

Organic & Biomolecular Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

Oxidative Activation of Dihydropyridine Amides to Reactive Acyl Donors

Cite this: DOI: 10.1039/x0xx00000x

Erik Daa Funder,^a Julie B. Trads,^a and Kurt V. Gothelf^aReceived 00th January 2012,
Accepted 00th January 2012

DOI: 10.1039/x0xx00000x

www.rsc.org/

Amides of 1,4-dihydropyridine (DHP) are activated by oxidation for acyl transfer to amines, alcohols and thiols. In the reduced form the DHP amide is stable towards reaction with amines at room temperature. However, upon oxidation with DDQ the acyl donor is activated via a proposed pyridinium intermediate. The activated intermediate reacts with various nucleophiles to give amides, esters, and thio-esters in moderate to high yields.

Introduction

The amide bond is among the most common and important functional groups in chemistry and biochemistry. Thus, it is an integral part of molecules such as proteins, peptides, pharmaceuticals, and polymers.^{1,2,3,4} The classical approach to the formation of an amide is to use a reactive acyl derivative or *in situ* activation of carboxylic acids for the reaction with an amine.⁵ A plethora of efficient reagents have been developed for this transformation, including the arsenal of carbodiimides for *in situ* carboxylic acid activation or the generation of activated esters taking advantage of electron-poor alcohols.^{6,7,8} Reactive acyl derivatives such as acyl chlorides and *N*-hydroxysuccinimides can be stored in pure form or in non-nucleophilic solvents, however they are highly sensitive to moisture and nucleophiles.

An alternative method to create an amide bond is to take advantage of transamidation strategies. In most cases these take advantage of primary amides using either high temperatures (100 to 160 °C) or added metals as essential activating reagents.^{9,10,11,12,13} Others have used tertiary amide derivatives, that can chelate metals such as copper, as protecting groups for carboxylic acids.^{14,15,16,17,18} The strategy resulted in either the free acid or a new ester bond. Also, tertiary and secondary amides, activated by Tf₂O and Zn(OTf)₂ respectively, have resulted in clever ways of creating intra- and inter-molecular ester bonds from amide bonds.^{19,20}

Herein, we present a new activatable acyl donor which in the “off-state” is stable towards nucleophilic attack by e.g. amines, whereas in the “on-state” it reacts with nucleophiles such as amines, alcohols, and thiols. The activation is orthogonal to most other known methods for amide bond formation as it exploits a metal free oxidation to form a highly reactive acyl transfer intermediate.

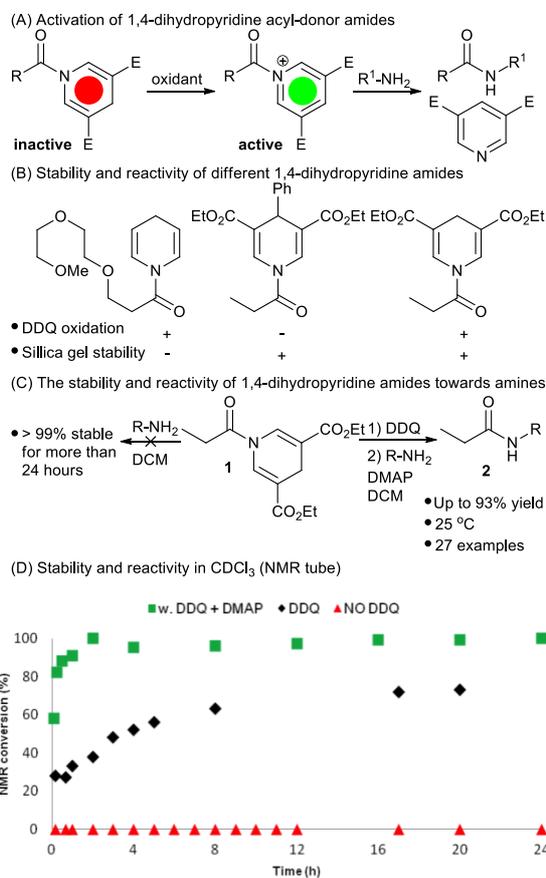
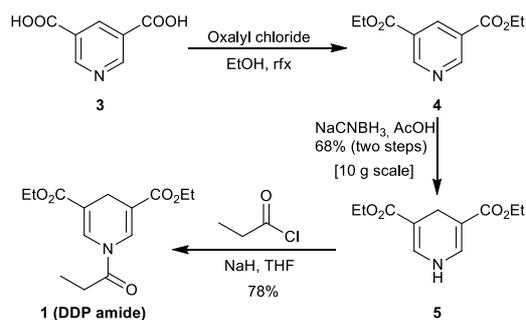


Figure 1. (A) Principle of activation for acyl dihydropyridine donors. By oxidation a reactive pyridinium intermediate is proposed which react with amines to form amides. (B) Three initial DHP-amides investigated for their stability and reactivity towards silica gel purification and DDQ-oxidation. (C) and (D) The stability (without DDQ) and reactivity (with DDQ and additive) of dihydropyridine amide **1** towards amine nucleophiles.

Amides are generally much more stable towards nucleophiles than esters.^{21,22} For this reason, amides are not immediately recognizable as acyl donors. A few examples for the release of quinoline and acridine amides from a solid support have been demonstrated.^{23,24,25} Acyl-pyridinium derivatives are known to be highly reactive acyl donors and molecules such as 4-dimethylaminopyridine (DMAP) are commonly used as acyl-transfer catalysts.^{26,27} Inspired by the redox relay of the pyridinium–1,4-dihydropyridine (DHP) redox couple known as NAD⁺ and NADH, found in all living systems, we speculated if it was feasible to prepare a stable reduced acyl DHP derivative that could be oxidized to form a reactive acyl pyridinium species.^{28,29,30} Thus capturing the inherent reactivity of e.g. an acid chloride in an inactive DHP amide.^{31,32}

Results and discussion

Only a few amides of 1,4-dihydropyridines are described in the literature³³ and their use as acyl donors has, to our knowledge, remained unexplored until the current study. Initially, we investigated the amide of an unsubstituted DHP (Figure 1B).³⁴ This derivative was stable towards hydrolysis and did not react with amines in CDCl₃. Oxidation with DDQ activated the DHP for acyl transfer with amines. However, the unsubstituted DHP amide was not stable towards silica gel flash column chromatography. It appears that it was oxidized by oxygen when exposed to silica gel which diminishes the usefulness of the described approach. In order to avoid degradation on silica gel we investigated electron deficient 3,5-diethoxycarbonyl-DHP derivatives which have higher oxidation potentials.^{35,36} For this reason, an electron deficient 4-phenyl DHP amide was prepared and this proved stable towards purification by silica gel chromatography (Figure 1B). However, this amide was inert towards oxidation by DDQ and CAN. Further fine-tuning of the properties was achieved with the propionyl 3,5-diethoxycarbonyl-DHP (DDP) amide **1**, which had the desired properties of stability and reactivity (Figure 1B and 1C).



Scheme 1. The synthesis of 1,4-dihydropyridine amide **1**

The synthesis of **1** is straight-forward (Scheme 1). It is performed by converting the commercially available, and inexpensive, pyridine-3,5-dicarboxylic acid **3** to the di-ethyl ester **4** by treatment with oxalyl chloride and ethanol. The resulting crude mixture was used directly in the reduction to the 1,4-dihydropyridine **5** using NaCNBH₃ in acetic acid.³⁷

Conveniently, the reaction could be performed on 10 g scale and upon the addition of water DHP **5** precipitates as a yellow powder to provide 68% yield over two steps. The DDP-amide was prepared by deprotonating DHP **5** with NaH followed by the addition of propionyl chloride to furnish DDP amide **1** in an isolated yield of 78%. In addition, amide bond formation with **5** could also be achieved by the use of EDC, HOBT, and amine-base albeit in lower yield of 73%.

The amide **1** was stable towards silica gel flash column chromatography as well as acidic and basic work-up conditions.

Table 1. Optimization of reaction conditions for the oxidation of **1** and subsequent reaction with phenylethylamine^{a,b}

Entry	Base	Amine (eq.)	Solvent	Time (h)	Conversion (%)
1	-	3	CDCl ₃	17	72
2	DIPEA	3	CDCl ₃	2	47
3	2,6-Lutidine	3	CDCl ₃	2	65
4	Pyridine	3	CDCl ₃	2	75
5	K ₂ CO ₃	3	CDCl ₃	2	77
6	CS ₂ CO ₃	3	CDCl ₃	2	100 (50)
7	DMAP	3	CDCl ₃	1	100 (86)
8	DMAP	3	DCM	1	100 (87)
9	DMAP	3	EtOAc	26	(44)
10	DMAP	3	Dioxane	26	(65)
11	DMAP (0.2 eq.)	3	DCM	4	(84)
12	DMAP (1 eq.)	3	DCM	1.5	(94)
13	DMAP (1 eq.)	1	DCM	6	(83)
14	DMAP (3 eq.)	1.1	DCM	1	(90)

^a Reaction conditions for entry 1-10: **1** (0.036 mmol) and DDQ (0.036 mmol) were stirred in 0.6 mL of solvent in a sealed vial (all reactions with isolated yields were performed on 0.18 mmol scale using 3.0 mL of solvent). After 30 minutes phenylethylamine (0.11 mmol) and base (0.11 mmol) were added. Reaction conditions for entry 11-14: **1** (0.18 mmol) and DDQ (0.18 mmol) were stirred in 3 mL DCM in a sealed vial (entry 14 was performed with DDQ (1 eq.) and **1** (1.1 eq.)). After 30 minutes phenylethylamine and base were added according to the table. ^b Conversions were determined by ¹H-NMR spectroscopy based on **1**. Isolated yields are reported in parenthesis.

With these preliminary results at hand, we tested the stability and reactivity of the DDP amide **1**. Without activation the amide was stable (>99% by ¹H-NMR in CDCl₃) towards aminolysis with 1 eq of phenylethylamine over a period of more than 48 h at rt, 40 and 60 °C. With 3 eq DMAP and 1 eq amine present at rt the stability in CDCl₃ was determined by ¹H-NMR to >99% after 24 h and >98% after 48 h. After raising the temperature to 40 and 60 °C and using a prolonged reaction time (48 h) the stability of the amide was determined to be >94% and >93% respectively. In addition, 14 other solvents were investigated in order to explore solvent effects on the amide bond formation and it was observed that the amide was slightly less stable in more polar solvents (see supporting information for specific details). For storage the DDP amide **1** was determined to be completely stable to oxidation in the freezer under argon for a period of more than 4 months.

ARTICLE

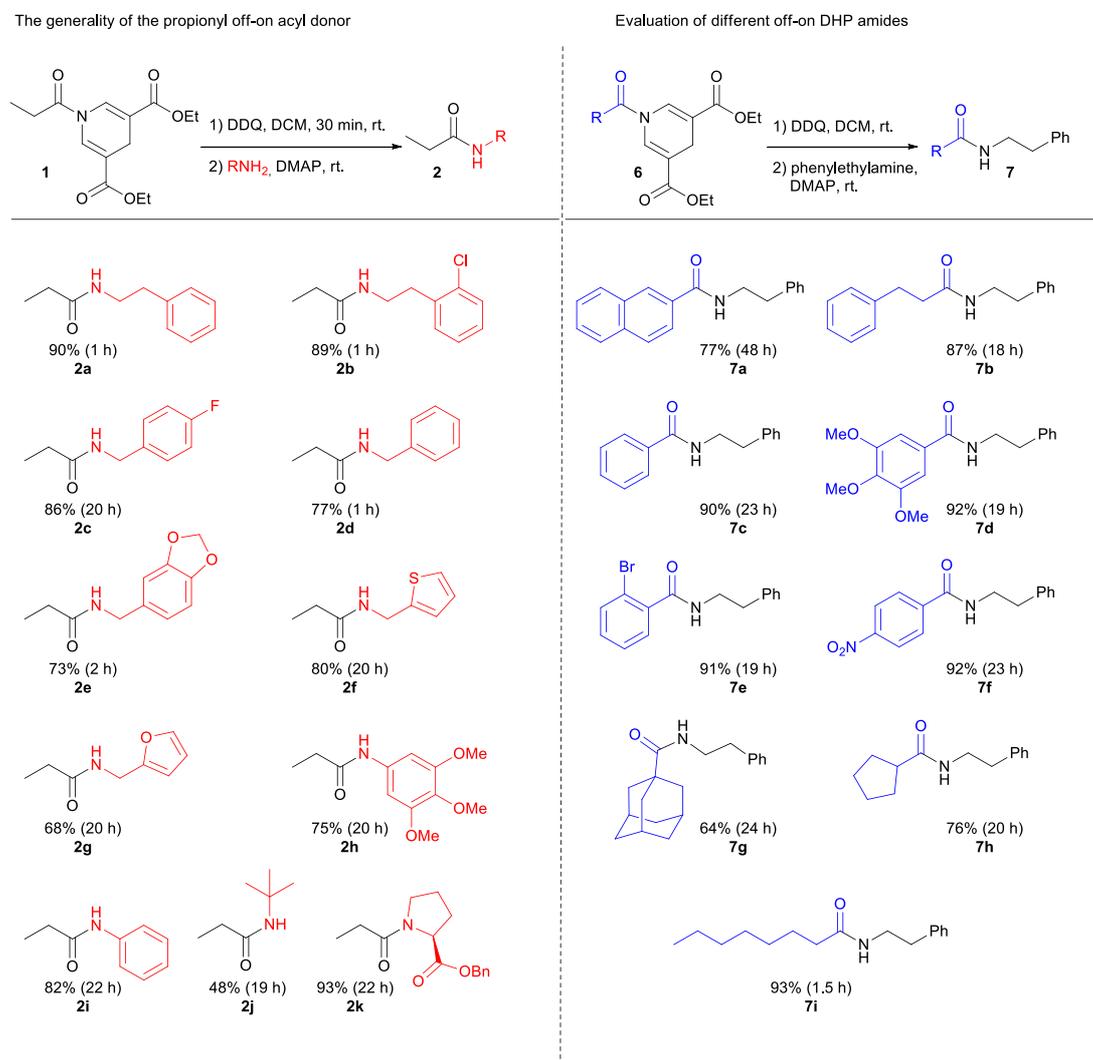


Figure 2. The generality of DDP acyl donor **1** with various amines and different DHP amides.

In the event of DDP amide **1** oxidation using DDQ with subsequent addition of amine, the reaction generates the desired amide **2a** (Table 1).³⁸ Initially, the reaction was, however, rather slow providing only 70% conversion after 17 h. After carefully investigating the mechanism using HPLC analysis of the crude-reaction mixture we identified the different intermediates in the reaction. From this we concluded that the proposed acyl pyridinium intermediate, formed after oxidation, is immediately trapped as the mono- or di-dichlorodicyano hydroquinone ester, since these intermediates were identified using HPLC analysis (see supporting information). Such electron-deficient esters are reactive towards acyl transfer, though less active than the acyl pyridinium salt. To increase the

rate of the reaction we have investigated the effect of adding various bases and acyl transfer catalysts. The results are presented in Table 1.

The addition of weak non-nucleophilic bases increased the rate of the reaction to some extent. Addition of DMAP increased the rate of the reaction significantly and complete conversion was observed after only 1 h (this included 30 min oxidation) as shown by ¹H-NMR. Also, carbonate bases increased the conversion significantly, however, the isolated yields were inferior to the yields obtained using DMAP in DCM or chloroform. As DCM is a less toxic alternative to chloroform while still preserving the reactivity of the system, it was used in the following optimization with regard to yield. Next the

stoichiometry of the amine and DMAP were systematically varied (Table 1). It was observed that the reaction proceeds well using only catalytic amounts of DMAP and 3 eq of amine, yielding the product in a satisfactory 84% yield after 4 h. However, in order to decrease the equivalents of amine used and increase the rate of the reaction, other conditions were investigated. The optimal conditions were identified using 3 eq DMAP and 1.1 eq amine. The optimized reaction conditions gave rise to an isolated yield of **2a** in 90% yield after a total reaction time of only 1 h.

To probe the generality of the reaction, different amine nucleophiles were employed in the reaction with **1** (Figure 2). Primary aliphatic amines furnished good yields using a total reaction time of only 1 h. Electron-rich and electron-poor benzylic amines provided yields from 73 to 86% with reaction times between 1 and 20 h.

Heteroaryl methylene amines such as furfuryl amine and 2-thiophenemethylamine also proved reactive giving rise to yields up to 80%.

In order to test the reactivity of the acyl donor's on-state, neutral and electron-rich anilines were investigated. Although slower, these amines proved viable in the amide bond forming reaction providing isolated yields between 75 and 82%.

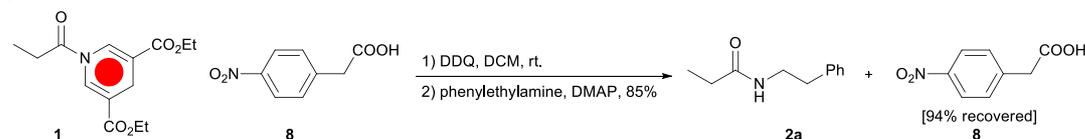
Furthermore, sterically hindered *tert*-butyl amine gave rise to 48% yield whereas the secondary amine in benzyl protected proline furnished an excellent yield of 93% after a total reaction time of 22 h.

To expand the generality of the reaction different 1,4-dihydropyridine amides were synthesized and investigated in the reaction with phenylethylamine. After oxidation, electron-rich, neutral, and electron-poor aromatic benzoyl DHP amides provided the desired amide product in excellent yields ranging from 90 to 92%. Primary and secondary aliphatics as well as sterically hindered adamantyl-DHP also provided good yields ranging from 64 to 93%. In addition, naphthalene-DHP amide gave rise to a yield of 77% after a total reaction time of 48 h. The oxidation with DDQ was in most cases complete within 30 minutes. However, in a few cases the oxidation was slower, as in the case with the naphthalene-DHP where the oxidation was complete after approximately 24 h.

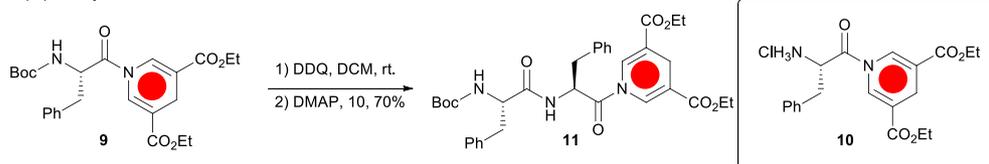
The most notable characteristic of the reaction is the off-on feature, which prevents aminolysis and hydrolysis without prior activation and permits the reaction only in the presence of an activator. Using this methodology it is possible to have a free acid moiety present during the activation step (Figure 3A). Thus, only one of the acids has the possibility of being activated and the free acid remains unscathed (94% recovery by flash column chromatography) in the amide bond forming step. It is not possible to obtain such off-on reactivity with conventional amide bond forming reagents without the use of protecting groups. We envision that this conceptually new reactivity can be used in directed chemistry thus enhancing specificity and suppressing unwanted reactions.

(A) Specific amide-bond activation in the presence of other carboxylic acids, off-on acyl donor amino acids, and the synthesis of 2,5-diketopiperazines.

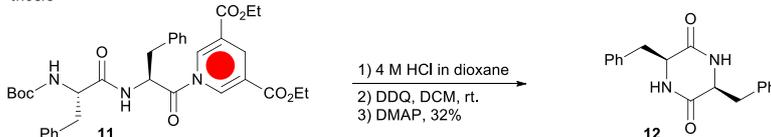
Specific amide-bond activation



Di-peptide synthesis



Diketopiperazine synthesis



(B) The off-on acyl donors reactivity with O and S-nucleophiles

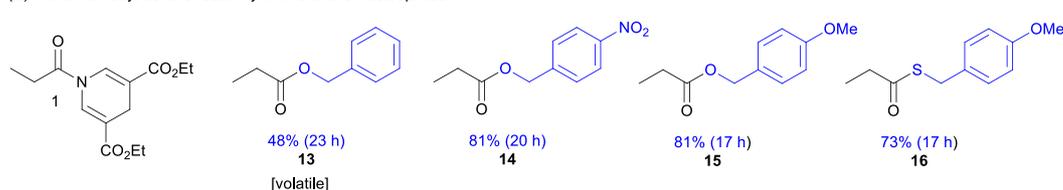


Figure 3. (A) The application of DHP amides in specific amide-bond activation, peptide synthesis, and the intramolecular synthesis of 2,5-diketopiperazines. (B) The reactivity of the DDP amide **1** with oxygen and sulfur nucleophiles.

ARTICLE

One of the drawbacks of the reaction is that the amine has to be added after the oxidation step as it will otherwise react with DDQ. This obstacle can however be overcome by using the ammonium salt of the amine during the oxidation step, which prevents unwanted side reactions with DDQ and gives rise to an isolated yield of the desired amide product in 87% (see supporting information).

To explore the applications of the optimized off-on acyl donor system, we first synthesized a Boc-protected phenyl-alanine DHP-amide (Figure 3A). Secondly, a phenylalanine DHP-amide with a free amine was obtained. We envisioned that this approach would allow for the iterative synthesis of amide bonds. Gratifyingly, the amide bond formation between the two fragments resulted in a phenylalanine DHP di-peptide in an isolated yield of 70%, which can be activated for further couplings. The crude ¹H-NMR analysis of the reaction mixture, after workup, indicated a 85:15 diastereomeric ratio of the dipeptide. Next, we also demonstrated the formation of a cyclic dipeptide by an analogous intramolecular reaction (Figure 3A).

Finally, other nucleophiles besides amines were investigated. Employing either alcohols or thiols provided the corresponding esters and thio-esters in yields between 48 and 81% yield. Thus the DHP-amide can function as a useful ester or thio-ester surrogate.³⁹ The off-on switchable DHP-acyl donor is the first example of its kind which provides an off-state that is non-reactive at room temperature and reactive when activated. The advantage of this method is two-fold. It provides shielding of the acyl group in the off-state and thus carboxylic acids located elsewhere in a molecule can undergo reactions selectively. Compared to the free carboxylic acid group it is non-acidic and provides better solubility of the compound in organic solvents. Compared to protected carboxylic acids such as methyl, benzyl, and tert-butyl esters it offers an orthogonal (oxidation) one-pot route to the activated acyl derivative. In terms of yield and reaction rate the DHP-acyl donor method presented here cannot compete with the traditional activation by carbodiimides and other reagents that have been explored and optimized exhaustively. Other drawbacks of the method is that the amine (unless protonated) cannot be present during the oxidation and that 3-fold of the amine is used to give an optimal yield. However, we believe that the DHP-acyl donor method can be further optimized, e.g. by improving the oxidation reagent and optimizing the substituents of the DHP. Furthermore, other methods for activation such as metal-catalyzed activation by oxygen or photochemical activation may become feasible. We are exploring the application of the DHP-acyl donor in aqueous media as we believe that next generation of the method may have potential for bioconjugation.

Conclusions

In summary, we have described the development of an off-on switchable acyl donor in the form of different 1,4-dihydropyridine amides. In the absence of activating reagent (DDQ) the amide is inert towards aminolysis and hydrolysis. In contrast, after activation by DDQ, the DHP-system is activated to a proposed pyridinium intermediate which rapidly is captured by reduced DDQ as either a mono- or di-ester. These then readily reacts with a variety of nitrogen, oxygen, and sulfur nucleophiles. Reaction times can be as short as one hour with yields up to 93% thus demonstrating the reactivity of the on-stage. In addition to these findings the DHP-amides have been utilized in amide couplings in the presence of a free carboxylic acid and for peptide couplings.

Experimental Section

General methods

For all water and/or oxygen sensitive reactions were used flame dried glassware, which was purged with argon and evacuated a total of three times. All reactions were monitored by thin layer chromatography (TLC) analysis on either Merck® silica gel 60 F254 TLC plates or TLC Silica gel 60 F254 glass plates. The TLC plates were visualized by exposure to UV (254 nm) or by staining with either a basic aq. KMnO₄ solution or *p*-anisaldehyde stain. Flash column chromatography was performed using Merck® silica gel 60 (230-400 mesh). Preparative TLC was carried out using TLC Silica gel 60 F254 glass plates. All solvents used were of HPLC grade quality. For inert reactions the solvents THF and DCM were dried prior to use by the use of a MBRAUN solvent purification system 800. Triethylamine was distilled from CaH₂ before use. Unless noted all reagents are commercially available. The reagents were purchased at the highest quality and used without further purification.

NMR spectra were recorded at 400 MHz (¹H-NMR) and at 100 MHz (¹³C-NMR) on a Bruker® 400 spectrometer and calibrated to the residual solvent peak. In the interpretation of the ¹H-NMR spectra the following abbreviations are used: s, singlet; d, doublet; t, triplet; q, quartet; qu, quintet; sext, sextet; m, multiplet; br, broad. Melting points are uncorrected. High-resolution mass spectrometry (HRMS) was recorded using electrospray ionization. LC-MS measurements were done on a Shimadzu LCMS-2020EV connected to a Shimadzu prominence RP-ULPC system equipped with a YMCbasic 50 x 4.6 mm i.d. S-3 μm column using a gradient from 20 to 95% MeCN in a solution of 0.1% formic acid in MilliQ purified water.

Diethyl 1,4-dihydropyridine-3,5-dicarboxylate (5)³⁷ A stirred suspension of pyridine-3,5-dicarboxylic acid (10 g, 60 mmol, 1 eq.) in ethanol (99.5%, 100 mL) was cooled to 0 °C after which oxalyl chloride (23 mL, 270 mmol, 4.7 eq.) was added. The solution was heated to reflux for 2h where after a clear solution was obtained. The solvent was removed *in vacuo* followed by three rounds of co-evaporation with toluene. The resulting crude mixture was then dissolved in glacial acetic acid (94 mL), cooled using an ice bath, and slowly added NaCNBH₃ (13.2 g, 209 mmol, 3.5 eq.). After completion by TLC analysis (approximately 5 hours) the reaction was quenched by the addition of water (520 mL). The resulting heterogeneous mixture was cooled using an ice bath after which the yellow precipitate was collected by filtration under suction and dried under high vacuum overnight, thus obtaining the product as a yellow solid (9.8 g, 68%). R_f = 0.18 (silica, NEt₃/EtOAc/DCM 2:1:97 v/v/v). mp (uncorr.) 124.4-132.3 °C (lit.⁴⁰ 122-123 °C). IR (neat) ν_{max} 2995, 2927, 1726 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 5.1 Hz, 2H), 5.76 – 5.61 (m, 1H), 4.19 (q, *J* = 7.2 Hz, 4H), 3.26 (s, 2H), 1.27 (t, *J* = 7.2 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 168.2, 135.0, 105.1, 60.1, 21.5, 14.2. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₁H₁₆NO₄: 226.1074. Found: 226.1074.

Diethyl 1-propionyl-1,4-dihydropyridine-3,5-dicarboxylate (1) To a 0 °C solution of 1,4-dihydropyridine **1** (3.06 g, 13.6 mmol, 1 eq.) in a minimum of dry THF (44 mL) was added NaH (1.22 g, 30.6 mmol, 2.25 eq.) from a 60% suspension in mineral oil. The reaction was left to stir for 30 minutes after which propionyl chloride (1.68 mL, 15.6 mmol, 1.15 eq.) was added. After 24 hours the reaction was diluted with water (10 mL) and extracted with EtOAc (3 x 20 mL). If the two phases in the separation funnel did not separate appropriately brine was added. The combined organic phases were washed with saturated NaHCO₃, dried (MgSO₄), filtered, and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography (silica gel) using EtOAc:DCM (gradient from 1% EtOAc to 5% EtOAc, all eluents with 2% NEt₃) as the eluent. The product was isolated as a light yellow solid (2.97 g, 78%) R_f = 0.50 (silica, NEt₃/EtOAc/DCM 2:1:97 v/v). mp (uncorr.) 107.8-109.3 °C. IR (neat) ν_{max} 2941, 2983, 1720, 1704 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.86 (bs, 2H), 4.23 (q, *J* = 7.1 Hz, 4H), 3.22 (s, 2H), 2.60 (q, *J* = 7.3 Hz, 2H), 1.30 (t, *J* = 7.1 Hz, 6H), 1.22 (t, *J* = 7.3 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 170.3, 166.6, 146.1, 112.4, 61.0, 27.1, 28.2, 14.5, 8.7. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₄H₂₀NO₅: 282.1336. Found: 282.1337.

Diethyl 1-propionyl-1,4-dihydropyridine-3,5-dicarboxylate (1) To a stirred solution of propanoic acid (0.107 g, 124 μL, 1.44 mmol, 1.3 eq.) in DMF 5 mL was added HOBt (0.300 g, 2.22 mmol, 2 eq.), EDC-Cl (0.318 g, 1.66 mmol, 1.5 eq.), DIPEA (0.4 mL, 2.22 mmol, 2 eq.), and DHP (0.250 g, 1.11 mmol, 1 eq.). The resulting mixture was stirred for two days under argon after which it was quenched with water and brine, transferred to a separation funnel, and extracted with EtOAc (x3). The combined organic phases were washed with 0.5 M

HCl and NaHCO₃ (sat. solution), dried (MgSO₄), and evaporated *in vacuo*. The desired compound was isolated using flash column chromatography (silica) using NEt₃:EtOAc:DCM 2:1:97. The product was isolated as a light yellow solid (228 mg, 73%).

General procedure A using various amines with DHP amide **1**

To a stirred solution of 1,4-dihydropyridine amide **1** (110 mg, 0.393 mmol, 1.1 eq.) in DCM (6 mL) was added DDQ (81 mg, 0.355 mmol, 1 eq.) after which the color changed from light yellow to a milky white color. After 30 minutes the amine nucleophile (0.393 mmol, 1.1 eq.) followed by DMAP (131 mg, 1.07 mmol, 3 eq.) were added. The reaction was monitored by TLC and crude ¹H-NMR until completion. Most amines provided full conversion after 0.5 to 1 hours (see individual experiments for exact reaction time) after which the mixture was diluted with EtOAc (20 mL). The mixture was transferred to a separation funnel and washed with 1 M HCl (2 x 10 mL) and saturated NaHCO₃ (2 x 10 mL). The remaining organic phase was dried with MgSO₄ or Na₂SO₄, filtered, and evaporated to dryness *in vacuo*. In most cases the remaining residue consisted exclusively of the amide product and the pyridine **4** (observed by ¹H-NMR in CDCl₃) which cannot be removed by acidic work-up using 1 M HCl. Increasing the acidity of the aqueous wash to 6 M HCl facilitates the protonation and removal of **4**, however the amide products are in many cases not stable towards these conditions or removed from the organic phase.

The products were purified by flash column chromatography using silica gel (see details in the following individual experimentals).

N-Phenethylpropionamide (2a)⁴¹ The general procedure A was followed with a total reaction time of 1 hour (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (57 mg, 90%). R_f = 0.28 (silica, EtOAc:pentane 50:50 v/v). mp (uncorr.) 58.6-60.6 °C (lit.⁴² 57-57.5 °C). IR (neat) ν_{max} 3273, 3070, 2970, 1643, 1552 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.33 – 7.26 (m, 2H), 7.24 – 7.12 (m, 3H), 5.78 (br s, 1H), 3.49 (q, *J* = 7.0 Hz, 2H), 2.80 (t, *J* = 7.1 Hz, 2H), 2.14 (q, *J* = 7.6 Hz, 2H), 1.10 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.5, 139.4, 129.1, 128.9, 126.8, 40.4, 35.5, 29.4, 9.5. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₁H₁₅NO: 178.1226 Found: 178.1230.

The synthesis of **2a** was repeated, however in this case the oxidation was performed in the presence of the amine as its hydrochloride salt. The DDP amide **1** and amine hydrochloride was first dissolved in DCM after which DDQ was added. After oxidation for 30 minutes in the presence of the hydrochloride salt of the amine, DMAP was added. Amounts and equivalents of reagents were used as described in the general procedure A. The desired amide **2a** was isolated using flash column chromatography using EtOAc:pentane (gradient from 20%

EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (55 mg, 87%). The characterization of **2a** matched the data given above.

¹H-NMR, ¹³C-NMR and IR are in accordance with literature values.

N-(2-Chlorophenethyl)propionamide (2b) The general procedure A was followed with a total reaction time of 1 hour (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (67 mg, 89%). *R_f* = 0.24 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 50.5-51.4 °C. IR (neat) *v_{max}* 3264, 3083, 2939, 1631, 1560 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.32 (d, *J* = 8.9 Hz, 1H), 7.23 – 7.07 (m, 3H), 5.85 (br s, 1H), 3.49 (q, *J* = 6.9 Hz, 2H), 2.94 (t, *J* = 7.0 Hz, 2H), 2.15 (q, *J* = 7.6 Hz, 2H), 1.10 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.0, 136.7, 134.1, 131.0, 129.6, 128.0, 127.0, 39.2, 33.4, 29.7, 9.9. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₁H₁₄ClNO: 212.0837 Found: 212.0840.

N-(4-Fluorobenzyl)propionamide (2c) The general procedure A was followed with a total reaction time of 20 hours (including oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (55.5 mg, 86%). *R_f* = 0.5 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 102.0-102.5 °C. IR (neat) *v_{max}* 3292, 2941, 1641, 1510 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.24 – 7.13 (m, 2H), 6.96 (t, *J* = 8.7 Hz, 2H), 6.21 (br s, 1H), 4.33 (d, *J* = 5.8 Hz, 2H), 2.20 (q, *J* = 7.6 Hz, 2H), 1.13 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.5, 162.7 (d, *J_{CF}* = 246), 134.8 (d, *J_{CF}* = 3.2), 129.8 (d, *J_{CF}* = 8.0), 115.7 (d, *J_{CF}* = 21.4), 42.6, 29.4, 9.5. ¹⁹F NMR (376 MHz, CDCl₃) δ -116.0. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₀H₁₃FNO: 182.0976 Found: 182.0978.

N-Benzylpropionamide (2d)⁴³ The general procedure A was followed with a total reaction time of 1 hour (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as white solid with trace amount of red (45 mg, 77%). *R_f* = 0.14 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 51.2-52.3 °C (lit.⁴⁴ 49-50 °C). IR (neat) *v_{max}* 3290, 3031, 2979, 1637, 1540 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.26 (m, 5H), 5.72 (s, 1H), 4.44 (d, *J* = 5.7 Hz, 2H), 2.25 (q, *J* = 7.6 Hz, 2H), 1.18 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 173.7, 138.5, 128.9, 128.0, 127.7, 43.8, 29.9, 10.0. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₀H₁₃NO: 164.1070 Found: 164.1070. ¹H-NMR, ¹³C-NMR and IR are in accordance with literature values.

N-(Benzo[d][1,3]dioxol-5-ylmethyl)propionamide (2e)⁴⁵ The general procedure A was followed with a total reaction time of 2 hours (including 30 min oxidation with DDQ). The product

was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (54 mg, 73%). *R_f* = 0.23 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 99.3-99.7 °C (lit.⁴⁶ 99 °C). IR (neat) *v_{max}* 3286, 1636, 1499, 1441 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 6.82 – 6.65 (m, 3H), 5.93 (s, 2H), 5.77 (br s, 1H), 4.32 (d, *J* = 5.6 Hz, 2H), 2.22 (q, *J* = 7.6 Hz, 2H), 1.16 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.3, 148.3, 147.4, 132.8, 121.3, 108.6, 108.5, 101.2, 43.2, 29.4, 9.5. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₁H₁₄NO₃: 208.0968 Found: 208.0969.

N-(Thiophen-2-ylmethyl)propionamide (2f) The general procedure A was followed with a total reaction time of 20 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a taint solid (48.5 mg, 80%). *R_f* = 0.33 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 45.4-46.5 °C. IR (neat) *v_{max}* 3262, 1551, 1633, 696 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.17 (dd, *J* = 4.6, 1.5 Hz, 1H), 6.99 – 6.75 (m, 2H), 6.31 (br s, 1H), 4.54 (d, *J* = 5.7 Hz, 2H), 2.19 (q, *J* = 7.6 Hz, 2H), 1.12 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.3, 141.7, 127.2, 126.2, 125.4, 38.0, 29.2, 9.4. HRMS (ES) *m/z* [M + H]⁺ calcd for C₈H₁₂NOS: 170.0634 Found: 170.0634.

N-(Furan-2-ylmethyl)propionamide (2g) The general procedure A was followed with a total reaction time of 20 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a light brown oil (37 mg, 68%). *R_f* = 0.33 (silica, EtOAc/pentane 50:50 v/v). IR (neat) *v_{max}* 3285, 1647, 1541 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.37 – 7.33 (m, 1H), 6.32 (dd, *J* = 2.9, 1.8 Hz, 1H), 6.23 (d, *J* = 3.2 Hz, 1H), 5.69 (br s, 1H), 4.44 (d, *J* = 5.4 Hz, 2H), 2.23 (q, *J* = 7.6 Hz, 2H), 1.17 (t, *J* = 7.6 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.1, 152.0, 142.7, 110.8, 107.7, 36.4, 29.4, 9.4. HRMS (ES) *m/z* [M + H]⁺ calcd for C₈H₁₂NO₂: 154.0863 Found: 154.0862.

N-(3,4,5-Trimethoxyphenyl)propionamide (2h)⁴⁷ The general procedure A was followed with a total reaction time of 20 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a yellow solid (64 mg, 75%). *R_f* = 0.23 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 120.0-122.0 °C. IR (neat) *v_{max}* 3262, 2941, 1610, 1505, 1122 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 6.84 (s, 2H), 3.82 (s, 6H), 3.80 (s, 3H), 2.37 (q, *J* = 7.5 Hz, 2H), 1.23 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 172.8, 153.9, 135.0, 134.7, 97.6, 61.0, 56.1, 30.6, 9.3. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₂H₁₈NO₄: 240.1230 Found: 240.1234.

N-Phenylpropionamide (2i)^{48,49} The general procedure A was followed with a total reaction time of 22 hours (including 30

min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (43.5 mg, 82%). $R_f = 0.23$ (silica, EtOAc/pentane 30:70 v/v). mp (uncorr.) 106.0-107.0 °C (lit.⁵⁰ 106-107 °C). IR (neat) ν_{max} 3196, 2975, 1600, 1495 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.62 (br s, 1H), 7.52 (d, $J = 7.9$ Hz, 2H), 7.29 (t, $J = 7.8$ Hz, 2H), 7.08 (t, $J = 7.3$ Hz, 1H), 2.37 (q, $J = 7.6$ Hz, 2H), 1.22 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 173.0, 138.5, 129.3, 124.5, 120.3, 30.5, 9.4. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_9\text{H}_{12}\text{NO}$: 150.0913 Found: 150.0915. ^1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

***N*-(tert-Butyl)propionamide (2j)**^{51,52} The general procedure A was followed with a total reaction time of 19 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a white solid (22 mg, 48%). $R_f = 0.47$ (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 50.5-51.4 °C (lit.⁵¹ 87-91 °C). IR (neat) ν_{max} 3315, 2967, 2930, 1643, 1542 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 5.21 (s, 1H), 2.11 (q, $J = 7.6$ Hz, 2H), 1.34 (s, 9H), 1.11 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 173.8, 50.9, 30.4, 28.6, 9.5. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_7\text{H}_{16}\text{NO}$: 130.1226 Found: 130.1228. ^1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

(S)-Benzyl 1-propionylpyrrolidine-2-carboxylate (2k)⁵³ The general procedure A was followed with a total reaction time of 22 hour (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 20% EtOAc in pentane to 1:1 EtOAc:pentane). The product was isolated as a clear oil (87 mg, 93%). $R_f = 0.23$ (silica, EtOAc/pentane 50:50 v/v). IR (neat) ν_{max} 2966, 2933, 2878, 1743, 1647, 1425 cm^{-1} . ^1H NMR 1:5 inseparable mixture of rotamers. ^1H NMR (400 MHz, CDCl_3) δ 7.38 – 7.27 (m, 5H), 5.27 – 5.00 (m, 2H), 4.53 (dd, $J = 8.6, 3.3$ Hz, 0.8H), 4.38 (dd, $J = 8.5, 2.3$ Hz, 0.2H)*, 3.67 – 3.36 (m, 2H), 2.43 – 1.75 (m, 6H), 1.13 (t, $J = 7.4$ Hz, 2.4H), 1.04 (t, $J = 7.4$ Hz, 0.6H)*. *minor rotamer. ^{13}C -NMR major rotamer: ^{13}C NMR (100 MHz, CDCl_3) δ 173.3, 173.0, 136.2, 128.9, 128.5, 128.4, 66.7, 58.7, 46.7, 28.9, 27.4, 24.5, 8.3. ^{13}C NMR minor rotamer: ^{13}C -NMR (100 MHz, CDCl_3) δ 173.3, 172.8, 135.7, 129.0, 129.0, 128.7, 67.2, 59.4, 46.2, 31.2, 27.2, 22.3, 8.6. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{15}\text{H}_{20}\text{NO}_3$: 262.1438 Found: 262.1439. ^1H -NMR is in accordance with literature values.

4-Nitrobenzyl propionate (14)⁵⁴ The general procedure A was followed with a total reaction time of 20 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 5% EtOAc:pentane to 10% EtOAc:pentane to 20% EtOAc:pentane). The product was isolated as a white solid (60.5 mg, 81%). $R_f = 0.34$ (silica, EtOAc/pentane 10:90 v/v).

mp (uncorr.) 31.9-33.0 °C (lit.⁵⁵ 33 °C). IR (neat) ν_{max} 3087, 2994, 2945, 1734, 1323 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 8.19 (d, $J = 8.5$ Hz, 2H), 7.50 (d, $J = 8.5$ Hz, 2H), 5.20 (s, 2H), 2.41 (q, $J = 7.6$ Hz, 2H), 1.16 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 174.6, 148.2, 144.0, 128.7, 124.1, 64.6, 27.2, 8.7. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{10}\text{H}_{12}\text{NO}_4$: 210.0761 Found: 210.0762.

S-4-Methoxybenzyl propanethioate (16) The general procedure A was followed with a total reaction time of 17 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 1% EtOAc in pentane to 2% EtOAc in pentane). The product was isolated as a clear oil (55 mg, 73%). $R_f = 0.62$ (silica, EtOAc/pentane 10:90 v/v). IR (neat) ν_{max} 2977, 2936, 1686, 1510, 1243 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.21 (d, $J = 8.5$ Hz, 2H), 6.83 (d, $J = 8.5$ Hz, 2H), 4.08 (s, 2H), 3.78 (s, 3H), 2.58 (q, $J = 7.5$ Hz, 2H), 1.18 (t, $J = 7.5$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 200.7, 159.4, 130.4, 130.2, 114.4, 55.3, 37.1, 32.4, 9.3. HRMS (ES) m/z $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{11}\text{H}_{14}\text{O}_2\text{SNa}$: 233.0607 Found: 233.0607.

Benzyl propionate (13)⁵⁶ The general procedure A was followed with a total reaction time of 23 hours (including 30 min oxidation with DDQ). The product was purified by flash column chromatography using EtOAc:pentane (gradient from 1% EtOAc:pentane to 2% EtOAc:pentane to 8% EtOAc:pentane). The product was isolated as a clear oil (28 mg, 48%). Care must be taken as the product is volatile at low pressure (10^{-2} mbar). $R_f = 0.66$ (silica, EtOAc/pentane 10:90 v/v). IR (neat) ν_{max} 2981, 2943, 1735, 1172, 696 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.44 – 7.28 (m, 5H), 5.13 (s, 2H), 2.39 (q, $J = 7.6$ Hz, 2H), 1.17 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 175.0, 136.6, 129.0, 128.6 (2C), 66.2, 27.4, 8.8. HRMS (ES) m/z $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{10}\text{H}_{12}\text{O}_2\text{Na}$: 187.0730 Found: 187.0728. ^1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

4-Methoxybenzyl propionate (15)⁵⁷ The general procedure A was followed with a total reaction time of 17 hours (including 30 min oxidation with DDQ). The residue was purified by flash column chromatography using EtOAc:pentane (5:95). The product was isolated as a clear oil (55.5 mg, 81%). $R_f = 0.59$ (silica, EtOAc/pentane 20:80 v/v). IR (neat) ν_{max} 2955, 2837, 1732, 1612, 1514, 1246 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.30 (d, $J = 8.7$ Hz, 2H), 6.89 (d, $J = 8.7$ Hz, 2H), 5.05 (s, 2H), 3.80 (s, 3H), 2.35 (q, $J = 7.6$ Hz, 2H), 1.15 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 174.5, 159.7, 130.1, 128.4, 114.0, 66.0, 55.3, 27.7, 9.2. HRMS (ES) m/z $[\text{M} + \text{Na}]^+$ calcd for $\text{C}_{11}\text{H}_{14}\text{NaO}_3$: 217.0835 Found: 217.0837. ^1H -NMR and IR are in accordance with literature values.

General procedure B for the synthesis of DHP amides:

To a 0 °C solution of 1,4-dihydropyridine **5** (1 eq.) in a minimum of dry THF was added NaH (2.2 eq.) from a 60%

suspension in mineral oil. After the addition of sodium hydride the color changed from yellow to a very distinct clear light yellow. The mixture was left to stir for 30 minutes after which the acid chloride (1.5 eq.) was added. After reacting overnight the reaction was diluted with water (10 mL) and extracted with EtOAc (3 x 15 mL). The combined organic phases were washed with saturated NaHCO₃, dried (Na₂SO₄), filtered, and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography using silica gel.

Diethyl 1-(3,4,5-trimethoxybenzoyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.8 g of 1,4-dihydropyridine **5**. The product was purified by flash column chromatography by first using EtOAc:pentane (10:90 to 20:80) and then a second column with the eluent EtOAc/NEt₃/DCM (1:2:97). The product was isolated as a white solid (0.909 g, 61%). *R_f* = 0.62 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 141.1-142.8 °C. IR (neat) ν_{max} 3116, 2983, 1710, 1620 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 2H), 6.81 (s, 2H), 4.24 (q, *J* = 7.0 Hz, 4H), 3.93 (s, 3H), 3.89 (s, 6H), 3.31 (s, 2H), 1.30 (t, *J* = 7.0 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 167.3, 166.3, 153.4, 141.6, 132.3, 126.6, 111.6, 106.5, 61.0, 60.9, 56.5, 23.0, 14.3. HRMS (ES) *m/z* [M + H]⁺ calcd for C₂₁H₂₆NO₈: 420.1653 Found: 420.1657.

Diethyl 1-(3-phenylpropanoyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.6 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using EtOAc/DCM (1:99). The product was isolated as yellow oil (0.617 g, 64%). *R_f* = 0.63 (silica, EtOAc/DCM 5:95 v/v). IR (neat) ν_{max} 2939, 1715, 1681, 1621 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.06 (br. s, 1H), 7.59 (br. s, 1H), 7.30 – 7.21 (m, 2H), 7.21 – 7.10 (m, 3H), 4.17 (q, *J* = 7.1 Hz, 4H), 3.15 (s, 2H), 2.97 (t, *J* = 7.5 Hz, 2H), 2.82 (t, *J* = 7.5 Hz, 2H), 1.24 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 168.8, 166.5, 140.1, 130.6, 128.8, 128.6, 126.7, 111.9, 61.0, 35.5, 30.5, 22.8, 14.5. HRMS (ES) *m/z* [M + H]⁺ calcd for C₂₀H₂₄NO₅: 358.1649 Found: 358.1652.

Diethyl 1-(2-bromobenzoyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.27 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using the eluent EtOAc/NEt₃/pentane (1:5:94). The solvent was removed using a flow of N₂ (g). The product was isolated as a yellow solid (363 mg, 74%). The product is light sensitive and therefore it should be stored in the dark in the freezer. *R_f* = 0.21 (silica, EtOAc:pentane 20:80). mp (uncorr.) 123.4-125.2 °C. IR (neat) ν_{max} 2984, 1713, 1624, 1231 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (br. s, 1H), 7.69 (d, *J* = 7.4 Hz, 1H), 7.48 (t, *J* = 7.4 Hz, 1H), 7.42 (d, *J* = 7.4 Hz, 1H), 7.38 (t, *J* = 7.4 Hz, 1H), 7.17 (br. s, 1H), 4.39 – 4.07 (m, 4H), 3.31 (s, 2H), 1.42 – 1.14 (m, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 165.9, 154.2, 134.6, 133.4, 132.3, 129.6, 129.1, 128.2, 119.7, 114.0, 111.9, 61.0, 23.2, 14.4. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₈H₁₉BrNO₅: 408.0441 Found: 408.0438.

Diethyl 1-(4-nitrobenzoyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.3 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using EtOAc/pentane (10:90 to 20:90) and then a second column with the eluent EtOAc/Pentane (10:90). The product was isolated as a yellow solid (238 mg, 58%). *R_f* = 0.64 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 150.3-150.8 °C. IR (neat) ν_{max} 2986, 1671, 1527, 1343, 1233 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.38 (d, *J* = 8.7 Hz, 2H), 7.83 (br. s, 2H), 7.76 (d, *J* = 8.7 Hz, 2H), 4.24 (q, *J* = 7.1 Hz, 4H), 3.32 (s, 2H), 1.30 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 166.0, 165.7, 149.8, 138.0, 131.1, 129.8, 124.3, 113.2, 61.2, 23.2, 14.4. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₈H₁₉N₂O₇: 375.1187 Found: 375.1187.

Diethyl 1-(2-naphthoyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.8 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using EtOAc/pentane (10:90) and then Et₃N/EtOAc/DCM (2:1:97). The product was isolated as a yellow solid (154 mg, 11%). *R_f* = 0.42 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 139.4-141.3 °C. IR (neat) ν_{max} 2982, 1713, 1621, 1191, 1223 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.12 (s, 1H), 7.97 (s, 1H), 7.95 – 7.81 (m, 4H), 7.63 – 7.52 (m, 3H), 4.20 (q, *J* = 7.1 Hz, 4H), 3.32 (s, 2H), 1.24 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 167.8, 166.2, 134.9, 132.4, 132.3, 130.0, 129.1, 129.0, 128.9, 128.5, 127.9, 127.3, 124.6, 111.7, 60.8, 23.0, 14.3. HRMS (ES) *m/z* [M + H]⁺ calcd for C₂₂H₂₂NO₅: 380.1492 Found: 380.1495.

Diethyl 1-octanoyl-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.8 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using EtOAc/pentane (10:90). The product was isolated as a yellow solid (0.951 g, 76%). *R_f* = 0.65 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 118.2-119.7 °C. IR (neat) ν_{max} 3055, 2921, 1782, 1722, 1257 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.97 (br. s, 2H), 4.25 (q, *J* = 7.2 Hz, 4H), 3.24 (s, 2H), 2.57 (t, *J* = 7.2 Hz, 2H), 1.76 – 1.63 (m, 2H), 1.41 – 1.18 (m, 14H), 0.94 – 0.82 (m, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 169.5, 166.3, 130.3, 111.3, 60.7, 33.3, 31.6, 28.95, 28.91, 24.3, 22.5, 22.5, 14.2, 14.0. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₉H₃₀NO₅: 352.2118 Found: 352.2123.

Diethyl 1-((3r,5r,7r)-adamantane-1-carbonyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.6 g of 1,4-dihydropyridine with a reaction time of 4 day. The product was purified by flash column chromatography using EtOAc/pentane (10:90 to 20:80). The product was isolated as yellow solid (0.365 g, 27%). *R_f* = 0.71 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 124.2-126.6 °C. IR (neat) ν_{max} 2907, 2852, 1723, 1450, 1236 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.21 (s, 2H), 4.24 (q, *J* = 7.1 Hz, 4H), 3.21 (s, 2H), 2.23 – 1.99 (m, 9H), 1.85 – 1.61 (m, 6H), 1.31 (t, *J* = 7.1 Hz, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 174.0, 166.8,

132.7, 110.5, 60.9, 43.3, 39.0, 36.5, 28.2, 22.4, 14.5. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{22}H_{30}NO_5$: 388.2118 Found: 388.2122.

Diethyl 1-(cyclopentanecarbonyl)-1,4-dihydropyridine-3,5-dicarboxylate The general procedure B was followed using 0.51 g of 1,4-dihydropyridine. The product was purified by flash column chromatography using EtOAc/pentane (10:90). The product was isolated as a yellow solid (0.582 g, 68%). R_f = 0.58 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 42.0–42.7 °C. IR (neat) ν_{max} 2961, 1721, 1702, 1621, 1233, 1189 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 7.95 (br. s, 2H), 4.30 – 4.17 (m, 4H), 3.22 (s, 2H), 3.10 (p, J = 7.2 Hz, 1H), 2.00 – 1.80 (m, 4H), 1.80 – 1.51 (m, 4H), 1.35 – 1.21 (m, 6H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 172.9, 166.6, 131.1, 111.3, 60.9, 41.6, 30.1, 26.1, 22.8, 14.4. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{17}H_{24}NO_5$: 322.1649 Found: 322.1652.

General procedure C for the synthesis of amides from various DHP-amides

To a solution of DHP amide (175 μ mol, 1.1 eq) in DCM (3 mL) was added DDQ (1 eq.). The reaction was left to stir until full oxidation by TLC analysis was observed. Hereafter the amine (1.1 eq.) and DMAP (3 eq.) was added. The reaction was followed by 1H NMR. The reaction was diluted with EtOAc (10 mL) and washed with saturated $NaHCO_3$, 1 M HCl, dried (Na_2SO_4), filtered, and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography.

3,4,5-Trimethoxy-*N*-phenethylbenzamide (7d)⁵⁸ The general procedure C was followed. Total reaction time was 19 h including 3 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (20:80 to 40:60). The product was isolated as a white solid (46.3 mg, 92%). R_f = 0.32 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 125.3–128.8 °C (lit.⁵⁸ 122–123 °C). IR (neat) ν_{max} 3297, 2938, 1631, 1580, 1537, 1501, 1337, 1233, 1127 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 7.39 – 7.30 (m, 2H), 7.30 – 7.21 (m, 3H), 6.94 (s, 2H), 6.30 (br. s, 1H), 3.88 (s, 3H), 3.86 (s, 6H), 3.70 (q, J = 6.8 Hz, 2H), 2.95 (t, J = 6.8 Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 167.3, 153.2, 140.8, 139.0, 130.2, 128.9, 128.8, 126.7, 104.3, 61.0, 56.3, 41.3, 35.7. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{18}H_{22}NO_4$: 316.1543 Found: 316.1545.

***N*-Phenethylbenzamide (7c)**⁵⁹ The general procedure C was followed. Total reaction time was 23 h including 15 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (10:90). The product was isolated as red solid (33.4 mg, 93%). R_f = 0.21 (silica, EtOAc/ pentane 20:80 v/v). mp 115.3–116.0 °C (lit.⁶⁰ 115–116 °C). IR (neat) ν_{max} 3341, 3059, 2968, 1637, 1534 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 7.61 (d, J = 7.4 Hz, 2H), 7.39 (t, J = 7.4 Hz, 1H), 7.31 (t, J = 7.4 Hz, 2H), 7.24 (t, J = 7.3 Hz, 2H), 7.20 – 7.10 (m, 3H), 6.17 (br. s, 1H), 3.63 (q, J = 6.7 Hz, 2H), 2.85 (t, J = 6.7 Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 167.6, 139.0, 134.8, 131.5, 128.9, 128.8, 128.7, 126.9, 126.7, 41.3,

35.8. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{15}H_{16}NO$: 266.1226 Found: 266.1667. 1H -NMR, ^{13}C -NMR and IR are in accordance with literature values

***N*-Phenethyl-3-phenylpropanamide (7b)**⁶¹ The general procedure C was followed. Total reaction time was 18 h including 3 h of oxidation. The product was purified by flash column chromatography using EtOAc/Pentane (20:80). The product was isolated as a white compound with trace amounts of a reddish taint (35.0mg, 87%). R_f = 0.62 (silica, EtOAc/pentane 50:50 v/v). mp (uncorr.) 94.8–95.4 °C (lit.⁶¹ 94–95 °C). IR (neat) ν_{max} 3297, 2929, 1635, 1541 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 7.26 – 7.16 (m, 4H), 7.16 – 7.07 (m, 4H), 7.01 (d, J = 7.3 Hz, 2H), 5.33 (br. s, 1H), 3.39 (q, J = 6.7 Hz, 2H), 2.86 (t, J = 7.6 Hz, 2H), 2.65 (t, J = 6.7 Hz, 2H), 2.34 (t, J = 7.6 Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 172.1, 141.0, 139.0, 128.9, 128.75, 128.65, 128.5, 126.6, 126.4, 40.7, 38.7, 35.8, 31.8. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{17}H_{20}NO$: 254.1539 Found: 254.1544. 1H -NMR, ^{13}C -NMR and IR are in accordance with literature values

4-Nitro-*N*-phenethylbenzamide (7f)⁶² The general procedure C was followed. Total reaction time was 23 h including 14 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (10:90 to 20:80). The product was isolated as white solid (39.8 mg, 92%). R_f = 0.26 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 149.8–150.6 °C (lit.⁶³ 149.5–150.5 °C). IR (neat) ν_{max} 3328, 1641, 1541, 1516 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 8.25 (d, J = 8.6 Hz, 2H), 7.83 (d, J = 8.6 Hz, 2H), 7.34 (t, J = 7.3 Hz, 2H), 7.29 – 7.21 (m, 3H), 6.23 (br. s, 1H), 3.75 (q, J = 6.7 Hz, 2H), 2.96 (t, J = 6.7 Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 165.6, 149.7, 140.3, 138.6, 129.0, 128.9, 128.1, 127.0, 124.0, 41.5, 35.6. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{15}H_{15}N_2O_3$: 271.1077 Found: 271.1079. 1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

2-Bromo-*N*-phenethylbenzamide (7e)⁶⁴ The general procedure C was followed. Total reaction time was 19 h including 10 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (20:80). The product was isolated as white solid (44.2 mg, 91%). R_f = 0.26 (silica, EtOAc/ pentane 20:80 v/v). mp (uncorr.) 120.9–121.1 °C. IR (neat) ν_{max} 3284, 2928, 1640, 1547 cm^{-1} . 1H NMR (400 MHz, $CDCl_3$) δ 7.56 (d, J = 8.8 Hz, 1H), 7.45 (dd, J = 7.6, 1.7 Hz, 1H), 7.39 – 7.29 (m, 3H), 7.29 – 7.19 (m, 4H), 6.06 (br. s, 1H), 3.74 (q, J = 6.9 Hz, 2H), 2.97 (t, J = 6.9 Hz, 2H). ^{13}C NMR (100 MHz, $CDCl_3$) δ 167.7, 138.8, 138.0, 133.5, 131.3, 129.6, 129.0, 128.8, 127.6, 126.8, 119.4, 41.3, 35.6. HRMS (ES) m/z $[M + H]^+$ calcd for $C_{15}H_{15}BrNO$: 304.0332 Found: 304.0330. 1H -NMR is in accordance with literature values.

***N*-Phenethylcyclopentanecarboxamide (7h)**⁶⁵ The general procedure C was followed. Total reaction time was 19.5 h including 2.5 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (10:90 to 20:80).

The product was isolated as white solid (26.4 mg, 76%). $R_f = 0.27$ (silica, EtOAc/ pentane 20:80 v/v). mp (uncorr.) 107.7–108.7 °C. IR (neat) ν_{max} 3293, 2949, 2866, 1635, 1548, 1236 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.31 (t, $J = 7.3$ Hz, 2H), 7.25 – 7.21 (m, 1H), 7.21 – 7.16 (m, 2H), 5.48 (br. s, 1H), 3.51 (q, $J = 6.9$ Hz, 2H), 2.81 (t, $J = 6.9$ Hz, 2H), 2.44 (q, $J = 7.9$ Hz, 1H), 1.85 – 1.64 (m, 6H), 1.61 – 1.45 (m, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 176.3, 139.2, 128.9, 128.7, 126.6, 46.0, 40.7, 35.9, 30.5, 26.0. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{14}\text{H}_{20}\text{NO}$: 218.1539 Found: 218.1544. ^1H -NMR is in accordance with literature values.

***N*-Phenethyl-2-naphthamide (7a)**⁶⁶ The general procedure C was followed. Total reaction time was 48 h including 25.5 h of oxidation. The product was purified by flash column chromatography using EtOAc/pentane (10:90 to 20:80). The product was isolated as red solid (33.6 mg, 77%). $R_f = 0.23$ (silica, EtOAc/ pentane 20:80 v/v). mp (uncorr.) 138.9–140.1 °C (lit.⁶⁶ 133–134 °C). IR (neat) ν_{max} 3334, 3057, 2936, 1640, 1543, 1305 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 8.14 (s, 1H), 7.86 – 7.75 (m, 3H), 7.69 (d, $J = 8.6$ Hz, 1H), 7.54 – 7.42 (m, 2H), 7.32 – 7.25 (m, 2H), 7.25–7.14 (m, 3H), 6.23 (br. s, 1H), 3.72 (q, $J = 6.7$ Hz, 2H), 2.92 (t, $J = 6.7$ Hz, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ 167.6, 139.1, 134.8, 132.8, 132.0, 129.04, 129.02, 128.9, 128.6, 127.9, 127.7, 127.4, 126.9, 126.8, 123.6, 41.4, 35.9. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{19}\text{H}_{18}\text{NO}$: 276.1383 Found: 276.1384. ^1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

***N*-Phenethyloctanamide (7i)**⁶⁷ The general procedure C was followed. Total reaction time was 1.5 h including 30 min of oxidation. The product was purified by flash column chromatography using EtOAc/Pentane (10:90 to 20:90). The product was isolated as a white solid (36.8 mg, 93%). $R_f = 0.17$ (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 54.9–55.4 °C (lit.⁴³ 54–54.5 °C). IR (neat) ν_{max} 3301, 2924, 1636, 1545 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.30 (t, $J = 7.3$ Hz, 2H), 7.25 – 7.21 (m, 1H), 7.21–7.14 (m, 2H), 5.57 (br. s, 1H), 3.51 (q, $J = 6.8$ Hz, 2H), 2.81 (t, $J = 6.8$ Hz, 2H), 2.11 (t, $J = 7.6$ Hz, 2H), 1.62 – 1.53 (m, 2H), 1.31 – 1.20 (m, 8H), 0.87 (t, $J = 6.7$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 173.3, 139.1, 128.9, 128.7, 126.6, 40.6, 36.9, 35.8, 31.8, 29.3, 29.1, 25.9, 22.7, 14.2. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{16}\text{H}_{26}\text{NO}$: 248.2009 Found: 248.2013. ^1H -NMR and IR are in accordance with literature values

(3r,5r,7r)-*N*-Phenethyladamantane-1-carboxamide (7g)⁶⁸ A modified version of the general procedure C was followed. After oxidation for 0.5 h at rt. the reaction was heated to 50 °C for 23.5 h giving a total reaction time of 24 h. The product was purified by flash column chromatography using EtOAc/Pentane (10:90 to 20:90). The product was isolated as a white solid (28.8 mg, 64%). $R_f = 0.29$ (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 132.2–134.4 °C (lit.⁶⁸ 133 °C). IR (neat) ν_{max} 3317, 2903, 2849, 1627, 1547, 1279 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 7.31 (t, $J = 7.3$ Hz, 2H), 7.27 – 7.14 (m, 3H), 5.59 (br.

s, 1H), 3.49 (q, $J = 6.9$ Hz, 2H), 2.80 (t, $J = 6.9$ Hz, 2H), 2.01 (s, 3H), 1.88 – 1.75 (m, 6H), 1.75 – 1.60 (m, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ 178.0, 139.2, 129.0, 128.7, 126.6, 40.7, 40.5, 39.3, 36.6, 35.8, 28.2. HRMS (ES) m/z $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{16}\text{H}_{26}\text{NO}$: 248.2009 Found: 248.2012. ^1H -NMR is in accordance with literature values

2-(4-Nitrophenyl)acetic acid (8)⁶⁹ Propionyl DHP (110 mg, 0.390 mmol), 2-(4-nitrophenyl)acetic acid (70 mg, 0.390 mmol), and DDQ (80 mg, 0.350 mmol) was dissolved in DCM (6 mL) and stirred for 30 min after which phenylethylamine (50 μL , 0.390 mmol) and DMAP (130 mg, 1.06 mmol) was added. After a total reaction time of 1.5 h the reaction mixture was diluted with EtOAc, washed with 1 M HCl, sat. NaHCO_3 , dried (Na_2SO_4), filtered, and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography using EtOAc/pentane (20:80 to 50:50) yielding the *N*-phenethylpropionamide as a white solid (55.5 mg, 89%).

In order to recover the acid the basic aqueous solution was acidified using 6 M HCl to pH = 1 and extracted with EtOAc (3 x 20 mL). The combined organic phases were washed with brine, dried with Na_2SO_4 , evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography using (formic acid/DCM 1:99) thus recovering the 2-(4-nitrophenyl)acetic acid as a white solid (66.4 mg, 94%). $R_f = 0.33$ (silica, Formic acid/DCM 1:99 v/v). mp (uncorr.) 153.7–154.3 °C (lit.⁷⁰ 153–155 °C). IR (neat) ν_{max} 2919, 2848, 2641, 1700, 1509, 1339, 1250 cm^{-1} . ^1H NMR (400 MHz, CDCl_3) δ 8.21 (d, $J = 8.7$ Hz, 1H), 7.47 (d, $J = 8.7$ Hz, 1H), 3.78 (s, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ 175.5, 147.5, 140.5, 130.6, 124.0, 40.6. HRMS (ES) m/z $[\text{M} - \text{H}]^-$ calcd for $\text{C}_8\text{H}_6\text{NO}_4$: 180.0302 Found: 180.0301. ^1H -NMR, ^{13}C -NMR and IR are in accordance with literature values.

(S)-Diethyl-1-(2-((tert-butoxycarbonyl)amino)-3-phenylpropanoyl)-1,4-dihydropyridine-3,5-dicarboxylate (9)

To a stirred solution of Boc-Phe-Ala (0.383 g, 1.44 mmol, 1.3 eq.) in DMF 5 mL was added HOBt (0.300 g, 2.22 mmol, 2 eq.), EDC-Cl (0.318 g, 1.66 mmol, 1.5 eq.), DIPEA (0.4 mL, 2.22 mmol, 2 eq.), and DHP (0.250 g, 1.11 mmol, 1 eq.). The resulting mixture was stirred for 2 two days under argon after which it was quenched with water and brine, transferred to a separation funnel and extracted with EtOAc (x3). The combined organic phases were washed with 0.5 M HCl and NaHCO_3 (sat. solution), dried (MgSO_4), and evaporated *in vacuo*. The desired compound was isolated using flash column chromatography (silica) with a gradient from (5% acetone:pentane to 10% acetone:pentane and finally 20% acetone:pentane). The compound was loaded onto the column using DCM. During the purification with flash column chromatography the compound precipitates out as a flaky white solid when the collected fractions are allowed to concentrate for a short while in the fume hood. The product was isolated as a flaky white solid (112 mg, 21%). $R_f = 0.33$ (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 144.5–147.5 °C. IR (neat) ν_{max} 3325, 2989, 1697, 1678, 1240 cm^{-1} . ^1H NMR (400

MHz, CDCl₃) δ 7.99 (br s, 1H), 7.47 (br s, 1H), 7.24 – 7.12 (m, 3H), 7.09 (d, *J* = 7.2 Hz, 2H), 5.25 (d, *J* = 8.2 Hz, 1H), 5.11 – 4.75 (m, 1H), 4.17 (q, *J* = 7.0 Hz, 4H), 3.08 (br s, 2H), 3.02 – 2.87 (m, 2H), 1.34 (s, 9H), 1.25 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 169.8, 166.6 (br s), 155.5, 135.5, 130.3 (br s), 129.8, 129.2, 127.7, 113.6, 112.4, 80.6, 60.9, 52.5, 39.4, 28.1, 22.3, 14.0. ¹³C NMR (100 MHz, CDCl₃) δ 169.8 (C=O), 166.6 (br s, (C=O)), 155.5 (C=O), 135.5 (Ar), 130.3 (br s, (-CHCCO₂CH₂CH₃)), 129.8 (Ar), 129.2 (Ar), 127.7 (Ar), 113.6 (CCO₂CH₂CH₃), 112.4 (CCO₂CH₂CH₃), 80.6 (C(CH₃)₃), 60.9 (-CO₂CH₂CH₃), 52.5 (-CHCH₂Ph), 39.4 (-CHCH₂Ph), 28.1 (-CO₂CH₂CH₃), 22.3 (DHP Sp³), 14.0 (C(CH₃)₃). HRMS (ES) *m/z* [M + Na]⁺ calcd for C₂₅H₃₂N₂O₇Na: 495.2102 Found: 495.2108.

Diethyl 1-((S)-2-((S)-2-((tert-butoxycarbonyl)amino)-3-phenylpropanamido)-3-phenylpropanoyl)-1,4-dihydropyridine-3,5-dicarboxylate (11) First, **9** (196 mg, 0.416 mmol, 2.1 eq.) was deprotected using HCl (4 mL) from a 4 M solution in dioxane. After deprotecting overnight the mixture was evaporated to dryness *in vacuo*. The resulting hydrochloride salt **10** was saved in flask A and saved to react with activated **9**.

Second, a solution of unprotected **9** (104 mg, 0.218 mmol, 1.1 eq.) in DCM (3 mL) was added DDQ (45 mg, 0.198 mmol, 1 eq.). After 6 hours activated **9** was added to flask A containing **10** followed by DMAP (73 mg, 0.595 mmol, 3 eq.). A clear change in color occurs after the addition of DMAP. The resulting mixture was left to stir overnight after which it was diluted with EtOAc. The resulting mixture was then transferred to a separation-funnel and washed with 1 M HCl (x2) and NaHCO₃ (x2). The organic phase was dried (MgSO₄), filtered, and evaporated *in vacuo*. The desired compound was isolated using flash column chromatography (silica) with a gradient from 5% acetone:pentane, 10% acetone:pentane, 20% acetone:pentane, and finally 50% acetone:pentane. The compound was loaded onto the column using Celite.

The product was isolated as a beige solid (86 mg, 70%). R_f = 0.3 (30% EtOAc:pentane). mp (uncorr.) 179.0-180.4 °C. IR (neat) *v*_{max} 3348, 3272, 2980, 1720, 1697, 1655, 1231, 1176 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.80 (br s, 1H), 7.27 (br s, 1H), 7.16 – 6.95 (m, 8H), 6.89 (d, *J* = 6.6 Hz, 2H), 6.47 (d, *J* = 7.6 Hz, 1H), 5.16 – 4.99 (m, 1H), 4.77 (br s, 1H), 4.18 (br s, 1H), 4.08 (q, *J* = 7.1 Hz, 4H), 3.06 – 2.90 (m, 2H), 2.90 – 2.74 (m, 4H), 1.24 (s, 9H), 1.16 (t, *J* = 6.8 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 171.5, 168.8, 166.8, 155.9, 136.8, 135.1, 130.4, 129.7, 129.7, 129.3, 129.2, 127.9, 127.5, 113.9, 112.5, 80.6, 61.0, 55.8, 51.0, 39.1, 38.1, 28.1, 22.3, 14.1. ¹³C NMR (100 MHz, CDCl₃) δ 171.5, 168.8, 166.8, 155.9, 136.8 (Ar), 135.1 (Ar), 130.4 (-CHCCO₂CH₂CH₃), 129.7 (Ar), 129.7 (Ar), 129.3 (Ar), 129.2 (Ar), 127.9 (Ar), 127.5 (Ar), 113.9 (-CHCCO₂CH₂CH₃), 112.5 (-CHCCO₂CH₂CH₃), 80.6 (-CH(CH₃)₃), 61.0 (-CO₂CH₂CH₃), 55.8 (-CHCH₂Ph), 51.0 (-CHCH₂Ph), 39.1(-CHCH₂Ph), 38.1(-CHCH₂Ph), 28.1(-CH(CH₃)₃), 22.3 (DHP Sp³), 14.1 (-CO₂CH₂CH₃). HRMS

(ES) *m/z* [M + Na]⁺ calcd for C₃₄H₄₁N₃O₈Na: 642.2786 Found: 642.2790.

(3S,6S)-3,6-Dibenzylpiperazine-2,5-dione (12)⁷¹ Compound **11** (91.6 mg, 0.148 mmol, 1.12 eq.) was dissolved in HCl (5 mL) from a 4 M solution in dioxane. After stirring overnight the reaction was evaporated to dryness *in vacuo* and dissolved in DCM (6 mL) after which DDQ (30 mg, 0.132 mmol, 1 eq.) was added. After 22 hours the mixture was diluted to 20 mL DCM, added DMAP (49 mg, 0.396 mmol, 3 eq.), and left to stir overnight. Hereafter, the reaction mixture was cooled to 0 °C and filtered under suction. The collected yellow precipitate was washed with DCM, pentane, MeOH, and pentane resulting in a white flaky solid (12.6 mg, 32%). mp (uncorr.) 308.5-309.2 °C (lit.⁷² 306-308 °C). IR (neat) *v*_{max} 3395 (br), 3206, 1673, 1660, 699 cm⁻¹. ¹H NMR (400 MHz, DMSO-d₆) δ 7.92 (s, 2H), 7.32 – 7.25 (m, *J* = 7.3 Hz, 4H), 7.24 – 7.17 (m, 2H), 7.06 – 7.01 (m, 4H), 4.05 – 3.87 (m, 2H), 2.57 (dd, *J* = 13.6, 4.9 Hz, 2H), 2.23 (dd, *J* = 13.5, 6.2 Hz, 2H). ¹³C NMR (100 MHz, DMSO-d₆) δ 166.9, 137.2, 130.4, 128.7, 127.0, 55.5 (one carbon missing). HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₈H₁₉N₂O₂: 295.1441 Found: 295.1439. ¹H-NMR, ¹³C-NMR and IR are in accordance with literature values.

2-(2-(2-Methoxyethoxy)ethoxy)-1-(pyridin-1(4H)-yl)ethanone

To a solution of lithium tri-*tert*-butoxyaluminium hydride (3.05 mL, 3.05 mmol, 3 eq. from a 1 M solution in THF) in THF (7.6 mL) was cooled to 0 °C after which CuBr (0.642 g, 4.48 mmol, 4.4 eq.) was added in one portion. After 30 minutes the mixture was cooled to -25 °C where after pyridine (0.123 mL, 1.53 mmol, 1.50 eq.) was added followed by the drop wise addition of 2-(2-(2-methoxyethoxy)ethoxy)acetyl chloride (0.200 g, 1.02 mmol, 1.0 eq.). After reacting at -25 °C for 1.5 hours the reaction was quenched by the addition of a 20% aqueous solution of NH₄Cl (20 mL). The resulting slurry was then filtered through Celite. The filtrate was extracted with EtOAc (3 x 20 mL) where after the combined organic phases were dried (MgSO₄), filtered, and evaporated to dryness *in vacuo*. The remaining residue was unstable towards flash column chromatography (silica gel). The reaction proceeded by NMR with full conversion into product. IR (neat) *v*_{max} 3353, 2922, 2853, 1668 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.08 (d, *J* = 8.5 Hz, 1H), 6.56 (d, *J* = 8.3 Hz, 1H), 5.13 – 5.04 (m, 1H), 4.99 – 4.86 (m, 1H), 4.29 (s, 2H), 3.76 – 3.60 (m, 6H), 3.57 – 3.50 (m, 2H), 3.37 (s, 3H), 2.90 – 2.80 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 164.9, 123.3, 122.3, 108.7, 107.5, 71.6, 70.4, 70.3, 70.2, 69.7, 58.7, 22.5. HRMS (ES) *m/z* [M + Na]⁺ calcd for C₁₂H₂₀NO₄: 242.1387. Found: 242.1387.

Diethyl 4-phenyl-1,4-dihydropyridine-3,5-dicarboxylate⁷³

To a solution of ethyl propiolate (0.924 g, 9.42 mmol, 2 eq.) and NH₄OAc (0.545 g, 7.07 mmol, 1.5 eq.) in glacial acetic acid (5 mL) was added benzaldehyde (0.50 mL, 4.7 mmol, 1 eq.). The resulting mixture was heated to 100 °C. After 24 hours the reaction was cooled to rt, diluted with EtOAc (10 mL)

and neutralized with a solution of saturated NaHCO₃. The mixture was then transferred to a separation funnel and washed with a solution of saturated NaHCO₃ (3 x 10 mL) after which the organic phase was dried (MgSO₄), filtered, and evaporated to dryness *in vacuo*. The remaining residue was purified by flash column chromatography (silica gel) using EtOAc:pentane (20:80) as the eluent. The product was isolated as a yellow solid (0.206 g, 14%). R_f = 0.29 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 114.6-117.0 °C (lit.⁷⁴ 117-118 °C). IR (neat) ν_{max} 3314, 2900, 1689, 1667 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 7.52 – 7.21 (m, 6H), 7.17 (t, *J* = 7.3 Hz, 1H), 6.75 (br s, 1H), 4.92 (s, 1H), 4.24 – 3.88 (m, 4H), 1.25 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 168.0, 147.5, 134.6, 128.6, 128.3, 126.7, 108.1, 59.9, 37.4, 13.8. HRMS (ES) *m/z* [M + H]⁺ calcd for C₁₇H₂₀NO₄: 302.1387. Found: 302.1390. ¹H-NMR, ¹³C-NMR and IR are in accordance with literature values.

Diethyl 4-phenyl-1-propionyl-1,4-dihydropyridine-3,5-dicarboxylate To a 0 °C solution of phenyl 1,4-dihydropyridine (diethyl 4-phenyl-1,4-dihydropyridine-3,5-dicarboxylate) (0.200 g, 0.663 mmol, 1 eq.) in a minimum of dry THF (4 mL) was added NaH (53 mg, 1.3 mmol, 2 eq.) from a 60% suspension in mineral oil. After the addition of sodium hydride the color changed from yellow to a very distinct clear light yellow. The reaction was left to stir for 30 minutes after which propionyl chloride (63 μ L, 0.73 mmol, 1.1 eq.) was added. After reacting overnight the reaction was diluted with water (10 mL) and extracted with EtOAc (3 x 15 mL). The combined organic phases were washed with saturated NaHCO₃, dried (MgSO₄), filtered, and evaporated to dryness *in vacuo*. The residue was purified by flash column chromatography (silica gel) using EtOAc:DCM (gradient starting with 1% EtOAc in DCM ending of at 5% EtOAc in DCM). The product was isolated as a white powder (0.162 g, 68%). R_f = 0.68 (silica, EtOAc/pentane 20:80 v/v). mp (uncorr.) 125.0-125.5 °C. IR (neat) ν_{max} 2986, 1697, 1637 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 8.09 (br. s, 2H), 7.27-7.25 (m, 4H), 7.21-7.15 (m, 1H), 4.89 (s, 1H), 4.20-4.05 (m, 4H), 2.75 (q, *J* = 7.2 Hz, 2H), 1.30 (t, *J* = 7.2 Hz, 3H); 1.21 (t, *J* = 7.1 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃) δ 170.7, 166.0, 143.8, 129.4, 128.7, 128.3, 127.1, 115.6, 60.9, 38.7, 27.3, 14.2, 8.7. HRMS (ES) *m/z* [M + H]⁺ calcd for C₂₀H₂₄NO₅: 358.1649. Found: 358.1653.

Acknowledgements

We thank the Danish National Research Foundation Center for DNA Nanotechnology (DNRF81), The Danish Council for Strategic Research, and The Danish Council for Independent Research for financial support.

Notes and references

^aInterdisciplinary Nanoscience Center and Department of Chemistry, Aarhus University, Gustav Wieds Vej 14, 8000 Aarhus C, Denmark. E-mail kvg@chem.au.dk

† Electronic Supplementary Information (ESI) available: Including additional spectroscopic data and solvent effect data. See DOI: 10.1039/b000000x/

- 1 E. Valeur and M. Bradley, *Chem. Soc. Rev.*, 2009, **38**, 606-631.
- 2 E. Armelin, L. Franco, A. Rodriguez-Galan and J. Puiggali, *Macromol. Chem. Phys.*, 2002, **203**, 48-58.
- 3 S. D. Roughley and A. M. Jordan, *J. Med. Chem.*, 2011, **54**, 3451-3479.
- 4 J. M. Chalker, *Chem. Biol. Drug Des.*, 2013, **81**, 122-135.
- 5 V. R. Pattabiraman and J. W. Bode, *Nature*, 2011, **480**, 471-479.
- 6 A. Williams and I. T. Ibrahim, *Chem. Rev.*, 1981, **81**, 589-636.
- 7 C. A. G. N. Montalbetti and V. Falque, *Tetrahedron*, 2005, **61**, 10827-10852.
- 8 R. M. Lanigan, P. Starkov and T. D. Sheppard, *J. Org. Chem.*, 2013, **78**, 4512-4523.
- 9 T. A. Dineen, M. A. Zajac and A. G. Myers, *J. Am. Chem. Soc.*, 2006, **128**, 16406-16409.
- 10 M. Zhang, S. Imm, S. Bahn, L. Neubert, H. Neumann and M. Beller, *Angew. Chem. Int. Ed.*, 2012, **51**, 3905-3909.
- 11 C. L. Allen, B. N. Atkinson and J. M. J. Williams, *Angew. Chem. Int. Ed.*, 2012, **51**, 1383-1386.
- 12 S. N. Rao, D. C. Mohan and S. Adimurthy, *Org. Lett.*, 2013, **15**, 1496-1499.
- 13 M. Tamura, T. Tonomura, K.-I. Shimizu and A. Satsuma, *Green Chem.*, 2012, **14**, 717-724.
- 14 S. Mundinger, U. Jakob and W. Bannwarth, *Chem. Eur. J.*, 2014, **20**, 1258-1262.
- 15 S. Mundinger, U. Jakob, P. Bichovski and W. Bannwarth, *J. Org. Chem.*, 2012, **77**, 8968-8979.
- 16 M. C. Bröhmer, S. Mundiger, S. Bräse and W. Bannwarth, *Angew. Chem. Int. Ed.*, 2011, **50**, 6175-6177.
- 17 R. A. Kramer, M. C. Bröhmer, N. V. Forkel and W. Bannwarth, *Eur. J. Org. Chem.*, 2009, **25**, 4273-4283.
- 18 M. C. Bröhmer and W. Bannwarth, *Eur. J. Org. Chem.*, 2008, **26**, 4412-4415.
- 19 V. Valerio, D. Petkova, C. Madelaine and N. Maulide, *Chem. Eur. J.* 2013, **19**, 2606-2610.
- 20 Y. Kita, Y. Nishii, Y. Higuchi and K. Mashima, *Angew. Chem.*, 2012, **124**, 5821-5824.
- 21 A. Radzicka and R. Wolfenden, *J. Am. Chem. Soc.*, 1996, **118**, 6105-6109.
- 22 R. M. Smith and D. E. Hansen, *J. Am. Chem. Soc.*, 1998, **120**, 8910-8913.
- 23 S. Arseniyadis, A. Wagner and C. Mioskowski, *Tetrahedron Lett.*, 2004, **45**, 2251-2253.
- 24 W.-R. Li, N.-M. Hsu, H.-H. Chou, S. T. Lin and Y.-S. Lin, *Chem. Commun.*, 2000, **5**, 401-402.
- 25 T. Kamijo, R. Yamamoto, H. Harada and K. Iizuka, *Chem. Pharm. Bull.*, 1982, **30**, 4242-4244.
- 26 W. Steglich and G. Höfle, *Angew. Chem. Int. Ed.*, 1969, **8**, 981.
- 27 U. Ragnarsson and L. Grehn, *Acc. Chem. Res.*, 1998, **31**, 494-501.
- 28 J. M. Berg, J. L. Tymoczko and L. Stryer, *Biochemistry*, W. H. Freeman and Company, USA, 5th edn., 2003.
- 29 P. Belenky, K. L. Bogan and C. Brenner, *Trends Biochem. Sci.*, 2007, **32**, 12-19.

- 30 J.-P. Cheng, Y. Lu, X. Zhu and L. Mu, *J. Org. Chem.*, 1998, **63**, 6108-6114.
- 31 A. R. Katritzky, K. Suzuki and Z. Wang, *Synlett.*, 2005, **11**, 1656-1665.
- 32 J. E. Tulor, M. D. Jones, J. M. J. Williams and S. D. Bull, *J. Org. Chem.*, 2012, **77**, 2808-2818.
- 33 To our knowledge only one example in the literature describes the acylating effect of a 4-substituted 1,4-dihydropyridine amide: J. Bergman, R. Carlsson and S. Misztal, *Acta. Chem. Scand.*, 1976, **30**, 853-862.
- 34 D. L. Comins and A. H. Abdullah, *J. Org. Chem.*, 1984, **49**, 3392-3394.
- 35 J. Arguello, L. J. Núñez-Vergara, J. C. Sturm and J. A. Squella, *Electrochim. Acta.*, 2004, **49**, 4849-4856.
- 36 Y. P. Stradyn, Y. I. Beilis, Y. R. Uldrikis, G. Y. Dubur, S. E. Sausin and B. S. Chekavichus, *Chem. Heterocycl. Compd.*, 1975, **11**, 1299-1303.
- 37 E. Booker and U. Eisner, *J. Chem. Soc. Perkin Trans. 1*, 1975, **10**, 929-931.
- 38 A variety of different reagents for oxidation were investigated. In detail these attempts can be seen in the supporting information
- 39 P. E. Dawson, T. W. Muir, I. Clark-Lewis and S. B. H. Kent, *Science*, 1994, **266**, 776-779.
- 40 P. J. Brignell, U. Eisner and P. G. J. Farrell, *Chem. Soc. (B)*, 1966, 1083-1089.
- 41 K. Singh and K. Singh, *Tetrahedron*, 2009, **65**, 10395-10399.
- 42 A. J. Hill and G. E. J. Hall, *J. Am. Chem. Soc.*, 1952, **74**, 666-668.
- 43 C. L. Allen, A. R. Chhatwal and J. M. J. Williams, *Chem. Commun.*, 2012, **48**, 666-668.
- 44 S. Y. Lee, C. Lee and D. Y. J. Oh, *Org. Chem.*, 1999, **64**, 7017-7022.
- 45 P. Capdevielle, A. Lavigne and M. Maumy, *Tetrahedron*, 1990, **46**, 2635-2844.
- 46 P. Borgna, L. Vicarini and C. Calderara, *Farmaco, Edizione Scientifica*, 1977, **32**, 813-820.
- 47 F. Belin, P. Barthélémy, K. Ruiz, J. M. Lacombe and B. Pucci, *Helv. Chim. Acta.*, 2003, **86**, 247-265.
- 48 P. S. Mahajan, J. P. Mahajan and S. B. Mhaske, *Synth. Commun.*, 2013, **43**, 2508-2516.
- 49 H.-J. Pi, J.-D. Dong, N. An. W. Du and W.-P. Deng, *Tetrahedron*, 2009, **65**, 7790-7793.
- 50 S. R. Thopate, S. R. Kote, S. V. Rohokale and N. M. Thorat, *J. Chem. Res.*, 2011, **35**, 124-125.
- 51 C. A. Goodman, J. B. Eagles, L. Rudahindwa, C. G. Hamaker and S. R. Hitchcock, *Synth. Commun.*, 2013, **43**, 2155-2164.
- 52 R. G. Kalkhambkar, S. N. Waters and K. K. Laali, *Tetrahedron Lett.*, 2011, **52**, 867-871.
- 53 L. Muley, B. Baum, M. Smolinski, M. Freindorf, A. Heine, G. Klebe and D. G. Hangauer, *J. Med. Chem.*, 2010, **53**, 2126-2135.
- 54 M. Bolton and J. Walker, *J. Chem. Soc. C.*, 1967, 2095-2096.
- 55 E. Reid, *J. Am. Chem. Soc.*, 1917, **39**, 124-136.
- 56 I. Dhimitruka, J. SantaLucia, *Org. Lett.*, 2006, **8**, 47-50.
- 57 R. Hirose, B. G. Kopek, Z.-H. Wang, R. Yusa and B. W. Baldwin, *Tetrahedron*, 2003, **9**, 1831-1833.
- 58 A. Lasslo and W. D. Jordan, *J. Org. Chem.*, 1956, **21**, 805-805.
- 59 S. A. Rossi, K. W. Shimkin, Q. Xu, L. M. Mori-Quiroz and D. A. Watson, *Org. Lett.*, 2013, **15**, 2314-2317.
- 60 B. R. Kim, H. Lee, S. Kang, G. H. Sung, J. Kim, J. K. Park, S. Lee and U. Yoon, *Synthesis*, 2012, **44**, 42-50.
- 61 J.-B. Gualtierotti, X. Schumancher, P. Fontaine, G. Masson, Q. Wang, J. Zhy, *Chem. Eur. J.*, 2012, **18**, 14812-14819.
- 62 V. Prasad, R. R. Kale, B. B. Mishra, D. Kumar and V. K. Tiwari, *Org. Lett.*, 2012, **14**, 2936-2939.
- 63 M. Kunishima, C. Kawachi, K. Hioki, K. Terao and S. Tani, *Tetrahedron*, 2001, **57**, 1551-1558.
- 64 M. Hellal and G. D. Cuny, *Tetrahedron Lett.*, 2011, **42**, 5508-5511.
- 65 M. D. Wendt, T. W. Rockway, A. Geyer, W. McClellan, M. Weitzberg, X. Zhao, R. Mantei, V. L. Nienaber, K. Stewart, V. Klinghofer and V. L. Giranda, *J. Med. Chem.*, 2004, **47**, 303-324.
- 66 T. Gonec, P. Bobal, J. Suján, M. Pesko, J. Guo, K. Kralova, L. Pavlacka, L. Vesely, E. Kreckova, J. Kos, A. Coffey, P. Kollar, A. Imramovsky, L. Placek and J. Jampilek, *Molecules*, 2012, **17**, 613-644.
- 67 T. Ogawa, T. Hikasa, T. Ikegami, N. Ono and H. Suzuki, *Chem. Lett.* 1993, **22**, 815-818.
- 68 I. Kim, Y. Park, B. D. Hammock and K. J. Nishi, *Med. Chem.*, 2011, **54**, 1752-1761.
- 69 G.-B. Zhou, P.-F. Zhangb and Y.-J. Pan, *Tetrahedron*, 2005, **61**, 5671-5677.
- 70 D. H. Huh, J. S. Jeong, H. B. Lee, H. Ryu and Y. G. Kim, *Tetrahedron*, 2002, **58**, 9925-9932.
- 71 J.-M. Wang, G.-Z. Ding, L. Fang, J.-G. Dai, S.-S. Yu, Y.-H. Wang, X.-G. Chen, S.-G. Ma, J. Qu, S. Xu and D. Du, *J. Nat. Prod.*, 2010, **73**, 1240-1249.
- 72 K. B. Joshi and S. Verma, *Tetrahedron Lett.*, 2008, **49**, 4231-4234.
- 73 N. A. Al-Awadi, M. R. Ibrahim, M. H. Elnagdi, E. John and Y. A. Ibrahim, *Beilstein J. Org. Chem.*, 2012, **8**, 441-447.
- 74 B. Loev, M. M. Goodman, K. M. Snader, R. Tedeschi and E. Macko, *J. Med. Chem.*, 1974, **17**, 956-965.