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An energy-optimization method to study gel-swelling in confinement†

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We recast the problem of hydrogel swelling under physical constraints as an energy optimization problem. We apply this approach to compute equilibrium shapes of hydrogel spheres confined within a jammed matrix of rigid beads, and interpret the results to determine how confinement modifies the mechanics of swollen hydrogels. In contrast to the unconfined case, we find a spatial separation of strains within the bulk of the hydrogel as strain becomes localized to an outer region. We also explore the contact mechanics of the gel, finding a transition from Hertzian behavior to non-Hertzian behavior as a function of swelling. Our model, implemented in the Morpho shape optimization environment, and validated against an experimentally demonstrated prototypical scenario, can be applied in any dimension, readily adapted to diverse swelling scenarios and extended to use other energies in conjunction.

1 Introduction

Hydrogels are polymer networks that have an incredible capacity to absorb water while remaining intact 1 . They are suitable for a variety of practical applications such as hygienic products and contact lenses and other areas². Their similarity to biological tissues have made them a promising material candidate for biomedical and bio-interface devices^{3,4} and drug delivery applications⁵. Additionally, hydrogels are used as soil conditioners to improve water retention and other desirable agricultural properties^{6,7}. The success of these improvements is known to depend on the size of the soil particles they are embedded in⁸, but the mechanisms by which confinement alters the behavior of the gel are challenging to study directly.

A recent experimental work on swelling of hydrogel confined in a granular medium 9 characterized the 3D swelling of a hydrogel sphere surrounded by transparent beads under confining pressure. If the confining pressure is weak, the hydrogel sphere tends to rearrange the surrounding matrix as it swells; for strong confinement the sphere deforms and tends to fill the interstices of the bead packing as is shown in Fig. 1A. The overall degree of swelling was found to be well described by a model that balanced swelling pressures and contact forces, making some assumptions about the distribution of deformation. While the deformation state and the swelling ratio of the hydrogel sphere can be measured, the internal stress and strain distributions are not directly accessible in the experiments.

Modeling provides a complementary approach — thermodynamic and kinetic models of hydrogel swelling have been successfully used to predict the evolution of the concentration and deformation of constrained hydrogels 10 and tested against experiment ^{11,12}. However, because hydrogels are highly deformable, they change shape considerably when swollen or indented. Due to the difficulty of capturing these dramatic deformations, modeling of hydrogels has often been restricted to simple geometries 13 . Incorporation of constraints that arise naturally in applications, such as the surrounding matrix of soil particles, is also a challenge for modeling ¹⁴.

In this paper, we aim to build a simulation methodology to predict the structure of a swollen hydrogel bead in a realistic matrix, and determine how confinement modifies the mechanical properties, such as distribution of strain, that are difficult to resolve experimentally. To do so, we recast hydrogel expansion as an optimization problem, discretize the gel using finite elements and solve the resulting problem to identify thermodynamic equilibrium states. Other finite element models of hydrogel swelling have enabled researchers to access stress and strain distributions 13,15. Variational approaches have been previously used to compute thermodynamic hydrogel profiles under certain class

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Fig. 1 Hydrogel swelling under 3D confinement. A Snapshots of an initially-spherical hydrogel (blue) embedded within a granular medium composed of glass beads (hazy transparent circles) packed within a transparent acrylic chamber. As the hydrogel swells, it deforms strongly due to confinement. Black circles show dyed beads used as tracers to quantify any deformations of the granular packing. The images are taken following the same experimental protocol as in Ref. ⁹. B Simulation snapshots of a swelling hydrogel surrounded by stationary hard-sphere beads (grey spheres) at various intermediate stages of energy minimization. C Corresponding simulated images obtained by rasterizing the configurations in subfigure B and projecting the the viewing plane.

of constraints^{14,16}. The mechanics of contact for constrained hydrogels have been also explored analytically for prototypical geometries 17 . However, there is presently no general purpose finite element scheme to solve for equilibrium hydrogel shapes with realistic constraints and/or additional energetic influence, such as surface tension, applied fields, etc. Here, we will use *Morpho*, a programmable environment for shape optimization $18,19$ to construct and solve the model.

The rest of the paper is organized as follows: In section 2, we review the Flory-Rehner theory of hydrogel swelling, formulate the equilibrium problem as a shape-driven energy optimization problem and describe the computational method. In section 3, we describe the resulting simulations for hydrogel spheres swelling in the presence of jammed beads and examine results. Finally, in section 4, we discuss other applications of our method and possible extensions to it.

2 Model

Theoretical modeling of hydrogel configurations in the literature often uses a pressure balancing approach, whereby the mixing pressure and entropic elasticity compete to determine the degree of swelling $20,21$. Here, we wish to instead pose the problem as optimizing a free energy to identify stationary states. In order to do so, we review the Flory-Rehner theory of hydrogel swelling $20,22-27$ and present it in a form amenable to discretization. The theory constructed reduces to conventional presentations of pressure balance as shown in the Appendix.

2.1 Theory

Consider a polymer hydrogel in a solvent at a fixed temperature *T* with an internal mesh of permanently crosslinked polymer chains. We note that hydrogels with dynamic or transient crosslinks are also of interest because they give rise to new relaxation dynamics and viscoelastic effects, but are not treated here. Let the number of polymers be N_p and the number of solvent molecules inside the hydrogel be *N*s. Let the volume occupied by one monomer/molecule be v_s , in the sense of Flory's lattice model $15,27$. Since v_s is fixed, along with the number of polymers *N*p in the hydrogel, the only free parameter during the hydrogel swelling is the number of solvent molecules *N*s. Since the swelling process occurs at a fixed total volume (hydrogel plus the external solvent) and temperature, the usual Helmholtz free energy for the mixing of a polymer with a solvent can be used:

$$
\Delta F_{\text{mix}} = k_{\text{B}} T \left[N_{\text{p}} \ln \phi + N_{\text{s}} \ln (1 - \phi) + \chi N_{\text{s}} \phi \right] \tag{1}
$$

where k_B is the Boltzmann constant and $\phi = xN_D/(N_s + xN_p)$ is the volume fraction of the polymer molecules, with *x* being the number of units per polymer. χ is the Flory-Huggins mixing parameter²⁷. Since we are considering highly swollen hydrogels, we can assume that $N_p \ll N_s$, thus simplifying the free energy as follows:

$$
\Delta F_{\text{mix}} = N_{\text{s}} k_{\text{B}} T \left[\ln(1 - \phi) + \chi \phi \right] \tag{2}
$$

As we noted above, this formalism can be connected to the osmotic pressure formalism by noting that the swelling process also occurs at a constant pressure 27 , and thus we can equate the Helmholtz free energy to the Gibbs free energy: $ΔF$ _{mix} = $\Delta G_{\rm mix}$ ^{27–30}. The mixing process alters the chemical potential μ of the solvent, resulting in an osmotic pressure, which can be derived from the Gibbs free energy:

$$
\Pi_{\text{mix}} = -\frac{\Delta\mu}{v_s} = -\frac{1}{v_s} \frac{\partial \Delta G_{\text{mix}}}{\partial N_s} = -\frac{1}{v_s} \frac{\partial \Delta F_{\text{mix}}}{\partial N_s}.
$$
 (3)

Similarly, the free energy associated with elasticity can be written like so:

$$
\Delta G_{\rm el} = \Delta F_{\rm el} = \frac{3k_{\rm B}T N_{\rm c}}{2} [\alpha^2 - 1 - \ln \alpha]
$$
 (4)

where N_c is the number of polymer chains, where a chain is defined as the polymer between two cross-link points²⁷, and $\alpha = (V/V_0)^{1/3} = (\phi_0/\phi)^{1/3}$ is the linear swelling ratio, with V_0 and ϕ_0 being a reference volume and fraction²⁰. The change in the free energy, under a separability approximation, can we written as

$$
\Delta F = \Delta F_{\text{mix}} + \Delta F_{\text{el}} \tag{5}
$$

Equilibrium is defined by the extremization of this free energy, which is equivalent to the balance of osmotic pressures, $\Pi =$ ∂∆*F* ∂*Ns* = 0. Note that due to the direct relationship between N_s and ϕ , the free energy can be written solely in terms of ϕ , and thus, we can cast hydrogel swelling as a free energy minimization problem with respect to ϕ .

2.2 Finite Element Modeling

We now consider a hydrogel where the volume fraction of the polymer can vary over space, defining x as the 3D spatial coordinate. Hence, we work with a free energy *density* ∆*f*mix(x), which is now a function of a spatially varying field $\phi(\mathbf{x})$. If this space is discretized using simplicial elements—in this work we use tetrahedra in 3D, but the theory is dimensionally independent and readily applicable to other kinds of elements—it is useful to consider the expression (1) for a single element. We will work in the deformed frame of reference¹⁵ as this is the most natural frame to express interpenetrability constraints as desired for the application. Hence, the energy density locally at a point x *in the deformed frame of reference* will be Eq. (1) evaluated at x divided by the volume of the element. Since this volume would also be given by $v_s(xN_p + N_s)$, we have,

$$
\Delta f_{\text{mix}} = \frac{N_{\text{s}}}{v_{\text{s}}(xN_{\text{p}} + N_{\text{s}})} k_{\text{B}} T \left[\ln(1 - \phi) + \chi \phi \right] \tag{6}
$$

$$
=\frac{(1-\phi)}{v_{\rm s}}k_{\rm B}T\left[\ln(1-\phi)+\chi\phi\right]
$$
\n(7)

This can be expressed in terms of an 'effective diameter' of the solvent molecule *d* such that $d^3 = v_s^9$. In terms of *d*, this reduces to,

$$
\Delta f_{\text{mix}} = \frac{k_{\text{B}} T}{d^3} \left[(1 - \phi) \ln(1 - \phi) + \chi \phi (1 - \phi) \right] \tag{8}
$$

Similarly for the elastic energy, we can compute the free energy density by dividing by the volume.

We wish to minimize $\Delta F = \int \Delta f(\mathbf{x}) d\mathbf{x}$, where dx is the volume

element. This shape optimization problem amounts to minimizing this free energy with respect to all the vertex positions x*i* of the mesh, $\partial \Delta F / \partial x_i = 0$. To compute these derivatives, we use the chain rule,

$$
\frac{\delta \Delta F}{\delta \mathbf{x}_i} = \frac{\delta \Delta F}{\delta N_s} \frac{\partial N_s}{\partial \mathbf{x}_i} \tag{9}
$$

and note that $\delta \Delta F / \delta N_s \propto -\Pi(x)$ as discussed in the previous section. Since each element's volume is given by $V = xN_p + N_s$, and *xN*p is a constant (total number of monomers in the system), $\partial N_s/\partial \mathbf{x}_i = \partial V/\partial \mathbf{x}_i$, and hence we find,

$$
\frac{\delta \Delta F}{\delta \mathbf{x}_i} \propto -\Pi(\mathbf{x}) \frac{\partial V}{\partial \mathbf{x}_i}
$$
 (10)

where $V(x)$ is the volume of the simplicial element and $\Pi(x)$ is the corresponding osmotic pressure. The volume is a known function of its vertices, and thus an analytical derivative of the free energy with respect to the shape of the hydrogel is obtained, facilitating high performance of the resulting code. We program this functional and its shape gradient in *Morpho* 19. Within this environment, we can now minimize this functional in the presence of additional energies and constraints¹⁹ for arbitrary geometries in any dimension. Details of the *Morpho* implementation are provided in the Appendix and codes are provided as Supplementary Information.

In this work, we assume that the chains are uniformly distributed throughout the hydrogel, so N_c does not depend on x , but the formulation above and the implementation in *Morpho* can be easily tweaked to allow a spatially varying initial *N*c. It can be seen that we have three non-dimensional parameters, namely, the Flory-Huggins mixing parameter χ , the relative strength of the elastic energy to the mixing energy $N_\mathrm{c} \alpha^3/V_0$ and the reference volume fraction $\phi_0{}^{20}$. Given an initial value of ϕ , we can vary these parameters to change the minima of the overall free energy. Thus, we can tune the volumetric swelling ratio, given by $r_{sw} = V_f/V_i = \phi_i/\phi_f = \phi_i/\phi_{eq}$, where the subscripts *i* and *f* refer to initial and final (equilibrium) states. Motivated by the experiments in Ref.⁹, we choose the values $\chi = 0.499$, $N_c \alpha^3/V_0 = 1$ and $\phi_0 \sim 0.036$, which together set the equilibrium value $\phi_{eq} \sim 0.1$. Varying the initial volume fraction ϕ_i between 0.1 and 1, we get volumetric swelling ratios $r_{sw} = \phi_i / \phi_{eq} \sim 1 - 10$, allowing us to access the range observed in the experiments.

We leverage a convenient hard constraint available in *Morpho*, whereby vertices are excluded from a boundary defined by the contours or level-sets of a scalar function. Inspired by the experiments described in the introduction⁹, we introduce hardsphere beads surrounding the hydrogel. To mimic the experimental geometry, N_b hard-spheres are distributed around the hydrogel sphere in contact with its surface. An illustrative example, Fig. 1B, shows snapshots as the minimization proceeds from an initial spherical state for $N_b = 30$, comparable to the experimental scenario, and depicts the final equilibrium state. To account for the projective imaging used in the experiment, we compute simulated images by rasterizing the configurations in 3D and summing them along a viewing axis as displayed in Fig. 1C.

3 Results

We perform simulations of the swelling hydrogel for swelling ratios ranging from *r*sw ∈ [2,6], inspired by experimental values, and with varying sizes (R_b) and numbers (N_b) of confining beads.

In Fig. 2A, B and C, we display the equilibrium configuration for a hydrogel sphere with 10, 20 and 30 adjacent beads respectively of $R_b = 0.7$. We observe that the resulting configurations strongly resemble the morphologies observed in experiments as shown in Fig. 1A and Ref.⁹. We also display corresponding cross sections of these configurations with the state of strain in Fig. 2D-F as will be discussed later. The final volume V_f of the hydrogel is less than that of the final volume $V_{f,u}$ of the unconstrained hydrogel with the same parameters. We define the percent swelling suppression due to the confinement then as $c = (\Delta V_{\text{f,u}} - \Delta V_{\text{f}})/\Delta V_{\text{f,u}} \times 100$, where ∆*V* = *V* −*V*ⁱ is the volume of the solvent absorbed. We plot this swelling suppression as a function of the number of beads N_b in Fig. 2G for a swelling ratio of $r_{sw} = 3$ and bead radius $R_b = 0.7$. The increase in the swelling suppression with confinement is consistent with the observations in the experiments in Ref.⁹

3.1 Internal strain

Next, we reconstruct the state of strain in the deformed gel as follows. First, we run a corresponding simulation without the bead constraints to obtain the unconfined swollen profiles. By comparing the elements and their vertex positions in the constrained and unconstrained swollen meshes, we can compute the Cauchy-Green strain tensor (CG) for each element as follows. First, we compute a Gram matrix for every element *V^k* in the confined mesh,

$$
G_{ij}^k = \mathbf{s}_i \cdot \mathbf{s}_j, \quad i \in \{1, 2, 3\} \tag{11}
$$

where, $\vec{s}_i = \vec{v}_i^{\,k} - \vec{v}_0^{\,k}$ is the vector connecting the 0^{th} and i^{th} vertex of the element. We also compute the corresponding Gram matrix for the reference unconfined element,

$$
(G_{\text{ref}}^k)_{ij} = \mathbf{s}_i^r \cdot \mathbf{s}_j^r, \quad i \in \{1, 2, 3\}.
$$
 (12)

From these quantities, we compute the Cauchy-Green tensor for the volume element V_k as,

$$
CG_{ij}^k = (G_{\text{ref}}^k)_{il}^{-1} G_{lj}^k, \quad i \in \{1, 2, 3\}.
$$
 (13)

In the present work, both constrained and unconstrained meshes have the same topology. It is however important that if refinement or element exchanges are performed during optimization that they be executed on both meshes to preserve an elementto-element map between the two final states for the CG tensor calculation to be valid.

In Fig. 2H, we plot the Trace of this tensor $(I_1 = Tr(CG))$, averaged over angular variables, as a function of the distance from the center, thus probing the strain due to confinement. We also display similar plots of the determinant of this tensor $(I_3 = det(CG))$, which corresponds to the local volume change, in Fig. 2I. We can reconstruct and visualize these quantities spatially. Example profiles of Tr(CG) sliced across the *x*−*y* plane for the simulations in Fig. 2A, B and C are shown in Fig. 2D, E and F respectively.

From this analysis, we observe that the strain is largely confined to the outer half of the hydrogel sphere, and is increasingly localized to the outer extremities as we increase the number of beads. For a small number of beads, the strain profiles are markedly different from those expected from a uniform spherical confinement, which would result in a constant value of $Tr(CG)$ throughout the hydrogel. Indeed, the localization of strain *a posteriori* justifies a key assumption of the model developed in Ref.⁹: that the sphere can be decomposed into an undeformed core and a highly deformed outer region.

3.2 Contact mechanics

To understand the contact mechanics of the gel-constraint interface, we compute the contact forces between the hydrogel and the beads as follows. We obtain the swelling force at each mesh vertex by taking the gradient of the free energy (Eq. 5) with respect to the vertex positions. The total contact pressure per bead for various swelling ratios and bead radii are shown in Fig. 3. We observe an increase in the pressure as a function of the swelling ratio and a steeper increase with the bead radius as confinement increases. A few examples of this force at the boundary are plotted in Fig. 4A, B and C. As expected, this force is largest at the center of the contacts due to the constraints. To probe the mechanics of the hydrogel-bead contact, we plot the 1D profiles of the contact pressure, averaged over 1D boundary slices of the forces and over all the beads. As shown in Fig. 4D, this profile follows the Hertzian contact mechanics estimate of $p = a\sqrt{1-(d/d_0)^2)}$ close to the contact point. As discussed in Ref.⁹, Hertzian contact mechanics is a good first approximation for this system provided a few requirements, one of them being that the linear strains remain small — $(R_{\rm f,u}-R_{\rm f})/R_{\rm f,u}\approx$ 50%. For the swelling ratios that we have used, our maximum value for $(R_{\rm f,u}-R_{\rm f})/R_{\rm f,u}$ is \approx 20%, and so we are well within the linear elastic regime. Our model also does not include any friction or adhesion between the hydrogel and the beads, which is another crucial requirement of the Hertzian theory. Lastly, we also assume that the gel does not phase separate 31. Our results thus provide further support for the applicability of Hertzian contact mechanics assumed in the model developed in Ref. 9 and confirmed experimentally. We also see in the right panel of Fig. 4D that the extent to which a Hertzian profile is followed decreases when the contacts overlap, echoing the analysis in Ref. ⁹ for stronger confinements, namely that the radii of curvature of the bead and the hydrogel must remain much larger than the radius of the area of contact between them for model to be applicable.

4 Discussion

In this paper, we formulated a general approach to determining the equilibrium configurations and properties of osmotically swollen hydrogels under arbitrary confinement in 3D. Recasting the state of the system as the solution to an energy optimization problem enables us to take advantage of optimization theory and permits convenient enforcement of constraints.

We applied this framework to a specific case of the swelling of a large grain of uncharged, macroscopically homogeneous,

Fig. 2 Hydrogel Swelling in 3D. A, B, C Swollen hydrogels with $N_b = 10$, 20 and 30 surrounding beads respectively of radius $R_b = 0.7$. The swelling ratio is *r*sw = 3. D, E, F Trace of the Cauchy-Green tensor Tr(CG) sliced along the *x*−*y* plane for the simulations in A, B and C respectively. G The % suppression of swelling as a function of *N*b. H Tr(CG), averaged along angular variables, as a function of distance from the center of the hydrogel, plotted for various number of beads. The distance is normalized by its maximum. I The corresponding plots for the determinant of the CG tensor.

Fig. 3 Average contact pressure. The average pressure experienced by a single bead for hydrogels with varying swelling ratios and radii of beads on the left, with the values along the cyan and magenta dashed lines shown on the linear plot on the right, for $N_b = 20$ (A,B) and $N_b = 30$ (C,D). Panel A shows a jump in the pressure, beyond a level of confinement.

Fig. 4 Contact forces for a swollen confined hydrogel. A, B, C Gradient of the hydrogel functional at the boundary of the hydrogel for number of beads *N*_b, swelling ratio r_{sw} and bead radius R_b equal to (20,2.4,0.5), (40,3.0,0.3) and (30,2.4,0.7) respectively. D The average contact force as a function of the normalized arc-length *d*/*R*^b away from the center of the contact (as illustrated in the inset) for the simulations in A, B and C. The dashed line shows a fit to the Hertzian contact pressure profile, $p\!=\!a\sqrt{1-(d/d_0)^2)}.$ We can see that the Hertzian model works well when the contacts are independent, but starts to diverge away from it at larger d/R_b values when the contacts overlap.

non-thermoresponsive hydrogel (i.e., polyacrylamide) as an experimentally-validated prototypical example. Our resulting numerically optimized configurations give fresh insight into the unusual mechanical properties of these gels that are not experimentally accessible. Notably, we observe a transition in the behavior as a function of confinement: for small enough confinements, the strain is localised on the outside of the sphere with the core of the hydrogel remaining relatively strain-free. At higher confinement, the strain becomes more and more uniformly distributed throughout the hydrogel matching the configuration expected for a sphere swelling with a fixed outer boundary. We also examined the contact mechanics of the gel-bead system, finding a transition from Hertzian to non-Hertzian behavior in the contact pressure distribution as a function of swelling.

We formulated and solved the problem using our open source shape optimization environment *Morpho*, which means that we can readily accommodate a number of experimentally relevant extensions to the model. We could easily incorporate other energies such as gravitational potentials, electric fields, surface tension and surface elasticity, for example. By allowing the constraining beads to move with a pinning energy, we could model situations where the confinement pressure is finite.

We note that our results are applicable for hydrogels with uniformly distributed chains, swelling in an ideal solvent with no ionic charge, pH variations or any other contaminants, which could occur in soil. Further, our results sought equilibrium configurations of the hydrogels, and thus do not account for the kinetics of the process. Lastly, we did not consider swelling extents high enough to cause fracture or other changes in the topology of the hydrogel. Studying such effects would allow further insights in the confined swelling process.

While we investigated homogeneous hydrogels, our framework can be easily used to impose a position-dependent swelling ratio, elasticity, etc. which can also be used in applications such as hydrogel bi-layers 32–35 or other functional hydrogels. Further, it can be readily extended to include ionic contributions 20. Our method could also be adapted to study the kinetics of swelling in the quasistatic limit where experiments typically take place by recasting the optimization problem as a gradient-flow problem with an appropriate time-stepping scheme. In addition, careful application of refinement could be used to accommodate topological changes, or strain dependent connectivity energies could be incorporated in order to study fracture of hydrogels^{36,37}.

Author Contributions

CJ, MQG and TJA developed the theoretical model and implemented the code. CJ obtained the simulation data and analyzed the results. J-FL and SSD designed the experiment and obtained the experimental images. All authors contributed to preparing the manuscript.

Conflicts of interest

There are no conflicts to declare.

Appendix

Osmotic Pressure

Conventional presentations of hydrogel swelling rely on a pressure-balance approach. In this appendix, we show that our optimization formalism reduces to the regular theory. To do so, begin by noting that the mixing contribution to the free energy, ∆*F*mix, is given by the Flory-Huggins theory. The osmotic pressure contribution from this energy is

$$
\Pi_{\text{mix}} = -\frac{1}{v_s} \frac{\partial \Delta F_{\text{mix}}}{\partial N_s} \tag{14}
$$

Because the volume fraction ϕ depends on N_s ,

$$
\phi = \frac{xN_p}{(xN_p + N_s)},\tag{15}
$$

1

we may re-express derivatives with respect to N_s using the chain rule,

$$
\frac{\partial}{\partial N_{\rm s}} = \frac{\partial \phi}{\partial N_{\rm s}} \frac{\partial}{\partial \phi}
$$

$$
= -\frac{xN_{\rm p}}{(xN_{\rm p} + N_{\rm s})^2} \frac{\partial}{\partial \phi}
$$

$$
= -\frac{1}{xN_{\rm p}} \phi^2 \frac{\partial}{\partial \phi}.
$$

Hence,

$$
\frac{1}{k_{\text{B}}T} \frac{\partial \Delta F_{\text{mix}}}{\partial N_{\text{s}}} = \frac{\partial}{\partial N_{\text{s}}} \{ N_{\text{s}} \left[\ln(1 - \phi) + \chi \phi \right] \}
$$

$$
= \left[\ln(1 - \phi) + \chi \phi \right]
$$

$$
+ N_{\text{s}} \left(-\frac{1}{xN_{\text{p}}} \phi^2 \right) \frac{\partial}{\partial \phi} \left[\ln(1 - \phi) + \chi \phi \right]
$$

$$
= \left[\ln(1 - \phi) + \chi \phi \right] - \left(\frac{N_{\text{s}}}{xN_{\text{p}}} \phi^2 \right) \left[\frac{-1}{(1 - \phi)} + \chi \phi \right]
$$

We rearrange Eq. (15),

$$
\frac{N_{\rm s}}{xN_{\rm p}}=\frac{1}{\phi}-1=\frac{(1-\phi)}{\phi},
$$

and use this to eliminate N_s/xN_p from the osmotic pressure,

$$
\frac{1}{k_{\text{B}}T} \frac{\partial \Delta F_{\text{mix}}}{\partial N_{\text{s}}} = \left[\ln(1 - \phi) + \chi \phi \right] - \left(\frac{N_{\text{s}}}{xN_{\text{p}}} \phi^2 \right) \left[\frac{-1}{(1 - \phi)} + \chi \right]
$$

$$
= \left[\ln(1 - \phi) + \chi \phi \right] - \left(\phi(1 - \phi) \right) \left[\frac{-1}{(1 - \phi)} + \chi \right]
$$

$$
= \left[\ln(1 - \phi) + \chi \phi \right] + \phi - \chi \phi(1 - \phi)
$$

$$
= \phi + \ln(1 - \phi) + \chi \phi^2.
$$

We hence recover the standard result, expressed for example as Eq. (8) from Ref. 20:

$$
\Pi_{\text{mix}} = -\frac{1}{v_s} \frac{\partial \Delta F_{\text{mix}}}{\partial N_s} = -\frac{k_B T}{v_s} \left[\phi + \ln(1 - \phi) + \chi \phi^2 \right] \tag{16}
$$

Note that in the literature, this osmotic pressure is sometimes expressed in terms of an 'effective diameter' of the solvent molecule⁹:

$$
\Pi_{\text{mix}} = -\frac{k_{\text{B}}T}{d^3} \left[\phi + \ln(1-\phi) + \chi \phi^2 \right],\tag{17}
$$

which can be readily understood, since it implies $d^3 = v_{\rm s}$.

Parameter	Symbol	Value
Flory-Huggins parameter		0.499
Relative elastic strength	$N_{\rm c}\alpha^3/V_0$	
Reference volume fraction	Φο	0.036
Confining bead radius	$R_{\rm h}$	[0.5, 1]
Swelling ratio	$r_{\rm sw}$	[2,6]
Number of beads		[5,10,40]

Table 1 Parameters used for the hydrogel swelling simulations.

Simulation details

To compute the structure of the hydrogel in *Morpho*, we start by constructing an initially spherical **Mesh** corresponding to the unit ball |x| ² < 1 with Morpho's *meshgen* module. An **OptimizationProblem** object is then defined and a **Hydrogel** functional, implementing the above discussed free energy density, is added to it. For hard confinements, we define level-set constraints corresponding to the objects (spheres, ellipsoids, planes, etc.) through the **ScalarPotential** object from the *functionals* module. A **ShapeOptimizer** object is then created to optimize the shape. We perform gradient descent with a fixed step size. A **Volume** object is used to keep track of the volume of the hydrogel during relaxation.

To initialize the positions of the hard spheres, we define a dummy shell mesh with radius $R + R_b$ with N_b number of vertices placed randomly. We first confine the vertices to lie on the shell by using a **ScalarPotential** object. We then define an electrostatic repulsive pairwise interaction between the vertices using a **PairwisePotential** object from the *functionals* module, thus proceeding to solve the Thomson problem. The resulting mesh vertex positions are used as the sphere centers for the level set constraints. We thus get equidistantly packed spheres on the outer shell.

All 3D visualizations are made using the *povray* module. The slices of the Cauchy Green strain tensor's trace are generated using the *meshslice* module.

The parameters used in the simulations are listed in Table 1.

Supplementary codes

Here, we briefly describe the codes provided in as Supplementary Information.:

sphere.mesh**:** This is the 3D sphere mesh used in all the simulations in this paper, in the Morpho .mesh format.

sphere.vtk**:** The same mesh as above in the .vtk format for convenience.

ConfinedHydrogel.morpho**:** This script defines the **Confined-Hydrogel** Class to set up the simulation discussed above.

main.morpho**:** This script uses the ConfinedHydrogel.morpho file to simulate the system for a given set of parameters. We equilibrate the confined hydrogel first, followed by the corresponding unconfined equilibration.

computeCauchyGreen.morpho**:** This script uses the results from the main.morpho file to compare the confined and unconfined hydrogel meshes to compute the Cauchy-Green strain tensor as discussed in the main text.

computeContactPressure.morpho**:** This script uses the results from the main.morpho file to compute the average contact pressure between the hydrogel and the beads.

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