



Lab on a Chip

A bioinspired, passive microfluidic lobe filtration system

Journal:	<i>Lab on a Chip</i>
Manuscript ID	LC-ART-05-2021-000449.R1
Article Type:	Paper
Date Submitted by the Author:	21-Jul-2021
Complete List of Authors:	Clark, Andrew; North Carolina State University, Chemical & Biomolecular Engineering San-Miguel, Adriana; North Carolina State University, Chemical & Biomolecular Engineering

SCHOLARONE™
Manuscripts

A bioinspired, passive microfluidic lobe filtration system

Andrew S. Clark¹, Adriana San-Miguel^{1,*}

Abstract:

Microparticle filtration plays an important role in many medical and biological applications. Size-based microfluidic filtration systems can be affected by clogging, which prevents their use in high-throughput and continuous applications. To address these concerns, we have developed two microfluidic lobe filters bioinspired by the filtration mechanism of two species of Manta Ray. These chips enable filtration of particles around 10 - 30 μm with precise control and high throughput by using two arrays of equally spaced filter lobes. For each filter design, we investigated multiple inlet flow rates and particle sizes to identify successful operational parameters. Filtration efficiency increases with fluid flow rate, suggesting that particle inertial effects play a key role in lobe filter separation. Microparticle filtration efficiencies up to 99% were obtainable with inlet flow rates of 20 mL/min. Each filter design successfully increased microparticle concentrations by a factor of two or greater at different inlet flow rates ranging from 6-16 mL/min. At higher inlet flow rates, ANSYS Fluent simulations of each device revealed a complex velocity profile that contains three local maxima and two inflection points. Ultimately, we show that distances from the lobe array to the closest local maxima and inflection point of the velocity profile can be used to successfully estimate lobe filtration efficiency at each operational flow rate.

¹Department of Chemical & Biomolecular Engineering, North Carolina State University, Raleigh, NC.

* correspondence to: asanmig@ncsu.edu

Electronic supplementary information (ESI) available.

26 1. Introduction

27 Size-based microparticle filtration is used in applications with widely different scales.
28 Research and clinical microparticle manipulation applications often separate biological samples
29 with volumes of approximately 1-1000 μL in size¹⁻³, while industrial applications often deal with
30 filtering with volumes greater than 1 L^{4,5}. Currently, size-based microparticle filters are made of a
31 mesh sieve, which intercept particles larger than the pore size. Due to the inherent nature of these
32 filters, they commonly clog and require an operator to either change or clean the filter⁶, which
33 ultimately decreases microparticle separation throughput.

34 Microfluidic devices offer promising advantages for microparticle filtration as they enable
35 precise manipulation of fluids, and therefore microparticle suspensions, within channels with
36 dimensions around 1-1000 μm ^{7,8}. Microfluidic filters are commonly split into two groups: active
37 and passive filtration. Active microfluidic filters connect the microfluidic device to external
38 equipment, which then relies on external force fields, such as acoustics^{9,10} or magnetics¹¹ to
39 manipulate particles. These technologies usually require particle pre-treatment, as well as complex
40 and expensive external hardware, making them less attractive for high-throughput applications.

41 Conversely, passive microfluidic filters do not rely on active external fields and are often
42 praised for their simplicity. These filters utilize different methods, such as deterministic lateral
43 displacement¹², cross-flow filtration^{13,14}, and membrane filtration¹⁵. These methods have all been
44 shown to perform microparticle filtration with adequate efficiency; however, each is limited by
45 throughput. For instance, deterministic lateral displacement must be operated at precise and slow
46 flow rates ($\sim 10 \mu\text{L}/\text{min}$) to reach efficient separation¹², while membrane filters and crossflow
47 filtration are plagued by the possibility of clogging since particles are larger than the filter pore
48 size^{15,16}.

49 Another option for microparticle filtration within microfluidic devices is inertial particle
50 separation. Unlike many microfluidic devices that operate at very low Reynolds Numbers ($Re =$
51 $\rho UH/\mu$; where ρ is fluid density, U is average flow velocity, H is hydraulic diameter, and μ is fluid
52 viscosity; $Re \rightarrow 0$), inertial microfluidics considers the nonlinear effects that fluid inertia has on
53 microfluidic systems that operate under intermediate laminar flow¹⁷⁻¹⁹. Inertial particle separation
54 relies on a balancing act of two main forces, the shear-induced lift and the wall-induced lift, to
55 precisely manipulate microparticles based on size⁸. The resulting net, inertial lift force is dependent
56 on Re and particle position within the channel, as well as directly proportional to the product of

57 shear rate and shear gradient^{20–23}. Thus, if the signs of shear rate and shear gradient are different,
58 the resulting inertial lift force could change direction^{20,21}. Since the inertial lift force is highly
59 dependent on fluid flow velocity (Re), inertial microfluidic filters are often limited by finding a
60 Goldie-locks flow rate (not too fast or slow)^{19,23–28}.

61 In a straight channel, particle equilibrium positions are determined by cross-section geometry,
62 particle size, and flow rate^{21,29,30}. Channel geometry and an introduction of a secondary flow can
63 thus significantly alter particle equilibrium position. Secondary flow, which is a minor flow
64 perpendicular to the primary flow, helps reduce the number of equilibrium positions by applying
65 an additional drag force to help control particle location¹⁸. Most state-of-the-art inertial particle
66 separation technologies utilize secondary flows to increase particle filtration efficiency. These
67 devices are commonly separated into groups based on their strategy of controlling secondary flow
68 (expansion-contraction arrays, spiral, or sinusoidal devices)^{15,21,24,26,30–35}. In all cases, smaller
69 particles experience greater effects from the secondary flow. Using these principles, inertial
70 microfluidics applications include continuous blood cell separation^{32,36}, bacteria filtration from
71 other particles^{34,37}, circulating tumor cell separation and filtration^{24,38–40}, and rare cell trapping^{41,42}.

72 Recently, biomimicry, or the emulation of elements of nature to solve complex problems, has
73 significantly advanced multiple technologies. Interestingly, Divi et al. recently explored the Manta
74 Ray, specifically *Manta birostris*' and *Mobula tarapacana*'s, filter feeding mechanism: lobe
75 filtration. These animals use an array of nonstick filter lobes to capture zooplankton while
76 swimming⁴³. The main difference between the two species includes a slight difference in lobe
77 design, which permits the *M. tarapacana* to feed at nearly seven times smaller pressure head⁴³.
78 Unlike most filter feeding marine life, these animals continuously feed on particles smaller than
79 their filter's pore size by using precisely spaced filter lobes⁴³. These lobes, which are separated by
80 $\sim 340 \mu\text{m}$ ⁴⁴, cause fluid to quickly change directions, creating a secondary flow. At adequate bulk
81 flow rates, larger particles diverge from fluid streamlines and continue their inertial path, resulting
82 in a non-clogging filtering mechanism with attributes similar to inertial particle separation, which
83 can be better visualized in **Fig. 1A**. Interestingly, both *M. birostris* and *M. tarapacana* can capture
84 zooplankton using this mechanism, where efficiency increases with particle size and bulk fluid
85 Re .^{44,45} Moreover, Divi et al. noted that increased swimming speeds with $Re > 1000$, do not affect
86 filtration efficiency⁴³. Nevertheless, obtaining $Re \sim 1000$ in a microfluidic device is often difficult
87 due to the proportional relationship between channel dimensions and Re . Thus, scaling down lobe

88 filtration to a microfluidic device capable of filtering smaller particles ($\sim 10\ \mu\text{m}$) with high
89 efficiencies may be difficult.

90 In this work, we sought to demonstrate that lobe filtration, bioinspired by both *M. birostris* and
91 *M. tarapacana*, can be scaled down to a microfluidic device to create a high throughput
92 microparticle filter capable of filtering particles on the order of $10\ \mu\text{m}$ with processing speeds up
93 to $20\ \text{mL}/\text{min}$ in a single device. We designed and characterized two filter designs based on the
94 lobe structures of *M. birostris* and *M. tarapacana* (named Oblong lobe and Bent lobe, respectively)
95 by running $25\ \mu\text{m}$ and $15\ \mu\text{m}$ particles through the devices at varying flow rates, showing passive
96 lobe filtration's potential for wide-ranged applications. We further explored the effect that particle
97 size has on lobe filtration efficiency by processing particle suspensions at various inlet flow rates
98 for both designs. Moreover, by utilizing ANSYS Fluent simulations, we revealed an unexpected,
99 complex velocity profile for microfluidic flow, which contains multiple velocity local maxima and
100 inflection points. In the region between the velocity local maxima and the inflection point, the
101 inertial lift force changes direction. We obtained the distances between the lobes and location in
102 the main channel of the velocity local maxima and the inflection point. Comparing these distances
103 to various particle sizes with experimentally obtained efficiencies revealed a simple and robust
104 explanation for microfluidic lobe filtration success.

105

106 **2. Materials and Methods**

107 **a. Design and fabrication of microfluidic devices**

108 Both microfluidic filter devices used in this study were designed and fabricated through
109 standard photo and soft-lithography techniques. Designs were drawn in AutoCAD 2018 drafting
110 software (Autodesk). Transparency films from the designs were printed by FineLine Imaging. SU-
111 8 2025 (Kayaku Advanced Materials, Inc.) negative photoresist was spun at $1500\ \text{rpm}$ to obtain a
112 $60\ \mu\text{m}$ layer on a 4-inch silicon wafer. We noticed that low and gradual bake times significantly
113 improved master mold resolution. Therefore, the wafer was then soft baked by gradually increasing
114 a room temperature hot plate to $65\ ^\circ\text{C}$, holding for 10 minutes, then increasing the hotplate to 95
115 $^\circ\text{C}$ and holding for 30 minutes. The wafer was then allowed to cool to room temperature on the hot
116 plate. Following the soft bake, the wafer was exposed to UV light masked by the photomask for 6
117 seconds in a Kloe UV-KUB 3 mask aligner. The wafer was then baked with the same procedure
118 as the previous soft bake to ensure complete cross-linking of exposed areas. The wafer was then

119 shaken in SU-8 developer for 20 minutes to remove unexposed SU-8. The device was hard baked
120 at 200 °C for 2 minutes then placed in a vacuum chamber with a few drops of trichloro-
121 perfluorooctyl-silane overnight to avoid irreversible adhesion of PDMS to SU-8 photoresist. A 9:1
122 ratio of polymer to crosslinker of polydimethylsiloxane (PDMS) was used for soft lithography.
123 The polymer/crosslinker mixture was mixed and degassed to remove bubbles prior to pouring on
124 the microfluidic mold. The PDMS was then cured at 80 °C for 2 hours prior to peeling. Individual
125 filters were then cut and punched with a sharpened 0.44 mm dispensing needle (McMaster-Carr).
126 Devices were bonded to 22x50 mm glass slides in an O₂ plasma chamber. Finally, tubing was
127 attached to each inlet and outlet on the devices.

128

129 **b. Preparation of particles**

130 Particle suspensions were made using various concentrations and particle sizes. Device
131 characterization experiments used 25 µm red fluorescent particles (Fisher Scientific, Inc.) (Ex: 542
132 nm, Em: 612 nm) and 15 µm green fluorescent particles (Fisher Scientific, Inc.) (Ex: 468 nm, Em:
133 508 nm), which were diluted using 0.1% w/v Triton TX100-water solution to ~10⁶ particles/mL
134 and ~5x10⁶ particles/mL, respectively. Low concentration experiments were conducted with ~10⁴
135 particles/mL of 25 and 15 µm particles, while high concentration experiments used 10⁷
136 particles/mL. Particle range experiments using green fluorescent particles (Cospheric LLC) (Ex:
137 468 nm, Em: 508 nm) 10-29 µm in size were diluted to ~10⁶ particles/mL with a 0.1% w/v Triton
138 X-100 water solution. Particle suspensions were mixed with a vortex mixer for 1 minute prior to
139 use within filter devices.

140

141 **c. Experimental set up**

142 Each microfluidic lobe filter was tested by flowing fluorescent particles through the device
143 and analyzing steady state operation, as well as particle concentrations in both outlets. Steady state
144 operation was achieved when there was no discernable change in particle tracks under operator
145 observation. Particle suspensions were inserted into the device through a syringe pump (Harvard
146 Apparatus) and a 10 mL syringe (BD Plastic). Inlet flow rates depended on experiment type and
147 filter design. Most experiments with the Oblong lobe device used flow rates of 1, 2, 4, 6, 8, 10, 12,
148 and 16 mL/min. Most experiments with the Bent lobe device used inlet flow rates of 1, 2, 4, 6, 8,
149 12, 16, and 20 mL/min. Inlet samples were taken before each experiment and outlets were collected

150 for later analysis of particle concentrations. Fluorescent images of steady state operation were
151 taken using Infinity Capture and a Lumenera Infinity3 color CCD camera on a Leica M165 FC
152 microscope using a dual band pass filter in fluorescence mode with a metal halide light source.

153

154 **d. Image processing and characterization**

155 Filtration experiments using 25 μm and 15 μm particles were characterized by obtaining
156 particle counts from the inlet and both outlets for each experimental parameter. Images of 1 μL
157 samples pressed between two glass slides were taken on a Leica M165 FC with Infinity Capture
158 software and Lumenera Infinity3 color CCD camera at 7.3 X magnification. A custom-written
159 MATLAB image processing code enabled particle counting to obtain concentrations at the inlet
160 and outlets. This code separated images into red and green channels to analyze 25 μm and 15 μm
161 particle counts separately. These images were then binarized using the “imbinarize” function in
162 MATLAB. The resulting binary object sizes were obtained using “regionprops” function. If a
163 binary object’s area (in pixels) was within the corresponding range for the current particle size
164 analysis, it was counted toward the particle count. Three images were processed for each
165 experimental condition.

166 Similarly, particle size range experiments utilized images of samples taken at the inlet and both
167 outlets. Samples were prepared by placing 1 μL droplets on a glass slide and imaging the droplets
168 with an inverted Leica DMI8 widefield fluorescence microscope equipped with a Lumencor
169 Spectra X fluorescent LED light source and Hamamatsu Orca-Flash4.0 camera at 10 X
170 magnification. Three images were taken for each inlet and outlet for each experiment. Images were
171 then processed using a custom written MATLAB image processing algorithm to find circles and
172 measure the radius. Particle counts were placed in 5 μm bins ranging from 10-30 μm . Each bin
173 size was then analyzed separately for efficiency.

174 Each filter design was characterized for particle filtration efficiency and particle concentration
175 ratio as others have done^{43,46,47} using particle counts obtained from the image processing
176 algorithms. In each case, efficiency was calculated as:

$$177 \quad \text{Efficiency} = \left(1 - \frac{(\text{Out 2 Concentration})}{(\text{Inlet Concentration})} \right) \times 100\% \#(1)$$

178 Where Out 2 refers to the peripheral device outlet, intended for the filtrate.

179 Concentration ratio was calculated as:

$$180 \quad \text{Concentration Ratio} = \frac{\text{Out 1 Concentration}}{\text{Inlet Concentration}} \#(2)$$

181 Where Out 1 refers to the center device outlet, intended for particle collection.

182

183 e. Velocity profile simulations and analysis

184 Velocity profiles within each filter design were obtained using ANSYS Fluent 19.1
185 computational fluid dynamics simulation software. Three-dimensional microfluidic filter designs
186 were exported from AutoCAD as ACIS files and subsequently loaded into the ANSYS Fluent
187 graphical user interface (GUI). Each design was split into five parts to permit finer meshing (7 μm
188 element size; cartesian meshing method in ANSYS Meshing) in the center channel (**Supplemental**
189 **S1.A and S1.B**). Steady state simulations (with 500 iterations) were conducted using standard
190 water properties for the fluid, as well as a laminar flow model with the SIMPLE solver. Velocity
191 inlet boundary conditions were used (1 mL/min to 20 mL/min) with zero-gauge pressure outlets.
192 A residual convergence criterion of 10^{-5} was used. A mesh independence study was conducted
193 using outlet mass flux to ensure mesh performance (**Supplemental S1.C**). Following simulations,
194 velocity values from the $z = 30 \mu\text{m}$ plane were exported as ASCII files and subsequently imported
195 into MATLAB. The imported velocity values (x, y, z, and magnitude) were then segregated by
196 their x-coordinate corresponding to each filter lobe in the design to obtain velocity profiles along
197 the device. Smooth profiles were obtained by interpolating between points within the main channel
198 using the “interp1” function in MATLAB. Velocity profiles were then analyzed using a custom
199 MATLAB algorithm.

200 All custom code is available at GitHub (<https://github.com/asanmiguel/MantarayFilter>).

201

202 3. Results and discussion

203 a. Device designs

204 The lobe structures of *M. birostris* and *M. tarapacana* inspired the designs of the Oblong
205 lobe and Bent lobe microfluidic devices, respectively. Using the dimensions listed by Divi et al.
206 as a basis⁴³, the lobe dimensions were scaled down by approximately 6 times to aim for a target
207 particle filtration size of 10-30 μm . The target filtration size was chosen for its multiple real-world
208 applications, such as cell separation^{48,49}, cell aggregation filtration^{38,40}, and microplastic
209 removal^{50,51}. Each design had similar features including one inlet that throttles to a center channel

210 with an array of equally spaced lobes on each side, and two outlets (**Fig. 1B, 1C**). Since lobe
211 filtration had not yet been conducted in a microfluidic device, there were many potential
212 parameters that could influence filtration success including lobe design, lobe angle, lobe width,
213 lobe separation, center channel dimension, among others. Thus, both the Oblong and Bent lobe
214 design dimensions were obtained by scaling down previously reported measurements of *M.*
215 *birostris* and *M. tarapacana*⁴³. The Oblong lobe device included lobes of 480 μm in length and 80
216 μm in width, separated by 50 μm , with a 30-degree orientation. Each array of lobes contained 31
217 individual lobes to provide ample opportunities for microparticle filtration (**Fig. 1B**). Since both
218 *M. birostris* and *M. tarapacana* feed successfully at moderate Reynold's number flow ($Re \sim 1000$),
219 we hypothesized that a similar Re would be necessary for microfluidic filtration success. Hence,
220 center channel dimensions were designed to permit high flow rates ($\sim 200 \mu\text{m}$ in width by 60 μm
221 in height).

222 The Bent lobe design has similar dimensions with the key design change being the shape
223 of the lobe. The lobe design, seen in **Fig. 1C**, features a bend approximately one third from the top
224 of the lobe, causing the angle the lobe to be closer to the horizontal of the center channel and the
225 minimum distance between lobes to be slightly closer ($\sim 4 \mu\text{m}$). Like the Oblong lobe design, the
226 other dimensions selected were intended to obtain moderate Re flow in the center channel.

227

228 **b. Lobe filtration operation**

229 Both filter lobe designs were tested for their ability to filter and/or concentrate large
230 particles of 25 μm and 15 μm at several inlet flow rates. Mixed particle suspensions were pumped
231 at different inlet flow rates through each filter design using a syringe pump to test both particle
232 sizes concurrently, removing the need for extra experiments. Since the 25 μm and 15 μm particles
233 were fluorescently labeled in different colors (red and green, respectively), size-based particle
234 tracks were visualized within the device using a fluorescent dissecting scope. At slow inlet flow
235 rates with $Re \sim 130$ (**Fig. 2A**), both 25 μm and 15 μm particles leave from the center channel
236 through the first few lobe pores and into the outer channel. We observed that particles appear to
237 return from the outer channels into the main channel and exit into Out 1 for both particle sizes. We
238 hypothesize that the particles that return into the main channel are simply following fluid path
239 lines, since it appears that only some particles closest to the lobes return to the main channel. This
240 phenomenon of particles returning into the main channel was observed in both device designs.

241 To test the filter at higher Re within each device, we gradually increased the inlet flow rates
242 until the syringe pump did not have enough power to flow fluid at the desired rate. These inlet
243 flow rates (16 mL/min for the Oblong lobe and 20 mL/min for the Bent lobe) were then determined
244 to be the maximum inlet flow rate for each device. It is important to note that neither filter broke
245 from too much pressure, suggesting that higher inlet flow rates could be achieved with a stronger
246 syringe pump. At higher inlet flow rates, the steady state particle tracks significantly changes. The
247 particle tracks at the maximum inlet flow rate for the Oblong lobe and Bent lobe devices can be
248 observed in **Fig. 2B** and **Fig. 2C**, respectively. In both cases, 25 μm particles (red channel) appear
249 to be enter evenly dispersed throughout the channel. As the particles travel along the devices, they
250 eventually stabilize near the edge of the main channel (by the lobe arrays) without exiting through
251 the lobe pores. By contrast, a portion of the 15 μm particles appear to exit through each of the filter
252 lobes. Once 15 μm particles exit through the filter pores, a majority stay in the outer channel and
253 exit through Out 2. Although, like particles in slow flow operation, a small portion of 15 μm
254 particles appear to return to the main channel at the last filter pore. We also ensured particle track
255 changes were a result of the lobes and not of solely inertial forces, as we tested the same channel
256 design with no lobes (**Supplemental S2**). As expected, no particle filtration was observed in the
257 design with no lobes. Notably, steady state operational images of both lobe filter designs
258 demonstrated successful filtration of 25 μm particles and partial filtration of 15 μm particles.

259

260 c. Lobe filtration characterization

261 To quantify filtration efficiency and concentration capability, samples of the inlet and both
262 outlets were collected and imaged for each experiment. Using these images, particle concentrations
263 could be obtained and filtration efficiencies for both particle sizes and lobe designs could be
264 calculated. Filtration efficiencies were grouped into three main categories for quick visualization
265 of filter performance. The three categories include low (0-60%), moderate (60-90%), and high
266 filtration (>90%), which are represented in **Fig. 3A** by the red, yellow, and green backgrounds,
267 respectively. A filtration efficiency of 0% indicates no change in particle concentration between
268 the inlet and Out 2 suspensions. The grey background and negative efficiencies in **Fig. 3A**
269 represent a higher concentration of particles in the filtrate (Out 2) compared to the inlet. At
270 common inertial particle flow rates (~ 1 mL/min), both filters perform poorly with low efficiencies
271 under 40%. Interestingly, both device designs have sharp increases in efficiency at a 4 mL/min

272 inlet flow rate, indicating a change in forces experienced by particles within each device. At inlet
273 flow rates higher than 4 mL/min, the Bent lobe device obtains much higher filtration efficiencies
274 for 25 μm particles compared to the Oblong lobe design. In this range (4 mL/min to 20 mL/min),
275 the Bent lobe device offers high filtration efficiencies with a maximum near 99%. Remarkably,
276 this design can successfully process up to 20 mL/min of a 25 μm particle suspension, which
277 correlates to a clean filtrate (Out 2) flow rate of approximately 10 mL/min. On the other hand, the
278 Oblong lobe design operates with moderate 25 μm filtration efficiencies over these flow rates (up
279 to 16 mL/min) with maximum filtration efficiency of 88%. Moreover, the Oblong lobe design
280 appears to experience a slight decrease filtration efficiency with inlet flow rates over 10 mL/min,
281 which is not observed with the Bent lobe design. Ultimately, the Oblong lobe design obtained
282 clean filtrate flow rates from approximately 3-8 mL/min leaving Out 2, as compared to clean
283 filtrate flows of 2-10 mL/min for the Bent lobe design.

284 As expected, both lobe filters designs performed worse with 15 μm particles. The Oblong lobe
285 design operated with low efficiencies throughout all inlet flow rates with a maximum efficiency
286 near 41%. In fact, the Oblong lobe design appears to slightly increase 15 μm particle concentration
287 in the filtrate outlet when operated at 1 mL/min. However, the Bent lobe design offered moderate
288 filtration efficiencies of 75% for 15 μm particles at flow rates over 6 mL/min, which provides
289 evidence that lobe filtration does not have a binary particle cutoff size for successful filtration.

290 Continuous microparticle filters are also commonly used to concentrate sample particles of
291 interest. Therefore, each lobe design was tested for its ability to concentrate particles within this
292 size range. **Fig. 3B** shows the concentration ratio (CR) of 25 μm particles for each device at various
293 flow rates (see experimental for calculation equation). Almost every flow rate tested successfully
294 concentrated particles. As inlet flow rates and particle filtration efficiencies increased,
295 concentration ratios increased until an eventual plateau of 2.05 at 12 mL/min for the Oblong lobe
296 design. Although filtration efficiencies steadied around 8 mL/min, proportionally more fluid exits
297 through Out 2 with increasing inlet flow rates (**Supplemental S3**), which ultimately increases
298 particle CR. The Bent lobe design offers similar CR at comparatively higher flow rates. We
299 hypothesize that a higher inlet flow rate is needed to obtain similar CR values with the Bent lobe
300 design, since it operates with proportionally more fluid exiting through Out 1 when holding the
301 inlet flow rates constant (**Supplemental S3**). At higher flow rates, the Bent lobe design achieves

302 >98% filtration efficiencies, permitting increased concentration ratios. In either case, microfluidic
303 lobe filtration may also be used to concentrate particles at high processing flow rates.

304 Since the Bent lobe design significantly improved particle filtration performance, we
305 investigated changing other lobe design parameters (**Supplemental S4**). We changed lobe spacing
306 to 30 μm , lobe length to 600 μm , or lobe width to 150 μm , and measured particle filtration
307 performance at an 8 mL/min inlet flow rate (**Supplemental S5**). Decreasing the lobe spacing to 30
308 μm was the only design change that offered improved filtration performance with an efficiency
309 comparable to the Bent lobe design. However, since the filter pores in this design are much closer
310 in size to the tested microparticles, the 30 μm spacing may be more prone to clogging and act
311 similarly to a crossflow filter. Therefore, compared to the original Oblong lobe design, the Bent
312 lobe design is the preferred modification for improved particle filtration performance.

313 Device performance across varying particle concentrations is important for potential filtration
314 applications. Therefore, we tested both the Oblong and Bent lobe designs at low (10^4 particles/mL)
315 and high (10^7 particles/mL) concentrations using previously determined successful operational
316 flow rates. In both cases, particle concentration has no effect on successful particle filtration at 6
317 and 16 mL/min for the Oblong lobe design and 6 and 20 mL/min for the Bent lobe design
318 (**Supplemental S6**). Thus, lobe filtration may be applied to applications with wide-ranging particle
319 concentrations.

320 Both the Oblong lobe filter and Bent lobe filter designs are successful at filtering and/or
321 concentrating 25 μm particles. The Bent lobe design offers slightly higher filtrate purity, while the
322 Oblong lobe design offers increased 15 μm particle filtrate recovery rates (**Supplemental S7**).
323 Moreover, the Bent lobe device excels at filtration with highly efficient operation from 4 mL/min
324 up to 20 mL/min. Given typical sizes of single cells obtained from tissue dissociation are around
325 15 μm , the high filtrate purity for this particle size (~99%) makes upstream processing for single
326 cell analysis a promising application of this device, such as MCF-7 cell aggregate filtration³⁸.
327 However, the Oblong lobe design offers increased 15 μm particle filtrate recovery rates and similar
328 concentration ratios at slower inlet flow rates, which may be useful for sensitive applications that
329 require operation with minimal shear forces.

330

331 **d. Particle size significantly effects lobe filtration efficiency**

332 To test how particle size affects lobe filter operation, we flowed various particle sizes (10-
333 29 μm particles) at different flow rates through each device and compared outlet concentrations
334 for each size. For each design, particle range suspensions were injected into the device at the
335 following flow rates: 2, 4, 6, 10, 14, and 18 (Bent lobe only) mL/min. Samples of the inlets and
336 both outlets were then imaged as detailed in the experimental section. A custom-written image
337 processing algorithm was then used to detect microparticles of various sizes (**Supplemental S8**).
338 In short, the algorithm binarized the fluorescent images and detected circles with radii within a
339 predetermined size range. Detected particles were then binned based on diameter into 5 μm bins
340 and counted for efficiency analysis.

341 The efficiency curves based on 5 μm particle size bins for the Oblong lobe can be
342 visualized in **Fig. 4A**. As expected, filtration efficiency increases with increasing particle size.
343 However, there is no apparent difference in efficiency between the 10-15 μm and 15-20 μm bins,
344 indicating that particle size may only affect filtration efficiency beyond a certain threshold size.
345 Moreover, holding particle size constant, filtration efficiency increases with increasing inlet flow
346 rates, which matches previous experimental observations (**Fig. 3A**). The low efficiency (0-60%),
347 moderate efficiency (60-90%), and high efficiency (>90%) regions are indicated by the red,
348 yellow, and green backgrounds in **Fig. 4**, respectively. For the Oblong lobe design, particles in the
349 low efficiency particle size range (10-20 μm) experience only slight increases in filtration
350 efficiencies with increasing inlet flow rates. We hypothesize that some particles in this size range
351 may never have an opportunity to leave through the filter pores due to small transverse velocities
352 compared to the bulk flow direction, and thus experience increased filtration efficiency with
353 increasing flow. Particles in the 20-25 μm size range can achieve moderate efficiencies, which
354 suggests that particles of this size are large enough to experience different hydrodynamic lift forces
355 within the filter. The increase in filtration efficiency of 25-30 μm particles to >90% provides more
356 evidence for this hypothesis. Interestingly, efficiencies near 100% were not reached with the tested
357 particle size range in the Oblong design.

358 Binned particle filtration efficiencies for the Bent lobe design can be seen in **Fig. 4B**. Like
359 the Oblong lobe design, particle filtration efficiency in the Bent lobe design increases with
360 increasing particle size and increasing flow rates. However, no particle sizes tested resulted in low
361 efficiencies, which suggests that even the smallest particles (10-15 μm) experience some
362 hydrodynamic lift forces keeping them in the main channel in this filter design. Moreover, 15-20

363 μm particles achieve mostly moderate efficiencies with a maximum of 93% at 10 mL/min, while
364 particles larger than 20 μm reach efficiencies near 99%, indicating these particles experience
365 strong lift forces keeping them from exiting through the filter lobe pores.

366

367 **e. Velocity field simulations reveal velocity profile with inflection points**

368 The multiple forces particles experience in microchannels can be estimated and explained
369 by various aspects of the velocity field, such as the boundary layer location⁵² and the saddle
370 point^{46–48} within the device. Since inertial lift coefficient, and thus the forces acting on the particles,
371 is proportional to the product of the shear rate and the shear gradient^{20,22}, estimating the velocity
372 profile within the device seemed a necessary first step to understand microfluidic lobe filtration.
373 We opted to obtain the velocity profile at the experimental inlet flow rates from computational
374 fluid dynamics simulations run in ANSYS Fluent 19.1 for both the Oblong and Bent lobe designs.
375 The mesh for each design was obtained by first splitting the design into five parts (inlet body, out
376 1 body, out 2 body, outer channel body, and main channel body) to obtain different element sizes
377 for each region (**Supplemental S1.A**). Since it was hypothesized that the main channel body would
378 have the most complex velocity profile, a 7 μm element size was used to obtain more data points
379 within this region. Moreover, a cartesian sweeping method was utilized within the main channel
380 body mesh to facilitate velocity field analysis at individual lobes by creating evenly spaced nodes
381 with a cartesian grid pattern. Default element sizes were used for the four remaining bodies for
382 ease of calculation. After ensuring mesh quality, a parametric study using various inlet flow rates
383 was conducted for each design. Simulation parameters can be found in the experimental section.

384 For each inlet flow rate, outlet flow rates were monitored to match experimental
385 observation. Prior to conducting a full parametric three-dimensional study on each device, two-
386 dimensional (2D) simulations were conducted to accelerate calculation speed. To determine if the
387 simulations roughly matched our experimental data, we first assessed the flow leaving through the
388 device as Out 1 proportional flow (Out 1 flow rate / Inlet flow rate). 2D simulations predicted
389 increased Out 1 proportional flow with increasing inlet flow rates, while experimental results
390 revealed a decreasing Out 1 proportional flow with increasing flow rates (**Supplemental S1.D**). It
391 was then hypothesized that this discrepancy could stem from the small height of the device (60
392 μm) significantly affecting the flow profile in the device, which 2D studies do not adequately

393 account for. Supporting this hypothesis, three-dimensional (3D) simulations matched experimental
394 proportional Out 1 flow split results, providing more evidence of simulation accuracy.

395 Using known coordinates of each filter geometry, a 2D velocity profile in the main channel
396 of the device was obtained for each flow rate across the x-y plane at $z = 30 \mu\text{m}$. This mid-point
397 plane was selected to avoid drastic ceiling and floor effects. Example x-velocity contours for 2
398 mL/min and 20 mL/min inlet flow rates for the Bent lobe design are shown in **Fig. 5**. As can be
399 seen, the inlet throttle significantly increases the fluid velocity over the beginning few filter lobes
400 from which most fluid leaves the main channel (depicted by dark blue in between the lobes).
401 Interestingly, the lobe pore where most fluid leaves the main channel changes with inlet flow rate,
402 likely resulting from changes in fluid inertia⁵³. Further down the device, all simulations for both
403 devices predict proportionally smaller transverse y-velocities between the inner and outer
404 channels.

405 The most interesting result from the simulations was obtained when analyzing the x-
406 velocity profile at x-coordinates at the edge of individual filter lobes before the downstream pore.
407 Here, the x- velocity profile was obtained at all points in the main channel along the y-axis keeping
408 the x-coordinate constant (portrayed by the thin, black box on velocity contour in **Fig. 5**). As
409 expected, at slower inlet flow rates, the x-velocity profile mimicked Poiseuille flow commonly
410 seen in most microfluidic flows (**Fig. 5A**). However, as inlet flow rates increased to greater than 4
411 mL/min, a new, complex velocity profile points emerged. At these flow rates, the x-velocity profile
412 at each lobe had three local maxima and two inflection points, which can be visualized in **Fig. 5B**.
413 Moreover, this complex velocity profile also appeared in the Oblong lobe device simulations
414 (**Supplemental S9**).

415

416 **f. Complex velocity profiles predict filtration success**

417 At moderate Reynold's numbers, microparticles in confined flow experience an inertial lift
418 force due to fluid shear gradient and wake asymmetry brought by a channel wall. These forces
419 point outward and inward from the center of the channel⁸, respectively. These forces are often
420 equated to a net, inertial lift force that is dependent on the sign of the shear rate and shear gradient,
421 among other factors^{8,17,20–23}. Therefore, the net, inertial lift force points outward from the center
422 channel in classic, confined Poiseuille flow. However, the complex channel design of a lobe filter
423 greatly changes the velocity profile along the x-axis of the device, thus significantly changing the

424 inertial lift forces a particle experiences. Moreover, the array of lobes causes the wall-induced lift
425 force to periodically disappear along the x-axis of the device. Without the wall-induced lift force,
426 a particle will experience a greater outward shear-induced inertial lift force, as well as increased
427 drag force from fluid flow in the y direction, which will cause it to pass through the filter lobes.
428 Therefore, poor filtration would be expected if the main channel velocity profile only showed
429 Poiseuille flow, which can be experimentally observed by the poor particle filtration with inlet
430 flow rates under 4 mL/min (**Figs. 3A,4**).

431 However, since the inertial lift force coefficient is proportional to the signs of shear rate
432 and shear gradient, the net inertial lift forces a particle experience could potentially change
433 directions in flows with inflection points^{20,21}. Based on this hypothesis, in the range where both
434 shear rate and shear gradient are negative, the net inertial lift force points toward the center of the
435 main channel. This region encompasses the location of the local maxima closest to the lobe (U^*)
436 to the location of the inflection point (D^*), as can be visualized in **Fig. 6A** (yellow shading).
437 Therefore, we hypothesize that if a particle's diameter (D_p) is larger than the distance from the
438 lobe to the inflection point (D^*), the particle will experience the lift force direction reversal. Thus,
439 the particle will remain in the same channel and achieve high filtration efficiencies. Likewise, if
440 D_p is less than the distance from the lobe to the height of the local maxima (U^*), the particle is
441 unable to experience the inertial force direction change. Hence, the particle will leave the main
442 channel and exit through following filter pore, obtaining only low filtration efficiencies.
443 Furthermore, if D_p is larger than U^* but smaller than D^* , the particle will not experience the full
444 lift force reversal region. Therefore, we expect some of the particles of this size will be filtered
445 while others will leave through filter pores, resulting in moderate filtration efficiencies.

446 Using this hypothesis, we were curious if we could predict lobe filtration success.
447 Accordingly, using the velocity profiles obtained from simulations, we found the location of U^*
448 and D^* for several inlet flow rates for both devices. For this analysis, the heights were measured
449 at lobe locations with the highest outward secondary flow to observe the region with the strongest
450 lateral force due to the y-velocity component, which varied by inlet flow rate. This is the location
451 where the particles experience the strongest y-velocity resistance to remain in the main channel.
452 **Fig. 6B** and **Fig. 6C** shows the locations of U^* (dashed line) and D^* (solid line) for the Oblong
453 lobe and Bent lobe devices across various inlet flow rates. The estimated filtration efficiencies
454 based on the previous hypothesis are depicted by the red, yellow, and green backgrounds.

455 Experimental filtration efficiencies based on particle size were compared to the simulation
456 predicted filtration success to test the viability of using U^* and D^* to explain lobe filtration results.
457 Hence, overlaid on **Figs. 6B,6C** are the respective experimental filtration efficiencies based on
458 particle size (D_p) and inlet flow rate. For ease of comparison, experimental particle size filtration
459 efficiencies were binned and categorized into low (0-60%), moderate (60-90%), and high (>90%)
460 efficiencies, which are depicted by a red x, black dash, and green circle, respectively.

461 As predicted by our theoretical analysis, the simulation-derived values for U^* and D^*
462 predict poor filtration under 4 mL/min for both devices due to the lack of inflection points in the
463 velocity profiles, which is recapitulated by the experimentally determined values. However, at
464 inlet flow rates above 4 mL/min, predicted filtration success varies between both devices. At each
465 inlet flow rate, D^* for the Oblong lobe device is higher than the D^* for the Bent lobe device with
466 a minimum D^* of 23 μm for the Oblong lobe device and 19 μm for the Bent lobe. Thus, these
467 differences in D^* predicted a larger D_p necessary for high efficiency filtration in the Oblong lobe
468 device. The estimated efficiencies are supported by the experimentally obtained filtration
469 efficiencies, as the Oblong lobe device only obtained high efficiencies with the 25-30 μm bin,
470 while the Bent lobe device obtained high efficiencies down to the 15-20 μm bin. Additionally, the
471 channel location where the inertial lift force points inward, or the area indicated by the yellow in
472 both figures, is predicted to be much smaller for the Oblong lobe device, which would predict
473 fewer particle sizes that are able to obtain moderate filtration efficiencies. Again, the experimental
474 values support the predicted values, as the Oblong device only obtained moderate efficiencies at
475 three inlet flow rates for the 20-25 μm particle size bin. Moreover, the Oblong lobe simulation
476 predicted a higher U^* at each inlet flow rate than the Bent lobe device with a minimum at 17 μm
477 compared to 10 μm for the Bent lobe device. The predicted values are further supported by the
478 low efficiencies obtained by all particles under 20 μm in the Oblong lobe device. Conversely, the
479 Bent lobe device obtained moderate efficiencies with the 10-15 μm bin for all inlet flow rates over
480 6 mL/min. Interestingly, both device simulations predicted a slight increase in both U^* and D^* , at
481 the filter's maximum inlet velocity, which may suggest decreased filtration success at inlet flow
482 rates higher than tested. Remarkably, experimental filtration efficiencies match very well with the
483 simulation-estimated efficiencies for both devices, which strongly supports that simulation-
484 derived distances for the inertial lift force reversal region can be used to predict microfluidic lobe
485 filtration success.

486 **4. Conclusions**

487 Microfluidic microparticle filtration is often a slow and tedious process plagued with filter
488 clogging and slow flow rates. Lobe filtration, bioinspired by the Manta Ray's filter feeding
489 mechanism, offers a unique solution for these issues. As a novel mechanism for microfluidic
490 applications, lobe filtration offers high throughput microparticle filtration with processing speeds
491 up to 20 mL/min. The high processing speeds open the possibility for various applications in which
492 large volumes of liquid need to be filtered. For example, microplastic removal, which has
493 concentrations of ~ 400 parts/L^{4,54}, would otherwise not be possible using a microfluidic device
494 without extensive parallelization. Moreover, lobe filtration offers high sample filtrate purity (>
495 98%), making it promising solution for applications such as tissue dissociation and filtration of
496 MCF-7 human cancer cells and murine kidney tissue cells³⁸. Lobe filtration also enables
497 microparticle concentration up to a factor of 2.05 at 10 mL/min, which would similarly increase
498 throughput of sample concentrations of dilute microparticle suspensions.

499 Remarkably, lobe filtration success can be estimated through a simple analysis of the velocity
500 profiles within the device. Understanding that the inertial lift force may change directions in the
501 presence of an inflection point in the bulk velocity profile, microparticle filtration success can be
502 estimated by comparing the particle size to the distance from a filter lobe with the highest
503 transverse velocity to the inflection point in bulk flow. We have shown that this method of
504 predicting filtration success works for both filter lobe designs over various inlet flow rates. Quick
505 visualization of U^* and D^* for both devices reveal that the Bent lobe device will provide better
506 filtration efficiencies compared to the Oblong lobe design since its high efficiency area (green
507 shading) in **Fig. 6** is larger and its low efficiency area (red shading) is smaller. Using this method,
508 lobe filter designs can be tuned to optimize the bulk flow inflection point location and thus filter
509 or concentrate particles of desired sizes at ultra-high throughputs.

510

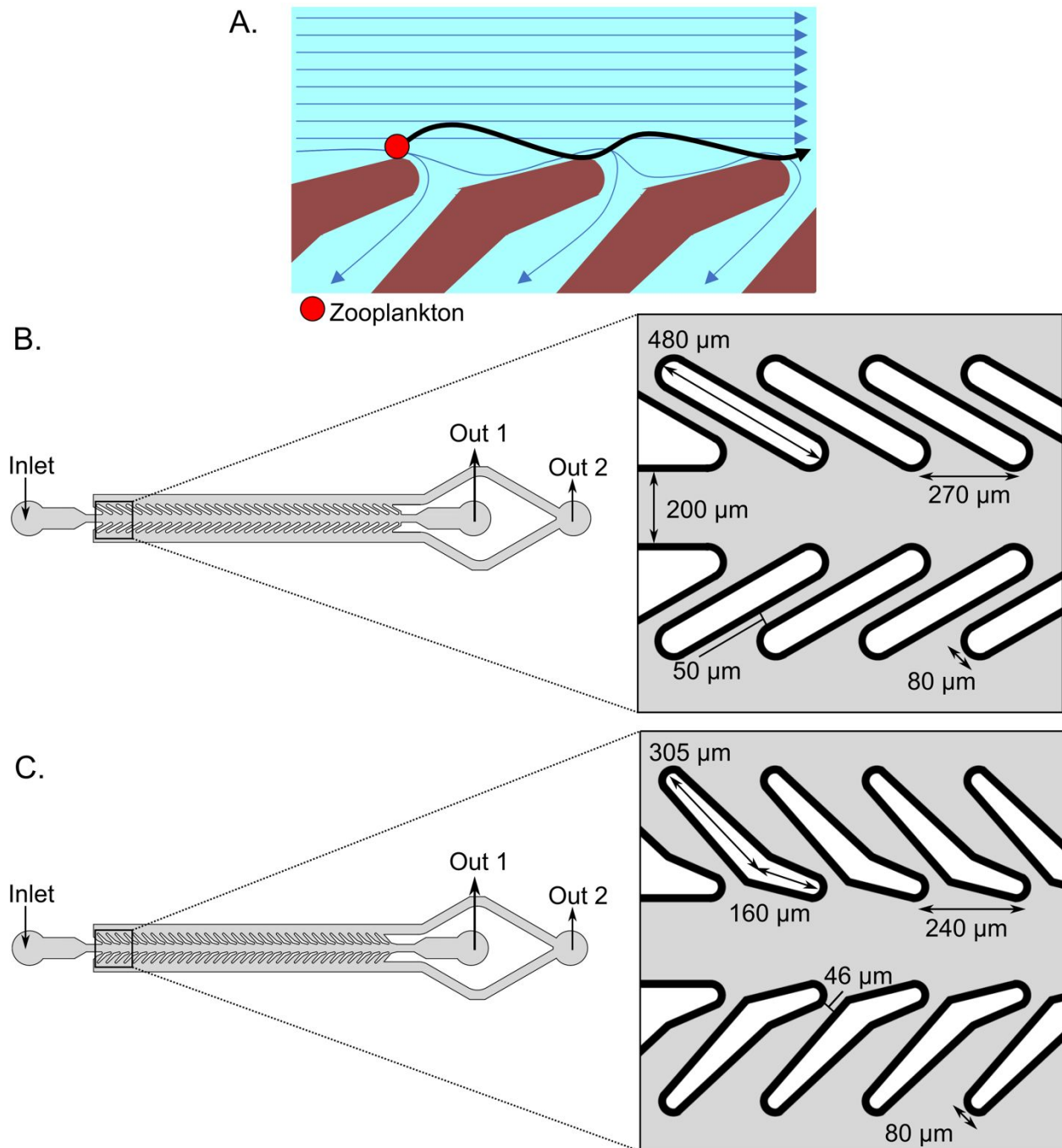
511 **5. Conflicts of interest**

512 There are no conflicts to declare.

513

514 **6. Acknowledgements**

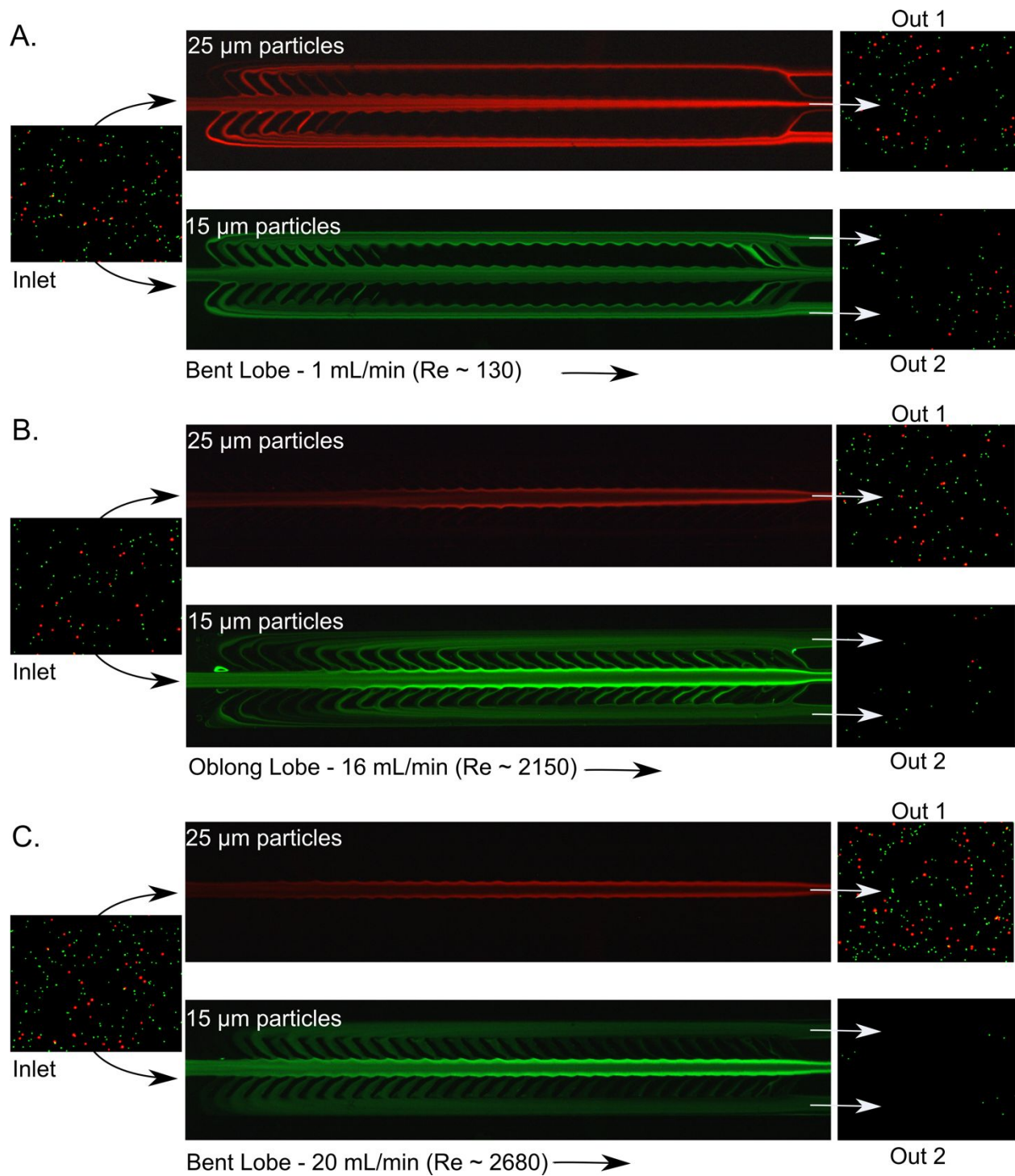
515 This work was supported by the U.S. National Science Foundation (IOS 1838314) and the NIH
516 (R21AG059099)

517 **Figures:**

518

519 **Figure 1. A.** Cartoon schematic portraying how both species of the Manta Ray feed on zooplankton. Blue
 520 arrows indicated fluid flow direction and the black arrow represents an example particle path. The lobe
 521 design shown is based on the *M. tarapacana*. **B.** Schematic of the Oblong lobe microfluidic device based
 522 on the *M. birostris* lobe design. Dimensions of the main channels are shown in the inset image with a total
 523 device height of 60 μm . **C.** Schematic of the Bent lobe microfluidic device based on the *M. tarapacana*

524 lobe design. Dimensions of the main channels are shown in the inset image with a total device height of 60
 525 μm . The main channel is 200 μm in width.



526

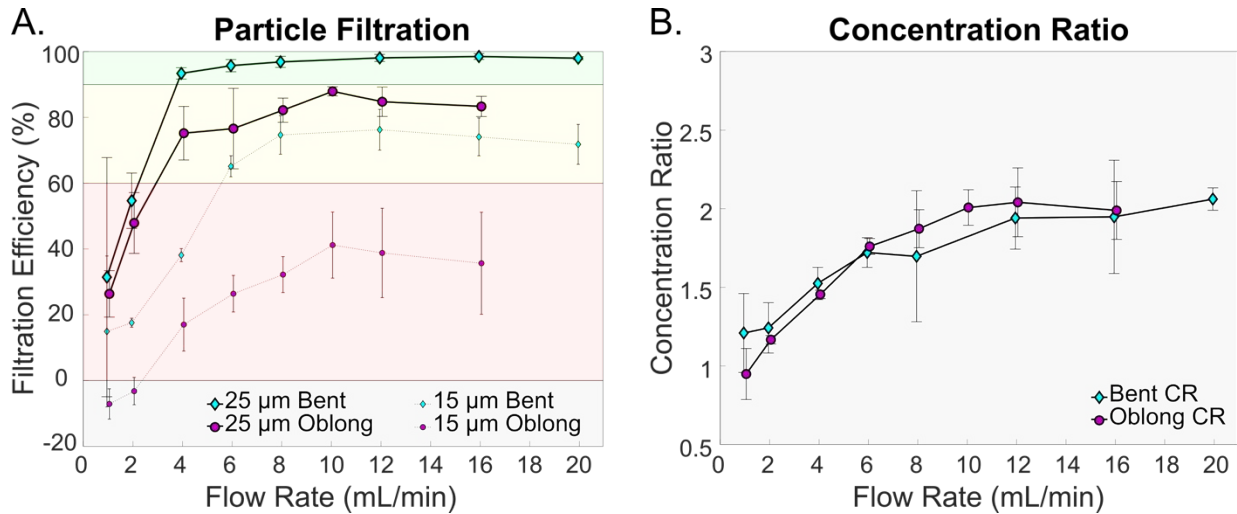
527 **Figure 2.** Representative steady state device operation with example inlet and outlet images for the A. Bent

528 lobe device at 1 mL/min, B. Oblong lobe device at 16 mL/min, and C. Bent lobe device at 20 mL/min. Red

529 and green channel images depict the particle tracks for 25 μm particles and 15 μm particles, respectively.
 530 Particle count images have both channels overlapped to easily compare particle concentrations.

531

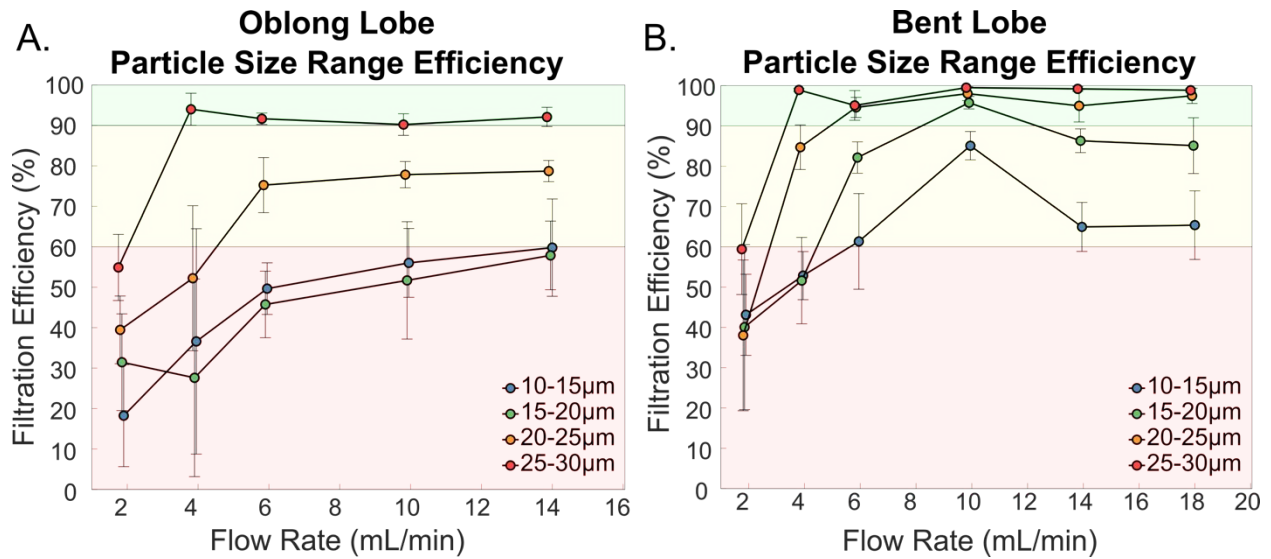
532



533

534 **Figure 3 A.** Microparticle filtration efficiencies for both the Oblong lobe (purple circle) and Bent lobe filter
 535 (blue diamond) designs over various inlet flow rates. The solid lines indicate 25 μm filtration efficiencies
 536 while the dashed line represents 15 μm filtration efficiencies (standard deviation as error bars, N=3). The
 537 red, yellow, and green shaded backgrounds represent low (0-60%), moderate (60-90%), and high (>90%)
 538 filtration efficiency regions, respectively. The grey background indicates a negative efficiency, meaning
 539 particle concentrations are higher in Out 2 then in the inlet. **B.** Concentration ratio results (standard
 540 deviation as error bars, N=3) for the Oblong lobe (purple circle) and Bent lobe (blue diamond) designs over
 541 various inlet flow rates. A CR greater than 1 indicates a higher concentration in Out 1 compared to the
 542 starting concentration.

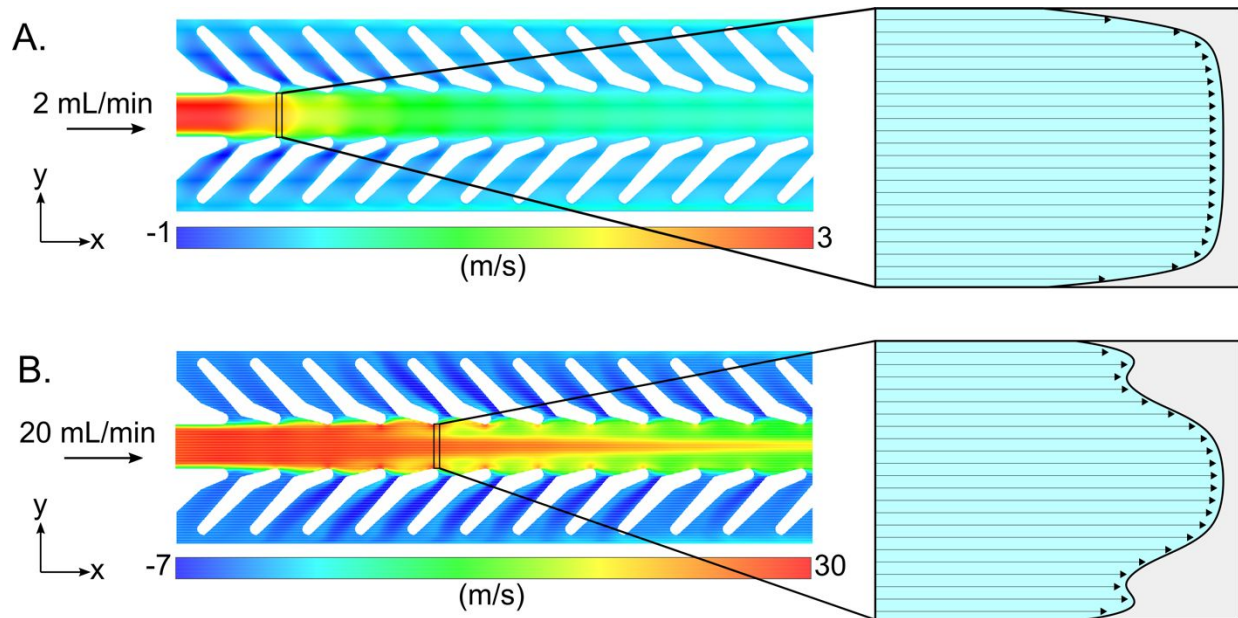
543



544

545 **Figure 4.** Filtration efficiency (Standard deviation error bars, N=3) for particles ranging from 10 to 30 μm
 546 in diameter for the **A.** Oblong lobe design and **B.** Bent lobe design. Particles were binned by size into groups
 547 of 5 μm . The red, yellow, and green backgrounds indicate low (0-60%), moderate (60-90%), and high
 548 (>90%) filtration efficiency regions, respectively.

549

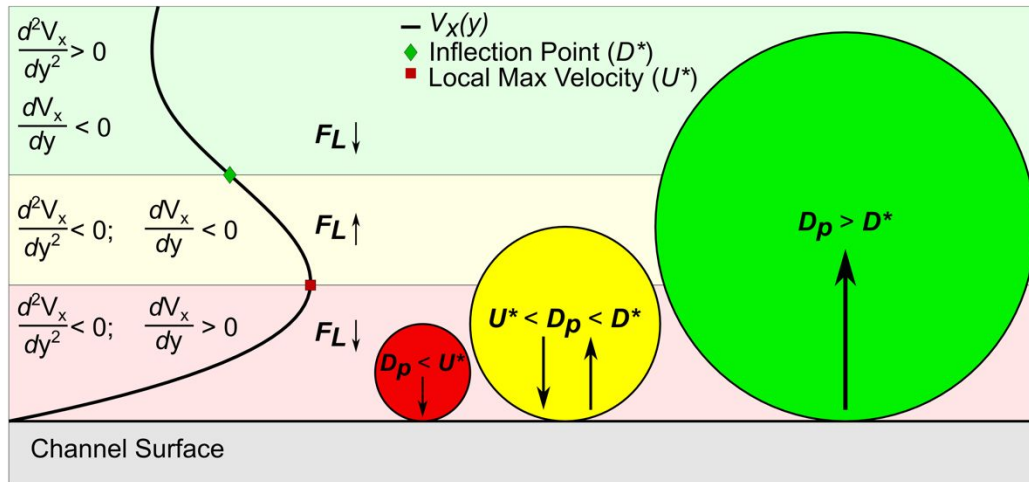


550

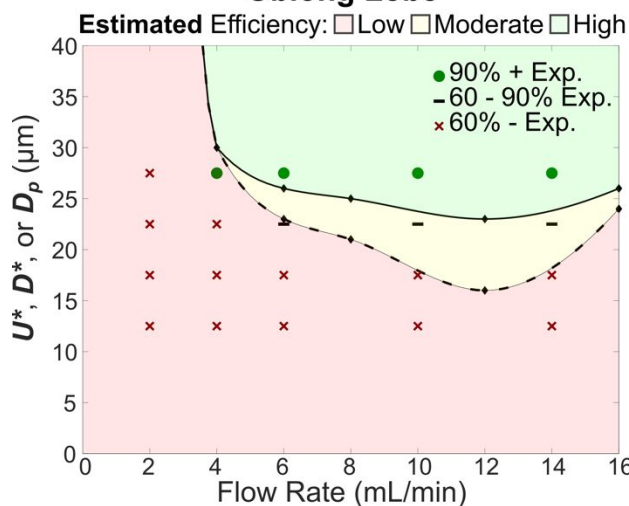
551 **Figure 5.** Example velocity contours with main channel velocity profiles shapes at individual x-coordinates
 552 obtained from Ansys Fluent simulations of the Bent lobe design. **A.** A 2 mL/min inlet flow rate showed a
 553 classic, Poiseuille flow profile at individual lobes within the device, while the **B.** 20 mL/min inlet flow rate
 554 revealed a complex velocity profile consisting of three local velocity maxima and two inflection points.

555

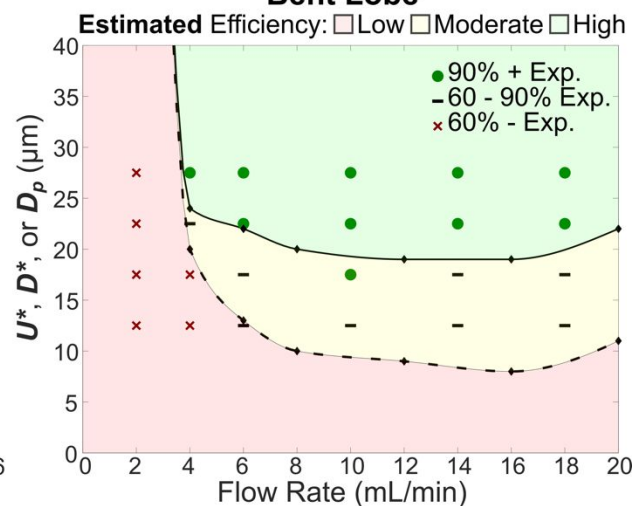
A.



B. **Estimated vs Experimental Filtration Oblong Lobe**



C. **Estimated vs Experimental Filtration Bent Lobe**



556

557 **Figure 6. A.** Cartoon representation of the inertial lift force reversal region, which is dependent on the
 558 locations of the nearest local velocity maxima and the inflection point in the velocity profile. From the
 559 channel surface to the nearest local maxima, the shear rate is positive and the shear gradient is negative,
 560 causing the inertial lift coefficient to point outward from the center of the channel. Particles small than this
 561 distance only experience outward lift force. In between the local velocity maxima (U^*) and the inflection
 562 point (D^*), the sign of the shear rate changes direction, which causes the inertial lift force to change
 563 directions in this region. Particles with diameters in this range may experience part of the inertial lift force
 564 direction region. Particles with diameters larger than the inflection point experience the entire lift force
 565 reversal region and thus, are filtered by the device at these lobes. Heights of the local max velocity (U^* -
 566 dashed line) and inflection point (D^* - solid line) at the lobe the lobe with the greatest outward y-velocity

567 with experimental filtration efficiencies based on particle size (D_p) for the **B**. Oblong lobe design and **C**.
568 Bent lobe design. Particle size efficiency data was binned by low, moderate, and high efficiency depicted
569 by the red x, dashed line, and green circle, respectively.

570

571 **References:**

- 572 1. Maria, M. S., Kumar, B. S., Chandra, T. S. & Sen, A. K. Development of a microfluidic
573 device for cell concentration and blood cell-plasma separation. *Biomed. Microdevices* **17**,
574 1–19 (2015).
- 575 2. Hou, H. W. *et al.* Microfluidic devices for blood fractionation. *Micromachines* **2**, 319–343
576 (2011).
- 577 3. Tachi, T., Kaji, N., Okamoto, Y., Tokeshi, M. & Baba, Y. Simultaneous separation,
578 metering and dilution of plasma from human whole blood using a microchip with an
579 interchannel microstructure. *Proc. Conf. MicroTAS 2009 - 13th Int. Conf. Miniaturized*
580 *Syst. Chem. Life Sci.* **81**, 427–429 (2009).
- 581 4. Li, J., Liu, H. & Paul Chen, J. Microplastics in freshwater systems: A review on
582 occurrence, environmental effects, and methods for microplastics detection. *Water Res.*
583 **137**, 362–374 (2018).
- 584 5. Talvitie, J., Heinonen, M., Koistinen, A. & Mikola, A. How well is microlitter purified
585 from wastewater? A detailed study on the stepwise removal of microlitter in a tertiary
586 level wastewater treatment plant. **109**, 164–172 (2017).
- 587 6. Dressaire, E. & Sauret, A. Clogging of microfluidic systems. *Soft Matter* **13**, 37–48
588 (2017).
- 589 7. Duffy, D. C., McDonald, J. C., Schueller, O. J. A. & Whitesides, G. M. Rapid prototyping
590 of microfluidic systems in poly(dimethylsiloxane). *Anal. Chem.* **70**, 4974–4984 (1998).
- 591 8. Di Carlo, D. Inertial microfluidics. *Lab Chip* **9**, 3038–3046 (2009).
- 592 9. Ding, X. *et al.* Cell separation using tilted-angle standing surface acoustic waves. (2014)
593 doi:10.1073/pnas.1413325111.
- 594 10. Akiyama, Y., Egawa, T., Koyano, K. & Moriwaki, H. Acoustic focusing of microplastics
595 in microchannels: A promising continuous collection approach. *Sensors Actuators, B*
596 *Chem.* **304**, (2020).
- 597 11. Hejazian, M., Li, W. & Nguyen, N. Lab on a chip for continuous-flow magnetic cell

- 598 separation. 959–970 (2015) doi:10.1039/c4lc01422g.
- 599 12. Huang, L. R., Cox, E. C., Austin, R. H. & Sturm, J. C. Continuous Particle Separation
600 Through Deterministic Lateral Displacement. *Science (80-.)*. **304**, 987–990 (2004).
- 601 13. Chen, X., Cui, D. F., Liu, C. C. & Li, H. Microfluidic chip for blood cell separation and
602 collection based on crossflow filtration. *Sensors Actuators, B Chem.* **130**, 216–221 (2008).
- 603 14. Li, H. Y., Bertram, C. D. & Wiley, D. E. Mechanisms by which pulsatile flow affects
604 cross-flow microfiltration. *AIChE J.* **44**, 1950–1961 (1998).
- 605 15. Yoon, Y. *et al.* Clogging-free microfluidics for continuous size-based separation of
606 microparticles. *Sci. Rep.* **6**, 1–8 (2016).
- 607 16. Lenshof, A. & Laurell, T. Continuous separation of cells and particles in microfluidic
608 systems. *Chem. Soc. Rev.* **39**, 1203–1217 (2010).
- 609 17. Stoecklein, D. & Di Carlo, D. Nonlinear Microfluidics. *Anal. Chem.* **91**, 296–314 (2019).
- 610 18. Zhao, Q., Yuan, D., Zhang, J. & Li, W. A review of secondary flow in inertial
611 microfluidics. *Micromachines* **11**, 1–23 (2020).
- 612 19. Wang, L. & Dandy, D. S. High-Throughput Inertial Focusing of Micrometer- and Sub-
613 Micrometer-Sized Particles Separation. *Adv. Sci.* **4**, (2017).
- 614 20. Ho, B. P. & Leal, L. G. Inertial migration of rigid spheres in two-dimensional
615 unidirectional flows. *J. Fluid Mech.* **65**, 365–400 (1974).
- 616 21. Amini, H., Lee, W. & Di Carlo, D. Inertial microfluidic physics. *Lab Chip* **14**, 2739–2761
617 (2014).
- 618 22. Liu, C., Xue, C., Sun, J. & Hu, G. A generalized formula for inertial lift on a sphere in
619 microchannels. *Lab Chip* **2016**, 884–892 (2016).
- 620 23. Asmolov, E. S. The inertial lift on a spherical particle in a plane poiseuille flow at large
621 channel Reynolds number. *J. Fluid Mech.* **381**, 63–87 (1999).
- 622 24. Kwak, B. *et al.* Hydrodynamic blood cell separation using fishbone shaped microchannel
623 for circulating tumor cells enrichment. *Sensors Actuators, B Chem.* **261**, 38–43 (2018).
- 624 25. Özbey, A., Karimzadehkhoei, M., Alijani, H. & Koşar, A. Microparticle Inertial
625 Focusing in an Asymmetric Curved Microchannel. *Fluids* **3**, 57 (2018).
- 626 26. Wang, X., Zhou, J. & Papautsky, I. Vortex-aided inertial microfluidic device for
627 continuous particle separation with high size-selectivity, efficiency, and purity.
628 *Biomicrofluidics* **7**, (2013).

- 629 27. Volpe, A., Paiè, P., Ancona, A. & Osellame, R. Polymeric fully inertial lab-on-a-chip with
630 enhanced-throughput sorting capabilities. *Microfluid. Nanofluidics* **23**, 1–10 (2019).
- 631 28. Warkiani, M. E. *et al.* Malaria detection using inertial microfluidics. *Lab Chip* **15**, 1101–
632 1109 (2015).
- 633 29. Di Carlo, D., Edd, J. F., Humphry, K. J., Stone, H. A. & Toner, M. Particle segregation
634 and dynamics in confined flows. *Phys. Rev. Lett.* **102**, 1–4 (2009).
- 635 30. Bhagat, A. A. S., Kuntaegowdanahalli, S. S. & Papautsky, I. Inertial microfluidics for
636 continuous particle filtration and extraction. *Microfluid. Nanofluidics* **7**, 217–226 (2009).
- 637 31. Shen, S. *et al.* Spiral microchannel with ordered micro-obstacles for continuous and
638 highly-efficient particle separation. *Lab Chip* **17**, 3578–3591 (2017).
- 639 32. Tasadduq, B., Lam, W., Alexeev, A., Sarioglu, A. F. & Sulchek, T. Enhancing size based
640 size separation through vertical focus microfluidics using secondary flow in a ridged
641 microchannel. *Sci. Rep.* **7**, 1–10 (2017).
- 642 33. Johnston, I. D. *et al.* Dean flow focusing and separation of small microspheres within a
643 narrow size range. *Microfluid. Nanofluidics* **17**, 509–518 (2014).
- 644 34. Wang, L. & Dandy, D. S. High-Throughput Inertial Focusing of Micrometer- and Sub-
645 Micrometer-Sized Particles Separation. *Adv. Sci.* **4**, (2017).
- 646 35. Mihandoust, A. *et al.* High-throughput particle concentration using complex cross-section
647 microchannels. *Micromachines* **11**, (2020).
- 648 36. Wu, Z., Chen, Y., Wang, M. & Chung, A. J. Continuous inertial microparticle and blood
649 cell separation in straight channels with local microstructures. *Lab Chip* **16**, 532–542
650 (2016).
- 651 37. Lee, J. H., Lee, S. K., Kim, J. H. & Park, J. H. Separation of particles with bacterial size
652 range using the control of sheath flow ratio in spiral microfluidic channel. *Sensors*
653 *Actuators, A Phys.* **286**, 211–219 (2019).
- 654 38. Qiu, X. *et al.* Microfluidic filter device with nylon mesh membranes efficiently dissociates
655 cell aggregates and digested tissue into single cells. *Lab Chip* **18**, 2776–2786 (2018).
- 656 39. Kulasinghe, A., Zhou, J., Kenny, L., Papautsky, I. & Punyadeera, C. Capture of
657 circulating tumour cell clusters using straight microfluidic chips. *Cancers (Basel)*. **11**, 1–
658 11 (2019).
- 659 40. Qiu, X. *et al.* Microfluidic channel optimization to improve hydrodynamic dissociation of

- 660 cell aggregates and tissue. *Sci. Rep.* **8**, 1–10 (2018).
- 661 41. Zhou, J., Kasper, S. & Papautsky, I. Enhanced size-dependent trapping of particles using
662 microvortices. *Microfluid. Nanofluidics* **15**, 611–623 (2013).
- 663 42. Wang, L. & Dandy, D. S. A microfluidic concentrator for cyanobacteria harvesting. *Algal*
664 *Res.* **26**, 481–489 (2017).
- 665 43. Divi, R. V., Strother, J. A. & Misty Paig-Tran, E. W. Manta rays feed using ricochet
666 separation, a novel nonclogging filtration mechanism. *Sci. Adv.* **4**, (2018).
- 667 44. Paig-Tran, E. W. M., Kleinteich, T. & Summers, A. P. The filter pads and filtration
668 mechanisms of the devil rays: Variation at macro and microscopic scales. *J. Morphol.*
669 **274**, 1026–1043 (2013).
- 670 45. Paig-tran, E. W. M., Bizzarro, J. J., Strother, J. A. & Summers, A. P. Bottles as models :
671 predicting the effects of varying swimming speed and morphology on size selectivity and
672 filtering efficiency in fishes. 1643–1654 (2011) doi:10.1242/jeb.048702.
- 673 46. Mossige, E. J., Jensen, A. & Mielnik, M. M. An experimental characterization of a tunable
674 separation device. *Microfluid. Nanofluidics* **20**, 1–10 (2016).
- 675 47. Mossige, E. J., Jensen, A. & Mielnik, M. M. Separation and Concentration without
676 Clogging Using a High-Throughput Tunable Filter. *Phys. Rev. Appl.* **9**, 54007 (2018).
- 677 48. Mossige, E. J., Edvardsen, B., Jensen, A. & Mielnik, M. M. A tunable, microfluidic filter
678 for clog-free concentration and separation of complex algal cells. *Microfluid. Nanofluidics*
679 **23**, 1–13 (2019).
- 680 49. Gossett, D. R. *et al.* Label-free cell separation and sorting in microfluidic systems. *Anal.*
681 *Bioanal. Chem.* **397**, 3249–3267 (2010).
- 682 50. Ding, J., Zhang, S., Razanajatovo, R. M., Zou, H. & Zhu, W. Accumulation, tissue
683 distribution, and biochemical effects of polystyrene microplastics in the freshwater fish
684 red tilapia (*Oreochromis niloticus*). *Environ. Pollut.* **238**, 1–9 (2018).
- 685 51. Frias, J. P. G. L. & Nash, R. Microplastics: Finding a consensus on the definition. *Mar.*
686 *Pollut. Bull.* **138**, 145–147 (2019).
- 687 52. Ozawa, R., Iwadate, H., Toyoda, H., Yamada, M. & Seki, M. A numbering-up strategy of
688 hydrodynamic microfluidic filters for continuous-flow high-throughput cell sorting. *Lab*
689 *Chip* **19**, 1828–1837 (2019).
- 690 53. Blonski, S. *et al.* Impact of inertia and channel angles on flow distribution in microfluidic

- 691 junctions. *Microfluid. Nanofluidics* **24**, (2020).
- 692 54. Browne, M. A. *et al.* Accumulation of microplastic on shorelines worldwide: Sources and
- 693 sinks. *Environ. Sci. Technol.* **45**, 9175–9179 (2011).
- 694