STAT-3, and ERK were found on MKN45 cells treated with **20**. Among the proteins, STAT3 and ERK were completely inhibited by **20** at the concentration of 100 nM.

Experimental

Chemistry

Reagents used purchased without further purification. were as 5-(2,6-Dichlorobenzyl)sulfonylindolin-2-one (6) was prepared according to literature procedures.^{22,23} Proton NMR spectra were recorded on a Bruker (500 MHz) spectrometer with CDCl₃ or DMSO- d_6 as solvent. Multiplicities are abbreviated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; J, coupling constant (hertz). Carbon-13 NMR spectra were obtained from a Bruker spectrometer (125 MHz) by use of CDCl₃ or DMSO- d_6 as solvent. Carbon-13 chemical shifts are referenced to the center peak of CDCl₃ (δ 77.0 ppm) or DMSO-d₆ (δ 39.5 ppm). ESI-MS spectra were recorded with an Applied Biosystems API 300 mass spectrometer. The purities of the compounds were greater than 95% as determined from reversed-phase HPLC. The synthesis of compounds 5, 13a-e, 17a-e, 18a-e, and 19a-e was followed our previously reported procedures.¹⁷

General procedure for the synthesis of *N*-phenysulfonylpyrroles 14a–e. A CH₂Cl₂ solution (250 mL) containing NaOH suspension (0.10 mol, 5.0 equiv) was added with compound 13a–e (~0.020 mol, 1.0 equiv). The mixture was stirred at 0 °C for 30 min. A solution of phenylsulfonyl chloride (0.030 mol, 1.5 equiv) in CH₂Cl₂ (50 mL) was slowly added to the reaction mixture in a period of 30 min. After the addition was completed, the reaction mixture was stirred for additional 30 min at 0 °C and for 12 h at room temperature. The solution was added with H₂O (100 mL) and the organic layer was separated. The aqueous layer was re-extracted with CH₂Cl₂ (3 × 50 mL), and the combined organic layer was dried over MgSO₄(s) and concentrated under reduced pressure to give the desired 14a–e in 86–94% yields.

N,*N*-Dimethyl-3-[4-oxo-1-(phenylsulfonyl)-4,5,6,7-tetrahydro-1*H*-indol-3-yl]propanamide (14a). Yield: 94%; ¹H NMR (CDCl₃) δ 7.85 (d, *J* = 7.6 Hz, 2 H), 7.66 (t, *J* = 7.5 Hz, 1 H), 7.55–7.58 (m, 2 H), 7.06 (s, 1 H), 3.02 (s, 3 H), 2.89–2.98 (m, 7 H), 2.56–2.61 (m, 2 H), 2.36–2.41 (m, 2 H), 2.02–2.09 (m, 2 H); ESI-MS *m*/*z* 374.7 (M + H)⁺.

N,*N*-Diethyl-3-[4-oxo-1-(phenylsulfonyl)-4,5,6,7-tetrahydro-1*H*-indol-3-yl]propanamide (14b). Yield: 93%; ¹H NMR (CDCl₃) δ 7.85 (d, *J* = 7.6 Hz, 2 H), 7.66 (t, *J* = 7.5 Hz, 1 H), 7.54–7.57 (m, 2 H), 7.06 (s, 1 H), 3.29–3.38 (m, 4 H), 2.96 (t, *J* = 6.8 Hz, 4 H), 2.54–2.59 (m, 2 H), 2.37–2.43 (m, 2 H), 2.03–2.10 (m, 2 H), 1.07–1.13 (m, 6 H); ESI-MS *m/z* 402.9 (M + H)⁺.

3-[3-Oxo-3-(pyrrolidin-1-yl)propyl]-1-(phenylsulfonyl)-6,7-dihydro-1*H***-indol-4(5***H***)-one (14c). Yield: 89%; ¹H NMR (CDCl₃) δ 7.85 (d,** *J* **= 7.5 Hz, 2 H), 7.65 (t,** *J* **= 7.5 Hz, 1 H), 7.53–7.56 (m, 2 H), 7.06 (s, 1 H), 3.38–3.45 (m, 4 H), 2.92–2.97 (m, 4 H), 2.50–2.55 (m, 2 H), 2.34–2.41 (m, 2 H), 2.02–2.09 (m, 2 H), 1.85–1.92 (m, 2 H), 1.77–1.84 (m, 2 H); ESI-MS** *m/z* **400.8 (M + H)⁺.**

3-(3-Morpholino-3-oxopropyl)-1-(phenylsulfonyl)-6,7-dihydro-1*H***-indol-4(5***H***)-one (14d).** Yield: 89%; ¹H NMR (CDCl₃) δ (d, *J* = 7.6 Hz, 2 H), 7.67 (t, *J* = 7.5 Hz, 1 H), 7.56–7.59 (m, 2 H), 7.07 (s, 1 H), 3.55–3.65 (m, 8 H), 2.90–2.99 (m, 4 H), 2.57–2.62 (m, 2 H), 2.37–2.42 (m, 2 H), 2.04–2.11 (m, 2 H); ESI-MS *m/z* 417.0 (M + H)⁺.

3-[3-(4-Methylpiperazin-1-yl)-3-oxopropyl]-1-(phenylsulfonyl)-6,7-dihydro-1*H***-indol-4(5***H***)-one (14e). Yield: 86%; ¹H NMR (CDCl₃) δ 7.86 (d,** *J* **= 7.5 Hz, 2 H), 7.67 (t,** *J* **= 7.5 Hz, 1 H), 7.56–7.59 (m, 2 H), 7.06 (s, 1 H), 3.50–3.64 (m, 4 H), 2.90–2.98 (m, 4 H), 2.57–2.62 (m, 2 H), 2.33–2.41 (m, 6 H), 2.28 (s, 3 H), 2.04–2.10 (m, 2 H) ESI-MS:** *m/z* **430.0 (M + H)⁺.**

General procedure for the synthesis of dithianes 15a–e. To a methanol solution containing compound 14a–e (~0.20 mol, 1.0 equiv) and ethane-1,2-dithiol (1.1 equiv) was added BF₃·OEt₂ (0.25 equiv). The reaction mixture was stirred at room temperature for 45 min. The solution was mixed with 10% aqueous NaOH (50 mL) and concentrated under reduced pressure to remove the methanol. The solution was extracted with CH₂Cl₂ (3 × 200 mL), and the organic layer was washed with H₂O, dried over MgSO₄(s), and concentrated under reduced pressure to give the desired 15a–e in 87–99% yields.

N,*N*-Dimethyl-3-[1'-(phenylsulfonyl)-1',5',6',7'-tetrahydrospiro[[1,3]dithiolane-2,4'-indole]-3'-y l]propanamide (15a). Yield: 87%; ¹H NMR (CDCl₃) δ 7.77 (d, *J* = 7.5 Hz, 2 H), 7.62 (t, *J* = 7.5 Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.42–3.50 (m, 2 H), 3.30–3.36 (m, 2 H), 3.06–3.11 (m, 2 H), 3.02 (s, 3 H), 2.97 (s, 3 H), 2.61–2.70 (m, 4 H), 2.15–2.19 (m, 2 H), 1.80–1.89 (m, 2 H); ESI-MS *m/z* 450.8 (M + H)⁺.

N,*N*-Diethyl-3-[1'-(phenylsulfonyl)-1',5',6',7'-tetrahydrospiro[[1,3]dithiolane-2,4'-indole]-3'-yl] propanamide (15b). Yield: 92%; ¹H NMR (CDCl₃) δ 7.77 (d, *J* = 7.6 Hz, 2 H), 7.61 (t, *J* = 7.4 Hz, 1 H), 7.50–7.54 (m, 2 H), 6.99 (s, 1 H), 3.44–3.51 (m, 2 H), 3.37–3.43 (m, 2 H), 3.30–3.36 (m, 4 H), 3.07–3.13 (m, 2 H), 2.61–2.69 (m, 4 H), 2.15–2.19 (m, 2 H), 1.82–1.88 (m, 2 H), 1.10–1.20 (m, 6 H); ESI-MS *m/z* 478.8 (M + H)⁺.

3-[1'-(Phenylsulfonyl)-1',5',6',7'-tetrahydrospiro[[1,3]dithiolane-2,4'-indole]-3'-yl]-1-(pyrrolidi n-1-yl)propan-1-one (15c). Yield: 90%; ¹H NMR (CDCl₃) δ 7.74–7.80 (m, 2 H), 7.62 (t, *J* = 7.5 Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.39–3.53 (m, 6 H), 3.28–3.38 (m, 2 H), 3.05–3.15 (m, 2 H), 2.57–2.68 (m, 4 H), 2.12–2.22 (m, 2 H), 1.92–2.00 (m, 2 H), 1.79–1.90 (m, 4 H); ESI-MS *m/z* 477.0 (M + H)⁺.

1-Morpholino-3-[1'-(phenylsulfonyl)-1',5',6',7'-tetrahydrospiro[[1,3]dithiolane-2,4'-indole]-3'-y I]propan-1-one (15d). Yield: 99%; ¹H NMR (CDCl₃) δ 7.77 (d, *J* = 7.4 Hz, 2 H), 7.62 (t, *J* = 7.4 Hz, 1 H), 7.51–7.54 (m, 2 H), 6.99 (s, 1 H), 3.61–3.69 (m, 6 H), 3.42–3.52 (m, 4 H), 3.31–3.38 (m, 2 H), 3.07–3.12 (m, 2 H), 2.61–2.71 (m, 4 H), 2.15–2.19 (m, 2 H), 1.83–1.87 (m, 2 H); ESI-MS *m/z* 492.9 (M + H)⁺.

1-(4-Methylpiperazin-1-yl)-3-[1'-(phenylsulfonyl)-1',5',6',7'-tetrahydrospiro[[1,3]dithiolane-2,4 '-indole]-3'-yl]propan-1-one (15e). Yield: 96%; ¹H NMR (CDCl₃) δ 7.77 (d, *J* = 7.5 Hz, 2 H), 7.62 (t, *J* = 7.5 Hz, 1 H), 7.50–7.53 (m, 2 H), 6.99 (s, 1 H), 3.64–3.68 (m, 2 H), 3.50–3.54 (m, 2 H), 3.42–3.49 (m, 2 H), 3.30–3.37 (m, 2 H), 3.07–3.12 (m, 2 H), 2.61–2.72 (m, 4 H), 2.36–2.40 (m, 4 H), 2.31 (s, 3 H), 2.16–2.20 (m, 2 H), 1.82–1.88 (m, 2 H); ESI-MS *m/z* 506.0 (M + H)⁺.

General procedure for the synthesis of pyrroles 16a–e. Compounds 15a–e (1.0 equiv) in a solution of 30% MeOH and 70% H₂O were added with NaOH (5.0 equiv). The reaction mixture was

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heated at reflux for 3.0 h. The solution was neutralized with 3.0 N HCl and the resulting precipitate was collected to provide the desired **16a–e** in 96% to >99% yields. The spectroscopic data of were consistence with our previous report.¹⁷

General procedure for the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7 and 20–24. 2-Formylpyrroles 5 or 19a–e (~0.50 mmol, 1.0 equiv) was mixed with 5-(2,6-dichlorobenzyl)sulfonylindolin-2-one (6, 1.0 equiv) and piperidine (0.10 mL) in EtOH (5.0 mL). The reaction mixture was heated at reflux for 12 h. The resulting precipitate was filtered and washed with EtOH to provide the corresponding pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 7 and 20–24 in 60–89% yields.

(*Z*)-3-(2-{[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl}-4-oxo-4,5,6,7-tetrahy dro-1*H*-indol-3-yl)propanoic Acid (7). Yield: 89%; ¹H NMR (DMSO-*d*₆) δ 13.65 (s, 1 H), 12.11 (brs, 1 H), 11.52 (s, 1 H), 8.21 (d, *J* = 1.6 Hz, 1 H), 7.94 (s, 1 H), 7.46–7.52 (m, 3 H), 7.39 (dd, *J* = 8.7, 7.4 Hz, 1 H), 7.06 (d, *J* = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.20 (t, *J* = 7.6 Hz, 2 H), 2.93 (t, *J* = 6.1 Hz, 2 H), 2.52 (t, *J* = 7.6 Hz, 2 H), 2.40–2.44 (m, 2 H), 2.02–2.10 (m, 2 H); ¹³C NMR (DMSO-*d*₆) δ 19.28, 21.83, 21.91, 33.87, 37.50, 56.89, 108.78, 114.70, 118.27, 118.77, 124.91, 124.99, 125.43, 126.16, 126.53, 127.81, 130.16, 131.02, 133.03, 135.51, 142.09, 147.59, 168.70, 172.97, 192.83; ESI-MS *m*/*z* 573.0 (M + H)⁺.

(*Z*)-5-(2,6-Dichlorobenzylsulfonyl)-3-({3-[3-(dimethylamino)propyl]-4-oxo-4,5,6,7-tetrahydro-1 *H*-indol-2-yl}methylene)indolin-2-one (20). Yield: 70%; ¹H NMR (DMSO-*d*₆) δ 13.51 (s, 1 H), 11.46 (brs, 1 H), 7.97 (s, 1 H), 7.42–7.57 (m, 3 H), 7.34–7.42 (m, 2 H), 7.07 (d, *J* = 6.9 Hz, 1 H), 4.87 (s, 2 H), 2.92–3.05 (m, 4 H), 2.35–2.45 (m, 4 H), 1.97–2.19 (m, 8 H), 1.57–1.72 (m, 2 H); ¹³C NMR (DMSO-*d*₆) δ 19.73, 21.84, 21.96, 26.58, 37.56, 43.98, 56.37, 56.86, 108.88, 114.08, 117.69, 118.80, 124.90, 124.95, 125.04, 125.77, 126.62, 127.83, 130.17, 130.82, 134.51, 135.52, 142.03, 147.73, 168.67, 192.72; ESI-MS *m/z* 586.0 (M + H)⁺.

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-({3-[3-(diethylamino)propyl]-4-oxo-4,5,6,7-tetrahydro-1*H* -indol-2-yl}methylene)indolin-2-one (21). Yield: 78%; ¹H NMR (DMSO-*d*₆) δ 13.62 (s, 1 H), 8.11 (s, 1 H), 7.89 (s, 1 H), 7.43–7.56 (m, 3 H), 7.37–7.42 (m, 1 H), 7.07 (d, *J* = 8.10 Hz, 1 H), 4.87 (s, 2

- 10 -

H), 2.90–3.02 (m, 4 H), 2.32–2.47 (m, 8 H), 2.01–2.12 (m, 2 H), 1.65–1.71 (m, 2 H), 0.91 (t, J = 7.1 Hz, 6 H); ¹³C NMR (DMSO- d_6) δ 10.36, 21.02, 21.85, 21.98, 26.59, 37.59, 44.85, 50.24, 56.88, 108.86, 114.10, 117.77, 118.83, 124.97, 125.12, 125.48, 126.33, 126.51, 127.81, 130.15, 130.95, 134.42, 135.50, 142.04, 147.72, 168.70, 192.67; ESI-MS *m*/*z* 614.0 (M + H)⁺.

(*Z*)-5-(2,6-Dichlorobenzylsulfonyl)-3-({4-oxo-3-[3-(pyrrolidin-1-yl)propyl]-4,5,6,7-tetrahydro-1 *H*-indol-2-yl}methylene)indolin-2-one (22). Yield: 66%; ¹H NMR (DMSO-*d*₆) δ 13.62 (s, 1 H), 11.52 (s, 1 H), 8.05 (s, 1 H), 7.86 (s, 1 H), 7.45–7.50 (m, 3 H), 7.34–7.41 (m, 1 H), 7.05 (d, *J* = 8.2 Hz, 1 H), 4.85 (s, 2 H), 3.03 (t, *J* = 6.8 Hz, 2 H), 2.93 (t, *J* = 5.9 Hz, 2 H), 2.27–2.42 (m, 8 H), 2.01–2.09 (m, 2 H), 1.63–1.73 (m, 6 H); ¹³C NMR (DMSO-*d*₆) δ 20.92, 21.84, 21.97, 22.17, 28.05, 37.58, 52.45, 53.26, 56.87, 108.87, 114.18, 117.66, 118.78, 124.93, 124.99, 125.04, 126.40, 126.51, 127.81, 130.17, 130.93, 134.71, 135.52, 142.03, 147.70, 168.70, 192.70; ESI-MS *m*/*z* 612.0 (M + H)⁺.

(*Z*)-5-(2,6-Dichlorobenzylsulfonyl)-3-{[3-(3-morpholinopropyl)-4-oxo-4,5,6,7-tetrahydro-1*H*-in dol-2-yl]methylene}indolin-2-one (23). Yield: 72%; ¹H NMR (DMSO-*d*₆) δ 13.64 (s, 1 H), 11.48 (brs, 1 H), 8.21 (s, 1 H), 7.88 (s, 1 H), 7.46–7.50 (m, 3 H), 7.33–7.41 (m, 1 H), 7.05 (d, *J* = 8.2 Hz, 1 H), 4.86 (s, 2 H), 3.51–3.57 (m, 4 H), 3.02 (t, *J* = 7.1 Hz, 2 H), 2.93 (t, *J* = 6.0 Hz, 2 H), 2.21–2.42 (m, 8 H), 1.99–2.09 (m, 2 H), 1.67–1.74 (m, 2 H); ¹³C NMR (DMSO-*d*₆) δ 21.12, 21.84, 21.98, 26.07, 37.58, 52.34, 56.35, 56.90, 65.22, 108.81, 114.20, 118.05, 118.84, 124.94, 125.02, 125.10, 126.34, 126.46, 127.81, 130.16, 131.03, 134.94, 135.52, 142.02, 147.70, 168.72, 192.70; ESI-MS *m*/*z* 628.0 (M + H)⁺.

(Z)-5-(2,6-Dichlorobenzylsulfonyl)-3-({3-[3-(4-methylpiperazin-1-yl)propyl]-4-oxo-4,5,6,7-tetra hydro-1*H*-indol-2-yl}methylene)indolin-2-one (24). Yield: 60%; ¹H NMR (DMSO- d_6) δ 13.67 (s, 1 H), 11.52 (brs, 1 H), 8.20 (s, 1 H), 7.85 (s, 1 H), 7.46–7.51 (m, 3 H), 7.37–7.42 (m, 1 H), 7.06 (d, J = 8.2 Hz, 1 H), 4.88 (s, 2 H), 2.91–3.03 (m, 4 H), 2.22–2.44 (m, 10 H), 2.01–2.14 (m, 7 H), 1.70 (d, J = 6.8 Hz, 2 H); ¹³C NMR (DMSO- d_6) δ 21.22, 21.84, 21.97, 26.59, 37.59, 44.66, 51.70, 53.72, 56.01, 56.95, 108.78, 114.22, 118.13, 118.83, 124.91, 125.06, 126.31, 126.48, 127.81, 130.16, 130.96, 135.03, 135.51, 142.00, 143.92, 147.67, 168.72, 192.68; ESI-MS *m/z* 641.0 (M + H)⁺.

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(*Z*)-Ethyl **3-(2-{[5-(2,6-dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl}-4-oxo-4,5,6,7-tetrahydro-1***H***-indol-3-yl)propanoate (8). To a solution of 7** (105.5 mg, 0.1840 mmole) in EtOH (2.0 mL) was added with SOCl₂ (0.10 mL). The reaction mixture was heated at reflux for 12 h. The solution was concentrated under reduced pressure. The residue was added with H₂O (5.0 mL) and heated. The solution was cooled to room temperature and the resultant solids were collected to give the desired **8** (101.8 mg, 0.1692 mmol) in 92% yield: ¹H NMR (DMSO-*d*₆) δ 13.68 (s, 1 H), 11.53 (brs, 1 H), 8.27 (d, *J* = 1.1 Hz, 1 H), 7.95 (s, 1 H), 7.46–7.53 (m, 3 H), 7.34–7.42 (m, 1 H), 7.06 (d, *J* = 8.2 Hz, 1 H), 4.88 (s, 2 H), 4.02 (q, *J* = 7.1 Hz, 2 H), 3.25 (t, *J* = 7.4 Hz, 2 H), 2.94 (t, *J* = 6.0 Hz, 2 H), 2.61 (t, *J* = 7.4 Hz, 2 H), 2.38–2.45 (m, 2 H), 2.01–2.11 (m, 2 H), 1.13 (t, *J* = 7.1 Hz, 3 H); ¹³C NMR (DMSO-*d*₆) δ 13.13, 19.12, 21.84, 21.88, 33.49, 37.47, 56.90, 58.83, 108.81, 114.87, 118.32, 118.77, 124.93, 124.99, 125.34, 126.23, 126.63, 127.81, 130.14, 131.08, 132.42, 135.50, 142.11, 147.61, 168.68, 171.27, 192.87; ESI-MS *m/z* 601.0 (M + H)⁺.

General procedure for the synthesis of pyrrole–5-(2,6-dichlorobenzyl)sulfonylindolin-2-ones 9–12. Compound 7 (~1.0 g, 1.0 equiv) was suspended in CH_2Cl_2 (20 mL) and mixed with CDI (1.2 equiv). The reaction mixture was stirred at room temperature for 3.0 h. The solution was added with methylamine, *N*,*N*-diethylethane-1,2-diamine, pyrrolidine, or 1-methylpiperazine (2.0 equiv), and the reaction mixture was stirred at room temperature for 12 h. The solution was diluted with CH_2Cl_2 (200 mL), and washed with 0.1 N HCl (50 mL), saturated Na_2CO_3 (50 mL), and brine (50 mL). The organic layer was dried over anhydrous $MgSO_4(s)$ and concentrated under reduced pressure to provide the desired 9–12 as yellow solids in 70–85% yields.

(*Z*)-3-(2-{[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl}-4-oxo-4,5,6,7-tetrahy dro-1*H*-indol-3-yl)-*N*-methylpropanamide (9). Yield: 85%; ¹H NMR (DMSO- d_6) δ 13.61 (s, 1 H), 11.49 (brs, 1 H), 8.22 (d, *J* = 1.6 Hz, 1 H), 7.90 (s, 1 H), 7.72 (dd, *J* = 4.2, 4.2 Hz, 1 H), 7.46–7.53 (m, 3 H), 7.39 (dd, *J* = 8.7, 7.4 Hz, 1 H), 7.06 (d, *J* = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.22 (t, *J* = 7.3 Hz, 2 H), 2.94 (t, *J* = 6.1 Hz, 2 H), 2.48 (s, 3 H), 2.45–2.40 (m, 2 H), 2.37 (t, *J* = 7.3 Hz, 2 H), 2.03–2.10 (m, 2 H); ¹³C NMR (DMSO- d_6) δ 19.67, 21.85, 21.93, 24.44, 35.34, 37.54, 56.90, 108.80, 114.46, 118.06, 118.74, 125.00, 125.37, 126.23, 126.55, 127.81, 130.18, 131.10, 133.66, 135.51, 142.06,

147.57, 168.67, 170.75, 192.81; ESI-MS m/z 586.0 (M + H)⁺.

(*Z*)-3-(2-{[5-(2,6-Dichlorobenzylsulfonyl)-2-oxoindolin-3-ylidene]methyl}-4-oxo-4,5,6,7-tetrahy dro-1*H*-indol-3-yl)-*N*-[2-(diethylamino)ethyl]propanamide (10). Yield: 72%; ¹H NMR (DMSO- d_6) δ 13.60 (s, 1 H), 11.49 (brs, 1 H), 8.19 (s, 1 H), 7.89 (s, 1 H), 7.64 (d, *J* = 5.5 Hz, 1 H), 7.46–7.54 (m, 3 H), 7.35–7.43 (m, 1 H), 7.06 (d, *J* = 8.2 Hz, 1 H), 4.87 (s, 2 H), 3.18–3.27 (m, 2 H), 2.89–3.03 (m, 4 H), 2.40–2.45 (m, 2 H), 2.37 (t, *J* = 7.2 Hz, 2 H), 2.28 (q, *J* = 7.0 Hz, 4 H), 2.22 (t, *J* = 7.2 Hz, 2 H), 2.01–2.12 (m, 2 H), 0.77 (t, *J* = 7.0 Hz, 6 H); ¹³C NMR (DMSO- d_6) δ 10.56, 19.74, 21.85, 21.93, 35.47, 35.93, 37.55, 45.51, 50.64, 56.89, 108.76, 114.41, 117.94, 118.73, 124.93, 125.05, 125.49, 126.34, 126.47, 127.80, 130.17, 131.13, 133.50, 135.52, 142.07, 147.50, 168.69, 170.21, 192.79; ESI-MS *m/z* 671.0 (M + H)⁺.

(*Z*)-5-(2,6-Dichlorobenzylsulfonyl)-3-({4-oxo-3-[3-oxo-3-(pyrrolidin-1-yl)propyl]-4,5,6,7-tetrahy dro-1*H*-indol-2-yl}methylene)indolin-2-one (11). Yield: 82%; ¹H NMR (DMSO-*d*₆) δ 13.64 (s, 1 H), 11.44 (s, 1 H), 8.22 (d, *J* = 1.3 Hz, 1 H), 7.94 (s, 1 H), 7.46–7.51 (m, 3 H), 7.35–7.42 (m, 1 H), 7.05 (d, *J* = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.19–3.39 (m, 6 H), 2.94 (t, *J* = 5.9 Hz, 2 H), 2.52–2.57 (m, 2 H), 2.40–2.45 (m, 2 H), 2.03–2.09 (m, 2 H), 1.79–1.86 (m, 2 H), 1.68–1.76 (m, 2 H); ¹³C NMR (DMSO-*d*₆) δ 19.35, 21.85, 21.93, 22.97, 24.65, 34.28, 37.54, 44.32, 44.83, 56.86, 108.75, 114.49, 118.19, 118.85, 124.99, 125.06, 125.41, 126.19, 126.48, 127.82, 130.15, 131.04, 133.81, 135.51, 142.04, 147.58, 168.70, 168.75, 192.86; ESI-MS *m/z* 626.0 (M + H)⁺.

(*Z*)-5-(2,6-Dichlorobenzylsulfonyl)-3-({3-[3-(4-methylpiperazin-1-yl)-3-oxopropyl]-4-oxo-4,5,6,7 -tetrahydro-1*H*-indol-2-yl}methylene)indolin-2-one (12). Yield: 70%; ¹H NMR (DMSO-*d*₆) δ 13.66 (s, 1 H), 11.51 (brs, 1 H), 8.21 (d, *J* = 0.93 Hz, 1 H), 7.91 (s, 1 H), 7.47–7.50 (m, 3 H), 7.35–7.42 (m, 1 H), 7.05 (d, *J* = 8.2 Hz, 1 H), 4.88 (s, 2 H), 3.37–3.45 (m, 4 H), 3.20 (t, *J* = 7.6 Hz, 2 H), 2.94 (t, *J* = 6.0 Hz, 2 H), 2.57 (t, *J* = 7.6 Hz, 2 H), 2.43 (t, *J* = 6.0 Hz, 2 H), 2.14–2.17 (m, 4 H), 2.12 (s, 3 H), 2.04–2.09 (m, 2 H); ¹³C NMR (DMSO-*d*₆) δ 19.85, 21.86, 21.93, 32.63, 37.53, 39.90, 43.75, 44.65, 53.31, 53.76, 56.89, 108.76, 114.63, 118.27, 118.89, 124.99, 125.39, 126.23, 126.54, 127.83, 130.17, 131.05, 133.42, 135.51, 142.06, 147.62, 168.70, 169.02, 192.98; ESI-MS *m*/*z* 655.0 (M + H)⁺.

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Biological and computational methods

Met and Aurora kinase assays

Inhibition of kinase activity by the test compound was measured by quantifying the amount of ^{33}P incorporation into the substrate in the presence of a test compound. Standard assay conditions utilized 5 ng of recombinant Met kinase (Upstate, Cat. No. 14-526), 1 µg Poly Glu-Tyr (Sigma, Product Number P0275), 100 µM ATP, 0.2 µCi [^{33}P]ATP (specific activity 3000 Ci/mmol, PerkinElmer), 8 mM MOPS-NaOH (pH 7.0), and 1 mM EDTA in a total volume of 25 µL. Reaction mixtures were incubated at 30 °C for 30 min and reactions stopped by addition of 3% phosphoric acid; mixtures were harvested onto a 96-well GF/B UniFilter (PerkinElmer) using a unifilter harvester (PerkinElmer), and counted with a TopCount microplate scintillation counter (PerkinElmer). The IC₅₀ values of inhibitors were determined after assays were conducted at 3-fold serially diluted concentrations of each compound in duplicate. The results were analyzed using linear regression software (GraphPad Prism version 4; GraphPad Software Inc., San Diego, CA). For Aurora assay, 1 ng of recombinant Aurora A or Aurora B kinase (Upstate Biotechnology), 2 µg peptide substrate (tetra-LRRLSLG, synthesized by Genesis Biotech Inc.), 10 µM ATP (for Aurora A) or 40 µM ATP (for Aurora B), 0.2 µCi [^{33}P]ATP (specific activity 3000 Ci/mmol, PerkinElmer), 8 mM MOPS-NaOH (pH 7.0), and 1 mM EDTA in a total volume of 25 µL were utilized.

Cell proliferation assay

The antiproliferative activity of the compounds with respect to human gastric carcinoma cell, MKN45 (Japanese Collection of Research Bioresources), was measured using the CellTiter96 assay kit (Promega) following the manufacturer's instruction. In brief, the cells were maintained in DMEM containing 0.2% FBS and incubated at 37 °C in 5% CO₂ atmosphere. Cells were plated at a density of 2500 cells/well of a 96-well plate for 24 h, then treated with different concentrations of the compounds, and incubated for another 72 hours. At the end of the incubation, CellTiter 96 Aqueous One Solution Reagent (Promega) was added and incubated for another 3.0 hours. Cell viability was determined by measuring absorbance at 490 nm using EMax[®] microplate reader (Molecular Devices). Data were processed and analyzed using GraphPad Prism version 4.

Western blot analysis

The effect of Met inhibitors on cellular protein phosphorylation was evaluated by Western blot analysis on MKN45 cells. Cells grown to subconfluency were grown overnight in 0.5% (FBS) and DMEM. Cells in low serum were treated with compound 20 in DMSO (20 mM stock) for 30 minutes. Drug-treated cells were then harvested and washed twice with PBS, and the cell pellets were boiled in 2× SDS sample buffer and subjected to SDS-PAGE and immunoblotting analysis. Western blots were done using standard procedures and probed with the following antibodies: anti-Met (Cell Signaling Technology), anti-phospho-Met (Tyr1234/1235), anti-phospho-Gab-1 (Tyr627, Thermo Scientific), anti-phospho-FAK (Tyr861, Thermo Scientific), (Ser473), anti-phospho-PLC- γ (Tyr783), anti-phospho-Akt anti-phospho-STAT3 (Tyr705), anti-phospho-p44/42 ERK mitogen-activated protein kinase (Thr202/Tyr204), and horseradish peroxidase-conjugated secondary antibodies (Cell Signaling).

Computational method

The three dimensional structure of Met complexed with **1** was obtained from the Protein Data Bank (entry: 2RFS).¹² Hydrogen atoms were added, and water molecules cocrystallized with the protein were removed from the original structure using Accelrys Discovery Studio 2.5 (Accelrys, Inc.). The modified crystal structure of Met was docked by a genetic algorithm based program (GOLD) using the Chem scoring (CCDC Software Limited, Cambridge, U.K.). The genetic algorithm was executed at the default settings, and the active site radius is 10 Å from inhibitor complex for the docking study. Two hundred genetic algorithm (GA) runs were performed for docking compounds **2** and **20**. GOLD was run to save up to best docking solutions for each ligands. The results were visually analyzed using PyMOL program (http://www.pymol.org/) and Ligplot.²¹

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Note and references

*Electronic Supplementary Information (ESI) available: Ligplot diagrams of the ATP binding site of

Met complexed with compounds 2 and 20, and the kinase profiling data of 20. See DOI: xxx/xxx

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Table1Theenzymaticandcellularactivitiesof5-(2,6-dichlorobenzyl)sulfonylpyrrole-indolin-2-ones7-12and20-24withdifferentC-3'sidechains as Met kinase inhibitors



compd	Х	Met IC ₅₀ $(nM)^a$	MKN45 IC ₅₀ (μM) ^{a,b}	%inhibition at 1.0 µM	
				Aurora A	Aurora B
7	0 НО-Ё—ѯ	30.40	>10	65.5% ^c	9.4%
8	O EtO−C̈—ѯ	20.20	3.25 ± 0.41	22.7%	16.3%
9	O MeHN−C̈—ફ	4.22	>10	13.0%	6.9%
10	Et ₂ N HN-C	0.15	2.34 ± 0.13	10.8%	9.2%
11	O N-C-Ş	7.17	9.25 ± 0.32	2.2%	5.5%
12	-N_N-C-\$	2.94	0.99 ± 0.10	8.7%	1.3%
20	$\begin{array}{c} H_2\\ Me_2N-C^2-\xi\end{array}$	0.37	0.22 ± 0.03	2.7%	34.3%
21	H_2 Et ₂ N-C -{	7.70	0.40 ± 0.05	1.9%	6.2%
22	$\mathbf{N} - \mathbf{C}^2 - \mathbf{\xi}$	0.31	0.82 ± 0.08	4.2%	22.3%
23	0 N-C ⁻ {	0.43	0.47 ± 0.05	23.7%	35.4%
24	$-N$ $N-C^{2}-\xi$	0.26	0.24 ± 0.02	6.0%	15.3%
2	_	9^d	0.10 ± 0.03	_	_

^{*a*} The IC₅₀ values were averaged from two independent dose–response curves. ^{*b*} A 72-h cell proliferation assay. ^{*c*} IC₅₀: 855 nM. ^{*d*} Obtained from literature¹⁹.



Figure 1Structures of SU11274 (1), PHA665752 (2), PF-2341066 (3), and5-(2,6-dichlorobenzyl)sulfonylpyrrole-indolin-2-one of scaffold 4.



Figure 2 Effect of 20 on Met phosphorylation and signal transduction in MKN45 cells.



Figure 3 Overlapping of the docked binding modes of compounds **2** and **20** in the ATP binding site of Met kinase. Compound **2** is shown as light blue sticks, **20** is shown as yellow sticks, hydrogen bonds are shown as dashed gray line, and the oxygen, nitrogen, and sulfur atoms are colored in red, blue, and orange, respectively. The C-3' side chain of **20** faced the region around Ile1084 and the C-4' side chain of **2** faced the region around Tyr1159 and His1094.



Scheme 1 Synthesis of 5-(2,6-dichlorobenzyl)sulfonylpyrrole-indolin-2-ones 7-12 bearing C-3' carboxyl-, ethoxycarbonyl-, or amido-ethyl side chains as Met kinase inhibitors. Reagents and conditions: (a) piperidine, EtOH, reflux, 12 h, 89%; (b) SOCl₂, EtOH, 92%; (c) 7, amine, CDI, CH_2Cl_2 , 70-85%.



Scheme 2 Synthesis of 5-(2,6-dichlorobenzyl)sulfonylpyrrole–indolin-2-ones 20–24 equipped with C-3' aminopropyl side chain. Reagents and conditions: (a) PhSO₂Cl, NaOH, CH_2Cl_2 , 86–94%; (b) ethane-1,2-dithiol, BF₃·OEt₂, MeOH, 87–99%; (c) NaOH, H₂O, MeOH, reflux, 96% to >99%; (d) LiAlH₄, THF, r.t., 60–96%; (e) POCl₃, DMF, 71–97%; (f) HgCl₂, CaCO₃, CH₃CN, H₂O, 66–93%; (g) 5-(2,6-dichlorobenzylsulfonyl)indolin-2-one (6), piperidine, EtOH, reflux, 60–78%.

Graphical abstract

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