

Organic & Biomolecular Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

COMMUNICATION

Cite this:
10.1039/x0xx00000x

DOI: **An Ebselen like Catalyst with Enhanced GPx Activity via a Selenol Intermediate**

Received 00th January 2012,
Accepted 00th January 2012

DOI: 10.1039/x0xx00000x

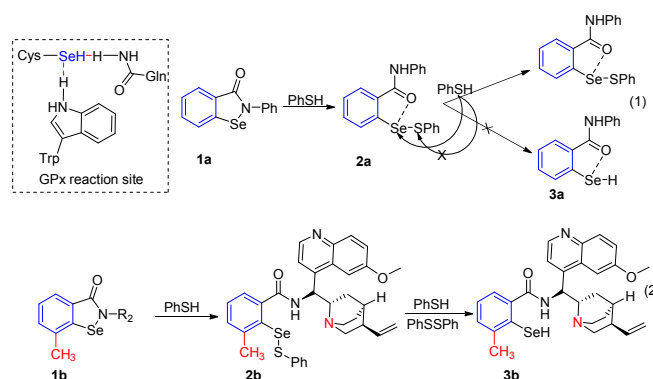
www.rsc.org/

Shah Jaimin Balkrishna,^a Shailesh Kumar,^a Gajendra Kumar Azad,^b Bhagat Singh Bhakuni,^a Piyush Panini,^a Navjeet Ahalawat,^a Raghuvir Singh Tomar,^b Michael R Detty,^c and Sangit Kumar^{a*}

The reaction of KSeO'Bu with 2-iodo-arylbenzamides gave benzamide ring-substituted, quinine-derived isoselenazones **1b-1d**. The reaction of PhSH with *ortho*-methyl-substituted isoselenazolone **1b** gave selenol **3b**, which is oxidized by H₂O₂ to regenerate **1b**. Isoselenazolone **1b** shows a high rate ($0.33 \times 10^3 \mu\text{M} \cdot \text{min}^{-1}$) of oxidation of PhSH with H₂O₂, which is $\sim 10^3$ -fold more active than ebselen (**1a**) and ≥ 30 -fold more active than the other isoselenazones of this study. Compound **1b** shows less inhibition of the growth of yeast cells than **1a**.

The selenoenzyme glutathione peroxidase (GPx) is present in the human body and functions as a catalytic antioxidant for the reduction of various hydroperoxides using glutathione as the stoichiometric co-reductant. The reactive site of GPx contains selenocysteine (Cys-SeH), which is responsible for the high catalytic activity for the reduction of hydroperoxides (Scheme 1).¹ The GPx mimic ebselen (**1a**) or 2-phenyl-1,2-benzisoselenazol-3(2H)-one (PZ 51) is a biologically non-toxic (LD₅₀ = 6.81 g/kg), well-studied organoselenium compound with anti-inflammatory and antioxidant therapeutic properties as well as the potential to treat indications such as bipolar disorder.^{2,3} The diverse remedial properties of **1a** are apparently due to its reduction of hydroperoxides together with its low toxicity. Unfortunately, **1a** is a relatively inefficient catalyst for the reduction of hydroperoxides, which encourages attempts to synthesize effective GPx-mimics with improved catalytic activity for the reduction of peroxides while maintaining low toxicity. The poor catalytic reactivity of **1a** is presumably due to the lack of reactivity of selenenylsulfide **2a** to produce selenol **3a** when it reacts with an additional molecule of thiol (eq. 1 Scheme 1).⁴ The generation of a selenol by the reaction of an organothiol with an organoselenium compound is a challenging task in general and from isoselenazones in particular.⁵⁻⁷ Several *N*-substituted isoselenazones have been described; however, none appear to form a stable selenol upon their reaction with organothiols.⁸⁻¹³ Therefore, these

isoselenazones are limited in their catalytic activity for the reduction of H₂O₂ via an ebselen-like pathway.

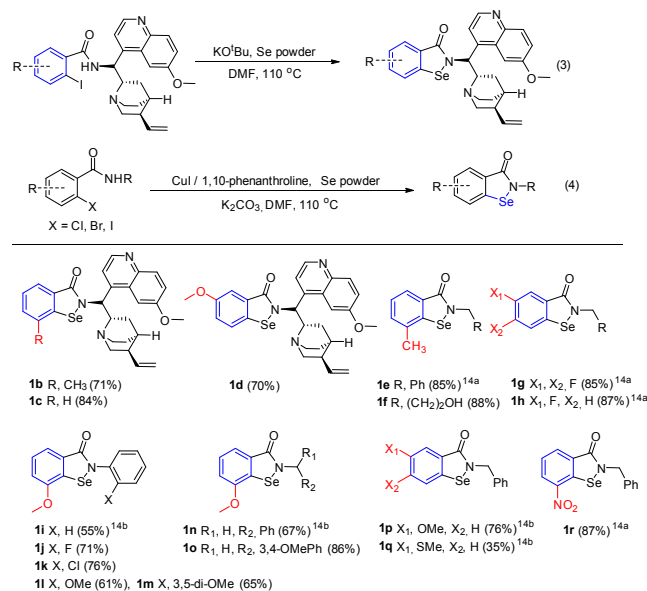


Scheme 1 GPx Reaction Site and Generation of Selenol

Isoselenazones with additional benzamide ring functionality have not been studied as GPx mimics presumably due to difficulties in their synthesis. Here, in continuation of our work on organochalcogen chemistry,¹⁴ we describe the synthesis of isoselenazones containing a quinine moiety and the facile generation of selenol **3b** by the reaction of isoselenazolone **1b** with PhSH (eq. 2). The catalytic activity of various isoselenazones as GPx mimics was also studied with **1b** showing much greater catalytic activity than ebselen (**1a**).

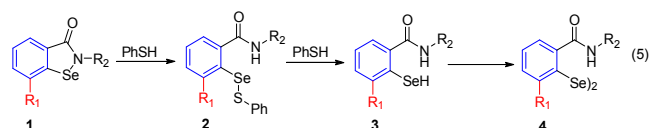
N-Quininamine-substituted [*N*-(1*S*)-(6-methoxyquinolin-4-yl)((2*S*,4*S*,5*R*)-5-vinyl-quinuclidin-2-yl)methyl] isoselenazones **1b-1d** were prepared using KSeO'Bu as the source of selenium¹⁵ and in higher yields (71% vs 55% for **1b**, 84% vs 72% for **1c** and 70% vs 60% for **1d**) in comparison to copper-catalyzed Se-N bond formation reactions (eq 3, Scheme 2). The use of KSeO'Bu as a selenium source is more efficient for the synthesis of isoselenazones bearing polar functionality. In the copper catalysed methodology, separation of the copper-1,10-phenanthroline catalyst from the polar quininamine-derived

isosenenazolones **1b-1d** was difficult. Using KSeO^tBu as the source of Se, quinamine-derived isosenenazolones **1b-1d** were obtained in higher yield with fewer side products. However, the use of KSeO^tBu was only applicable to 2-iodo- and selected 2-bromobenzamides. KSeO^tBu was unreactive with 2-chlorobenzamides as substrates. The isosenenazolones ebselen (**1a**) and **1e-1r** were prepared by the Cu-catalysed Se-N bond forming reaction (eq 4, Scheme 2).^{14a,b} The isosenenazolone structure was confirmed by ^{77}Se NMR (Table 1) and X-ray structural studies of **1b**, **1c** and **1e**.



Scheme 2 Synthesis of Isosenenazolones

Table 1 Reaction of Isosenenazolones with PhSH^a



Entry	1 (δ)	[PhSH]	2 (δ)	3 (δ)	4 (δ)
1	1a (912)	1	2a (591)	3a (-)	4b (-)
1	1b (851)	1	2b (-)	3b (-0.2)	4b (-)
2	1b (851)	2	2b (-)	3b (4.8)	4b (-)
3, 4	1c (858)	2	2c (589)	3c (-)	4c (-)
5	1e (859)	1	2e (496)	3e (-)	4e (416)
6	1e (859)	2	2e (496)	3e (-)	4e (416)
7	1n (889)	1	2n (454)	3n (-)	4n (405)
8	1n (889)	2	2n (-)	3n (-)	4n (401)

^a Reaction was carried out in CD₃OD using 1 equiv of PhSH with 1. (-) Not observed.

The reaction between PhSH and isosenenazolones was monitored by ^{77}Se NMR spectroscopy and mass spectrometry.¹⁶ An equimolar mixture of isosenenazolone **1b** and PhSH in CD₃OD gave a small peak at -0.2 ppm due to selenol **3b** (Table

1). Surprisingly, formation of the expected selenenylsulfide **2b** was not observed by ^{77}Se NMR. Addition of a second equiv of PhSH to the stoichiometric reaction mixture of **1b** and PhSH gave a sharp peak at 4.8 ppm. A freshly prepared solution of **1b** and PhSH was monitored by mass spectrometry and showed formation of selenenylsulfide **2b** (m/z 629.1615+H⁺) which suggests that the reaction proceeds *via* the formation of selenenylsulfide **2b**. However, this is a transient intermediate in the formation of selenol **3b**. Selenol **3b** is stable for at least two weeks in solution as ^{77}Se NMR shows a constant signal at -0.9 ppm. Signals due to the formation of diselenide **4b** or isosenenazolone **1b** were not observed.

The ^{77}Se NMR chemical shift of **3b** is similar to reported ^{77}Se NMR chemical shifts of *N,N*-dimethylbenzylamine selenol (9.9 ppm),^{6b} the aryl-selenol BmtSeH (6 ppm),^{7a} and a camphor-derived selenol (-49 ppm).^{9a} However, the ^{77}Se NMR chemical shift of **3b** is upfield relative to PhSeH (145 ppm).

Several other isosenenazolones (**1a-1c**, **1e**, and **1n**) were reacted with PhSH and results are summarized in Table 1. Isosenenazolone **1c** having a quinamine moiety and lacking the *ortho*-CH₃ substituent forms only selenenylsulfide **2c** in the presence of 1-3 equivalents of PhSH (entries 3-4, Table 1). However, *ortho* CH₃-substituted isosenenazolones **1e** gave selenenylsulfides **2e** and diselenide **4e** when reacted with one and two equivalents of PhSH. Similarly, **1n** gave **2n** and **4n** with one equivalent of PhSH (entry 7, Table 1). In contrast, addition of a second equivalent of PhSH led to complete conversion of **2n** into diselenide **4n** (entry 8, Table 1).

The formation of diselenides **4e** and **4n** could occur *via* either of two processes: oxidation of the respective selenols **3e** and **3n** or disproportionation of selenenylsulfides **2e** and **2n**.¹⁷ Mixtures of isosenenazolones **1b**, **1e**, or **1n** and PhSH (1:2 molar ratios) were reacted with CH₃I. Indeed, complete conversion of *in-situ* generated selenol **3b** into the corresponding methylselenide was observed. In contrast, isosenenazolones **1e** and **1n** failed to produce the corresponding methylselenides under similar conditions. This implies that the formation of diselenides **4e** and **4n** occurred by the disproportionation of selenenylsulfides **2e** and **2n** rather than *via* the formation of the corresponding selenols.

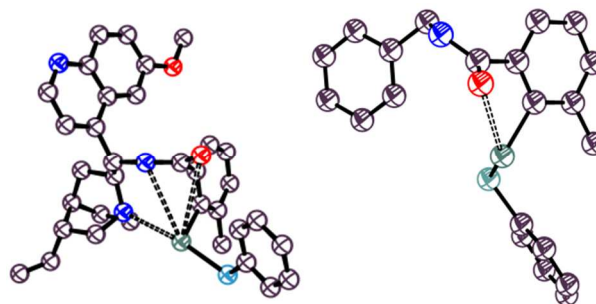
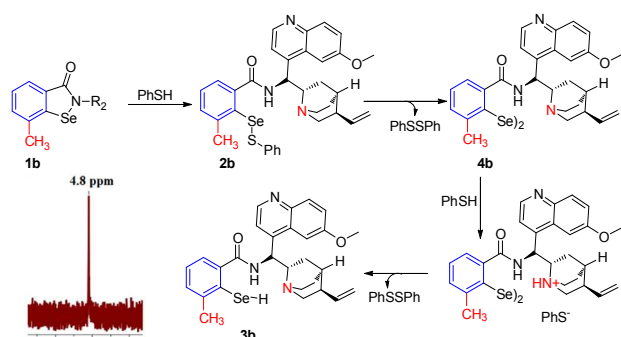


Figure 1 Optimized Structures of **2b** and **2e**. Se...O 3.811 Å (C=O), Se...N 2.825 Å (tert-N quinamine), Se...N (NH) in **2b**. Se...O 2.42 Å (C=O) in **2e**. Se...O 4.083 Å (C=O), Se...N 2.79 Å (tert-N), Se...N (NH) in **2c**.

Scheme 3 Proposed Generation of Selenol **3b** from **1b**

DFT calculations on the related selenenylsulfides **2b**, **2c**, **2e**, **2f** and **2n** were performed to study the nature of Se...O/N interactions (Figure 1) as these interactions appear to be important, not only in the stabilization of the selenol functionality (Scheme 1), but also in its formation from the corresponding selenenylsulfide. Short Se...O/N interactions increase electron density on selenium, which should then favor the nucleophilic attack of PhS⁻ at sulfur rather than at selenium in the selenenylsulfide intermediate leading to the selenol and disulfide.^{6a,c,9a,18} However, other factors must also be considered. In selenenylsulfide **2c**, intramolecular Se...O and Se...N distances are favorable for the generation of selenol **3c**,¹⁹ but neither **3c** or diselenide **4c** is observed.

We next examined the antioxidant properties of isoselenazolones **1a-1r** for the reduction of H₂O₂ in the presence of PhSH as co-reductant in CH₃OH according to eq 6. From the data in Table 2 and Figure 2, isoselenazalone **1b**, which forms selenol **3b**, shows a much higher initial rate of oxidation of PhSH ($v_o = 331 \mu\text{M}\cdot\text{min}^{-1}$) when compared to the remainder of the isoselenazolones of this study.²⁰ Isoselenazalone **1b** is 10³-fold more active than ebselen **1a** and 30-fold more active than quininamine-based isoselenazolones **1c** and **1e**. Also, **1b** is 475 times more active than the diphenyl diselenide. We have also evaluated the influence of an external base triethyl amine (0.05 mM) on the GPx activity of **1b** and **1c**. However, the external base gave no significant change in the catalytic activity of **1b** and **1c** (entries 5 and 7, Table 2). In other systems, the introduction of a methoxy group in an aromatic ring enhances the antioxidant property of catalysts.²¹ The methoxy-substituted isoselenazolones **1d** and **1i-1p**. However, these catalysts showed $\leq 5\%$ of the antioxidant activity of **1b**.

Isoselenazolones **1a**, **1c-1d**, **1g**, **1h**, **1p** and **1q**, which form only selenenylsulfides with PhSH via ⁷⁷Se NMR studies, are poor catalysts for this reaction based on v_o , whereas isoselenazolones **1e**, **1f**, **1i-1o** which form diselenides with PhSH are intermediate in catalytic activity relative to **1b**.

To further understand the mechanism of GPx activity of the isoselenazalone **1b**, the effects of catalyst and H₂O₂ concentration on v_o were examined. Initial values of v_o increased linearly with respect to catalyst concentration indicating a first-order dependence of GPx activity on catalyst

concentration. As the concentration of H₂O₂ was increased, values of v_o increased, but became constant with no further increase in v_o with increasing H₂O₂ concentration. The effect of changing peroxide concentration is consistent with the formation of an oxidized intermediate whose subsequent reduction limits the rate of turnover in the system.

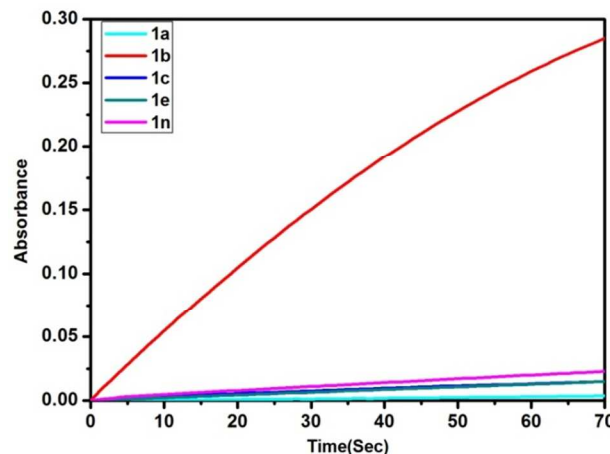
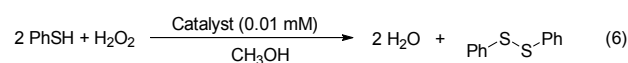
Figure 2 Initial Rates of PhSSPh Formation in the Presence of Catalysts **1a**, **1b**, **1c**, **1e** and **1n**

Table 2 GPx Like Activity of Isoselenazolones

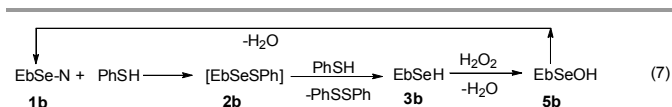


Entry	Cat.	$v_o \mu\text{M}\cdot\text{min}^{-1}$	Entry	Cat.	$v_o \mu\text{M}\cdot\text{min}^{-1}$
1	(PhSe) ₂	0.7±0.1	12	1h	13±1
2	1a	0.4±0.02	13	1i	17±2
3	1b	331±2 ^b	14	1j	13±1
4	Et ₃ N ^c	1.6±0.1	15	1k	17±1
5	1b +Et ₃ N ^c	332±2 ^b	16	1l	13±1
6	1c	11±1	17	1m	14±2
7	1c +Et ₃ N ^c	12.2±0.7	18	1n	11.7±0.3
8	1d	4.0±0.2	19	1o	15±1
9	1e	11±2	20	1p	1.2±0.3
10	1f	21±1	21	1q	0.20±0.02
11	1g	0.40±0.03	22	1r	10±3

The initial rates (v_o) for the oxidation of PhSH (1 mM) with H₂O₂ (3.75mM) in the presence of catalyst (0.01 mM) were determined in CH₃OH by monitoring the UV absorption at 305 nm due to the formation of phenyl disulfide.^b v_o obtained by Lineweaver Burk plot. ^c concentration of Et₃N was 0.05mM

To establish the catalytic cycle, selenol **3b** was treated with one equivalent of H₂O₂ in CD₃OD and the resulting mixture was examined by ⁷⁷Se NMR and mass spectrometry. A signal was observed at 1093 ppm in the ⁷⁷Se NMR and is attributed to selenenic acid (R-SeOH) **5b** (based on the observed m/z 536.1489) and a second signal at 851 ppm corresponding to

isosenenazalone **1b**. The addition of a second equivalent of PhSH to selenenic acid **5b** gave formation of **1b** and **2b**. The catalytic cycle for **3b** reacting with PhSH and H₂O₂ is summarized in Scheme 4.



Scheme 4 Catalytic cycle for **1b**

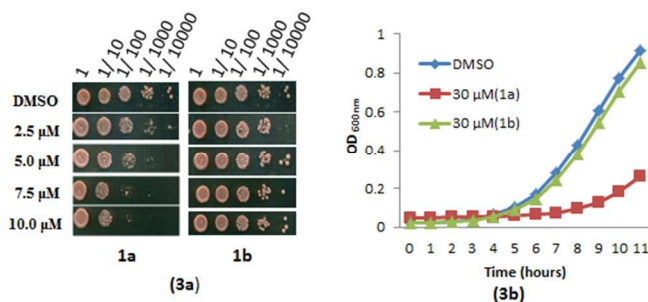


Figure 3 Yeast cells growth in the presence of catalysts

Ebselen (**1a**) is the most studied GPx-mimic in organoselenium chemistry and has demonstrated low toxicity in biological studies. We compared the effect of **1a** and **1b** on the growth of yeast cells to compare relative toxicities.²² The dose-dependent effect of various concentrations of isosenenazolones on the growth of yeast cells was over a 72 h time period (for more details, please see SI, S174-S175).²³ Figure 3a compares ebselen (**1a**) with isosenenazalone **1b** and shows a higher growth of cells in the presence of **1b** compared to **1a**.

These results were validated by growth curve analysis in liquid culture as shown in Figure 3b, (please see SI, S175-S179 for experimental data).²⁴ Yeast cells were treated with an increasing dose of either **1a** or **1b** (10, 20 and 30 μM) and growth was monitored at OD_{600nm} for 11 h at regular interval. It is apparent that the OD_{600nm} value for **1b** (0.85±0.01) which is significantly higher than **1a** (0.26±0.01). From growth curve analysis, doubling time for the growth of yeast cells was also calculated in presence of **1a** and **1b**. It was significantly higher for **1a** (278±12 min) compared to **1b** (124±3 min) further suggesting that **1b** inhibits cell growth to a lesser extent than ebselen (**1a**).

Conclusions

In summary, the reaction of isosenenazolones with additional substituents on the benzamide ring with PhSH has been investigated. Isosenenazalone **1b** bearing an *ortho*-methyl group on the benzamide ring and an *N*-quininamine group gave selenol **3b** in the presence of PhSH. The remainder of the isosenenazolones of this study formed either selenenylsulfides or diselenides upon reaction with PhSH. We have also shown that short intramolecular Se...N/O interactions are not sufficient for the generation of selenol from selenenylsulfide. The high GPx-like activity of isosenenazalone **1b**, which forms

selenol intermediate **3b**, suggests that the presence of the bulky *N*-quininamine substituent and the *ortho*-CH₃ benzamide substituent stabilizes **3b**, which regenerates **1b** following reaction with H₂O₂. In a comparison of ebselen (**1a**) and **1b**, growth of yeast cells in the presence of **1b** was comparable to the DMSO control and was significantly higher than in the presence of **1a**. Currently, we are investigating catalytic role of selenol for various biological activities involving thioredoxin reductase and deiodinase enzymes in which selenol functionality is critical for activity.

ACKNOWLEDGMENT

We are thankful to DAE-BRNS, Mumbai, DST-New Delhi, DRDO-New Delhi, IISER Bhopal for funding. SJB, SK, BSB thank to IISER Bhopal for fellowship. MRD thanks US Office of Naval Research (award N0014-09-1-0217) for partial support of this research.

Notes and references

^a Department of Chemistry, ^bDepartment of Biological Sciences, Indian Institute of Science Education and Research Bhopal (IISER), Indore bypass Road, Bhopal, MP 462 066, India.

^c Department of Chemistry, University at Buffalo, The State University of New York, Buffalo, New York 14260, United States.

Electronic Supplementary Information (ESI) available: [Characterization data, NMR and mass spectra on **1a-1r** and CIF files for **1b**, **1c**, and **1e** (CCDC no. 930741, 930742 and 953729), DFT calculations and geometry optimization on **2b**, **2c** and **2e**, GPx-activities, kinetic studies, toxicity tests.]. See DOI: 10.1039/c000000x/

- O. Epp, R. Ladenstein, A. Wendel, *Eur. J. Biochem.*, 1983, **133**, 51.
- (a) A. Müller, E. Cadenas, P. Graf, H. Sies, *Biochem. Pharmacol.*, 1984, **33**, 3235; (b) "Oxidative Stress: Introductory Remarks" Sies, H. in *Oxidative Stress* (Ed.: H. Sies) Academic Press, London, 1985, pp. 1; (c) H. Sies, *Angew. Chem. Int. Ed. Engl.*, 1986, **25**, 1058; (d) H. Sies, *Free Radical Biol. Med.* 1993, **14**, 313; (e) H. Sies, H. Masumoto, *Adv. Pharmacol.*, 1997, **38**, 229; (f) G. I. Giles, F. H. Fry, K. M. Tasker, A. L. Holme, C. Peers, K. N. Green, L.-O. Klotz, H. Sies, C. Jacob, *Org. Biomol. Chem.*, 2003, **1**, 4317; (g) C. Jacob, G. I. Giles, N. M. Giles, H. Sies, *Angew. Chem. Int. Ed.*, 2003, **42**, 4742; (h) T. Kalai, G. Magesh, G. Roy, H. Sies, Z. Berente, K. Hideg, *Org. Biomol. Chem.*, 2005, **3**, 3564; (i) H. Steinbrenner, E. Bilgic, L. Alili, H. Sies, P. Brenneisen, *Free Radical Res.*, 2006, **40**, 936. (j) B. J. Bhuyan, D. S. Lamani, G. Magesh and T. Wirth, in *Current Research on Mimics and Models of Selenium-Containing Antioxidants*, ed. F. A. Devillanova, W.-W. duMont, Royal Society of Chemistry, 2013, Vol. 2, pp. 25-46 (k) B. J. Bhuyan and G. Magesh, in *Biological and Biochemical Aspects of Selenium Compounds*, ed. T. Wirth, Wiley-VCH, Weinheim, Germany, 2012, 361-396 (l) G. Magesh, *Current Chemical Biology* 2013, **7**, 47-56 (m) L. Orian, S. Toppo, *Free Radical Biology and Medicine*, 2014, **66**, 65-74
- (a) N. Singh, A. C. Halliday, J. M. Thomas, O. V. Kuznetsova, R. Baldwin, E. C. Y. Woon, P. K. Aley, I. Antoniadou, T. Sharp, S. R. Vasudevan, G. C. Churchill, *Nat. Commun.*, 2013, **4**, 1332; (b) L. Favrot, A. E. Grzegorzewicz, D. H. Lajiness, R. K. Marvin, J. Boucau, D. Isailovic, M. Jackson, D. R. Ronning, *Nat. Commun.*, 2013, **4**, 2748.
- B. K. Sarma, G. Magesh, *J. Am. Chem. Soc.*, 2005, **127**, 11477.
- (a) G. Magesh, W. -W. du Mont, H. Sies, *Chem. Rev.*, 2001, **101**, 2125; (b) G. Magesh, H. B. Singh, *Chem. Soc. Rev.*, 2000, **29**, 347; (c) C. W. Nogueira, G. Zeni, J. B. T. Rocha, *Chem. Rev.*, 2004, **104**, 6255.
- (a) S. R. Wilson, P. A. Zucker, R.-R. C. Huang, A. Spector, *J. Am. Chem. Soc.*, 1989, **111**, 5936; (b) M. Iwaoka, S. Tomoda, *J. Am. Chem. Soc.*, 1994, **116**, 2557; (c) G. Magesh, A. Panda, H. B. Singh, N. S. Puneekar, R. J. Butcher, *Chem. Commun.*, 1998, **20**, 2227; (d) G. Magesh, A. Panda, H. B. Singh, N. S. Puneekar, R. J. Butcher, *J. Am. Chem. Soc.*, 2001, **123**, 839.

7. (a) K. Goto, M. Nagahama, T. Mizushima, K. Shimada, T. Kawashima, R. Okazaki, *Org. Lett.*, 2001, **3**, 3569; (b) K. Goto, D. Sonoda, K. Shimada, S. Sase, T. Kawashima, *Angew. Chem., Int. Ed.*, 2010, **49**, 545; (c) K. Shimada, K. Goto, T. Kawashima, N. Takagi, Y. -K. Choe, S. Nagase, *J. Am. Chem. Soc.*, 2004, **126**, 13238.
8. H. J. Reich, C. P. Jasperse, *J. Am. Chem. Soc.*, 1987, **109**, 5549.
9. (a) T. G. Back, B. P. Dyck, *J. Am. Chem. Soc.*, 1997, **119**, 2079. For different GPx-mimics, please see (b) T. G. Back, Z. Moussa, *J. Am. Chem. Soc.*, 2002, **124**, 12104; (c) T. G. Back, Z. Moussa, *J. Am. Chem. Soc.*, 2003, **125**, 13455; (d) T. G. Back, Z. Moussa, M. Parvez, *Angew. Chem. Int. Ed.*, 2004, **43**, 1268; (e) T. G. Back, D. Kuzma, M. Parvez, *J. Org. Chem.*, 2005, **70**, 9230; (f) D. J. Press, E. A. Mercier, D. Kuzma, T. G. Back, *J. Org. Chem.*, 2008, **73**, 4252; (g) T. G. Back, *Can. J. Chem.*, 2009, **87**, 1657; (h) D. J. Press, T. G. Back, *Org. Lett.*, 2011, **13**, 4104.
10. R. S. Glass, F. Farooqui, M. Sabahi, K. W. Ehler, *J. Org. Chem.*, 1989, **54**, 1092.
11. (a) I. A. Cotgreave, R. Morgenstern, L. Engman, J. Ahokas, *Chem- Biol. Interactions.*, 1992, **84**, 69; (b) I. A. Cotgreave, P. Moldus, R. Brattsand, A. Hallberg, C. M. Anderson, L. Engman, *Biochem. Pharmacol.*, 1992, **43**, 793.
12. K. Satheeshkumar, G. Muges, *Chem. Eur. J.*, 2011, **17**, 4849.
13. (a) K. P. Bhabak, G. Muges, *Chem. Eur. J.*, 2007, **13**, 4594; (b) K. P. Bhabak, G. Muges, *Acc. Chem. Res.*, 2010, **43**, 1408; (c) S. S. Zade, S. Panda, S. K. Tripathi, H. B. Singh, G. Wolmershäuser, *Eur. J. Org. Chem.*, 2004, **18**, 3857; (d) V. P. Singh, H. B. Singh, R. J. Butcher, *Chem. Asian J.*, 2011, **6**, 1431; (e) V. P. Singh, H. B. Singh, R. J. Butcher, *Eur. J. Org. Chem.*, 2011, 5485; (f) K. Selvakumar, P. Shah, H. B. Singh, R. J. Butcher, *Chem. Eur. J.*, 2011, **17**, 12741.
14. (a) S. J. Balkrishna, B. S. Bhakuni, D. Chopra, S. Kumar, *Org. Lett.*, 2010, **12**, 5394; (b) S. J. Balkrishna, B. S. Bhakuni, S. Kumar, *Tetrahedron.*, 2011, **67**, 9565; (c) S. J. Balkrishna, C. D. Prasad, P. Panini, M. R. Detty, D. Chopra, S. Kumar, *J. Org. Chem.*, 2012, **77**, 9541.
15. Potassium tert-butoxide base reacted smoothly with selenium powder and produce brown-greenish mixture of potassium tert-butoxyselenolate (KSeOtBu) which was characterized by ¹H, ¹³C and ⁷⁷Se NMR (please see Supporting Information pages S180-S183).
16. The ⁷⁷Se NMR is very important technique for the characterization of organoselenol in solution. Mass spectrometry provides distinct isotopic pattern due to presence of six selenium isotopes.
17. ⁷⁷Se NMR experiments were conducted for 10 h which is substantial time for the oxidation of selenol into diselenide. To preclude this, immediate capture of selenol by electrophile was carried out.
18. Optimized geometries and DFT calculations of **1b**, **1c**, **1e**, **1f**, **1n**, and **2b**, **2c**, **2e**, **2f**, **2n** correlate well with experimentally obtained ⁷⁷Se NMR chemical shifts, Se...X distances (please see SI S148-S160).
19. Intramolecular Se...N/O (if heteroatom is in conjugation with selenium) interaction decreases electron density around selenium as the case with Se...O (C=O), heteroatom is not in conjugation with Se, interaction (Se...N (quinine N)) enhances overall electron density around selenium. See references: (a) M. Iwaoka, S. Tomoda, *J. Am. Chem. Soc.*, 1996, **118**, 8077; (b) M. Iwaoka, H. Komatsu, T. Katsuda, S. Tomoda, *J. Am. Chem. Soc.*, 2004, **126**, 5309; (c) A. J. Mukherjee, S. S. Zade, H. B. Singh, R. B. Sunoj, *Chem. Rev.*, 2010, **110**, 4357.
20. GPx-activities of 13 more isoselenazolones were tested and presented in SI page S173.
21. (a) T. Wirth, *Molecules* 1998, **3**, 164-166; (b) M. L. Jauslin, T. Wirth, T. Meier, F. Schoumacher, *Hum. Mol. Genet.* 2002, **11**, 3055-3063.
22. Yeast cells serves as a eukaryotic cell model to assess biotransformation related study. See references. (a) A. Buschini, P. Poli, C. Rossi, *Mutagenesis.*, 2003, **18**, 25-36; (b) A. Chatterjee, C. T. Jurgenson, F. C. Schroeder, S. E. Ealick, T. P. Begley, *J. Am. Chem. Soc.*, 2006, **128**, 7158-7159; (c) J. S. van Leeuwen, N. P. E. Vermeulen, J. Chris Vos, *Curr. Drug Metab.*, 2012, **13**, 1464-1475; (d) J. E. Dicarlo, A. J. Conley, M. Penttilä, J. Jäntti, H. H. Wang, G. M. Church, *ACS Synth. Biol.*, 2013, **2**, 741-749.
23. G. K. Azad, S. J. Balkrishna, S. Narayanan, S. Kumar, R. S. Tomar, *Biochem. Pharmacol.*, 2012, **83**, 296.
24. A. A. Hostetter, M. F. Osborn, V. J. DeRose, *ACS Chem. Biol.*, 2012, **7**, 218-225.