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ARTICLE TYPE

## Convenient Detection of Thiol Functional Group Using H/D Isotope Sensitive Raman Spectroscopy

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Raman spectra of several thiols (amino acids, peptides and organic) show that the C-S-H bending mode ( $\beta_{\text{CSH}}$ ) shifts from  $\sim 850 \text{ cm}^{-1}$  to  $\sim 620 \text{ cm}^{-1}$  on deuteration of the thiol proton by simply dissolving them in  $\text{D}_2\text{O}/\text{CD}_3\text{OD}$  where detection by  $^1\text{H}$  NMR is not possible. A nondestructive analytical tool for the detection of thiols in solid/neat and solution is developed.

Thiols form an important subclass of molecules in nature.<sup>1</sup> Naturally occurring thiols like glutathione (GSH), which is a tripeptide containing a cysteine residue (Fig. 1),<sup>2</sup> is responsible for maintaining cellular redox balance.<sup>3-5</sup> GSH plays an important role in toxicology, drug metabolism, protein, DNA synthesis and transport<sup>4, 6</sup> and in the development/progression of cancer.<sup>1, 7</sup> It plays a protecting role against cellular damage by the reactive oxygen species (ROS)<sup>5</sup> generated during metabolism.<sup>7, 8</sup> Thiol containing pharmaceuticals such as penicillamine (Fig. 1), mercaptopurine, and captopril are effective in the treatment of many serious diseases like arthritis, hypertension, skin disease and cancer.<sup>9</sup> Several natural products containing thiol or disulfide such as coenzyme A,<sup>10</sup> thioterpineol, lipoic acid,<sup>4, 10</sup> and N-2-mercaptopropionyl glycine<sup>11, 12</sup> are known. Apart from these, thiol has widely used to prepare self assembled monolayer (SAM) on Au, Ag and other coinage metal surfaces.<sup>13, 14</sup> SAM modified surfaces are important for nanoscience, nanotechnology, molecular electronics and nonlinear optics.<sup>15-20</sup> Protein, cell and other biological species can be easily immobilized on SAMs,<sup>21-23</sup> which can be extensively used for biotechnological applications such as in tissue engineering and biosensing. Investigations of these areas of science require synthesis of thiols and their adequate characterization.<sup>9</sup>

Thiol group in a molecule is detected with the help of several colorimetric or fluorescence assays.<sup>3, 24, 25</sup> In such cases the detection is achieved at the cost of the material *i.e.* the thiol used can not be easily recovered. The most commonly used spectroscopic tools for the detection of thiols are FTIR and  $^1\text{H}$  NMR. The S-H stretching frequency appears at  $\sim 2550 \text{ cm}^{-1}$  in the FTIR spectrum of a thiol containing molecule. However, the intensity of this peak is very weak (S-H is a weak dipole) and is difficult to detect in dilute solutions of thiols. For alkyl thiols the thiol proton resonance is observed between 1.2-1.8 ppm in  $^1\text{H}$  NMR.<sup>26</sup> This peak is often masked by alkyl protons in the case of long chain thiols<sup>27, 28</sup> and is not observed when measured in a protic solvent like  $\text{CD}_3\text{OD}$  or  $\text{D}_2\text{O}$  (Fig. S1-S7 in SI) as the thiol proton is readily exchanged with that of the solvent. This poses a

practical problem in the detection of the thiol functional group, in particular, in water soluble molecules in spite of their importance in chemistry and biology. Hence a convenient non-destructive spectroscopic tool for the detection of a thiol functional group is extremely desirable.

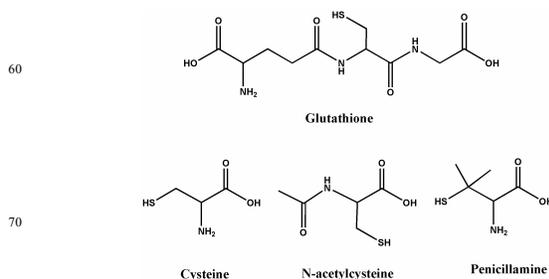
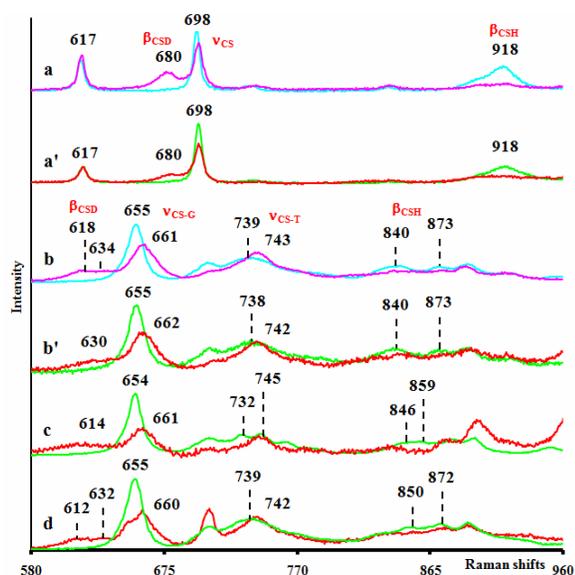


Fig. 1. Representation of biologically important thiols.

The Raman spectrum of thiol containing molecules show the C-S vibrations ( $\nu_{\text{CS}}$ ) in between  $650\text{-}700 \text{ cm}^{-1}$ , *i.e.* in a finger print region where most other functional groups do not interfere.<sup>29, 30</sup> However this fails to serve as a diagnostic feature as this vibration varies in position depending on the thiol and is also present in disulfides and thioethers.<sup>31, 32</sup> A while back it was reported that the C-S-H bending mode ( $\beta_{\text{CSH}}$ ) in ethanethiol shifts from  $870 \text{ cm}^{-1}$  to  $625 \text{ cm}^{-1}$ , in the gas phase, on deuteration of the thiol proton.<sup>33</sup> In this manuscript we show that the  $\beta_{\text{CSH}}$  observed around  $850\text{-}900 \text{ cm}^{-1}$  for different thiols (neat or in solution), shifts to  $600\text{-}630 \text{ cm}^{-1}$  on deuteration of the thiol proton (resulting in RSD). Further, the deuteration can be performed by simply dissolving thiols in protic solvents like  $\text{CD}_3\text{OD}$  and  $\text{D}_2\text{O}$ . Using this technique the RSH functionality has been detected in a series of organic molecules, amino acids and peptides known to bear a thiol group. The  $\sim 250 \text{ cm}^{-1}$  isotope shift observed in the  $\beta_{\text{CSH}}$  mode on deuteration of the thiol proton is absent in disulfides.



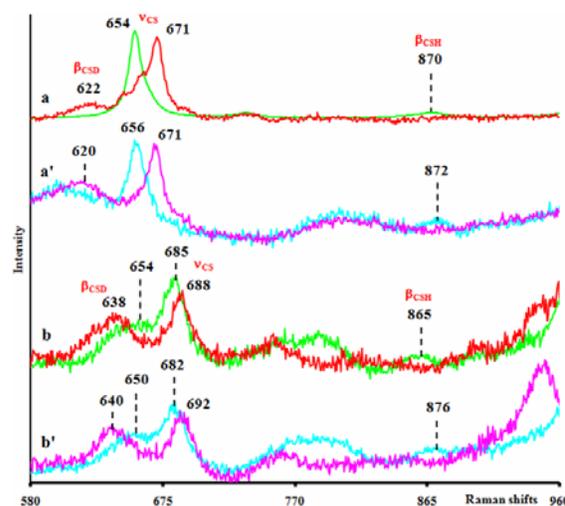
**Fig. 2.** The Raman spectra of (a) neat PhSH and PhSD; (a') PhSH in CH<sub>3</sub>OH and CD<sub>3</sub>OD; (b) neat C<sub>8</sub>H<sub>17</sub>SH and C<sub>8</sub>H<sub>17</sub>SD (b') C<sub>8</sub>H<sub>17</sub>SH in CH<sub>3</sub>OH and CD<sub>3</sub>OD; (c) C<sub>4</sub>H<sub>9</sub>SH in CH<sub>3</sub>OH and CD<sub>3</sub>OD and (d) C<sub>12</sub>H<sub>25</sub>SH in CH<sub>3</sub>OH and CD<sub>3</sub>OD. Sky blue and green spectra are for protonated species or solutions and pink and red spectra are for deuterated species or solutions. In solution spectra concentration was around 2-3 mM. Raman shifts are in cm<sup>-1</sup>.  $\nu$  is stretching mode,  $\beta$  is bending mode, G is gauche and T is trans conformation.

The  $\beta_{\text{CSH}}$  of neat PhSH occurs at 918 cm<sup>-1</sup> and the  $\nu_{\text{CS}}$  is observed at 698 cm<sup>-1</sup> (Fig. 2a, sky blue).<sup>34, 35</sup> On deuterating the thiol proton (PhSD, Fig. 2a, pink), the peak at 918 cm<sup>-1</sup> disappears and the  $\beta_{\text{CSD}}$  appears at 680 cm<sup>-1</sup> i.e. 238 cm<sup>-1</sup> shift is observed on replacing the thiol proton with deuterium. H/D isotope shifts of similar magnitude (i.e. 200-250 cm<sup>-1</sup>) are also reported for the H-X-H bending modes of H<sub>2</sub>O and H<sub>2</sub>S.<sup>36, 37</sup> The H/D exchange can easily be performed by dissolving PhSH in CH<sub>3</sub>OH (for H) or CD<sub>3</sub>OD (for D). The data obtained in CH<sub>3</sub>OH (Fig. 2a', green) and CD<sub>3</sub>OD (Fig. 2a', red) solutions of PhSH indicate that the changes observed in the data (H/D isotope shifts) are identical to those obtained between neat PhSH and PhSD. In neat octanethiol (C<sub>8</sub>H<sub>17</sub>SH), two  $\beta_{\text{CSH}}$  modes are observed due to the presence of gauche (G) and trans (T) conformations across the C<sub>1</sub>-C<sub>2</sub> bond.<sup>38</sup> These vibrations shift from 840 cm<sup>-1</sup> and 873 cm<sup>-1</sup> in neat C<sub>8</sub>H<sub>17</sub>SH (Fig. 2b, sky blue) to 618 cm<sup>-1</sup> and 634 cm<sup>-1</sup> in neat C<sub>8</sub>H<sub>17</sub>SD (Fig. 2b, pink). The same H/D isotope shifts are observed in the data obtained in CH<sub>3</sub>OH (Fig. 2b', green) and CD<sub>3</sub>OD (Fig. 2b', red) solutions of C<sub>8</sub>H<sub>17</sub>SH. Note that in the case of alkyl thiols the  $\nu_{\text{CS}}$  for the G and T conformations, which are at 655 cm<sup>-1</sup> and 739 cm<sup>-1</sup> for C<sub>8</sub>H<sub>17</sub>SH (both in neat and in CH<sub>3</sub>OH solution), shifts to 661 cm<sup>-1</sup> and 743 cm<sup>-1</sup>, respectively, in C<sub>8</sub>H<sub>17</sub>SD (both in neat and CD<sub>3</sub>OD solutions). The > 200 cm<sup>-1</sup> lowering on the  $\beta_{\text{CSH}}$  mode and ~6-7 cm<sup>-1</sup> increase in the  $\nu_{\text{CS}}$  mode is observed for a series of alkyl thiols (which includes butanethiol, dodecanethiol and ethanethiol, Fig. 2c, 2d, 3a and 3a'). In particular, the appearance of a new peak in the 600-630 cm<sup>-1</sup> (at a lower energy than the C-S stretching vibration) on deuteration is clearly observed for all the thiols and provides a

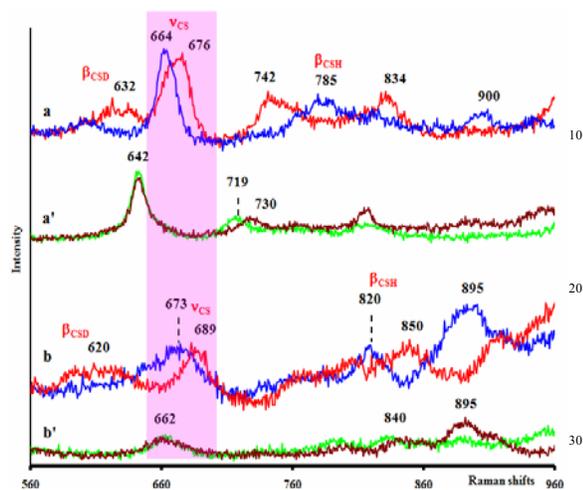
clear spectroscopic signature for the presence of thiol group in long chain alkyl thiols and in aromatic thiols in neat or in methanolic solution.<sup>29, 35</sup>

The trend continues to hold for thiol molecules soluble in water. The  $\beta_{\text{CSH}}$  mode in C<sub>2</sub>H<sub>5</sub>SH shifts from ~870 cm<sup>-1</sup> to ~620 cm<sup>-1</sup> after deuteration<sup>33, 39</sup> in both CH<sub>3</sub>OH/CD<sub>3</sub>OD (Fig. 3a, green/red) and H<sub>2</sub>O/D<sub>2</sub>O (Fig. 3a', blue/pink) solvents. The increase in the  $\nu_{\text{CS}}$  on deuteration of the thiol group, in this case, is ~16 cm<sup>-1</sup> which is significantly more than those observed for the long chain thiols. Since the thiol protons are easily exchangeable in H<sub>2</sub>O/D<sub>2</sub>O, this analytical technique may be extended to several biologically relevant molecules bearing thiol groups.

N-acetyl cysteine (Fig. 1) is possibly the smallest analogue of a cysteine containing peptide. The C-S region in the Raman spectrum in methanolic solution shows two vibrations at 654 cm<sup>-1</sup> and 685 cm<sup>-1</sup> representing the C=O deformation (of amide group) and  $\nu_{\text{CS}}$ , respectively<sup>40</sup> (Fig. 3b, green). In CD<sub>3</sub>OD (Fig. 3b, red) the  $\nu_{\text{CS}}$  appears at 688 cm<sup>-1</sup> and the  $\beta_{\text{CSH}}$  appears at 638 cm<sup>-1</sup>. Similarly the  $\nu_{\text{CS}}$  and  $\beta_{\text{CSH}}$  shifts from 682 cm<sup>-1</sup> and 876 cm<sup>-1</sup> in H<sub>2</sub>O (Fig. 3b' blue) to 692 cm<sup>-1</sup> and 640 cm<sup>-1</sup> in D<sub>2</sub>O (Fig. 3b', pink), respectively. These shifts are consistent with the trend observed for alkyl and aromatic thiols so far. The  $\nu_{\text{CS}}$  and  $\beta_{\text{CSH}}$  of cysteamine solubilized in water occurs at 664 cm<sup>-1</sup> and 785 cm<sup>-1</sup>, respectively.<sup>41</sup> These vibrations shift to 676 cm<sup>-1</sup> and 632 cm<sup>-1</sup>, respectively, (Fig. 4a, blue to red) when dissolved in D<sub>2</sub>O. The corresponding disulfide shows no H/D isotope sensitive band in this region (Fig. 4a', green to brown). GSH show the  $\nu_{\text{CS}}$  at 673 cm<sup>-1</sup> and the  $\beta_{\text{CSH}}$  mode at 820 cm<sup>-1</sup> in H<sub>2</sub>O<sup>42</sup> (Fig. 4b, green) which shift to 689 cm<sup>-1</sup> and 620 cm<sup>-1</sup>, respectively, in D<sub>2</sub>O (Fig. 4b, red). GSSG the corresponding disulfide does not show any H/D isotope sensitive band in this region (Fig. 4b', green to brown). The difference spectra of these experimental spectra are shown in supporting information (Fig. S8-S10)



**Fig. 3.** The Raman spectra of C<sub>2</sub>H<sub>5</sub>SH in (a) CH<sub>3</sub>OH (green) and CD<sub>3</sub>OD (red) and in (a') H<sub>2</sub>O (sky blue) and D<sub>2</sub>O (pink), and of N-acetyl cysteine in (b) CH<sub>3</sub>OH and CD<sub>3</sub>OD and in (b') H<sub>2</sub>O and D<sub>2</sub>O (same colour code). Concentration was around 2 mM. Raman shifts are in cm<sup>-1</sup>.  $\nu$  is stretching mode and  $\beta$  is bending mode.

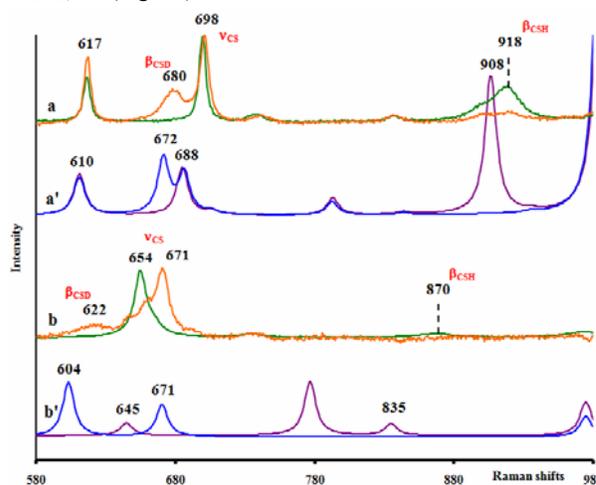


**Fig. 4.** The Raman spectra of (a) cysteamine, (a') cystamine and of (b) glutathione, (b') glutathione (S-S) solubilized in H<sub>2</sub>O and D<sub>2</sub>O. Blue and red spectra correspond to free thiols dissolved in H<sub>2</sub>O and D<sub>2</sub>O, respectively. The brown and green lines represent Raman data of the disulphides dissolved in H<sub>2</sub>O and D<sub>2</sub>O, respectively. The  $\nu_{CS}$  stretching region is highlighted. Concentration was around 2 mM. Raman shifts are in cm<sup>-1</sup>.  $\nu$  is stretching mode and  $\beta$  is bending mode.

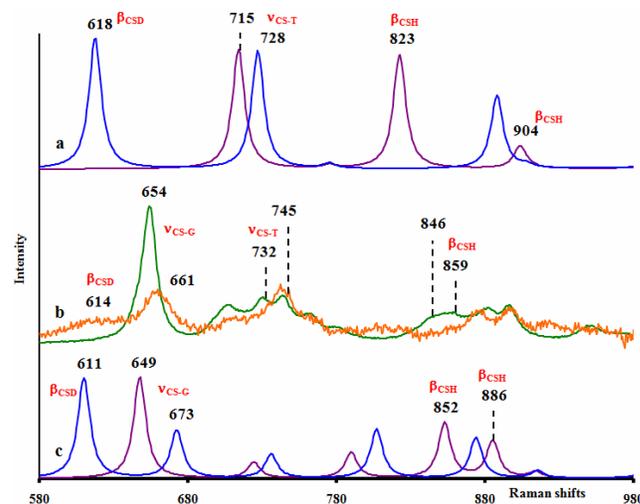
The experimentally observed H/D shifts in the Raman spectra of thiols can be corroborated to density functional theory (DFT, using BP86 functional, 6-311g\* basis set in Gaussian 03 ver. C02)<sup>43-45</sup> calculated H/D shifts. DFT calculations on PhSH indicate that the  $\beta_{CSH}$  is at 908 cm<sup>-1</sup> and it shifts to 672 cm<sup>-1</sup> in PhSD (Fig. 5a', purple to blue). These values are in close agreement with the experimental data which shows the  $\beta_{CSH}$  shift from 918 cm<sup>-1</sup> in PhSH to 680 cm<sup>-1</sup> in PhSD (Fig. 5a, green to orange). Similarly, the  $\beta_{CSH}$  in C<sub>2</sub>H<sub>5</sub>SH is calculated to shift by 231 cm<sup>-1</sup>, from 835 cm<sup>-1</sup> to 604 cm<sup>-1</sup>, on deuterating the thiol proton (Fig. 5b', purple to blue). The experimental data show a shift of 250 cm<sup>-1</sup> in the  $\beta_{CSH}$  mode on deuteration (Fig. 5b, green to orange). Furthermore the DFT calculations successfully reproduce the shift of the C-S stretching vibration of C<sub>2</sub>H<sub>5</sub>SH to higher energies on deuterating the thiol proton.

DFT calculations are also used to calculate the effect of H/D isotopic substitution on the  $\beta_{CSH}$  and  $\nu_{CS}$  modes of alkyl thiols having T or G conformations of the C<sub>1</sub>-C<sub>2</sub> bond (Fig. 6). In the case of C<sub>4</sub>H<sub>9</sub>SH having a T conformation, the  $\nu_{CS}$  and the  $\beta_{CSH}$  shifts from 715 cm<sup>-1</sup> and 823 cm<sup>-1</sup> to 728 cm<sup>-1</sup> and 618 cm<sup>-1</sup>, respectively, on deuterating the SH group (Fig. 6a, purple to blue). Alternatively, in G conformation, the  $\nu_{CS}$  and the  $\beta_{CSH}$  shifts from 649 cm<sup>-1</sup> and 852 cm<sup>-1</sup>, 886 cm<sup>-1</sup> to 673 cm<sup>-1</sup> and 611 cm<sup>-1</sup>, respectively, on deuterating the SH group (Fig. 6c). These calculations a) reproduce the relative energies of C-S vibration of thiols having G and T orientations and b) reproduce the shifts in  $\beta_{CSH}$  and  $\nu_{CS}$  on deuteration of the thiol. These calculations also indicate the presence of the  $\beta_{CSD}$  modes of both G and T conformers energetically close to each other explaining in the broad  $\beta_{CSD}$  peak observed in the the experimental spectrum of C<sub>4</sub>H<sub>9</sub>SD at 614 cm<sup>-1</sup> (Fig. 6b, orange). Similar broadening of the

$\beta_{CSD}$  band is also observed for both C<sub>8</sub>H<sub>9</sub>SD (Fig. 2b and 2b') and <sup>75</sup>C<sub>12</sub>H<sub>25</sub>SD (Fig. 2d).



**Fig. 5.** The Raman spectra of (a) PhSH in CH<sub>3</sub>OH (green) and CD<sub>3</sub>OD (orange), of (a') PhSH and PhSD by theoretical predictions, of (b) C<sub>2</sub>H<sub>5</sub>SH in CH<sub>3</sub>OH (green) and CD<sub>3</sub>OD (orange) and of (b') C<sub>2</sub>H<sub>5</sub>SH and C<sub>2</sub>H<sub>5</sub>SD by theoretical predictions. In theoretical spectra violet and blue 80 represents the protonated and deuterated results, respectively. Raman shifts are in cm<sup>-1</sup>.



**Fig. 6.** The Raman spectra of (b) C<sub>4</sub>H<sub>9</sub>SH in CH<sub>3</sub>OH (green) and CD<sub>3</sub>OD (orange). Theoretically predicted results in (a) T conformation and in (c) G conformation. The purple and blue lines represent the calculated Raman spectrum for protonated and deuterated thiols, respectively. Raman shifts are in cm<sup>-1</sup>.  $\nu$  is stretching mode,  $\beta$  is bending mode, G is gauche and T is trans conformation.

In summary, observation of a new peak below the C-S stretch 90 and the shift of the C-S vibration to higher energies on deuteration of the thiol proton is found to be true for a broad range of thiol group containing molecules and may qualify a Raman signature of a thiol group. The protonated and deuterated thiols can be probed in neat or in solution resulting in similar H/D 95 isotopic shifts. The deuteration can be performed in-situ by dissolving the thiol group containing molecule in CD<sub>3</sub>OD or in

D<sub>2</sub>O. The difference spectra of these theoretical results are shown in supporting information (Fig. S11-S12)

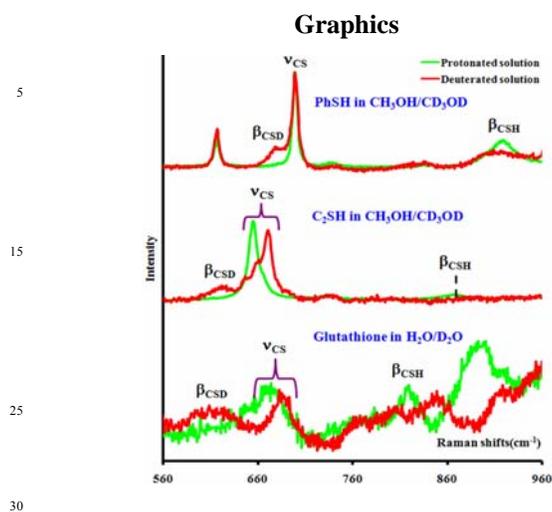
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## Notes and references

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<sup>†</sup> Electronic Supplementary Information (ESI) available: Experimental procedure, computational details, <sup>1</sup>H NMR spectra of thiols, the Raman spectra of above mentioned thiols along with their difference spectra and table of the spectroscopic data. See DOI: 10.1039/b000000x/

1. C. Jacob, *Natural Product Reports*, 2006, **23**, 851-863.
2. R. Requejo, T. R. Hurd, N. J. Costa and M. P. Murphy, *The FEBS journal*, 2010, **277**, 1465-1480.
3. X. Chen, Y. Zhou, X. Peng and J. Yoon, *Chemical Society Reviews*, 2010, **39**, 2120-2135.
4. T. Toyooka, *Journal of Chromatography B*, 2009, **877**, 3318-3330.
5. M. A. Baker and R. J. Aitken, *Molecular and Cellular Endocrinology*, 2004, **216**, 47-54.
6. A. Meister and M. E. Anderson, *Annual Review of Biochemistry*, 1983, **52**, 711-760.
7. G. K. Balendiran, R. Dabur and D. Fraser, *Cell Biochemistry and Function*, 2004, **22**, 343-352.
8. P. J. Dean, *American Journal of Physiology - Cell Physiology*, 2008, **295**, C849-C868.
9. T. J. O'Shea and S. M. Lunte, *Analytical Chemistry*, 1993, **65**, 247-250.
10. N. Haugaard, *Ann N Y Acad Sci*, 2000, **899**, 148-58., 2000.
11. M.-o. Date, T. Morita, N. Yamashita, K. Nishida, O. Yamaguchi, Y. Higuchi, S. Hirotsu, Y. Matsumura, M. Horii, M. Tada and K. Otsu, *Journal of the American College of Cardiology*, 2002, **39**, 907-912.
12. J. C. Fantinelli, L. F. Gonzalez Arbelaez, I. A. Perez Nanez and S. M. Mosca, *Experimental and Molecular Pathology*, 2013, **94**, 277-284.
13. A. Ulman, *Chemical Reviews*, 1996, **96**, 1533-1554.
14. C. Vericat, M. E. Vela, G. Benitez, P. Carro and R. C. Salvarezza, *Chemical Society Reviews*, 2010, **39**, 1805-1834.
15. J. C. Love, L. A. Estroff, J. K. Kriebel, R. G. Nuzzo and G. M. Whitesides, *Chemical Reviews*, 2005, **105**, 1103-1170.
16. Y. Arikuma, H. Nakayama, T. Morita and S. Kimura, *Langmuir*, 2011, **27**, 1530-1535.
17. N. K. Devaraj, R. A. Decreau, W. Ebina, J. P. Collman and C. E. D. Chidsey, *The Journal of Physical Chemistry B*, 2006, **110**, 15955-15962.
18. G. G. Huang, M. K. Hossain, X. X. Han and Y. Ozaki, *Analyst*, 2009, **134**, 2468-2474.
19. J. F. Smalley, H. O. Finklea, C. E. D. Chidsey, M. R. Linford, S. E. Creager, J. P. Ferraris, K. Chalfant, T. Zawodzinski, S. W. Feldberg and M. D. Newton, *Journal of the American Chemical Society*, 2003, **125**, 2004-2013.
20. D. Qin, Y. Xia and G. M. Whitesides, *Nat. Protocols*, 2010, **5**, 491-502.
21. Q. Yu and G. Golden, *Langmuir*, 2007, **23**, 8659-8662.
22. I. F. Gallardo and L. J. Webb, *Langmuir*, 2012, **28**, 3510-3515.
23. G. A. Hudalla and W. L. Murphy, *Langmuir*, 2010, **26**, 6449-6456.
24. H. S. Jung, X. Chen, J. S. Kim and J. Yoon, *Chemical Society Reviews*, 2013, **42**, 6019-6031.
25. Q. Qian, J. Deng, D. Wang, L. Yang, P. Yu and L. Mao, *Analytical Chemistry*, 2012, **84**, 9579-9584.
26. M. Hasan, D. Bethell and M. Brust, *Journal of the American Chemical Society*, 2002, **124**, 1132-1133.
27. J. P. Collman, N. K. Devaraj and C. E. D. Chidsey, *Langmuir*, 2004, **20**, 1051-1053.
28. D. Wishart, C. Bigam, A. Holm, R. Hodges and B. Sykes, *Journal of Biomolecular NMR*, 1995, **5**, 67-81.
29. M. A. Bryant and J. E. Pemberton, *Journal of the American Chemical Society*, 1991, **113**, 3629-3637.
30. M. A. Bryant and J. E. Pemberton, *Journal of the American Chemical Society*, 1991, **113**, 8284-8293.
31. A. L. Jenkins, R. A. Larsen and T. B. Williams, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2005, **61**, 1585-1594.
32. G. Zhu, X. Zhu, Q. Fan and X. Wan, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2011, **78**, 1187-1195.
33. J. R. Durig, W. E. Bucy, C. J. Wurrey and L. A. Carreira, *The Journal of Physical Chemistry*, 1975, **79**, 988-993.
34. T. H. Joo, M. S. Kim and K. Kim, *Journal of Raman Spectroscopy*, 1987, **18**, 57-60.
35. K. Kim, Y. M. Lee, H. S. Lee and K. S. Shin, *Journal of Raman Spectroscopy*, 2008, **39**, 1840-1847.
36. J. C. DeÅ k, S. T. Rhea, L. K. Iwaki and D. D. Dlott, *The Journal of Physical Chemistry A*, 2000, **104**, 4866-4875.
37. R. E. Miller and G. E. Leroi, *The Journal of Chemical Physics*, 1968, **49**, 2789-2797.
38. T. Ha Joo, K. Kim and M. Soo Kim, *Journal of Molecular Structure*, 1987, **158**, 265-274.
39. H. Wolff and J. Szydłowski, *Canadian Journal of Chemistry*, 1985, **63**, 1708-1712.
40. B. Koleva, M. Spiteller and T. Kolev, *Amino Acids*, 2010, **38**, 295-304.
41. A. Kudelski and W. Hill, *Langmuir*, 1999, **15**, 3162-3168.
42. G. G. Huang, X. X. Han, M. K. Hossain and Y. Ozaki, *Analytical Chemistry*, 2009, **81**, 5881-5888.
43. A. D. Becke, *Phys. Rev. A*, 1988, **38**, 3098-3100.
44. J. P. Perdew, *Physical Review B*, 1986, **33**, 8822-8824.
45. M. J. T. e. a. Frisch, *Gaussian 03*, 2004, C.02.



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