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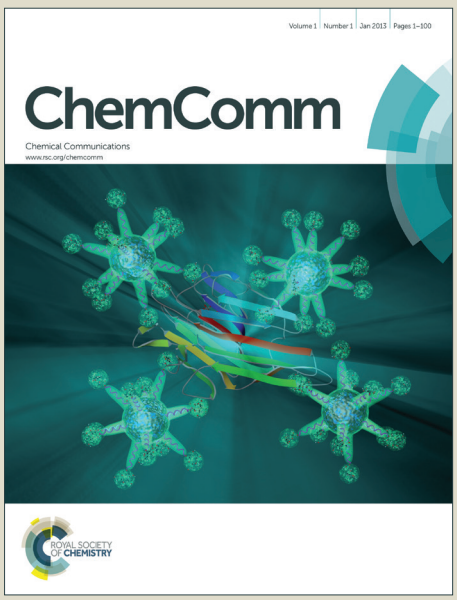

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ARTICLE TYPE

Switchable and selective detection for Zn^{2+} or Cd^{2+} in living cells based on 3'-O-substituted arrangement benzoxazole-derived fluorescent probes

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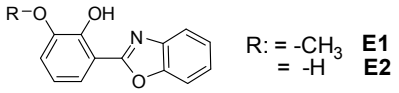
Two benzoxazole-derived ESIPT fluorescent sensors E1 and E2 show highly selectively detect Zn^{2+} and Cd^{2+}, respectively, in aqueous solution and living cells. The selectivity switching from Zn^{2+} to Cd^{2+} is attributed to the different binding mode which is dependent on the 3'-O-substituted arrangement.

Zinc ions (Zn^{2+}), as the second most abundant transition metal ion in the human body, plays a very important role in biological processes such as gene transcription, immune function, pathology, neural signal transmission, as well as catalysis of proteins.[1] Zinc imbalance is associated with a number of pathological disorders including Alzheimer's disease, epilepsy, Parkinson's disease, ischemic stroke and infantile diarrhea.[2] On the other hand, cadmium ions (Cd^{2+}), being in the same group of zinc ions in the periodic table, is a heavy metal and widely used in industry and agriculture including the production of electroplating, metallurgy, batteries, etc.[3] Cd^{2+} is highly toxic and can be easily absorbed and accumulated by plants and other organisms, leading to renal dysfunction, calcium metabolism disorders and cancers.[3c] Thereby the measurement of Zn^{2+} or Cd^{2+} in physiological media has been considered as an essential factor and addressed target in these relative diseases. Among the methods developed for Zn^{2+} or Cd^{2+} sensing, fluorescent probes have attracted considerable attention due to their simple operation, high sensitivity, noninvasiveness and real time detection.[4] Hitherto, significant effort has been devoted to the development of fluorescent sensors Zn^{2+} or Cd^{2+} with successful applications to image in living cells.[5] However, most of them suffered from limitative selectivity over biologically abundant metal ions like Fe^{2+}/Fe^{3+} and Cu^{2+} due to moderate coordination nature of Zn^{2+} and the use of di-2-picolyamine (DPA) as chelator.[6] In addition, Zn^{2+} and Cd^{2+} have similar properties to lead to similar binding mode to acceptor of sensors, providing identical spectral changes that disenable to discriminate them. Thus it is a large challenge to design the sensors for highly selective detection Zn^{2+} or Cd^{2+}. Although many fluorescent sensors with modified receptors can efficiently and highly detect Zn^{2+} or Cd^{2+}, there is no reliable design guide for selective recognition for Zn^{2+} or Cd^{2+}. Recently, Xu *et al* reported a fluorescent sensor with tautomerization-based transformable receptor.[10] Although the sensor shows distinct fluorescence change to discriminate Zn^{2+} and Cd^{2+} because of different binding mode (Zn^{2+} in an imidic acid form, Cd^{2+} in an amide tautomeric form), it can only recognize Zn^{2+} in the presence of both Zn^{2+} and Cd^{2+} due to different binding affinity. In order to study and understand the binding properties and diversity of Zn^{2+} and Cd^{2+}, it is necessary to conduct structure-tuned receptor to bring switchable selectivity for Zn^{2+} or Cd^{2+}.

Recently, fluorescent sensors based on excited-state intramolecular proton transfer (ESIPT), as seen from 2-(2'-hydroxyphenyl)benzoxazole (HBO), have been attracted more interests.[7] ESIPT sensors exhibit dual emissions from both the excited *enol* and *keto* tautomers. Fluorescent sensing of metal ions could realize by prohibiting ESIPT through the coordination of metal ion with ESIPT centers, resulting in detectable spectral change.[7] Compared with the widely used photoinduced electron transfer (PET) mechanism for Zn^{2+} or Cd^{2+}, the fluorescent sensors based on ESIPT can afford many advantages including dual fluorescence intensity changes and large Stokes shift.[9] They can detect metal ions with ratiometric fluorescent response and near infrared (NIR) fluorescent signal (700-900 nm), providing the self-calibration function and avoiding photodamage, scattering light and strong interference derived from short wavelength emission in biological media.[8]

Herein, we present two ESIPT fluorescent sensors **E1** and **E2** for Zn^{2+} and Cd^{2+} sensing, respectively (Scheme 1). Both of them show high sensitivity and selectivity in aqueous solution. Compared with **E1**, **E2** has no methyl group at 3'-hydroxyl position. With and without methyl group, the selectivity of sensor switches from Zn^{2+} to Cd^{2+}. As a proof-of-principle method, substituent arrangement-induced selectivity switching has been proved, which would be helpful to design of fluorescent sensors for efficient discrimination Zn^{2+} and Cd^{2+}.



Scheme 1 Structure of **E1** and **E2**

The detailed synthetic procedure was described in the Supporting Information. Briefly, **E1** was readily synthesized by condensation reaction between O-vanillin and 2-aminophenol. Subsequently, **E2** could be obtained from the demethylation of **E1** in the presence of BBr_3. All these compounds were fully characterized by ^1H NMR, ^{13}C NMR and HRMS.

In CH_3CN/HEPES buffer (1:1, v/v, pH 7.4), **E1** exhibited one absorption peak at 325 nm, while upon addition of Zn^{2+}, the absorption at 325 nm gradually decreased, whereas a new absorption peak appeared at 370 nm with a well-defined isosbestic point at 340 nm (Fig. S1a). Accordingly, upon excitation at 305 nm, the maximum emission at 380 nm stabilized and a new emission peak at 455 nm appeared, which evidently increased in intensity upon successive addition of Zn^{2+} (Fig. S2). The two emission bands at 380 and 455 nm can be attributed to the normal isomer (N* emission) and tautomer (T* emission) of **E1**, respectively.[7] The observed fluorescence increasing of tautomer indicated that the Zn^{2+} binding is beneficial for the stable of tautomer. The new emission peak can be used for the ratiometric fluorescent measurement of Zn^{2+}. Interestingly, upon excitation at 360 nm, except for emission band at 455 nm, a new NIR emission band at 880 nm occurred (Fig. 1). Their fluorescence intensities linearly gradually increased with addition of Zn^{2+} (Fig. S3). After addition of 1 equivalent of Zn^{2+}, the fluorescence quantum yield of **E1** changes from 0.0206 to 0.34. The Job's plot revealed 1:1 stoichiometry for the binding between **E1** and Zn^{2+} (Fig. S4). The binding constant was calculated to be 6.51×10^4 M^{-1} with the detection limit of 1.63×10^{-8} M. The selectivity of **E1** to various metal ions was further examined. As shown in Fig. S6, only Zn^{2+} promotes significant fluorescence intensity enhancement at 458 and 880 nm, whereas other metal ions cause no detectable spectra change except that Cu^{2+} induces somewhat fluorescence quenching. To explore the possible utility of **E1** as fluorescent sensor for Zn^{2+}, competitive experiments were carried out in the presence of 20 equivalents of Zn^{2+} and 20 equivalents of various other cations (Fig 2). Although Cd^{2+} exerts a weak increasing effect on the probe, the little interference can be eliminated by cysteine (Cys) that is abundant in vivo.[10] The **E1**+Zn^{2+} complex is stable in the presence of Cys while **E1**+Cd^{2+} complex is dissociated owing to the competition of Cys (Fig. S7). These results suggested that **E1** shows excellent binding selectivity for Zn^{2+} and can detect Zn^{2+} with NIR emission.

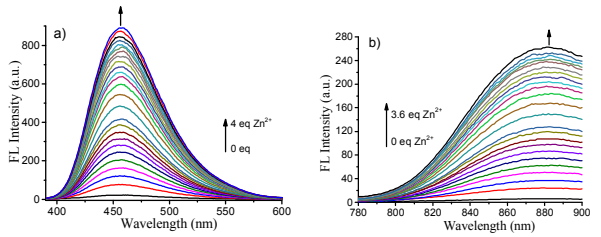


Fig. 1 The fluorescent spectra change of **E1** (10 μM) at short and long wavelength region upon addition of Zn^{2+} in CH_3CN/HEPES Buffer (1:1, v/v, pH 7.4), λ_{ex}=360 nm.

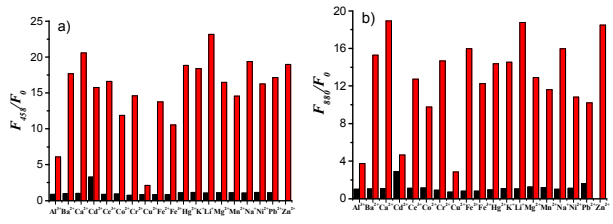


Fig. 2 Emission intensity of **E1** (10 μM) at 458 (a) and 880 nm (b) in CH_3CN/ HEPES Buffer (1:1, v/v, pH 7.4) in the presence of different metal ions (200 μM) with the excitation at 360 nm (blank bar). Red bars represent the intensity with subsequent addition of Zn^{2+} ions (200 μM).

Compared with **E1**, **E2** only lost the methyl group on the 3'-hydroxyphenyl position. In identical condition, **E2** has similar UV-Vis spectra change upon addition of Cd^{2+} with a well-defined isosbestic point at 350 nm (Fig. S1b). Upon excitation at 305 nm, **E2** has two maximum emission bands at 360 and 468 nm (Fig. S8), assigned to normal isomer (N* emission) and tautomer (T* emission), respectively. As excited at 360 nm, the fluorescence intensity at 468 nm evidently increased with successive addition of Cd^{2+}. The fluorescence quantum yield (Φ) was increased from 0.0012 to 0.032 in the presence of 1 equivalent of Cd^{2+}. Its fluorescence intensity linearly gradually increased with addition of Cd^{2+} (Fig. 3a and S9). The Job's plot revealed 1:1 stoichiometry for the binding between **E2** and Cd^{2+} (Fig. S10). The binding constant was calculated to be 1.99×10^4 M^{-1} and the detection limit was 1.33×10^{-7} M. In sharp contrast to **E1**, **E2** showed excellent selectivity to Cd^{2+}. Competitive experiments were carried out in the presence of 20 equivalents of Cd^{2+} and 20 equivalents of various other cations (Fig. 3b). Except that Mg^{2+} induced a little fluorescence increasing, other various metal ions including Zn^{2+} ions caused no detectable spectra change at 469 nm. The result suggested that **E2** can highly detect Cd^{2+} without interference by Zn^{2+}. Moreover, competitive experiments were carried out in the presence of Cys, showing that there is no any interference from Cys for the probe **E2** to detect Cd^{2+} ions possibly due to the stronger binding capability between **E2** and Cd^{2+} (Fig. S21-22). Both of them are pH independent in the range of 7-8, demonstrating that they can detect metal ions in biological environment (Fig. S11-12).

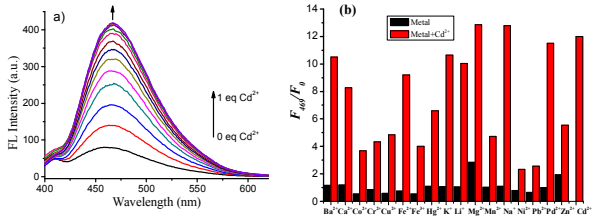


Fig. 3 (a) Fluorescence emission spectra of **E2** (10 μM) upon addition of Cd^{2+} in CH_3CN/HEPES Buffer (1:1, v/v, pH 7.4), λ_{ex}=360 nm; (b) Emission intensity of **E2** (10 μM) at 469 nm in CH_3CN/HEPES Buffer (1:1, v/v, pH 7.4) in the presence of different metal ions (200 μM) with the excitation at 360 nm (blank bar). Red bars represent the intensity with subsequent addition of Cd^{2+} ions (200 μM).

Interestingly, the selectivity can be switched from Zn^{2+} to Cd^{2+} through facial substituent effect of benzoxazole derivatives. To further evaluate the response nature and gain the insight into the recognition mechanism toward Zn^{2+} and Cd^{2+}. The ^1H NMR spectra titration of **E1** with Zn^{2+} and **E2** with Cd^{2+} were investigated. The chemical shift of the hydroxyl –OH can be used to value whether metal ion is bond to the hydroxyl oxygen. For **E1**, the Zn-O bond results in the disappearance of -OH resonance peak at 11.20 with addition of 2 equivalents of Zn^{2+} in DMSO-d_6 (Fig. S13). This result indicates that –OH group was involved in the binding with Zn^{2+}. In case of **E2**, the Cd^{2+} binding results in the –OH proton at 2' position upfield shift from 11.10 to 11.07

and the –OH proton at 3' position downfield shift from 9.56 to 9.60 (Fig. S14). The two different effects on the –OH proton could be considered as a result of the Cd^{2+} binding. Through-bond propagation increases the electron density on the hydroxyl group at 2' position and produces a shielding effect. While through-space effect increases the polarization of the hydroxyl group at 3' position, the partial positive charge causes a deshielding effect and downfield shift of its proton.[11] The non-vanishing of –OH protons at 2' and 3' position after addition of Cd^{2+} suggested that the two hydroxyl groups do not participate in the binding with Cd^{2+}. The strong intramolecular hydrogen bonding possibly prevents Cd^{2+} ions from binding with hydroxyl groups. The proposal binding modes of **E1** with Zn^{2+} and **E2** with Cd^{2+} were conducted, from which ions-induced different binding profiles are attributed to different selectivity (Scheme S1). More direct evidences were obtained by the ESI mass spectra, where the ion peak at *m/z* 510.31 (Fig. S15) corresponded to the molecular ion peak of [**E1**-H+Zn^{2+}+2CH_3OH+H_2O+ClO_4^-+Na^+] (calcd=510.15). For **E2**, the peak at *m/z* 451.74 (Fig. S16) corresponded to the molecular ion peak of [**E2**+Cd^{2+}+ CH_3OH+H_2O+NO_3^-] (calcd=451.69).

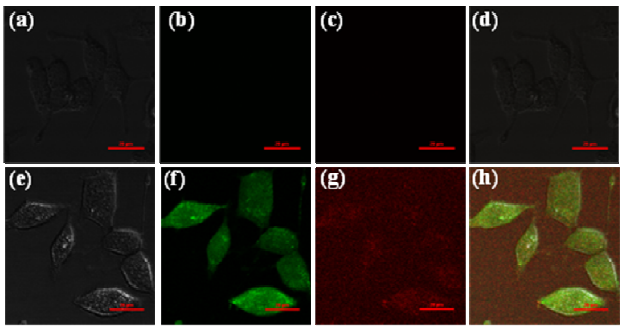


Fig. 4 Fluorescence images of SKOV-3 cells. (a-d) SKOV-3 cells incubated with probe **E1** (10 μM) for 30 min; (e-h) images of cells after treatment with probe **E1** (10 μM) for 30 min and subsequent treatment of the cells with 50 μM Zn^{2+} for 20 min. (a and e) Bright-field images of the SKOV-3 cells in samples; b) and f) images taken in green field; (c and g) images taken in red field; and (d and h) is the overlap of brightfield and fluorescence. Scare bar: 20 μm.

To further investigate the biological application of **E1** and **E2**, the fluorescence microscopy experiment in living cells was carried out. When ovarian cancer cells (SKOV-3) were incubated with 10 μM **E1** and **E2** in culture medium at 37 °C for 1 h, relatively, no detectable emission were observed. After incubation with Zn^{2+} and Cd^{2+} for **E1** and **E2**, respectively, strong green emission can been clearly showed, indicating a very good cellular uptake and efficiently fluorescent detection in living cells (Fig. 4 and Fig. S17). Moreover, NIR red emission can be detected in the case of **E1** treated with Zn^{2+}.

In summary, two kinds of benzoxazole-derived ligands **E1** and **E2**, being different at a methyl substituent, have been presented. For **E1**, it can selectively detect Zn^{2+} in buffer solution and living cells with fluorescence intensity increasing at 455 and 880 nm. The selectivity can be further improved without interference with Cd^{2+} in the presence of biological Cys. For **E2**, it shows excellent selectivity toward Cd^{2+} and can be applied for living cells image. The possible binding modes between them were investigated by ^{1}H NMR titration spectra, from which the reasons and recognition mechanisms were interpreted. As a proof-of-principle method, substituent arrangement-induced selectivity switching would be helpful in the design of fluorescent sensors for other metal ions.

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Notes and references

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† Electronic Supplementary Information (ESI) available: [details of synthetic procedure and additional spectra]. See DOI: 10.1039/b000000x/

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Graphic abstract: