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## ARTICLE

# The Pentafluorosulfanyl Group in Cannabinoid Receptor Ligands: Synthesis and Comparison with Trifluoromethyl and *tert*-Butyl Analogues

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An array of cannabinoid ligands, bearing *meta*- and *para*-substituted pentafluorosulfanyl (SF<sub>5</sub>) aniline groups in position 3 of the pyrazole ring, was efficiently synthesised and compared with the exact trifluoromethyl and *tert*-butyl analogues. In general, the SF<sub>5</sub> substituted ligands showed higher lipophilicity (i.e. LogP values) than the CF<sub>3</sub> counterparts and lower than the *tert*-butyl ones. In terms of pharmacological activity, SF<sub>5</sub> pyrazoles generally showed slightly higher or equivalent CB<sub>1</sub> receptor affinity (K<sub>i</sub>), always in the nanomolar range, and selectivity vs. the CB<sub>2</sub> relative to both CF<sub>3</sub> and *tert*-butyl analogues. Functional β-arrestin recruitment assays were used to determine equilibrium dissociation constants (K<sub>b</sub>) and showed that all of the tested SF<sub>5</sub> and CF<sub>3</sub> compounds are CB<sub>1</sub> neutral antagonists. These results confirm the possibility of successfully using an aromatic SF<sub>5</sub> group as a stable, synthetically accessible and effective bioisosteric analogue of the electron-withdrawing CF<sub>3</sub> group, and possibly also of bulky aliphatic groups, for drug discovery and development applications.

## Introduction

### Pentafluorosulfanyl group

Although only a few fluorinated natural compounds have been isolated<sup>1</sup>, it is well known that introduction of one or more fluorine atoms into a molecule can have profound effects on the binding to a receptor, and improve both its metabolic stability and bioavailability<sup>2</sup>. Fluorine could be incorporated by fluorination or alternatively *via* building-block approach. Among the fluorinated motifs, the trifluoromethyl group occupies a prominent role in drug discovery<sup>3</sup>, and several blockbuster drugs display a CF<sub>3</sub> substituent<sup>4</sup>.

In 1960 Sheppard reported for the first time the synthesis of an aromatic compound bearing a pentafluorosulfanyl group, SF<sub>5</sub><sup>5</sup>. However, due to of the inconvenient synthetic access to pentafluorosulfanyl arenes, the breakthrough in the commercialization first and then in the application of SF<sub>5</sub>-compounds in drug discovery and materials science came in the late 90s, with the improvement of their synthesis.

The synthesis of pentafluorosulfanyl compounds, their biological applications and properties have been reviewed<sup>6–9</sup>. Importantly, the pentafluorosulfanyl group is often compared to the trifluoromethyl group, and because of its higher lipophilicity<sup>10</sup>, electronegativity<sup>11</sup>, chemical stability and greater steric demand, which is only slightly lower than that of

the *tert*-Butyl group<sup>12</sup>, the SF<sub>5</sub> group is often referred to as a “super-trifluoromethyl” group (Figure 1)<sup>13–15</sup>.

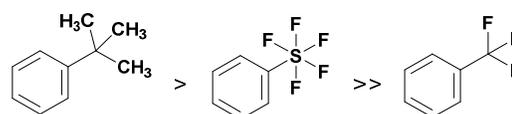


Figure 1: Steric demand of the three groups compared in this work: *t*-Bu > SF<sub>5</sub> >> CF<sub>3</sub>.

### Cannabinoid receptors

Cannabinoid receptors belong to the G-protein coupled receptors family (GPCRs)<sup>16,17</sup>. At least two cannabinoid receptor subtypes have been identified: CB<sub>1</sub> and CB<sub>2</sub><sup>18</sup>, furthermore, the CB<sub>1</sub> type has two splice variants, denominated CB<sub>1A</sub> and CB<sub>1B</sub><sup>19,20</sup>. The distribution of CB<sub>1</sub> receptors is localised predominantly in the brain<sup>21</sup> whereas the CB<sub>2</sub> are present in the peripheral nervous system (PNS) cells<sup>22</sup>. However, recent studies have demonstrated the presence of CB<sub>1</sub> in the PNS<sup>23</sup> and, on the other hand, of the CB<sub>2</sub> in the central nervous system, albeit in low density<sup>24</sup>. Since CB<sub>1</sub> receptors are associated with several disorders, such as depression<sup>25</sup>, anxiety<sup>26</sup>, stress<sup>27</sup>, schizophrenia<sup>28</sup>, chronic pain<sup>29</sup> and obesity<sup>30</sup>,

several cannabinoid ligands were developed. Among these ligands, the most studied is probably SR141716 (Rimonabant)<sup>31</sup>, a pyrazole-core inverse agonist which was discovered by Sanofi-Synthelabo (now Sanofi-Aventis) in 1994 (Figure 2), marketed in Europe as anti-obesity drug and subsequently withdrawn from the market owing to its side-effects.

The scientific question we wanted to address in this work was about the position occupied by the SF<sub>5</sub> group relative to its closest bioisosteric substituents, namely CF<sub>3</sub> and *tert*-butyl groups, in terms of its effect on key pharmacological and physico-chemical properties, such as lipophilicity and solubility. To answer, we decided to use a Rimonabant-type scaffold as a model bioactive structure for incorporating an SF<sub>5</sub>-group, as well as CF<sub>3</sub> and the *tert*-butyl groups, and directly compare the SF<sub>5</sub> derivatives with their CF<sub>3</sub> and *tert*-Bu counterparts from the pharmacological viewpoint. SF<sub>5</sub>, CF<sub>3</sub> or *tert*-butyl groups were incorporated on a carboxy-aniline residue in position 3 of the pyrazole ring, since (1) 3-carboxy-aniline Rimonabant analogues were shown to have excellent CB<sub>1</sub>-affinity and selectivity vs. the CB<sub>2</sub><sup>32</sup> and (2) SF<sub>5</sub>-substituted anilines or nitro-anilines are accessible starting materials (see below).

Since, to the best of our knowledge, SF<sub>5</sub>-substituted cannabinoid receptor ligands have never been described in the literature, we decided to synthesise two different classes of pyrazole-core CB<sub>1</sub> receptor ligands: the former based on a Rimonabant-type structure and the latter based on ligands described in a Pharmaness' patent<sup>33</sup>, where the 4-chloro-phenyl ring is replaced by a 2-bromo-thiophene.

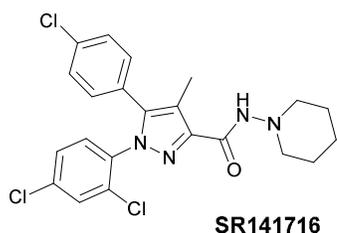
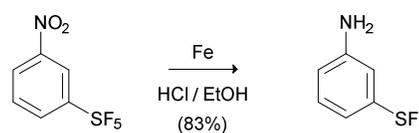


Figure 2: Chemical structure of Rimonabant (SR141716).

## Results and Discussion

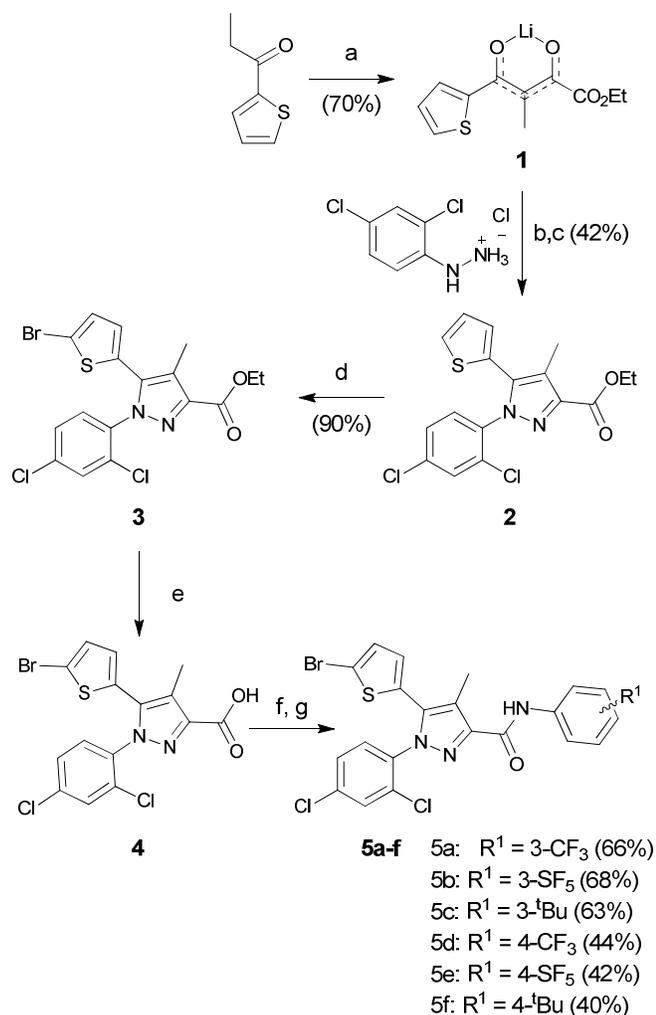
### Chemistry

The starting *para*-SF<sub>5</sub>-substituted aniline is commercially available whereas *meta*-SF<sub>5</sub>-aniline was synthesised from the commercially available 3-nitro-SF<sub>5</sub> benzene (Scheme 1). 3-Nitro-SF<sub>5</sub>-benzene was treated with iron-powder in a refluxing HCl/Ethanol solution<sup>34</sup>, affording the corresponding 3-(pentafluoro-λ<sup>6</sup>-sulfanyl)aniline in good yield (83%). The SF<sub>5</sub> group remained unreactive under these conditions, confirming the high chemical stability of this group.



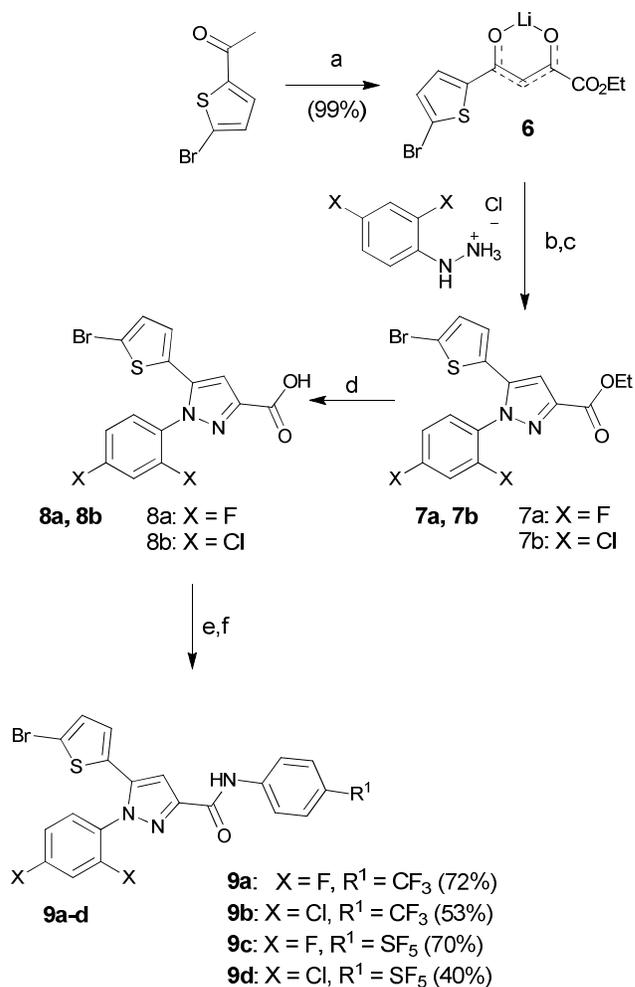
Scheme 1: Reagents and conditions: Fe, HCl/EtOH, reflux, 2 h.

Thiophenyl-compounds **5a-f** and **9a-d** were prepared according to the general synthetic methods shown in Schemes 2 and 3, respectively<sup>35</sup>. 4-Methyl-pyrazole-substituted compounds **5** were synthesised starting with a reaction of 2-propionyl-thiophene with diethyl oxalate using LHMDS as a base which provided the stable lithium salt **1** in moderate yields. The latter was allowed to react first with 2,4-dihalo-phenylhydrazine hydrochloride in ethanol, followed by intramolecular cyclization in refluxing acetic acid to provide the pyrazole ester **2**. Treatment of **2** with NBS afforded the 2-bromothiophene **3** in 90% yield *via* regioselective bromination in position 5 of the thiophene ring. The ester **3** was hydrolysed under basic conditions to give the carboxylic acid **4** in very good yield. The acid **4** was first converted into the corresponding acyl chloride with thionyl chloride and then reacted with several anilines to afford the desired products **5**.



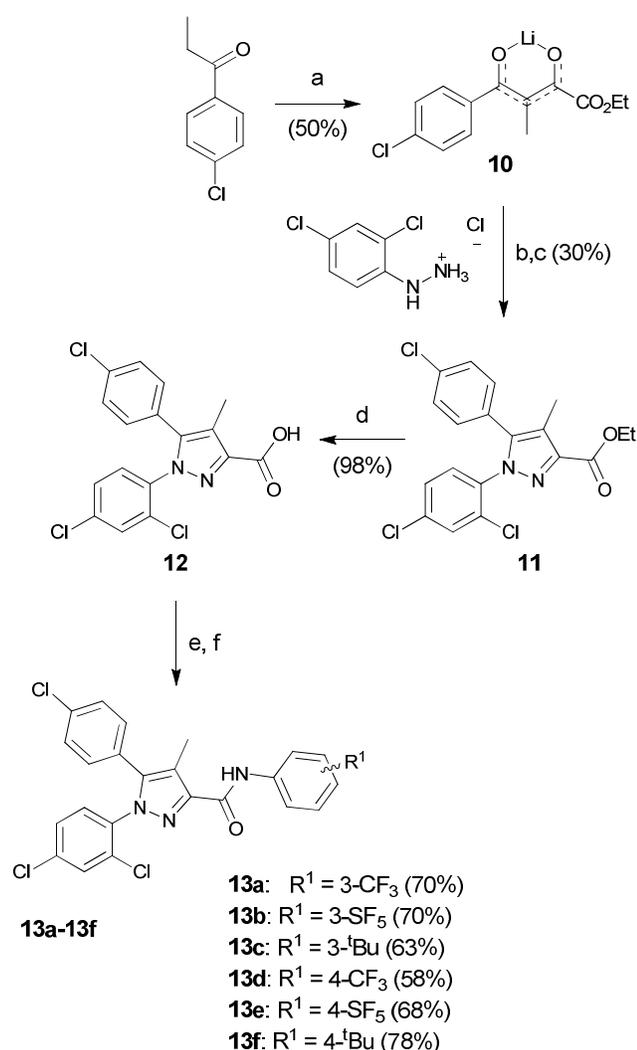
Scheme 2: Reagents and conditions: (a) Diethyl oxalate, LiHMDS, THF/Et<sub>2</sub>O (2/1), -78 °C to r.t., 16 h; (b) EtOH, r.t., 24 h; (c) AcOH, 120 °C, 16 h; (d) NBS, CH<sub>3</sub>CN, 0 °C to r.t., 16 h; (e) KOH, MeOH, reflux, 3 h; (f) thionyl chloride, toluene, reflux, 3 h; (g) Et<sub>3</sub>N, DCM, 0 °C to r.t., 16 h.

Similar procedure was used for the synthesis of the pyrazoles **9** (Scheme 3), having no substitution in position 4. In this case, however, the overall synthesis was one-step shorter thanks to the commercial availability of 2-bromo-5-acetyl-thiophene, which allowed us to skip the bromination reaction.



Scheme 3: Reagents and conditions: (a) Diethyl oxalate, LiHMDS, THF/Et<sub>2</sub>O (2/1), -78 °C to room temp, 16 h; (b) EtOH, room temp, 24 h; (c) AcOH, 120 °C, 16 h; (d) KOH, MeOH, reflux, 3 h; (e) thionyl chloride, toluene, reflux, 3 h; (f) Et<sub>3</sub>N, DCM, 0 °C to room temp, 16 h

The synthesis of the Rimonabant-like derivatives **13**, shown in Scheme 4, was analogously accomplished following the strategy described by Lan et al.<sup>32</sup>



Scheme 4: Reagents and conditions: (a) Diethyl oxalate, LiHMDS, THF/Et<sub>2</sub>O (2/1), -78 °C to room temp, 16 h; (b) EtOH, room temp, 24 h; (c) AcOH, 120 °C, 16 h; d) KOH, MeOH, reflux, 3 h; (e) thionyl chloride, toluene, reflux, 3 h; (f) Et<sub>3</sub>N, DCM, 0 °C to room temp, 16 h

### LogP measurement

Streich et al. showed that an SF<sub>5</sub> substituent on phenoxyacetic acid imparts higher lipophilicity than a CF<sub>3</sub> group<sup>10</sup>. We therefore investigated whether this was the case for our pyrazole cannabinoid ligands too.

Several software packages allow the prediction of physicochemical molecular properties, including the LogP (octanol/water partition coefficient). However, there are often significant discrepancies among the calculated values. We therefore decided to set up an experimental method for determining the LogP of these CB<sub>1</sub> ligands *via* reverse phase-HPLC analysis.

The retention times obtained (which are proportional to the lipophilic character of the molecules) confirmed that molecules bearing the SF<sub>5</sub> moiety are more lipophilic than the CF<sub>3</sub> counterparts and, in general, a substituent in *para* position influences the hydrophobicity of the entire molecule to a

greater extent ( $\Delta\text{LogP}_{(\text{SF}_5\text{-CF}_3)_p} = 0.3$ ) than in *meta* ( $\Delta\text{LogP}_{(\text{SF}_5\text{-CF}_3)_m} = 0.2$ ) (Tables 1 and 2). Not surprisingly, replacement of these fluorinated functions with a strongly lipophilic *tert*-butyl group further increased the LogP of the molecules.

Replacement of the 4-methyl group on the pyrazole, as in compounds **5** and **13**, with a hydrogen, as in compounds **9**, reduced the lipophilicity of both SF<sub>5</sub> and CF<sub>3</sub> derivatives by  $\Delta\text{LogP}_{(\text{CH}_3\text{-H})\text{SF}_5} = 0.7$  and  $\Delta\text{LogP}_{(\text{CH}_3\text{-H})\text{CF}_3} = 0.6$  units respectively. A further hydrophilicity enhancement was observed by substitution of the 2,4 difluoro aryl group in **9a,c** with the chlorinated analogue in **9b,d** ( $\Delta\text{LogP}_{(\text{F-Cl})} = 0.9$ ).

When a 4-chloro-phenyl moiety (compounds **13**) was replaced by a 5-bromo-thiophene group (compounds **5**), the LogP decreased by  $\Delta\text{LogP}_{(\text{Thy-Ph})} = 0.2$ , despite the benzene group (LogP benzene = 2.15)<sup>36</sup> is reported to be more hydrophobic than the thiophene group (LogP thiophene = 1.81)<sup>36</sup>. These results are presumably influenced by the presence on the thiophene ring of the bromine atom, which is softer and more hydrophobic than chlorine.

Most of the tabulated LogP values for the CB<sub>1</sub> inverse agonist Rimonabant (SR141716) were determined *in silico*, and the two experimental values reported in literature, obtained through the flask-shake technique, are quite different (LogD<sub>7.4} = 4.6 ± 0.8<sup>37</sup> and LogD<sub>7.4} = 3.8<sup>38</sup>). In order to obtain a new experimental value we decided to test SR141716 using the previously described RP-HPLC method, which provided a LogP value of 4.73 ± 0.20 for Rimonabant. This confirmed that all the new ligands presented in this article exhibit higher hydrophobicity than SR141716.</sub></sub>

### Binding affinity and SAR

#### EQUILIBRIUM BINDING ASSAYS

The binding affinities for the cannabinoid receptors of all the compounds **5**, **9** and **13** were determined by radio-receptor binding assays using the protocol previously described<sup>39</sup>. In this assay compounds **5**, **9** and **13** were subjected to equilibrium binding studies with the orthosteric agonist probe [<sup>3</sup>H]CP55940, and the ligands were assayed for their capacity to displace [<sup>3</sup>H]CP55940 from mouse brain membranes which express high levels of CB<sub>1</sub> receptors and from hCB<sub>2</sub> transfected CHO cells.

Compounds **9a-d** lack a substituent in position 4 of the pyrazole ring, incorporate a (5-bromo-thiophene) residue in position 5 and carry different aryl residues in position 1 (2,4-difluorophenyl for **9a,c** and 2,4-dichlorophenyl for **9b,d**). In this series, we compared the *para*-phenyl substituted compounds **9a** and **9b**, bearing a CF<sub>3</sub> group, with, respectively, **9c** and **9d**, carrying an SF<sub>5</sub> group (Table 1). In both cases, the SF<sub>5</sub> compounds **9c,d** showed higher affinity (lower K<sub>i</sub>) than the CF<sub>3</sub> counterparts. In terms of CB<sub>1</sub>/CB<sub>2</sub> selectivity, while **9a,c** were comparable, **9d** showed modest but higher CB<sub>1</sub>/CB<sub>2</sub> selectivity (10-fold) than **9b** (4-fold).

It has been previously demonstrated that an aliphatic substituent in position 4 of the pyrazole ring imparts higher CB<sub>1</sub> selectivity in pyrazole-based ligands; in particular Chen et al. performed a

3D quantitative structure-activity relationship (QSAR) of 5-aryl pyrazole structures using the comparative molecular field analysis (CoMFA)<sup>40</sup>, highlighting the importance of the 4-methyl group on pyrazole ring to achieve a better CB<sub>1</sub>/CB<sub>2</sub> ratio.

We therefore investigated also the effect of SF<sub>5</sub>, CF<sub>3</sub> and *tert*-butyl groups as 3-phenyl-carboxamide substituents in two series of 4-methyl-pyrazole cannabinoid ligands: 5-(5-bromothiophene)-pyrazoles **5a-f** and Rimonabant-type 5-(4-chlorophenyl)-pyrazoles **13a-f**.

As expected, the lower apparent K<sub>i</sub>s of all compounds **5** and **13** (Table 2) relative to **9a-d** confirmed that introduction of a methyl group in position 4 of the pyrazole ring results in an affinity increase *versus* CB<sub>1</sub> and, on the other hand, decreased the E<sub>max</sub> measured by the displacement assay on CB<sub>2</sub> CHO cells (see Table S1, Supporting Information).

In the *para*-phenyl substituted series of compounds **5d-f**, the SF<sub>5</sub>-compound **5e** showed the highest CB<sub>1</sub> affinity, whereas the presence of the *tert*-butyl group in **5f** caused a substantial drop both in affinity and CB<sub>1</sub>/CB<sub>2</sub> selectivity. Interestingly, the CB<sub>1</sub>/CB<sub>2</sub> selectivity was higher for the SF<sub>5</sub> derivative **5e** relative to the CF<sub>3</sub> analogue **5d**. In the *meta*-substituted series of compounds **5a-c**, the *tert*-butyl derivative **5c** featured the highest CB<sub>1</sub> affinity, whereas the SF<sub>5</sub> compound **5b** and the CF<sub>3</sub>-analogue **5a** showed comparable CB<sub>1</sub> affinities.

For the 4-chlorophenyl Rimonabant-type series of compounds **13a-f** (Table 2), we also observed nanomolar affinities for the CB<sub>1</sub>, in the same range of compounds **5a-f**, but the CB<sub>1</sub>/CB<sub>2</sub> selectivity was generally higher. In the *meta*-substituted series of compounds **13a-c**, the SF<sub>5</sub> derivative **13b** and the *tert*-butyl **13c** displayed the highest CB<sub>1</sub> affinity, and **13b** showed 2-fold higher affinity than the CF<sub>3</sub> analogue. The situation was somewhat reversed for the *para*-substituted compounds **13d-f** where the SF<sub>5</sub> compound **13e** displayed the lowest apparent CB<sub>1</sub> K<sub>i</sub>, whereas **13d** and **13f** showed similar affinities.

However, importantly, all of the 4-methyl-substituted pyrazoles bearing the SF<sub>5</sub> or <sup>1</sup>Bu groups in *para* position, namely **5e**, **5f**, **9d**, **9c** and **13f**, displaced [<sup>3</sup>H]CP55940 only partially at the maximum concentration of 10<sup>-5</sup> M with E<sub>max</sub> values ranging from 35.3 to 43.2 (see Figure 3 and Table S1, Supporting Information). Considering that the 4-trifluomethyl arene analogue **5d** produced a nearly full displacement of [<sup>3</sup>H]CP55940, we initially hypothesised that this behaviour could be explained by the binding of the compound to a topographically distinct site on CB<sub>1</sub> (an 'allosteric binding site'). We therefore investigated the effect of thiophenyl **5e** on the dissociation of [<sup>3</sup>H]CP55940 from mouse brain membranes; as this is the gold standard method of assessing an allosteric interaction<sup>41</sup>. However, compound **5e** had no significant effects on CB<sub>1</sub> agonist dissociation indicating that it is not an allosteric modulator.

At that point, we hypothesised that the partial displacement might be due to a solubility issue at the highest concentrations tested, i.e. 10<sup>-5</sup> M. In this context, Jackson et al. had previously shown that, although the S-F bond is longer and more polarizable than C-F one, the entropic cost that derives from

dissolving in water a larger group played a major role, resulting in lower S<sub>w</sub> (Water solubility) values for most of the pentafluorosulfanyl analytes relative to their CF<sub>3</sub> analogues<sup>42</sup>. Laser nephelometry has become the method of choice for measuring solubility of molecules in a drug discovery setting<sup>43</sup>.

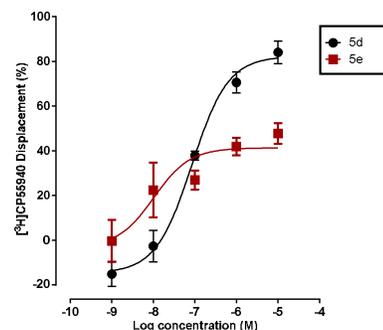


Figure 3: Effect of compounds **5d,e** on equilibrium binding of [<sup>3</sup>H]CP55940 to mouse brain membranes. Data shown are mean ± SEM 3-4 independent experiments. Data were best fitted by a sigmoidal concentration-response curve.

We therefore submitted compounds **5e**, **5d** and **5b** to a laser nephelometry assay, for assessing the solubility of each compound at different concentrations. However, these experiments showed that all of the compounds above were soluble at the highest concentration of 10<sup>-5</sup> M (99.9% aqueous buffer, 0.1% DMSO) (see Table S2, Supporting Information, for details).

#### FUNCTIONAL ASSAYS

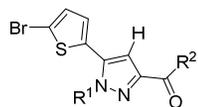
β-Arrestin recruitment assays, such as the PathHunter® β-arrestin assay, can be used for the identification of compounds with an agonistic, antagonistic, inverse agonistic, or allosteric modulation profile for GPCR ligands<sup>44</sup>. Increasing concentrations of an agonist binding to the orthosteric binding site on the receptor will result in a dose response curve where the EC<sub>50</sub> value is determined as the half maximal response. The addition of a competitive antagonist will result in a significant rightward shift in this dose response curve. This is due to the antagonist competing for the same site on the receptor as the agonist; therefore a higher concentration of agonist is required to reach the same maximal response. This will result in a significant increase in the EC<sub>50</sub> value obtained.

Using the PathHunter® β-arrestin assay in hCB1 cells, [<sup>3</sup>H]CP55940 stimulated β-arrestin recruitment with an EC<sub>50</sub> of 16.9 nM (11-27 nM) (95% confidence limits for all of the β-arrestin experiments herein described). The fluorinated *para*-substituted CB<sub>1</sub> receptor ligands, **5e**, **5d**, **13e** and **13d** (Table 2) caused a significant rightward shift in the dose response curve of [<sup>3</sup>H]CP55940, with an EC<sub>50</sub> of 111.0 nM (82-151 nM), 235.8 nM (157-355 nM), 245.8 nM (194-311 nM) and 439.6 nM (298-649 nM) respectively (see Supporting Information for the graphics). Analogously, with the fluorinated *meta*-substituted CB<sub>1</sub> receptor ligands **13b**, **13a**, **5b**, and **5a**, [<sup>3</sup>H]CP55940 stimulated beta-arrestin recruitment with an EC<sub>50</sub> of 17.2 nM

(14-22 nM) and the SF<sub>5</sub> compounds again, caused a significant rightward shift in the dose response curve of [<sup>3</sup>H]CP55940 with EC<sub>50</sub> values of 932.6 nM (451-1927 nM), 1528 nM (336-6953 nM), 158.6 nM (116-218 nM) and 57.0 nM (42-77 nM), respectively.

All of the tested compounds produced a significant increase in the EC<sub>50</sub> values with a rank order of efficacy of **13a**, **13b**, **13d**, **13e**, **5d**, **5b**, **5e**, and **5a**, and should be therefore considered competitive antagonists of [<sup>3</sup>H]CP55940 for the CB<sub>1</sub> receptor.

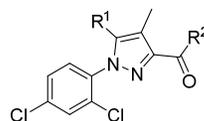
Table 1:



Compound	R <sup>1</sup>	R <sup>2</sup>	LogP <sup>a</sup> (±SEM)	CB <sub>1</sub> K <sub>i</sub> (nM) (±SEM)	CB <sub>2</sub> K <sub>i</sub> (nM) (±SEM)	Compound	R <sup>1</sup>	R <sup>2</sup>	LogP <sup>a</sup> (±SEM)	CB <sub>1</sub> K <sub>i</sub> (nM) (±SEM)	CB <sub>2</sub> K <sub>i</sub> (nM) (±SEM)
<b>9a</b>			4.67 (±0.20)	817.8 (±247.2)	2991.0 (±988.2)	<b>9c</b>			4.97 (±0.22)	302.7 (±72.1)	1009.0 (±349.2)
<b>9b</b>			5.60 (±0.27)	137.7 (±36.9)	528.4 (±204.8)	<b>9d</b>			5.89 (±0.30)	58.8 (±13.4)	588.2 (±229.9)

<sup>a</sup> Determined experimentally by means of RP-HPLC (see experimental section for details)

Table 2



Compound	R <sup>1</sup>	R <sup>2</sup>	LogP <sup>a</sup> (±SEM)	CB <sub>1</sub> K <sub>i</sub> (nM) (±SEM)	CB <sub>2</sub> K <sub>i</sub> (nM) (±SEM)	Compound	R <sup>1</sup>	R <sup>2</sup>	LogP <sup>a</sup> (±SEM)	CB <sub>1</sub> K <sub>i</sub> (nM) (±SEM)	CB <sub>2</sub> K <sub>i</sub> (nM) (±SEM)	Compound	R <sup>1</sup>	R <sup>2</sup>	LogP <sup>a</sup> (±SEM)	CB <sub>1</sub> K <sub>i</sub> (nM) (±SEM)	CB <sub>2</sub> K <sub>i</sub> (nM) (±SEM)
<b>5a</b>			6.19 (±0.32)	50.9 (±6.1)	362.2 (±67.6)	<b>5b</b>			6.42 (±0.35)	66.1 (±15.5)	991.2 (±231.7)	<b>5c</b>			7.09 (±0.40)	19.9 (±4.6)	102.1 (± 25.0)
<b>5d</b>			6.34 (±0.34)	65.3 (±13.0)	706.6 (±309.7)	<b>5e</b>			6.61 (±0.37)	7.9 (±3.3)	306.4 (±140.3)	<b>5f</b>			7.04 (±0.41)	106.9 (±30.1)	906.2 (± 361.0)
<b>13a</b>			5.97 (±0.30)	26.5 (±6.4)	609.6 (±195.5)	<b>13b</b>			6.21 (±0.33)	11.2 (±2.4)	637.3 (±205.2)	<b>13c</b>			6.95 (±0.38)	8.5 (±1.5)	1950.0 (±852.6)
<b>13d</b>			6.14 (±0.32)	31.3 (±9.8)	392.8 (±91.4)	<b>13e</b>			6.41 (±0.35)	17.1 (±3.4)	537.2 (±210.3)	<b>13f</b>			6.81 (±0.40)	47.9 (±19.8)	2053.0 (±949.8)

<sup>a</sup> Determined experimentally by means of RP-HPLC (see experimental section for details)

## Summary and Conclusions

We have shown that the pentafluorosulfanyl group can effectively replace a trifluoromethyl group in pyrazole-type cannabinoid ligands. The resulting SF<sub>5</sub>-compounds behaved as competitive CB<sub>1</sub> receptor antagonists, which is an interesting property since CB<sub>1</sub> inverse agonists have been reported to produce severe adverse effects that limit their clinical applications, whereas CB<sub>1</sub> neutral antagonists might have improved clinical utility<sup>45</sup>. Furthermore SF<sub>5</sub>-compounds showed nanomolar inhibition (K<sub>i</sub>) and equilibrium dissociation constants (K<sub>b</sub>) for the CB<sub>1</sub> receptor, with moderate CB<sub>1</sub>/CB<sub>2</sub> selectivity.

Both affinity and selectivity of SF<sub>5</sub>-compounds were generally slightly superior or comparable to that of CF<sub>3</sub> and *tert*-butyl analogues. Experimental lipophilicity (LogP) was found to follow the trend *tert*-butyl > SF<sub>5</sub> > CF<sub>3</sub>. Functional assays showed that all of the tested compounds, belonging to the SF<sub>5</sub> and CF<sub>3</sub> series, are CB<sub>1</sub> neutral antagonists. It is worth noting that some of the compounds, incorporating *para*-SF<sub>5</sub>- or *tert*-butyl-aryl groups on the C-3 pyrazole ring, displayed an apparent partial displacement of [<sup>3</sup>H]CP55940 in the functional assays (E<sub>max</sub> values ranging from 35.3 to 43.2), while ligands binding to the orthosteric binding pocket would be expected to fully displace the radioligand. One explanation for this observation could be that the compounds are binding to an allosteric pocket. However, this seems unlikely as we observed no change in agonist dissociation. Solubility problems were also excluded by means of nephelometry assays, so we are currently unclear as to the explanation for this observation. Overall, the data collected in this work confirm that (1) an aromatic SF<sub>5</sub> group is an effective bioisosteric analogue of the CF<sub>3</sub> group, and possibly also of bulky aliphatic groups like the *tert*-butyl, (2) it can be successfully used as a substituent in biologically active compounds and drug candidates.

## Experimental section

### Chemistry

*Solvents, reagents and apparatus.* Reagent-grade commercially available solvents and reagents were used without further purification.

NMR data were recorded on Bruker ADVANCE III for <sup>1</sup>H at 400.13 MHz, for <sup>13</sup>C at 100.58 MHz and for <sup>19</sup>F at 376.45 MHz. <sup>1</sup>H NMR chemical shifts are reported relative to TMS, and the solvent resonance was employed as the internal standard (CDCl<sub>3</sub> δ = 7.26). <sup>13</sup>C NMR spectra were recorded with complete proton decoupling, and the chemical shifts are reported relative to TMS with the solvent resonance as internal standard (CDCl<sub>3</sub> δ = 77.0). <sup>19</sup>F NMR spectra were referenced to CFC<sub>3</sub> as the external standard. All chemical shift (δ) are reported in parts per million (ppm) downfield from TMS and coupling constant (J) in Hertz. Splitting patterns are reported as follows: s, singlet; d, doublet; dd, doublet of doublets; ddd, doublet of doublet of doublets; t, triplet; td, triplet of doublets;

appt, apparent triplet; q, quadruplet; qd, quadruplet of doublets; m, multiplet; br, broad signal.

Mass Analysis was performed using an Agilent 1200 HPLC system coupled to an Agilent G6120 single quadrupole detector equipped with Electrospray ionization (ESI) source in direct infusion modality.

Lipophilicities were determined using a Reverse Phase (RP)-HPLC with an Agilent 1200 HPLC system equipped with a DAD, analytical Phenomenex Luna C-18 column (250 x 4.60 mm L x ID, particle size: 5 μ) and an ESI-MS detector.

HRMS analysis were performed using an LTQ Orbitrap XL MS spectrometer.

All reactions were carried out in oven- or flame-dried glassware under nitrogen atmosphere, unless stated otherwise, and were magnetically stirred and monitored by TLC on silica gel (60 F254 pre-coated glass plates, 0.25 mm thickness).

Visualization was accomplished using irradiation with a UV lamp (λ = 254 nm or λ = 365 nm), and/or staining with potassium permanganate or ceric ammonium molybdate solution.

Purification of reaction products was performed using flash chromatography on silica gel (60 Å, particle size 40-63 μm) according to the procedure of Still and co-workers<sup>46</sup>.

Yields refer to chromatographically and spectroscopically pure compounds, unless stated otherwise.

### 3-(Pentafluoro-λ<sup>6</sup>-sulfanyl)aniline

To a stirred acidic warm solution (50 °C) of pentafluoro(3-nitrophenyl)-λ<sup>6</sup>-sulfane (1.00 g, 3.93 mmol) in Ethanol:HCl (11 ml:0.8 ml, 37% v/v), iron powder (1.22 g, 21.63 mmol) was added portionwise. The reaction was refluxed for 2 h. After cooling, the solid was removed by means of filtration; the filtered was diluted with dichloromethane. The organic phase was acidified with HCl 2N. The aqueous phase was separated, basified and extracted with dichloromethane (3 X 50 ml). The combined extracts were washed with brine, dried over anhydrous sodium sulfate, filtered, and evaporated to give the crude aniline as a yellow oil (0.72 g, 83% yield): The obtained <sup>1</sup>H, <sup>12</sup>C and <sup>19</sup>F NMR spectra matched to those reported by Bowden et al<sup>34</sup>.

### Lithium Salt of Ethyl 3-Methyl-2,4-dioxo-4-(thiophen-2-yl)butanoate (1).

To a magnetically stirred solution of lithium bis(trimethylsilyl)amide (61 mL, 60.60 mmol, 1.0 M in THF) in diethyl ether (110 mL) at -78 °C, 1-(2-thienyl)-1-propanone (7 mL, 55.10 mmol) in diethyl ether (42 mL) was added dropwise under a nitrogen atmosphere. After the mixture was stirred at the same temperature for an additional 45 min, diethyl oxalate (9 mL, 66.11 mmol) was added dropwise. The reaction mixture was allowed to warm to room temperature and stirred for another 16 h. The reaction precipitate was filtered, washed with diethyl ether, and dried under vacuum to afford the crude lithium salt **1** (9.50 g, 70%) as a pale-yellow solid. The product was used without further purification. ESMS, calculated *m/z*

$C_{11}H_{11}LiO_4S$  246.05  $[M]^+$ , found  $m/z$  (relative intensity) 247.0  $[M+H]^+$  (100), 265.0  $[M+H+Na-Li]^+$  (100), 279.0  $[M+H+K-Li]^+$  (100).

### 1-(2,4-Dichlorophenyl)-4-methyl-5-thiophen-2-yl-1H-pyrazole-3-carboxylic Acid Ethyl Ester (2).

To a solution of lithium salt **1** (8.50 g, 33.83 mmol) in ethanol (26 mL) was added 2,4-dichlorophenylhydrazine hydrochloride (8.85 g, 40.60 mmol) in one portion at room temperature under nitrogen. The resulting mixture was stirred at the same temperature for 24 h. After the reaction was complete, the precipitate was filtered, washed with ethanol and diethyl ether, and dried under vacuum to give a light-yellow solid (7.47 g). This crude solid, without purification, was dissolved in acetic acid (68 mL) and heated to reflux for 16 h. The reaction mixture was poured into ice-water and extracted with ethyl acetate ( $3 \times 50$  mL). The combined extracts were washed with water, saturated aqueous sodium bicarbonate, and brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Purification by flash column chromatography on silica gel with DCM gave ester **2** (5.33 g, 42% over two steps) as a white solid: The obtained  $^1H$ ,  $^{13}C$  and spectra matched to those reported by Tseng et al.<sup>35</sup>

### 5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxylic Acid Ethyl Ester (3).

To a magnetically stirred solution of **2** (2.60 g, 6.61 mmol) in acetonitrile (23 mL) was added NBS (1.43 g, 7.94 mmol) in small portions under nitrogen at 0 °C. The resulting mixture was then warmed to room temperature and stirred for 16 h. The reaction was quenched with saturated aqueous sodium thiosulfate and concentrated under reduced pressure to remove acetonitrile. The aqueous layer was extracted with ethyl acetate ( $2 \times 40$  mL). The organic layers were combined, washed with water, brine, dried over anhydrous sodium sulfate, and concentrated to give the crude residue, which was subjected to purification by flash chromatography on silica gel with n-hexane/ethyl acetate (8:2) to afford bromo ester **3** (2.70 g, 90%) as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  7.46 (d,  $J = 2.0$  Hz, 1H), 7.36 (d,  $J = 8.5$  Hz, 1H), 7.33 (dd,  $J = 8.4$ , 2.0 Hz, 1H), 6.95 (dd,  $J = 3.9$ , 0.8 Hz, 1H), 6.64 (dd,  $J = 3.9$ , 0.8 Hz, 1H), 4.44 (q,  $J = 7.1$  Hz, 2H), 2.42 (s, 3H), 1.42 (t,  $J = 7.1$  Hz, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  162.5, 142.9, 136.9, 136.6, 135.7, 133.8, 130.9, 130.2, 130.1, 130.1, 129.3, 127.9, 120.4, 115.0, 61.1, 14.5, 10.0. ESMS, calculated  $m/z$   $C_{17}H_{13}BrCl_2N_2O_2S$  459.92  $[M]^+$ , found  $m/z$  (relative intensity) 460.9  $[M+H]^+$  (100), 482.9  $[M+Na]^+$  (100).

### 5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxylic acid (4).

To a solution of bromo ester **3** (2.64 g, 5.62 mmol) in methanol (26 mL) was added potassium hydroxide (0.73 g, 11.24 mmol) in methanol (8 mL) dropwise at room temperature. The resulting mixture was heated to reflux for 3 h. After hydrolysis was complete, the reaction mixture was cooled to room

temperature, poured into ice-water, and acidified with 2N hydrochloric acid. The precipitate was filtered, washed with water, and dried under vacuum to give the thiophene carboxylic acid **4** (2.19 g, 90%) as a white solid:  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  13.05 (s, 1H), 7.89 (d,  $J = 2.3$  Hz, 1H), 7.73 (d,  $J = 8.5$  Hz, 1H), 7.63 (dd,  $J = 8.5$ , 2.3 Hz, 1H), 7.23 (d,  $J = 3.9$  Hz, 1H), 6.88 (d,  $J = 3.9$  Hz, 1H), 2.32 (s, 3H);  $^{13}C$  NMR (101 MHz, DMSO)  $\delta$  163.9, 143.3, 136.7, 136.2, 135.9, 133.1, 132.3, 131.5, 130.8, 130.2, 130.0, 129.0, 119.6, 114.6, 10.2. ESMS, calculated  $m/z$   $C_{15}H_9BrCl_2N_2O_2S$  431.9  $[M]^+$ , found  $m/z$  (relative intensity) 470.9  $[M+K]^+$  (100)

### General Procedure for the Synthesis of Compounds 5a – 5f.

The general procedure is illustrated below for compound **5a**.

### 5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxylic acid chloride

A solution of the crude acid **4** (0.80 g, 1.81 mmol) and thionyl chloride (399  $\mu$ l, 5.44 mmol) in toluene (12 mL) was refluxed for 3 h. Solvent was evaporated under reduced pressure, and the residue was then re-dissolved in toluene (8 mL) first and then in Hexane (5 mL); after evaporation the crude acyl chloride (0.80 g, 98% yield) was obtained as a white solid.

### 5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (5a)

A solution in dichloromethane of the carboxylic chloride (1 mL, 0.36 M) obtained previously from **4**, was added dropwise to a solution of 3-(trifluoromethyl)aniline (50  $\mu$ l, 0.40 mmol) and triethylamine (50  $\mu$ l, 0.36 mmol) in dichloromethane (0.5 mL) at 0 °C. After stirring at room temperature for 16 h, to the reaction mixture was added water and the organic phase was extracted with dichloromethane ( $3 \times 3$  mL). The combined extracts were washed with brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Flash column chromatography on silica gel with n-Hexane / ethyl acetate (8:2) gave carboxamide **5a** (150 mg, 66% yield) as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.87 (s, 1H), 8.01 (s, 1H), 7.85 (d,  $J = 7.8$  Hz, 1H), 7.54 (d,  $J = 2.1$  Hz, 1H), 7.46 (t,  $J = 8.0$  Hz, 1H), 7.39 (dd,  $J = 8.5$ , 2.1 Hz, 1H), 7.36 (d,  $J = 8.4$  Hz, 2H), 6.98 (d,  $J = 3.9$  Hz, 1H), 6.68 (d,  $J = 3.9$  Hz, 1H), 2.52 (s, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.4, 144.3, 138.3, 137.7, 136.8, 135.5, 133.7, 131.42 (q,  $J = 32.4$  Hz), 130.6, 130.4, 130.2, 129.9, 129.5, 129.4, 128.1, 125.2, 123.87 (q,  $J = 272.6$  Hz), 122.57, 120.55 (q,  $J = 3.9$  Hz), 119.6, 116.34 (q,  $J = 4.0$  Hz), 115.2;  $^{19}F$  NMR (376 MHz, Chloroform-d)  $\delta$  -62.72. ESMS, calculated  $m/z$   $C_{22}H_{13}BrCl_2F_3N_3OS$  574.93  $[M]^+$ , found  $m/z$  (relative intensity) 575.9  $[M+H]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{22}H_{13}BrCl_2F_3N_3OS$  : 573.9370; found: 573.9361

**5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (5b)**

**5b** (163 mg, 68%yield) was obtained as a white solid:  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.87 (s, 1H), 8.13 (t, *J* = 2.0 Hz, 1H), 7.87 (dd, *J* = 8.1, 1.9 Hz, 1H), 7.54 (d, *J* = 2.1 Hz, 1H), 7.50 (d, *J* = 8.3 Hz, 1H), 7.44 (d, *J* = 8.1 Hz, 1H), 7.40 (dd, *J* = 8.5, 2.1 Hz, 1H), 7.36 (d, *J* = 8.4 Hz, 1H), 6.98 (d, *J* = 3.9 Hz, 1H), 6.68 (d, *J* = 3.9 Hz, 1H), 2.52 (s, 3H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  160.4, 154.25 (appt, *J* = 17.1 Hz), 144.2, 138.2, 137.8, 136.9, 135.4, 133.8, 130.6, 130.4, 130.3, 129.8, 129.4, 129.2, 128.1, 122.5, 121.28 (p, *J* = 4.7 Hz), 119.6, 117.25 (p, *J* = 4.0 Hz), 115.2, 9.7;  $^{19}\text{F}$  NMR (377 MHz, Chloroform-*d*)  $\delta$  84.17 (p, *J* = 149.2 Hz), 62.73 (d, *J* = 150.0 Hz). ESMS, calculated *m/z*  $\text{C}_{21}\text{H}_{13}\text{BrCl}_2\text{F}_5\text{N}_3\text{OS}_2$  632.90  $[\text{M}]^+$ , found *m/z* (relative intensity) 631.9  $[\text{M}-\text{H}]^-$  (100). HRMS *m/z*  $\text{M}^+\text{H}^+$  calcd. for  $\text{C}_{21}\text{H}_{13}\text{BrCl}_2\text{F}_5\text{N}_3\text{OS}_2$  : 631.9059; found: 631.9052

**5-(5-Bromothiophen-2-yl)-N-(3-(tert-butyl)phenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (5c)**

**5c** (87 mg, 63%yield) was obtained as a white solid  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.71 (s, 1H), 7.64 – 7.57 (m, 1H), 7.59 (d, *J* = 2.0 Hz, 1H), 7.54 (dd, *J* = 1.9, 0.7 Hz, 1H), 7.38 (dd, *J* = 1.9, 1.3 Hz, 2H), 7.30 (d, *J* = 8.6 Hz, 1H), 7.18 – 7.12 (m, 1H), 6.97 (d, *J* = 3.9 Hz, 1H), 6.67 (d, *J* = 3.8 Hz, 1H), 2.52 (s, 3H), 1.33 (s, 9H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  160.2, 152.2, 144.9, 137.5, 137.4, 136.7, 135.6, 133.8, 130.7, 130.4, 130.2, 130.1, 129.2, 128.6, 128.0, 121.2, 119.4, 117.0, 116.9, 115.0, 34.8, 31.3 (x 3), 9.8. ESMS, calculated *m/z*  $\text{C}_{25}\text{H}_{22}\text{BrCl}_2\text{N}_3\text{OS}$  563.00  $[\text{M}]^+$ , found *m/z* (relative intensity) 564.0  $[\text{M}+\text{H}]^+$  (100). HRMS *m/z*  $\text{M}^+\text{H}^+$  calcd. for  $\text{C}_{25}\text{H}_{22}\text{BrCl}_2\text{N}_3\text{OS}$  : 562.0122; found: 562.0111

**5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-N-(4-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (5d)**

**5d** (97 mg, 44%yield) was obtained as a white solid:  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.89 (s, 1H), 7.80 (d, *J* = 8.5 Hz, 2H), 7.60 (d, *J* = 8.6 Hz, 2H), 7.54 (d, *J* = 2.1 Hz, 1H), 7.40 (dd, *J* = 8.5, 2.1 Hz, 1H), 7.36 (d, *J* = 8.4 Hz, 1H), 6.98 (d, *J* = 3.9 Hz, 1H), 6.68 (d, *J* = 3.9 Hz, 1H), 2.52 (d, *J* = 0.5 Hz, 3H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  160.4, 144.3, 140.9, 137.7, 136.9, 135.4, 133.7, 130.6, 130.4, 130.2, 129.8, 129.4, 128.2, 126.25 (q, *J* = 3.6 Hz), 125.57 (q, *J* = 32.5 Hz, x2), 124.13 (q, *J* = 27.2), 119.6, 119.2 (x2), 115.2, 9.7;  $^{19}\text{F}$  NMR (376 MHz, Chloroform-*d*)  $\delta$  -62.07. ESMS, calculated *m/z*  $\text{C}_{22}\text{H}_{13}\text{BrCl}_2\text{F}_3\text{N}_3\text{OS}$  574.93  $[\text{M}]^+$ , found *m/z* (relative intensity) 575.8  $[\text{M}+\text{H}]^+$  (100). HRMS *m/z*  $\text{M}^+\text{H}^+$  calcd. for  $\text{C}_{22}\text{H}_{13}\text{BrCl}_2\text{F}_3\text{N}_3\text{OS}$  : 573.9370; found: 573.9361

**5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-4-methyl-N-(4-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (5e)**

**5e** (100 mg, 42%yield) was obtained as a white solid:  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.92 (s, 1H), 7.81 – 7.68 (m, 4H),

7.53 (d, *J* = 2.1 Hz, 1H), 7.42 – 7.31 (m, 2H), 6.98 (d, *J* = 3.9 Hz, 1H), 6.68 (d, *J* = 3.9 Hz, 1H), 2.51 (s, 3H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  160.4, 149.03 (appt, *J* = 17.6 Hz), 144.1, 140.5, 137.8, 136.9, 135.4, 133.7, 130.6, 130.4, 130.2, 129.7, 129.4, 128.1, 126.99 (p, *J* = 4.7 Hz, x2), 119.7, 118.8 (x2), 115.2, 9.7;  $^{19}\text{F}$  NMR (377 MHz, Chloroform-*d*)  $\delta$  85.35 (p, *J* = 150.1, 149.4 Hz), 63.55 (d, *J* = 150.0 Hz). ESMS, calculated *m/z*  $\text{C}_{21}\text{H}_{13}\text{BrCl}_2\text{F}_5\text{N}_3\text{OS}_2$  632.90  $[\text{M}]^+$ , found *m/z* (relative intensity) 631.8  $[\text{M}-\text{H}]^-$ . HRMS *m/z*  $\text{M}^+\text{H}^+$  calcd. for  $\text{C}_{21}\text{H}_{13}\text{BrCl}_2\text{F}_5\text{N}_3\text{OS}_2$  : 631.9059; found: 631.9048.

**5-(5-Bromothiophen-2-yl)-N-(4-(tert-butyl)phenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (5f)**

**5f** (84 mg, 40%yield) was obtained as a white solid:  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  8.67 (s, 1H), 7.58 (d, *J* = 8.7 Hz, 2H), 7.53 (dd, *J* = 1.9, 0.6 Hz, 1H), 7.41 – 7.32 (m, 4H), 6.97 (d, *J* = 3.9 Hz, 1H), 6.67 (d, *J* = 3.8 Hz, 1H), 2.52 (s, 3H), 1.32 (s, 9H);  $^{13}\text{C}$  NMR (101 MHz,  $\text{CDCl}_3$ )  $\delta$  160.2, 147.0, 144.9, 137.4, 136.7, 135.6, 135.2 (x2), 133.8, 130.7, 130.4, 130.2, 129.2, 128.0, 125.8 (x2), 119.6 (x2), 119.4, 115.0, 34.4, 31.4 (x3) 9.8. ESMS, calculated *m/z*  $\text{C}_{25}\text{H}_{22}\text{BrCl}_2\text{N}_3\text{OS}$  563.0  $[\text{M}]^+$ , found *m/z* (relative intensity) 564.0  $[\text{M}+\text{H}]^+$  (100). HRMS *m/z*  $\text{M}^+\text{H}^+$  calcd. for  $\text{C}_{25}\text{H}_{22}\text{BrCl}_2\text{N}_3\text{OS}$  : 562.0122; found: 562.0113.

**Lithium Salt of ethyl 4-(5-bromothiophen-2-yl)-2,4-dioxobutanoate (6).**

To a magnetically stirred solution of lithium bis(trimethylsilyl)amide (23 mL, 22.56 mmol, 1.0 M in THF) in diethyl ether (41 mL) at -78 °C, 1-(5-bromothiophen-2-yl)ethanone (4.25 g, 20.51 mmol) in diethyl ether (16 mL) was added dropwise under a nitrogen atmosphere. After the mixture was stirred at the same temperature for an additional 45 min, diethyl oxalate (3.4 mL, 24.62 mmol) was added dropwise. The reaction mixture was allowed to warm to room temperature and stirred for another 16 h. The reaction precipitate was filtered, washed with diethyl ether, and dried under vacuum to afford the crude lithium salt **6** (6.31 g, 99%) as a pale-yellow solid. The product was used without further purification. ESMS, calculated *m/z*  $\text{C}_{10}\text{H}_8\text{BrLiO}_4\text{S}$  309.9  $[\text{M}]^+$ , found *m/z* (relative intensity) 326.9  $[\text{M}-\text{Li}+\text{H}+\text{Na}]^+$  (63), 343  $[\text{M}-\text{Li}+\text{H}+\text{K}]^+$  (31)

**General Procedure for the Synthesis of Compounds 7a,7b.**

The general procedure is illustrated below for compound **7a**.

**Ethyl 5-(5-bromothiophen-2-yl)-1-(2,4-difluorophenyl)-1H-pyrazole-3-carboxylate (7a)**

To a solution of lithium salt **6** (3 g, 9.55 mmol) in ethanol (26 mL) was added 2,4-difluorophenylhydrazine hydrochloride (2.143 g, 11.46 mmol) in one portion at room temperature under nitrogen. The resulting mixture was stirred at the same temperature for 24 h. After reaction was complete, the precipitate was filtered, washed with ethanol and diethyl ether, and dried under vacuum to give a light-yellow solid (2.66 g).

This crude solid, without purification, was dissolved in acetic acid (20 mL) and heated to reflux for 16 h. The reaction mixture was poured into ice-water and extracted with ethyl acetate (3 × 70 mL). The combined extracts were washed with water, saturated aqueous sodium bicarbonate, and brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Purification by flash column chromatography on silica gel with DCM gave ester **7a** (2.08 g, 58 % over two steps) as a white solid. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.51 (td, J = 8.5, 5.7 Hz, 1H), 7.08 (s, 1H), 7.08 – 6.92 (m, 2H), 6.93 (d, J = 3.9 Hz, 1H), 6.69 (d, J = 3.9 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 1.41 (t, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 163.54 (dd, J = 254.1, 11.0 Hz), 161.6, 157.88 (dd, J = 256.9, 12.8 Hz), 145.3, 139.5, 130.83 (d, J = 10.1 Hz), 130.8, 130.4, 127.4, 123.29 (dd, J = 12.7, 4.1 Hz), 114.4, 112.25 (dd, J = 22.7, 3.8 Hz), 108.4, 105.29 (dd, J = 26.5, 23.0 Hz), 61.3, 14.3; <sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -104.59 (qd, J = 8.4, 5.7 Hz), -115.10 – -115.23 (m). ESMS, calculated *m/z* C<sub>16</sub>H<sub>11</sub>BrF<sub>2</sub>N<sub>2</sub>O<sub>2</sub>S 414.0 [M]<sup>+</sup>, found *m/z* (relative intensity) 415.0 [M+H]<sup>+</sup> (100), 435.0 [M+Na]<sup>+</sup> (100)

#### Ethyl 5-(5-bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-1H-pyrazole-3-carboxylate (**7b**)

**7b** (2.36 g, 60 % over two steps) was obtained as a white solid: <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.54 (d, J = 2.1 Hz, 1H), 7.45 (d, J = 8.4 Hz, 1H), 7.41 (dd, J = 8.5, 2.1 Hz, 1H), 7.08 (s, 1H), 6.92 (d, J = 3.9 Hz, 1H), 6.66 (d, J = 3.9 Hz, 1H), 4.44 (q, J = 7.1 Hz, 2H), 1.41 (t, J = 7.1 Hz, 3H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 161.7, 145.2, 139.5, 137.2, 135.3, 134.0, 131.0, 130.8, 130.5, 130.4, 128.2, 127.3, 114.5, 108.1, 61.4, 14.4. ESMS, calculated *m/z* 16 H<sub>11</sub>BrCl<sub>2</sub>N<sub>2</sub>O<sub>2</sub>S 445.9 [M]<sup>+</sup>, found *m/z* (relative intensity) 446.9 [M+H]<sup>+</sup> (100).

#### General Procedure for the Synthesis of Compounds **8a,8b**.

The general procedure is illustrated below for compound **8a**.

#### 5-(5-Bromothiophen-2-yl)-1-(2,4-difluorophenyl)-1H-pyrazole-3-carboxylic acid (**8a**).

To a solution of bromo ester **7a** (1.72 g, 4.07 mmol) in methanol (19 mL) was added potassium hydroxide (0.80 g, 12.21 mmol) in methanol (9 mL) dropwise at room temperature. The resulting mixture was heated to reflux for 3 h. After hydrolysis was complete, the reaction mixture was cooled to room temperature, poured into ice-water, and acidified with 2N hydrochloric acid. The precipitate was filtered, washed with water, and dried under vacuum to give thiophene carboxylic acid **8a** (1.34 g, 85%) as a white solid: <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ 13.18 (bs, 1H), 7.83 (td, J = 8.8, 5.9 Hz, 1H), 7.64 (ddd, J = 10.3, 9.0, 2.8 Hz, 1H), 7.42 – 7.32 (m, 1H), 7.28 (s, 1H), 7.23 (d, J = 4.0 Hz, 1H), 7.11 (d, J = 4.0 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO) δ 163.66 (dd, J = 251.4, 11.7 Hz), 163.0, 157.92 (dd, J = 253.6, 13.7 Hz), 146.0, 139.7, 132.28 (d, J = 10.7 Hz), 131.6, 130.8, 129.3, 123.43 (dd, J = 12.6, 3.8 Hz), 114.0, 113.45 (dd, J = 22.9, 3.6 Hz), 108.6, 106.16 (dd, J =

27.4, 23.6 Hz); <sup>19</sup>F NMR (376 MHz, DMSO-d<sub>6</sub>) δ -104.80 (qd, J = 8.7, 5.8 Hz), -117.33 (q, J = 9.4 Hz). ESMS, calculated *m/z* C<sub>14</sub>H<sub>7</sub>BrF<sub>2</sub>N<sub>2</sub>O<sub>2</sub>S 383.9 [M]<sup>+</sup>, found *m/z* (relative intensity) 422.9 [M+K]<sup>+</sup> (100).

#### 5-(5-bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-1H-pyrazole-3-carboxylic acid (**8b**)

**8b** (1.42 g, 79 %) was obtained as a white solid: <sup>1</sup>H NMR (400 MHz, DMSO-d<sub>6</sub>) δ 13.18 (s, 1H), 8.00 (d, J = 2.3 Hz, 1H), 7.82 (d, J = 8.5 Hz, 1H), 7.71 (dd, J = 8.5, 2.3 Hz, 1H), 7.31 (s, 1H), 7.22 (d, J = 4.0 Hz, 1H), 7.09 (d, J = 4.0 Hz, 1H); <sup>13</sup>C NMR (101 MHz, DMSO) δ 163.0, 145.8, 139.5, 136.9, 135.5, 133.6, 132.5, 131.6, 130.8, 130.6, 129.4, 129.1, 114.0, 108.3. ESMS, calculated *m/z* C<sub>14</sub>H<sub>7</sub>BrCl<sub>2</sub>N<sub>2</sub>O<sub>2</sub>S 417.9 [M]<sup>+</sup>, found *m/z* (relative intensity) 456.9 [M+K]<sup>+</sup> (100)

#### General Procedure for the Synthesis of Compounds **9a – 9d**.

The general procedure is illustrated below for compound **9a**.

#### 5-(5-Bromothiophen-2-yl)-1-(2,4-difluorophenyl)-1H-pyrazole-3-carboxyl chloride

A solution of the crude acid **8** (0.20 g, 0.51 mmol) and thionyl chloride (112 μl, 1.53 mmol) in toluene (3 mL) was refluxed for 3 h. The solvent was evaporated under reduced pressure, and the residue was then re-dissolved in toluene (3 mL) first and then in Hexane (4 ml); after evaporation the crude acyl chloride (0.19 g, 95% yield) was obtained as a white solid.

#### 5-(5-Bromothiophen-2-yl)-1-(2,4-difluorophenyl)-N-(4-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (**9a**)

A solution in dichloromethane of the acyl chloride obtained previously (0.9 ml, 0.24 M), was added dropwise to a solution of 4-(trifluoromethyl)aniline (30 μl, 0.23 mmol) and triethylamine (33 μl, 0.23 mmol) in dichloromethane (0.2 ml) at 0 °C. After stirring at room temperature for 16 h, to the reaction mixture was added water and the organic phase was extracted with dichloromethane (3 × 3 mL). The combined extracts were washed with brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Flash column chromatography on silica gel with n-Hexane / ethyl acetate (9:1) gave carboxamide **9a** (88 mg, 72% yield) as a white solid: <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 8.81 (s, 1H), 7.81 (d, J = 8.5 Hz, 2H), 7.61 (d, J = 8.6 Hz, 2H), 7.51 (td, J = 8.5, 5.7 Hz, 1H), 7.17 (s, 1H), 7.12 – 7.00 (m, 2H), 6.95 (d, J = 3.9 Hz, 1H), 6.74 (d, J = 3.9 Hz, 1H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 163.74 (dd, J = 254.8, 10.9 Hz), 159.1, 158.03 (dd, J = 257.3, 12.8 Hz), 147.8, 140.6, 130.7, 130.64 (d, J = 10.2 Hz), 130.5, 127.7, 126.32 (q, J = 3.8 Hz, x2), 125.99 (q, J = 32.6 Hz), 124.09 (q, J = 271.8 Hz), 123.15 (dd, J = 13.0, 4.2 Hz), 122.74, 119.3 (x2), 114.8, 112.48 (dd, J = 22.8, 3.9 Hz), 107.0, 105.66 (dd, J = 26.5, 23.0 Hz); <sup>19</sup>F NMR (376 MHz, Chloroform-d) δ -62.11, -104.02 (qd, J = 8.3, 5.7 Hz), -115.07 – -115.16 (m). ESMS, calculated *m/z* C<sub>21</sub>H<sub>11</sub>BrF<sub>5</sub>N<sub>3</sub>OS 529 [M]<sup>+</sup>, found *m/z* (relative intensity) 530

$[M+H]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{21}H_{11}BrF_5N_3OS$  : 527.9805; found: 527.9794

**5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-N-(4-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (9b)**

**9b** (69 mg, 53 %) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.81 (s, 1H), 7.80 (d,  $J$  = 8.5 Hz, 2H), 7.65 – 7.56 (m, 3H), 7.46 (d,  $J$  = 1.9 Hz, 2H), 7.18 (s, 1H), 6.94 (d,  $J$  = 3.9 Hz, 1H), 6.71 (d,  $J$  = 3.9 Hz, 1H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  159.1, 147.7, 140.6, 140.5, 137.5, 135.1, 134.1, 130.7, 130.7, 130.7, 130.5, 128.4, 127.6, 126.33 (q,  $J$  = 3.7 Hz, x2), 125.98 (q,  $J$  = 32.8 Hz), 124.08 (q,  $J$  = 271.3 Hz), 119.3 (x2), 114.8, 106.6;  $^{19}F$  NMR (376 MHz, Chloroform- $d$ )  $\delta$  -62.10. ESMS, calculated  $m/z$   $C_{21}H_{11}BrCl_2F_3N_3OS$  560.9  $[M]^+$ , found  $m/z$  (relative intensity) 583.8  $[M+Na]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{21}H_{11}BrCl_2F_3N_3OS$  : 559.9214; found: 559.9203

**5-(5-Bromothiophen-2-yl)-1-(2,4-difluorophenyl)-N-(4-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (9c)**

**9c** (110 mg, 70 %) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.81 (s, 1H), 7.81 – 7.72 (m, 4H), 7.51 (td,  $J$  = 8.5, 5.7 Hz, 1H), 7.18 (s, 1H), 7.13 – 7.01 (m, 2H), 6.96 (d,  $J$  = 3.9 Hz, 1H), 6.74 (d,  $J$  = 3.9 Hz, 1H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  163.76 (dd,  $J$  = 254.8, 11.0 Hz), 159.1, 158.02 (dd,  $J$  = 257.2, 12.8 Hz), 149.14 (appt,  $J$  = 17.5 Hz), 147.6, 140.7, 140.3, 130.63 (d,  $J$  = 10.4 Hz), 130.5, 127.7, 127.04 (p,  $J$  = 4.6 Hz, x2), 123.11 (dd,  $J$  = 12.6, 4.2 Hz), 118.9 (x2), 114.9, 112.50 (dd,  $J$  = 22.8, 3.9 Hz), 107.0, 105.67 (dd,  $J$  = 26.4, 22.9 Hz), 100;  $^{19}F$  NMR (377 MHz, Chloroform- $d$ )  $\delta$  85.18 (p,  $J$  = 150.6, 149.7 Hz), 63.49 (d,  $J$  = 150.0 Hz), -103.91 (qd,  $J$  = 8.3, 5.7 Hz), -115.12 (q,  $J$  = 8.9 Hz). ESMS, calculated  $m/z$   $C_{20}H_{11}BrF_7N_3OS_2$  586.9  $[M]^+$ , found  $m/z$  (relative intensity) 585.9  $[M-H]^-$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{20}H_{11}BrF_7N_3OS_2$  : 585.9493; found: 585.9484

**5-(5-Bromothiophen-2-yl)-1-(2,4-dichlorophenyl)-N-(4-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (9d)**

**9d** (60 mg, 40 %) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  8.82 (s, 1H), 7.82 – 7.70 (m, 4H), 7.61 (d,  $J$  = 1.9 Hz, 1H), 7.52 – 7.40 (m, 2H), 7.19 (s, 1H), 6.95 (d,  $J$  = 3.9 Hz, 1H), 6.71 (d,  $J$  = 3.9 Hz, 1H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  159.1, 149.13 (p,  $J$  = 17.9 Hz), 147.5, 140.6, 140.2, 137.6, 135.1, 134.1, 130.7, 130.7, 130.6, 130.5, 128.4, 127.6, 127.07 (p,  $J$  = 4.3 Hz, x2), 118.9 (x2), 114.9, 106.6;  $^{19}F$  NMR (377 MHz, Chloroform- $d$ )  $\delta$  85.17 (p,  $J$  = 150.7, 149.7 Hz), 63.48 (d,  $J$  = 150.0 Hz). ESMS, calculated  $m/z$   $C_{20}H_{11}BrCl_2F_5N_3OS_2$  618.9  $[M]^+$ , found  $m/z$  (relative intensity) 617.8  $[M-H]^-$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{20}H_{11}BrCl_2F_5N_3OS_2$  : 617.8902; found: 617.8894

**Lithium Salt of ethyl 4-(4-chlorophenyl)-3-methyl-2,4-dioxobutanoate (10).**

To a magnetically stirred solution of lithium bis(trimethylsilyl)amide (64 mL, 63.93 mmol, 1.0 M in THF) in diethyl ether (63 mL) at -78 °C, 1-(2-thienyl)-1-propanone (10 g, 58.12 mmol) in diethyl ether (73 mL) was added dropwise under a nitrogen atmosphere. After the mixture was stirred at the same temperature for an additional 45 min, diethyl oxalate (9 mL, 63.93 mmol) was added dropwise. The reaction mixture was allowed to warm to room temperature and stirred for another 16 h. The reaction precipitate was filtered, washed with diethyl ether, and dried under vacuum to afford the crude lithium salt **10** (8.05 g, 50%) as a pale-yellow solid. The product was used without further purification.

**Ethyl 5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxylate (11).**

To a solution of lithium salt **10** (8.05 g, 28.73 mmol) in ethanol (99 mL) was added 2,4-dichlorophenylhydrazine hydrochloride (6.88 g, 31.60 mmol) in one portion at room temperature under nitrogen. The resulting mixture was stirred at the same temperature for 24 h. After reaction was complete, the precipitate was filtered, washed with ethanol and diethyl ether, and dried under vacuum to give a light-yellow solid (6.1 g). This crude solid, without purification, was dissolved in acetic acid (45 mL) and heated to reflux for 16 h. The reaction mixture was poured into ice-water and extracted with ethyl acetate (3  $\times$  50 mL). The combined extracts were washed with water, saturated aqueous sodium bicarbonate, and brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Purification by flash column chromatography on silica gel with Hexane:Acetate (8:2) gave ester **11** (3.44 g, 30% over two steps) as a white solid:  $^1H$  NMR (400 MHz, Chloroform- $d$ )  $\delta$  7.38 (d,  $J$  = 2.2 Hz, 1H), 7.34 (d,  $J$  = 8.4 Hz, 1H), 7.32 – 7.27 (m, 3H), 7.07 (d,  $J$  = 8.6 Hz, 2H), 4.45 (q,  $J$  = 7.1 Hz, 2H), 2.33 (s, 3H), 1.43 (t,  $J$  = 7.1 Hz, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  163.2, 143.4, 143.3, 136.5, 136.3, 135.4, 133.5, 131.3 (x2), 131.2, 130.5, 129.3(x2), 128.2, 127.5, 119.6, 61.4, 14.9, 10.1. ESMS, calculated  $m/z$   $C_{19}H_{15}Cl_3N_2O_2$  408.0  $[M]^+$ , found  $m/z$  (relative intensity) 409.0  $[M+H]^+$  (100)

**5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxylic acid (12).**

To a solution of bromo ester **3** (2.32 g, 5.54 mmol) in methanol (26 mL) was added potassium hydroxide (0.72 g, 11.09 mmol) in methanol (8 mL) dropwise at room temperature. The resulting mixture was heated to reflux for 3 h. After hydrolysis was complete, the reaction mixture was cooled to room temperature, poured into ice-water, and acidified with 2N hydrochloric acid. The precipitate was filtered, washed with water, and dried under vacuum to give the carboxylic acid **12** (2.10 g, 98%) as a white solid:  $^1H$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  7.73 (d,  $J$  = 2.3 Hz, 1H), 7.67 (d,  $J$  = 8.5 Hz, 1H), 7.54 (dd,  $J$  = 8.5, 2.3 Hz, 1H), 7.42 (d,  $J$  = 8.4 Hz, 2H), 7.21 (d,  $J$  = 8.4 Hz, 2H), 2.20 (s, 3H);  $^{13}C$  NMR (101 MHz, DMSO)  $\delta$  164.1, 143.4,

142.9, 136.1, 135.6, 134.3, 132.3, 132.1, 131.8 (x2), 130.0, 129.2(x2), 128.8, 127.4, 118.4, 9.9. ESMS, calculated  $m/z$   $C_{17}H_{11}Cl_3N_2O_2$  380.0  $[M]^+$ , found  $m/z$  (relative intensity) 381.0  $[M+H]^+$  (100)

### General Procedure for the Synthesis of Compounds 13a – 13g.

The general procedure is illustrated below for compound 13a.

#### 5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carbonyl chloride

A solution of the crude acid **12** (1 g, 2.57 mmol) and thionyl chloride (565  $\mu$ l, 7.70 mmol) in toluene (17.1 mL) was refluxed for 3 h. Solvent was evaporated under reduced pressure, and the residue was then re-dissolved in toluene (10 mL) first and then in Hexane (10 ml); after evaporation the crude acyl chloride (0.98 g, 95% yield) was obtained as a white solid.

#### 5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (13a)

A solution in dichloromethane of the acyl chloride obtained previously (0.96 ml, 0.41 M), was added dropwise to a solution of 3-(trifluoromethyl)aniline (55  $\mu$ l, 0.44 mmol) and triethylamine (61  $\mu$ l, 0.44 mmol) in dichloromethane (0.4 ml) at 0 °C. After stirring at room temperature for 16 h, to the reaction mixture was added water and the organic phase was extracted with dichloromethane (3 x 3 mL). The combined extracts were washed with brine, dried over anhydrous sodium sulfate, filtered, and evaporated. Flash column chromatography on silica gel with n-Hexane / ethyl acetate (8:2) gave carboxamide **13a** (160 mg, 70% yield) as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.91 (s, 1H), 8.03 (t, J = 1.9 Hz, 1H), 7.86 (d, J = 8.3 Hz, 1H), 7.51 – 7.41 (m, 2H), 7.40 – 7.27 (m, 5H), 7.12 – 7.07 (d, J = 8.5 Hz, 2H), 2.43 (s, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.6, 144.4, 143.7, 138.4, 136.3, 135.7, 135.2, 133.0, 131.40 (q, J = 32.3 Hz), 130.8 (x2), 130.5, 130.4, 129.5, 129.0 (x2), 128.0, 126.8, 122.55, 122.53 (q, J = 272.0 Hz), 120.50 (q, J = 3.9 Hz), 118.4, 116.32 (q, J = 4.0 Hz), 9.5;  $^{19}F$  NMR (376 MHz, Chloroform-d)  $\delta$  -62.74. ESMS, calculated  $m/z$   $C_{24}H_{15}Cl_3F_3N_3O$  523  $[M]^+$ , found  $m/z$  (relative intensity) 546.0  $[M+Na]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{24}H_{15}Cl_3F_3N_3O$  : 524.0311; found: 524.0301

#### 5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(3-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (13b)

**13b** (182 mg, 70% yield) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.92 (s, 1H), 8.16 (t, J = 2.1 Hz, 1H), 7.91 (d, J = 9.0 Hz, 1H), 7.55 – 7.43 (m, 3H), 7.39 – 7.31 (m, 4H), 7.12 (d, J = 8.5 Hz, 2H), 2.45 (s, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.6, 154.25 (appt, J = 17.4 Hz), 144.2, 143.7, 138.3, 136.3, 135.7, 135.2, 133.0, 130.8 (x2), 130.5, 130.4, 129.2, 129.0 (x2), 128.0, 126.8, 122.4, 121.24 (p, J = 3.9, 3.4 Hz), 118.4, 117.22 (p, J = 4.9 Hz), 9.5;  $^{19}F$  NMR (377 MHz, Chloroform-d)  $\delta$  84.17 (p, J = 150.4 Hz), 62.72 (d, J =

150.0 Hz). ESMS, calculated  $m/z$   $C_{23}H_{15}Cl_3F_5N_3OS$  583.0  $[M]^+$ , found  $m/z$  (relative intensity) 582.0  $[M-H]^-$  (56). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{23}H_{15}Cl_3F_5N_3OS$  : 582.0000; found: 581.9991

#### N-(3-(tert-butyl)phenyl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (13c)

**13c** (88 mg, 63% yield) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.76 (s, 1H), 7.64 – 7.59 (m, 2H), 7.46 (t, J = 1.3 Hz, 1H), 7.35 – 7.25 (m, 5H), 7.18 – 7.13 (m, 1H), 7.10 (d, J = 8.5 Hz, 2H), 2.44 (s, 3H), 1.33 (s, 9H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.4, 152.2, 145.0, 143.4, 137.6, 136.2, 135.8, 135.0, 133.1, 130.9 (x2), 130.6, 130.4, 128.9 (x2), 128.6, 128.0, 127.1, 121.1, 118.2, 117.1, 116.9, 34.8, 31.4 (x3), 9.6. ESMS, calculated  $m/z$   $C_{27}H_{24}Cl_3N_3O$  511.1  $[M]^+$ , found  $m/z$  (relative intensity) 512.1  $[M+H]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{27}H_{24}Cl_3N_3O$  : 512.1063; found: 512.1052

#### 5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(4-(trifluoromethyl)phenyl)-1H-pyrazole-3-carboxamide (13d)

**13d** (141 mg, 58% yield) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.93 (s, 1H), 7.82 (d, J = 8.3 Hz, 2H), 7.60 (d, J = 8.3 Hz, 2H), 7.47 (s, 1H), 7.37 – 7.28 (m, 4H), 7.09 (d, J = 8.5 Hz, 2H), 2.43 (s, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.6, 144.4, 143.7, 141.0, 136.3, 135.7, 135.2, 133.0, 130.8 (x2), 130.5, 129.0 (x2), 128.2, 128.0, 126.8, 126.27 (q, J = 3.7 Hz, x2), 125.55 (q, J = 31.9 Hz), 124.93 (q, J = 272.0 Hz), 119.2 (x2), 118.5, 9.5. ESMS, calculated  $m/z$   $C_{24}H_{15}Cl_3F_3N_3O$  523  $[M]^+$ , found  $m/z$  (relative intensity) 524.0  $[M+H]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{24}H_{15}Cl_3F_3N_3O$  : 524.0311; found: 524.0300

#### 5-(4-Chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-N-(4-(pentafluoro- $\lambda$ 6-sulfanyl)phenyl)-1H-pyrazole-3-carboxamide (13e)

**13e** (177 mg, 68% yield) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.93 (s, 1H), 7.79 (d, J = 9.0 Hz, 2H), 7.77 – 7.69 (m, 2H), 7.47 (d, J = 2.1 Hz, 1H), 7.37 – 7.28 (m, 4H), 7.09 (d, J = 8.4 Hz, 2H), 2.43 (s, 3H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.6, 148.97 (appt, J = 17.0 Hz), 144.2, 143.8, 140.6, 136.4, 135.6, 135.2, 133.0, 130.8 (x2), 130.43, 130.42 129.0 (x2), 128.0, 126.97 (p, J = 4.6 Hz, x2), 126.72, 118.8 (x2), 118.5, 9.5;  $^{19}F$  NMR (377 MHz, Chloroform-d)  $\delta$  85.34 (p, J = 150.1, 149.1 Hz), 63.53 (d, J = 149.9 Hz). ESMS, calculated  $m/z$   $C_{23}H_{15}Cl_3F_5N_3OS$  582.0  $[M]^+$ , found  $m/z$  (relative intensity) 582.0  $[M-H]^-$  (64). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{23}H_{15}Cl_3F_5N_3OS$  : 582.0000; found: 581.9990

#### N-(4-(tert-butyl)phenyl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-1H-pyrazole-3-carboxamide (13f)

**13f** (162 mg, 78% yield) was obtained as a white solid:  $^1H$  NMR (400 MHz, Chloroform-d)  $\delta$  8.71 (s, 1H), 7.60 (d, J = 8.7 Hz, 2H), 7.46 (dd, J = 1.8, 0.8 Hz, 1H), 7.36 (d, J = 8.7 Hz, 2H), 7.33 – 7.29 (m, 4H), 7.09 (d, J = 8.5 Hz, 2H), 2.43 (s, 3H), 1.32 (s, 9H);  $^{13}C$  NMR (101 MHz,  $CDCl_3$ )  $\delta$  160.4, 147.0,

145.0, 143.4, 136.1, 135.9, 135.2, 135.0, 133.0, 130.9 (x2), 130.6, 130.4, 128.9 (x2), 127.9, 127.1, 125.8 (x2), 119.5 (x2), 118.2, 34.4, 31.4 (x3), 9.5. ESMS, calculated  $m/z$   $C_{27}H_{24}Cl_3N_3O$  511.1  $[M]^+$ , found  $m/z$  (relative intensity) 534.1  $[M+Na]^+$  (100). HRMS  $m/z$   $M^+H^+$  calcd. for  $C_{27}H_{24}Cl_3N_3O$  : 512.1063; found: 512.1053

### Determination of the partition coefficients (LogP)

Measurement of chromatographic capacity factor ( $k'$ ) by C-18 HPLC method was performed using the equation  $k' = (t_r - t_0)/t_0$ ,<sup>47,48</sup> where  $t_0$  is the retention time of unretained substance and  $t_r$  is the compound's retention time. The mobile phase was prepared mixing methanol with water in proportions of 85:15 and the flow rate was 1ml/min. A solution of urea in a methanol – water solvent (85:15) was used for measurement of the column dead time ( $t_0 = 2,673 \pm 0,002$ ,  $n=3$ ). Seven compounds having known LogP values, tabulated and in the range from 1.10 to 5.70 (Benzyl alcohol, LogP 1.10; Benzene, LogP 2.10; Toluene, LogP 2.70; Naphtalene, LogP 3.60; Biphenyl, LogP 4; Phenanthrene, LogP 4.50; Triphenylamine, LogP 5.70) were chosen as a “standard” calibration mixture for the determination of retention times ( $t_r$ ). Every measure was obtained in triplicate and at controlled temperature of 25 °C. Capacity factors ( $k'$ ) were calculated. The Log $k'$  value for each of seven compounds was plotted against its relative lipophilicity value reported in literature, based on the established linear relationship (LogP = 3.64 Log $k'$  + 3.39, correlation coefficient = 0.95). The capacity factor of each compound was determined (the value was obtained on average of three experiments) and the relative LogP value was obtained by extrapolation.

### In Vitro assays

#### Equilibrium Binding Assays

Binding assays were performed with the CB1 receptor agonist, [ $^3H$ ]CP55940 (0.7 nM), 1 mg ml<sup>-1</sup> bovine serum albumin (BSA) and 50 mM Tris buffer containing 0.1mM EDTA and 0.5mM MgCl<sub>2</sub> (pH 7.4), total assay volume 500  $\mu$ l. Binding was initiated by the addition of mouse brain membranes (30  $\mu$ g) or CB2 transfected CHO cells (5 $\mu$ g). Assays were carried out at 37 °C for 60 minutes before termination by addition of ice-cold wash buffer (50 mM Tris buffer, 1 mg ml<sup>-1</sup> BSA) and vacuum filtration using a 24-well sampling manifold (Brandel Cell Harvester) and Whatman GF/B glass-fibre filters that had been soaked in wash buffer at 4°C for 24 h. Each reaction tube was washed five times with a 4-ml aliquot of buffer. The filters were oven-dried for 60 min and then placed in 5 ml of scintillation fluid (Ultima Gold XR, Packard), and radioactivity quantitated by liquid scintillation spectrometry. Specific binding was defined as the difference between the binding that occurred in the presence and absence of 1  $\mu$ M of the corresponding unlabelled ligand and was 70 - 80% of the total binding.

### $\beta$ -Arrestin assays

PathHunter® HEK293 CB<sub>1</sub> Beta-arrestin cells were plated 48 hours before use and incubated at 37°C, 5% CO<sub>2</sub> in a humidified incubator. Compounds were dissolved in DMSO and diluted in OCC media to the required concentrations. 5 $\mu$ l of compound or vehicle solution was added to each well and incubated for 60 minutes at 37°C, 5% CO<sub>2</sub> in a humidified incubator. 5 $\mu$ l of increasing concentrations of CP55940 was added to each well followed by a 90 minute incubation at 37°C, 5% CO<sub>2</sub> in a humidified incubator. 55 $\mu$ l of detection reagent is then added followed by a further 90 minute incubation at room temperature in the dark. Chemiluminescence, indicated as RLU, was measured on a standard luminescence plate reader.

### Solubility tests

The aqueous solubility was measured using laser nephelometry (BMG Labtech Nephelometer) following serial dilution of DMSO stocks into Tris Buffer (50mM Tris HCL, 50mM Tris base and 0.1% BSA) to give final concentrations of 0.01, 0.1, 1 and 10 $\mu$ M and a final DMSO concentration of 0.1%. The amount of laser scatter caused by insoluble particulates (relative nephelometry units) was measured. RFU values 3-fold greater than control (0 $\mu$ M) indicate insolubility.

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### Notes and references

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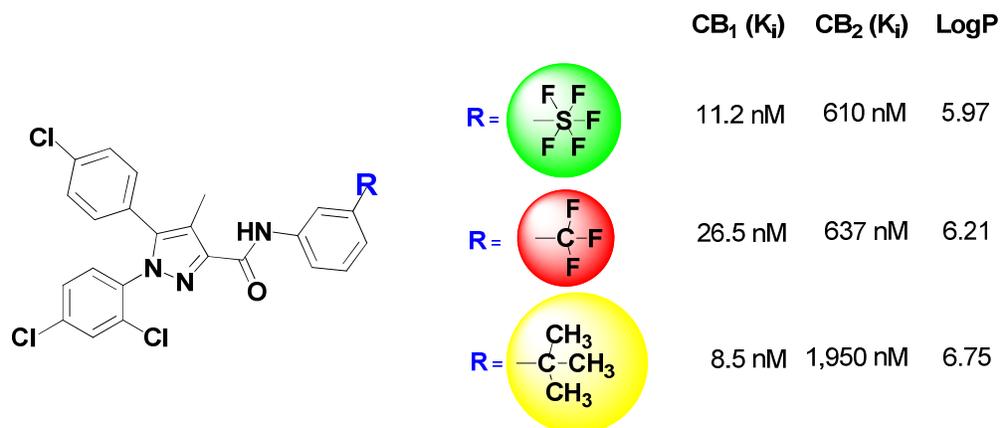
- O'Hagan, David and D. B. Harper, *J. Fluorine Chem.*, 1999, **100**, 127–133.
- S. Purser, P. R. Moore, S. Swallow, and V. Gouverneur, *Chem. Soc. Rev.*, 2008, **37**, 320–330.
- M. Zanda, *New J. Chem.*, 2004, **28**, 1401–1411.
- D. O'Hagan, *J. Fluorine Chem.*, 2010, **131**, 1071–1081.

5. W. A. Sheppard, *J. Am. Chem. Soc.*, 1960, **82**, 4751–4752.
6. J. T. Welch, in *Fluorine In Pharmaceutical And Medicinal Chemistry*, Imperial college press, 2012, vol. 6, pp. 175–207.
7. S. Altomonte and M. Zanda, *J. Fluorine Chem.*, 2012, **143**, 57–93.
8. R. W. Winter, R. A. Dodean, and G. L. Gard, in *Fluorine-Containing Synthons*, ed. V. A. Soloshonok, American Chemical Society, Washington, DC, 2005, vol. 911, pp. 87–118.
9. W. A. Sheppard, *J. Am. Chem. Soc.*, 1965, **87**, 2410–2420.
10. C. Hansch, R. M. Muir, T. Fujita, P. P. Maloney, F. Geiger, and M. Streich, *J. A. Chem. Soc.*, 1963, **85**, 2817–2824.
11. W. A. Sheppard, *J. Am. Chem. Soc.*, 1962, **84**, 3072–3076.
12. J. T. Welch and D. S. Lim, *Bioorg. Med. Chem.*, 2007, **15**, 6659–6666.
13. P. Kirsch, *Modern fluoroorganic chemistry synthesis, reactivity, applications*, Wiley-VCH, Weinheim, 2004.
14. D. Lentz and K. Seppelt, in *Chemistry of Hypervalent Compounds*, ed. K. Akiba, Wiley-VCH, New York, 1999, pp. 295–323.
15. L. J. Sæthre, N. Berrah, J. D. Bozek, K. J. Børve, T. X. Carroll, E. Kuk, G. L. Gard, R. Winter, and T. D. Thomas, *J. A. Chem. Soc.*, 2001, **123**, 10729–10737.
16. A. Kapur, P. Samaniego, G. A. Thakur, A. Makriyannis, and M. E. Abood, *J. Pharmacol. Exp. Ther.*, 2008, **325**, 341–348.
17. S. Munro, K. L. Thomas, and M. Abu-Shaar, *Nature*, 1993, **365**, 61–65.
18. A. C. Howlett, F. Barth, T. I. Bonner, G. Cabral, P. Casellas, W. A. Devane, C. C. Felder, M. Herkenham, K. Mackie, B. R. Martin, R. Mechoulam, and R. G. Pertwee, *Pharmacol. Rev.*, 2002, **54**, 161–202.
19. D. Shire, C. Carillon, M. Kaghad, B. Calandra, M. Rinaldi-Carmona, G. L. Fur, D. Caput, and P. Ferrara, *J. Biol. Chem.*, 1995, **270**, 3726–3731.
20. E. Ryberg, H. K. Vu, N. Larsson, T. Groblewski, S. Hjorth, T. Elebring, S. Sjögren, and P. J. Greasley, *FEBS Lett.*, 2005, **579**, 259–264.
21. M. Herkenham, A. B. Lynn, M. D. Little, M. R. Johnson, L. S. Melvin, B. R. de Costa, and K. C. Rice, *Proc. Natl. Acad. Sci. USA*, 1990, **87**, 1932–1936.
22. G. Griffin, S. R. Fernando, R. A. Ross, N. G. McKay, M. L. J. Ashford, D. Shire, J. W. Huffman, S. Yu, J. A. H. Lainton, and R. G. Pertwee, *Eur. J. Pharmacol.*, 1997, **339**, 53–61.
23. R. G. Pertwee, *Life Sci.*, 1999, **65**, 597–605.
24. J. C. Ashton, D. Friberg, C. L. Darlington, and P. F. Smith, *Neurosci. Lett.*, 2006, **396**, 113–116.
25. J. Horder, M. Browning, M. Di Simplicio, P. J. Cowen, and C. J. Harmer, *J. Psychopharmacol.*, 2012, **26**, 125–132.
26. G. Kunos, D. Osei-Hyiaman, S. Bátkai, K. A. Sharkey, and A. Makriyannis, *Trends Pharmacol. Sci.*, 2009, **30**, 1–7.
27. E. Kirilly, X. Gonda, and G. Bagdy, *Acta Physiol.*, 2012, **205**, 41–60.
28. B.-C. Ho, T. H. Wassink, S. Ziebell, and N. C. Andreasen, *Schizophr. Res.*, 2011, **128**, 66–75.
29. B. Costa, A. E. Trovato, M. Colleoni, G. Giagnoni, E. Zarini, and T. Croci, *Pain*, 2005, **116**, 52–61.
30. P. Gazzerò, M. G. Caruso, M. Notarnicola, G. Misciagna, V. Guerra, C. Laezza, and M. Bifulco, *Int. J. Obesity*, 2006, **31**, 908–912.
31. M. Rinaldi-Carmona, F. Barth, M. Héaulme, D. Shire, B. Calandra, C. Congy, S. Martinez, J. Maruani, G. Néliat, D. Caput, P. Ferrara, P. Soubrié, J. C. Brelière, and G. Le Fur, *FEBS Lett.*, 1994, **350**, 240–244.
32. R. Lan, Q. Liu, P. Fan, S. Lin, S. R. Fernando, D. McCallion, R. Pertwee, and A. Makriyannis, *J. Med. Chem.*, 1999, **42**, 769–776.
33. P. Lazzari, S. Ruiu, G. A. Pinna, and G. Murineddu, 2005.
34. R. D. Bowden, P. J. Comina, M. P. Greenhall, B. M. Kariuki, A. Loveday, and D. Philp, *Tetrahedron*, 2000, **56**, 3399–3408.
35. S.-L. Tseng, M.-S. Hung, C.-P. Chang, J.-S. Song, C.-L. Tai, H.-H. Chiu, W.-P. Hsieh, Y. Lin, W.-L. Chung, C.-W. Kuo, C.-H. Wu, C.-M. Chu, Y.-S. Tung, Y.-S. Chao, and K.-S. Shia, *J. Med. Chem.*, 2008, **51**, 5397–5412.
36. A. Leo, C. Hansch, and D. Elkins, *Chem. Rev.*, 1971, **71**, 525–616.
37. H. Fan, H. T. Ravert, D. P. Holt, R. F. Dannals, and A. G. Horti, *J. Labelled Compd. Radiopharm.*, 2006, **49**, 1021–1036.
38. S. Tobishi, T. Sasada, Y. Nojiri, F. Yamamoto, T. Mukai, K. Ishiwata, and M. Maeda, *Chem. Pharm. Bull.*, 2007, **55**, 1213–1217.
39. R. A. Ross, H. C. Brockie, L. A. Stevenson, V. L. Murphy, F. Templeton, A. Makriyannis, and R. G. Pertwee, *Br. J. Pharmacol.*, 1999, **126**, 665–672.
40. J.-Z. Chen, X.-W. Han, Q. Liu, A. Makriyannis, J. Wang, and X.-Q. Xie, *J. Med. Chem.*, 2006, **49**, 625–636.
41. M. R. Price, G. L. Baillie, A. Thomas, L. A. Stevenson, M. Easson, R. Goodwin, A. McLean, L. McIntosh, G. Goodwin, G. Walker, P. Westwood, J. Marrs, F. Thomson, P. Cowley, A. Christopoulos, R. G. Pertwee, and R. A. Ross, *Mol. Pharmacol.*, 2005, **68**, 1484–1495.
42. D. A. Jackson and S. A. Mabury, *Environ. Toxicol. Chem.*, 2009, **28**, 1866.
43. B. Hoelke, S. Gieringer, M. Arlt, and C. Saal, *Anal. Chem.*, 2009, **81**, 3165–3172.
44. M. M. C. van D. Lee, M. Bras, C. J. van Koppen, and G. J. R. Zaman, *J. Biomol. Screen.*, 2008, **13**, 986–998.
45. B. D. Kangas, M. S. Delatte, V. K. Vemuri, G. A. Thakur, S. P. Nikas, K. V. Subramanian, V. G. Shukla, A. Makriyannis, and J. Bergman, *J. Pharmacol. Exp. Ther.*, 2013, **344**, 561–567.
46. W. C. Still, M. Kahn, and A. Mitra, *J. Org. Chem.*, 1978, **43**, 2923–2925.
47. Organisation for Economic Co-operation and Development, *Test No. 117: Partition Coefficient (n-octanol/water), HPLC Method*, OECD Publishing, Paris, 2004.
48. X. Fei and Q. Zheng, *J. Liq. Chromatogr. Relat. Technol.*, 2005, **28**, 939–945.

Graphical abstract

## The Pentafluorosulfanyl Group in Cannabinoid Receptor Ligands: Synthesis and Comparison with Trifluoromethyl and *tert*-Butyl Analogues

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Competitive CB<sub>1</sub> receptor antagonists carrying an aromatic SF<sub>5</sub> group in position 3 of a pyrazole ring were synthesised and compared with their CF<sub>3</sub> and *tert*-butyl analogues, showing (1) LogP values in the order *tert*-butyl > SF<sub>5</sub> > CF<sub>3</sub>; (2) CB<sub>1</sub> K<sub>i</sub>s (in the nanomolar range) and CB<sub>1</sub>/CB<sub>2</sub> selectivities slightly higher or equivalent than the CF<sub>3</sub> and *tert*-butyl counterparts. This confirms that an aromatic SF<sub>5</sub> group can be used as a bioisosteric analogue of a CF<sub>3</sub> group and possibly of a bulky aliphatic group too.