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ARTICLE

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Understanding how to control the nucleation and growth rates is crucial for designing nanoparticles with specific sizes and shapes. In this study, we show that the nucleation and growth rates are correlated with the thermodynamics of metal-ligand/solvent binding for the pre-reduction complex and the surface of the nanoparticle, respectively. To obtain these correlations, we measured the nucleation and growth rates by in-situ small angle X-ray scattering during the synthesis of colloidal Pd nanoparticles in the presence of trioctylphosphine in solvents of varying coordinating ability. The results show that the nucleation rate decreased, while the growth rate increased in the following order, toluene, piperidine, 3,4-lutidine and pyridine, leading to a large increase in the final nanoparticle size (from 1.4 nm in toluene to 5.0 nm in pyridine). Using density functional theory (DFT), complemented by ³¹P nuclear magnetic resonance and X-ray absorption spectroscopy, we calculated the reduction Gibbs free energies of the solvent-dependent dominant pre-reduction complex and the solvent-nanoparticle binding energy. The results indicate that lower nucleation rates originate from solvent coordination which stabilizes the pre-reduction complex and increases its reduction free energy. At the same time, DFT calculations suggest that the solvent coordination affects the effective capping of the surface where stronger binding solvents slow the nanoparticle growth by lowering the number of active sites (not already bound by trioctylphosphine). The findings represent a promising advancement towards understanding the microscopic connection between the metal-ligand thermodynamic interactions and the kinetics of nucleation and growth to control the size of colloidal metal nanoparticles.

Introduction

Strategies for predictably controlling the synthesis of colloidal metal nanoparticles are of extreme importance because their properties (catalytic, thermal, optical) vary with size, shape, and composition.¹⁻⁶ Many studies have focused on controlling the size and shape of colloidal nanoparticles by changing synthesis reagents such as the precursors,⁷⁻¹⁰ ligands¹¹⁻¹⁷ or solvents,¹⁸⁻²⁰ or by changing the synthesis conditions^{21, 22} such as precursor/ligand concentrations²³⁻³⁰ and reaction temperature.³¹⁻³³ Among these factors, ligands have gained

significant attention due to their wide variety in terms of steric effect and binding strength with the metal precursor and nanoparticle surface. It has been demonstrated that the nanoparticle size and shape can be affected by the ligand type^{11, 13, 16, 34, 35} and concentration,^{12, 29, 36} which have been attributed to differences in the precursor reactivity, or the ligand capping (strength or density) on the nanoparticle surface. These differences in sizes and shapes which are largely dependent on the nature of metal-ligand interactions (binding), give evidences that ligands play an important role in controlling the nucleation and growth rates. We have recently shown³⁷⁻⁴⁰ that the ligand-metal interaction could be responsible for the synthesis mechanism of colloidal metal nanoparticles (e.g. Pd,^{37, 38} Au,⁴¹ Rh⁴² and Ir⁴³) not following the LaMer⁴⁴ nucleation and growth model. Specifically, instead of a short burst of nucleation followed by diffusion controlled growth (temporal separation), slow nucleation and temporal overlap of growth have been reported by several groups^{12, 37, 38, 42, 43} and alternative mechanisms/models have been proposed.^{37, 43, 45-48} Depending on whether the ligand has a higher affinity to the metal monomers or to the nanoparticle surface, it could either inhibit the nucleation resulting in fast growth and large sizes of nanoparticles, or block the surface sites for growth resulting in a continuous nucleation and smaller sizes.⁴⁹⁻⁵¹

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Organic solvents have been investigated to vary the size and shape of colloidal nanoparticles.^{18-20, 34, 52, 53} The large amount of solvent molecules in the synthetic phase may modify the ligand binding strength on the nanoparticle surface through Van der Waals interaction with the ligand carbon chains.^{20, 52} This steric interaction has been proposed to change the ligand capping density through different mechanisms, i.e., either occupying space between the surface ligands,⁵² or modifying the electronic property of the ligand binding functional group/atom.²⁰ Additionally, solvents can also act as ligands if they have considerable direct interactions with the metal complex or the nanoparticle surface. Solvent's binding to the metal center can lower the precursor reactivity^{16, 35} while binding to the nanoparticle surface can inhibit surface growth and the ability to bind to both can affect the nucleation and growth rates.⁵⁴ Therefore, the solvent can serve as a promising variable for controlling the size and shape of colloidal metal nanoparticles. We note that the effects of ligands, solvents, and other synthesis parameters on kinetics have been mostly shown indirectly, for example, by correlating known or calculated trend of metal-ligand/solvent binding energies/affinities with only the final size of the nanoparticles. However, the final nanoparticle size is the result of an interplay between nucleation and growth rates and the solvents and ligands can affect both. Therefore, without in-situ kinetics measurements the effects of solvents and ligands on kinetics and size are difficult to predict.

In this work, we investigated the effect of different solvents, i.e., toluene, and nitrogen coordinating (N-coordinating) solvents, piperidine, 3,4-lutidine and pyridine on the synthesis kinetics of Pd nanoparticles in the presence of trioctylphosphine (TOP). We directly correlate the metal-solvent interactions (structure and reducibility of the pre-reduction complexes, and solvent binding with the nanoparticle surface) with the nucleation and growth rates using in-situ small angle X-ray scattering (SAXS), ³¹P nuclear magnetic resonance (³¹P NMR), X-ray absorption fine structure (XAFS) and density functional theory (DFT) calculations. In this study, we directly measure the nucleation and growth rates to determine the effect of solvent properties on metal-ligand/solvent interactions and kinetics of metal colloidal nanoparticle synthesis. The results show that the nucleation and growth rates are correlated with the stability/reducibility of the TOP-Pd pre-reduction complexes and binding strength of the solvent/TOP-nanoparticle surface, respectively. The findings provide a promising route to systematically identify ligands/solvents to tune the synthesis kinetics and final nanoparticle size.

Experimental Section

Colloidal Pd Nanoparticle Synthesis and Characterization

The heat-up method⁸⁴ was used in the colloidal Pd nanoparticle synthesis. Palladium (II) acetate (Aldrich 99.9 %) was used after recrystallization. Trioctylphosphine (Aldrich 97 %) was used after further purification (distillation) and stored in a nitrogen (N₂) glovebox. The procedures for recrystallization and

distillation have been reported in our previous study^{48, 55}. Anhydrous toluene (Sial 99.8 %), 3,4-lutidine (Aldrich, 98 %), piperidine (Aldrich, redistilled 99.5 %) and pyridine (EMD Millipore 99.8 %) were used as solvents. Anhydrous hexanol (Sial 99 %) was used as the reducing agent. All the solvents and reducing agents were degassed using nitrogen (99.999% from liquid N₂ boil off) at a flow rate of 10 mL / min for 30 min before use. 10 mM recrystallized Pd acetate was dissolved in solvent : hexanol = 1:1 solution, with TOP : Pd = 1 or 2 (molar), and heated to 100 °C under 300 rpm stirring. All the agents were mixed in an inert gas glove box with both moisture and oxygen levels below 1.0 ppm. The syntheses have been repeated multiple times with 2 mL solution in 7 mL glass vials and 6 mL solution in 25 mL round bottom glass flask under 0 – 1000 rpm stirring, and the sizes were similar.

In-situ and Ex-situ Small-angle X-ray Scattering (SAXS) Data Acquisition and Analysis

The in-situ SAXS experiments were conducted at Argonne National Laboratory's Advanced Photon Source at beamline 12-ID-C. The incident X-ray energy is 18 keV. All the agents were mixed in the glove box at wet labs in sectors 16 and sector 20 and transferred immediately to the beamline under inert gas atmosphere. The reaction was conducted in a 25 mL round bottom flask (with 6 mL solution) with 300 rpm stirring. The synthesis setup is similar to the one used by Kwon, et al⁸⁵, which minimizes possible reactions triggered by the X-ray beam. A syringe needle was punched through the capping septum to avoid pressure build-up from the solvent evaporation during the synthesis. In each measurement, 300 µL was taken out of 6 mL solution into a quartz capillary for SAXS data acquisition using a syringe pump, and then returned into the flask. The pumping process took 7 s, and the fastest time resolution was set as 8 s. Each SAXS measurement was done when the solution flowed slowly through the capillary, with 0.1 s exposure time. The two-dimensional scattering pattern was averaged and normalized to get the curve of differential scattering cross section I(q) to the scattering vector q. The pure mixture of solvent and reducing agent at 50:50 volume ratio was used as background. To get the absolute scale intensity so that the nanoparticle concentration can be obtained, water was used as a primary standard due to its well-studied differential scattering cross section⁸⁶. The ex-situ SAXS was conducted on N8 Horizon (Bruker AXS GmbH, Germany) equipped with Cu (K α radiation, λ = 1.54 Å) source, 2-dimensional VANTEC-500 TM detector and SCATEX TM scatter-free pinholes. The colloidal nanoparticle solution samples were loaded and measured in a sealed in quartz capillaries (d = 1.5 mm or 2.0 mm) at room temperature in vacuum. Water was used as the standard for absolute calibration.

Data analysis was done using IGOR pro⁸⁷, Bruker SAXS software and simSAXS⁸⁸ package⁸⁸ on Matlab. The scattering curves were fitted using Schultz polydisperse spherical nanoparticles model, based on the assumption of spherical nanoparticle shape (justified from TEM images, see below), dilute solution

and homogeneous electron density. The Schulz-Zimm distribution function is:

$$f(r) = \frac{r^z}{\Gamma(z+1)} \left[\frac{z+1}{r_{avg}} \right]^{z+1} \exp\left(-\frac{(z+1)r}{r_{avg}}\right) \quad (5)$$

where $z = (r_{avg}/\sigma)^2 - 1$. r_{avg} is the average radius of nanoparticles; σ is the standard deviation and Γ is the gamma function.

If $I(q)$ is in absolute intensity, it is related to both of the size and number of nanoparticles in the solution:

$$I(q) = N_p \int_0^\infty f(r) V_p^2 P(q) dr \quad (6)$$

$$P(q) = [\Delta\rho \frac{3(\sin(qr) - qr\cos(qr))}{qr}]^2 \quad (7)$$

$P(q)$ is the form factor of a specific size of particles. For spherical particles, at $q = 0$, $P(q) = \Delta\rho^2$. N_p is the number of particles, V_p is the particle volume ($\frac{4}{3}\pi r^3$, and r is the radius), $\Delta\rho$ is the scattering length density difference between palladium and solvent. At $q = 0$, a relation can be used to calculate the number of particles:

$$I(0) = N_p \langle V_p^2 \rangle (\Delta\rho)^2 \quad (8)$$

From equation (8) the number of particles N_p , and further the extent of reaction (i.e. conversion of precursor into nanoparticles) can be calculated (see details in SI).

Transmission Electron Microscopy (TEM)

TEM images of synthesized nanoparticles were taken to confirm the nanoparticle size and shape. Images for the toluene, piperidine and 3,4-lutidine samples were acquired in scanning transmission electron microscopy (STEM) mode on FEI TITAN 80 – 300, equipped with the CEOS GmbH double-hexapole aberration corrector. The images were taken in high angle annular dark field (HAADF) mode. Images for pyridine samples were taken on JEOL 2100 TEM operated at 200 kV, which is equipped with a thermionic emission source and possesses a high-resolution objective pole piece. The resolution can reach 0.1 nm on both microscopes. The reaction solution was diluted 10 times with the respective anhydrous solvent. One drop of the solution was added on the lacey carbon copper grid and dried on top of a hotplate set up at 50 °C. The image analysis was performed with ImageJ⁸⁹. At least 300 nanoparticles were measured to calculate the number-average diameter and the polydispersity.

Nuclear Magnetic Resonance

Deuterated solvents, toluene-d8 (Alfa Aesar, 99.6 %) and pyridine-d5 (Alfa Aesar, 99.5 %) were used for NMR measurements. All the NMR measurements were taken at room temperature. The deuterated solvents were degassed with nitrogen at 10 mL/min before dissolving any agents. For each sample, 0.6 mL 20 mM freshly made palladium acetate solution was loaded in NMR tubes in the nitrogen glovebox, and sealed with rubber septa. All the measurements were taken immediately after the appropriate amount of ligand was added and mixed well in the NMR tube. One-dimensional ³¹P NMR spectra with proton decoupling were acquired at 25 °C on a

Bruker Avance II 500 MHz system equipped with a 5 mm broad band prodigy cryo probe. ³¹P NMR parameters included a total of 256 transients, 64K data points, spectral width of 59.5 kHz and a relaxation delay of 5.0 s between the transients. Apodization corresponding to a line broadening of 1.0 Hz was applied before Fourier Transformation. Resonances were assigned based on the chemical shift position of external standard 85% H₃PO₄ singlet at 0 ppm.

XAFS Data Collection and Analysis

XAFS experiments were performed at beamline BL 2-2 at the Stanford Synchrotron Radiation Light Source at SLAC National Accelerator Laboratory. 25 mM and 20 mM Pd acetate were dissolved in toluene and pyridine in 20 mL capped glass vial in an inert atmosphere glovebox and transferred to the synchrotron. The beam size was 200 μm vertical by 3 mm horizontal. The spectra were collected at the Pd K-edge in transmission mode. Four scans (20 minutes each) were collected then merged and aligned using a Pd foil spectrum collected simultaneously for each scan. All the measurements were taken at room temperature. Both XANES and EXAFS data processing and analysis were performed using Athena and Artemis programs of the IFEFFIT data analysis package^{90, 91}. After the normalization of the absorption coefficient, the smooth atomic background was subtracted using the AUTOBKG code Athena to obtain $\chi(k)$ (where k is the photoelectron wavenumber). FEFF6 code was used for constructing the theoretical EXAFS signal for Pd-Pd, Pd-P, Pd-C, and Pd-O scattering paths. For the scattering paths, structure of Pd acetate trimer (Pd-O and long Pd-C) was used in toluene, and Pd acetate monomer with terminal acetate (Pd-O and short Pd-C) was used in pyridine. The theoretical EXAFS signals fitting was conducted using Artemis in r -space. The spectra were fitted by changing the following parameters for the scattering paths of Pd-Pd, Pd-O, Pd-C: bond length disorder, σ_{Pd-Pd}^2 , σ_{Pd-O}^2 , σ_{Pd-C}^2 ; coordination numbers of the single scattering paths, N_{Pd-Pd} , N_{Pd-O} , N_{Pd-C} ; the effective scattering lengths (R_{Pd-Pd} , R_{Pd-O} , R_{Pd-C}); and the correction to the threshold energy, ΔE_0 (ΔE_0 for metallic Pd-Pd was assumed to be the same as other paths in the model to limit the number of parameters). The best fit of the passive electron reduction factor (0.83), S_0^2 , was fixed during the fitting based on analyzing the Pd foil spectrum. The k -range for Fourier Transform of the $\chi(k)$ and the r -range for the fitting were 2.5–12.5 Å⁻¹ and 1.1–3.1 Å, respectively.

Density Functional Theory (DFT) Calculations

DFT calculations were performed to capture the complexation of Pd species using the BP-86^{92, 93} functional combined with the def2-SV(P) basis set⁹⁴ and the resolution of identities (RI) approximation⁹⁵ as implemented in the Turbomole package.⁹⁶ To capture dispersion interactions between carbon chains we used the Grimme D3 dispersion correction.⁹⁷ Similar DFT methods have previously been used to study palladium complexes and nanoparticles, showing reasonable accuracy.^{98, 99} Implicit solvation effects (Fig. S17) were accounted for using the conductor-like screening model (COSMO)¹⁰⁰ with dielectric

constants taken, when available, from the Minnesota Solvent Descriptor Database¹⁰¹ or from other^{102, 103} databases. Different

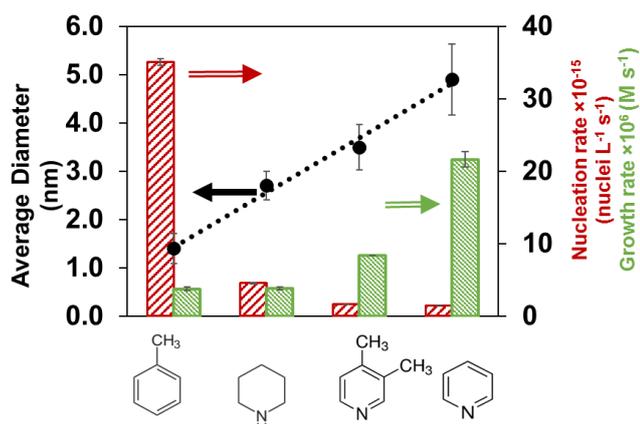


Figure 1 Final average nanoparticle size, and initial nucleation and growth rates (at 10 % Pd precursor conversion) measured in the different solvents. The error bars represent the standard deviation of the size distribution as obtained from the SAXS fits. error bars of the rates were calculated by propagating the error from the SAXS fits. The rates in toluene and pyridine were reproduced within 15% difference from two different in-situ experiments. The dotted lines show the observed size trend but do not represent a theoretical or empirical fit. Reaction conditions: Pd(OAc)₂ = 10 mM, TOP : Pd = 1 (molar), solv : hexanol = 50 : 50, T = 100 °C.

initial configurations were tested for each precursor complex and the lowest-energy fully relaxed structure is reported. Geometry relaxations were carried out using the quasi-Newton-Raphson method without any symmetry constraints. Multiple spin states were tested for each Pd-containing complex and the lowest-energy electron configuration always corresponded to the lowest possible spin state of the complex (i.e. singlet or doublet spin states). Vibrational analysis was performed to verify the relaxed structures as local minima and structures exhibiting imaginary frequencies were displaced (along the imaginary mode) and re-relaxed until no imaginary frequencies remained. Gibbs free energies were calculated (at 298.15 K) using the ideal gas rigid rotator harmonic oscillator approach applied to the vibrational modes for each system^{67, 104}. For each complex at least two initial configurations were tested and we report the values and structures of the lowest-energy configurations. Periodic DFT calculations were performed using the CP2K package to simulate adsorption on nanoparticle surface.¹⁰⁵ The PBE functional¹⁰⁶ was used with Grimme's D3 dispersion corrections. DZVP basis sets with the Goedecker, Teter, and Hutter (GTH) pseudopotentials were used with a kinetic energy cutoff of 500 Ry. To obtain the bulk unitcell parameters of Pd, a 2×2×2 supercell of the conventional unitcell (space group Fm-3m) was fully relaxed. The periodic geometries were optimized using the Broyden-Fletcher-Goldfarb-Shanno (BFGS) minimization algorithm with convergence criteria of 4.0×10⁻⁴ E_h per Bohr and 10⁻⁷ au for force and energy, respectively. To model the (111) surface facet of Pd, a p5×5 supercell (representing 1.649 nm² of surface area) was created using the primitive unitcell (of the relaxed unitcell) with four atomic layers and a vacuum of 12 Å was added for the construction of slab models. For surface calculations, the atoms of two bottom layers were kept frozen at their bulk positions and two top layers were allowed to relax. To reduce the

computational cost of periodic DFT calculations, triethylphosphine (TEP) was used as a model to represent the TOP ligand. We note that four TEP molecules sufficiently cover the full supercell of Pd (111) (Fig. S22) leaving no space for further ligands to adsorb on the surface. The adsorption binding energy of ligands (TEP or solvent) at $N_{lig}/4$ monolayer coverage (Fig. 5a) is defined as:

$$BE_{lig} = \frac{E[N_{lig}@Pd(111)] - E[Pd(111)] - N_{lig}E_{lig}}{N_{lig}} \quad (9)$$

Where $E[N_{lig}@Pd(111)]$ is the energy of N_{lig} ligand molecules adsorbed on the Pd(111) surface, $E[Pd(111)]$ is the energy of Pd(111) surface, and E_{lig} is the energy of ligand in the gas phase. Similarly, the binding energy of solvent at $N_{solv}/4$ monolayer coverage with TEP occupying remaining sites (Fig. 5c) is defined as:

$$BE_{solv} = \frac{E[N_{solv} + (4 - N_{solv})_{TEP}@Pd(111)] - E[(4 - N_{solv})_{TEP}@Pd(111)] - N_{solv}E_{solv}}{N_{solv}} \quad (10)$$

Where $E[(N_{solv} + (4 - N_{solv})_{TEP})@Pd(111)]$ is the energy of N_{solv} solvent and $(4 - N_{solv})$ TEP molecules adsorbed on Pd(111), $E[(4 - N_{solv})_{TEP}@Pd(111)]$ is the energy of $(4 - N_{solv})$ TEP molecules adsorbed on Pd(111), and E_{solv} is the energy of solvent in the gas phase. Additionally, the combined binding energy of solvent and TEP on Pd(111) (Fig. 5e) is defined as:

$$BE_{solv} = \frac{E[N_{solv} + (4 - N_{solv})_{TEP}@Pd(111)] - E[Pd(111)] - N_{solv}E_{solv} - (4 - N_{solv})E_{TEP}}{4} \quad (11)$$

Where E_{TEP} is the energy of TEP in the gas phase and other quantities are as defined previously.

Results and Discussion

Solvent coordination effects on synthesis kinetics using in-situ SAXS

To provide insights on the effect of the solvents on the synthesis kinetics and final nanoparticle size, we conducted in-situ SAXS measurements in all solvents. The first observation is that the final Pd nanoparticle size increased in the order of toluene < piperidine < 3,4-lutidine < pyridine for both TOP : Pd = 1 and 2 as shown in Fig. 1 and S1, respectively (see representative SAXS spectra in Fig. S2 and S3 for TOP : Pd = 1, TEM images and SAXS spectra in Fig. S4 and Fig. S5 for TOP : Pd = 2 in ESI). We note that the effect of exposure to the X-ray beam was negligible since no reaction was triggered under beam exposure (see Fig. S6), and the final nanoparticle size from in-situ and ex-situ SAXS measurements was similar (see Fig. S7). The results in Fig. 1 show that the nanoparticle size in toluene (1.4 nm) was much smaller than in N-coordinating solvents, piperidine (2.7 nm), 3,4-lutidine (3.5 nm) and pyridine (4.8 nm), indicating that the solvents have a strong effect on the nucleation and growth rates.

An advantage of the in-situ SAXS measurements is the ability to capture both the time evolutions of the number of nanoparticles and the nanoparticle size (Fig. S8), which allows the extraction of both nucleation and growth rates (Fig. S9, see details in ESI and previous studies^{48, 55}). We note that based on our estimation of the rates of reaction and diffusion (see details in ESI and our previous studies⁵⁶), the nanoparticle synthesis

reactions in our specific system are kinetically controlled rather than diffusion controlled. To directly identify the effect of

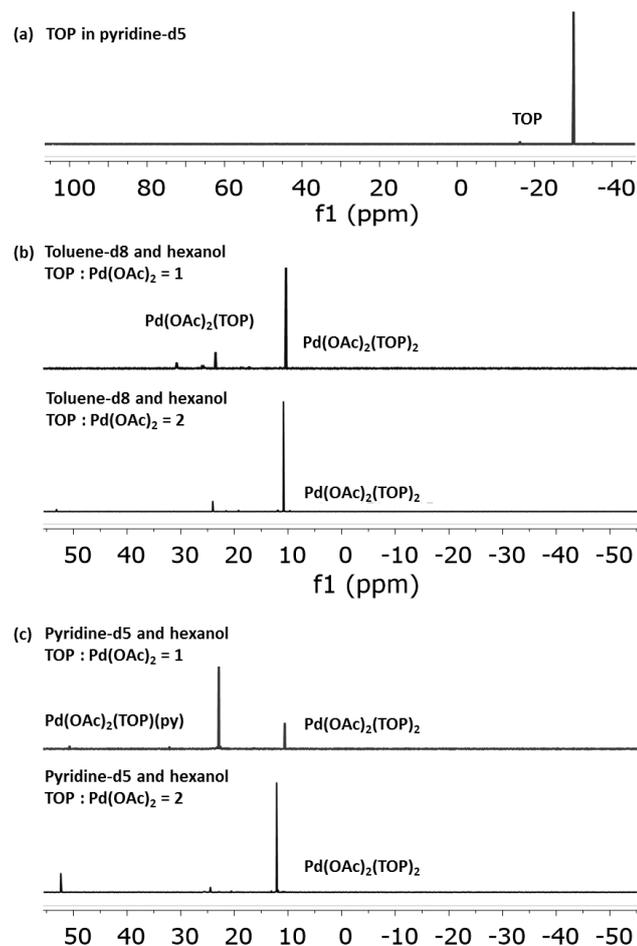


Figure 2 Gibbs free energy of different Pd complexes from DFT calculations. Example pyridine-containing precursor structures are shown as insets. Color code of atoms: Pd - teal, N - light blue, H - white, O - red, P - magenta, and C - gray. All Pd complex images are in Fig. S18.

solvents on the synthesis kinetics, it is important to compare the nucleation and growth rates in the kinetic regime at similar extent of reaction (i.e. low conversion of Pd acetate into nanoparticles, see Methods for details on how conversion is calculated) instead of at the same reaction time. At 5% and 10% extents of reaction (Fig. S10 and Fig. 1, respectively), the nucleation rate decreases and the growth rate increases in the following order: toluene, piperidine, 3,4-lutidine and pyridine which is consistent with the increasing trend of the final nanoparticle size.

The results in Fig. 1 indicate that in the order of toluene < piperidine < 3,4-lutidine < pyridine, growth on the existing nanoparticles becomes more preferred compared with forming new nuclei. This can be better seen in Fig. S11 where the ratio of growth-to-nucleation rates increases in the same solvent order of toluene < piperidine < 3,4-lutidine < pyridine up to 40% extent of reaction (highest conversion for reliable extraction of rates, in particular nucleation). The inhibited nucleation, and enhanced growth (likely autocatalytic as shown previously for toluene and pyridine^{48, 55}) can be due to solvent-dependent

precursor reactivity and/or surface properties of the nanoparticles (including coverage of TOP and/or solvent). To determine the possible role(s) of the solvent, we investigated the interactions of coordinating species (TOP and solvents) with Pd acetate and Pd nanoparticle surface under different solvent environments as discussed below.

Pd pre-reduction complexes in different solvents: ³¹P NMR and XAFS

To determine the initial Pd-acetate-TOP pre-reduction complex formed in the different solvents, we characterized Pd acetate in toluene and pyridine (without hexanol due to the instability of Pd acetate in hexanol without TOP), and the binding complexes of Pd(OAc)₂ with TOP in solvent-hexanol mixtures using ³¹P NMR and EXAFS. The TOP-Pd acetate binding complexes identified from the NMR and EXAFS experiments are representative of the initial pre-reduction precursor present during the synthesis, since the solutions (for all characterizations and synthesis) were freshly prepared and used within minutes. Modeling of the EXAFS spectra of Pd acetate dissolved in pyridine show that Pd is present, on average, as Pd(OAc)₂(py)₂, while in toluene it is present as trimers, Pd₃(OAc)₆ (see EXAFS results in Fig. S12 and Table S1).

³¹P NMR spectra of Pd(OAc)₂ dissolved in toluene-d₈ and pyridine-d₅, each with 1:1 hexanol at TOP : Pd molar ratio of 1 and 2 (Fig. 2), indicate that all the added TOP is bound with Pd(OAc)₂ or is converted to other species as evidenced by the absence of a free TOP peak (Fig. 2a). At TOP : Pd molar ratio of 2, the major peaks in toluene-d₈-hexanol at 10.4 ppm and pyridine-d₅-hexanol at 10.6 ppm can be assigned to the trans-Pd(OAc)₂(TOP)₂ complex as we previously reported for Pd(OAc)₂ dissolved in toluene-d₈ (peak at 10.3 ppm)⁵⁷. The small shift is due to the different solvents as previously reported for the divalent trans-Pd(OAc)₂(PPh₃)₂ complex in THF (14.48 ppm)⁵⁸ and in DMF (15.08 ppm)⁵⁹. At TOP : Pd molar ratio of 1, the major peak in pyridine-d₅-hexanol is at 22.9 ppm while in toluene-d₈-hexanol the major peak remains at 10.4 ppm with a

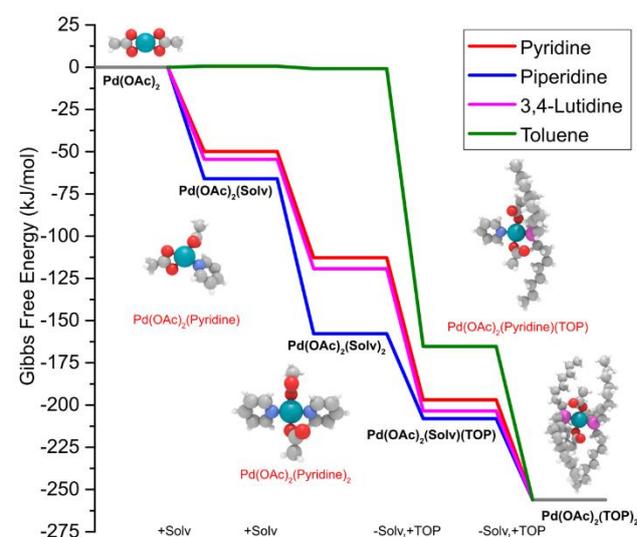


Figure 3 ³¹P NMR spectra of (a) pure TOP in pyridine-d₅ and Pd(OAc)₂ with 1 and 2 equivalent moles of TOP in (b) 10 mM Pd(OAc)₂, toluene-d₈ : hexanol = 1 : 1 and (c) 10 mM Pd(OAc)₂, pyridine-d₅ : hexanol = 1 : 1. The chemical shift of TOP in toluene-d₈ is at -31.5 ppm and in pyridine-d₅ is at -30.0 ppm.

small peak at 23.5 ppm. Moschetta et al.⁶⁰ reported that, in the same solvent, the peak for trans-PdCl₂(PPh₃)(solv) was at ca. 8-10 ppm higher (downfield) than the peak for the divalent trans-PdCl₂(PPh₃)₂ (e.g. in acetonitrile, 31.2 ppm for PdCl₂(PPh₃)(MeCN) vs. 23.4 ppm for PdCl₂(PPh₃)₂). Similarly, our ³¹P NMR spectra (Fig. S13) of the binding of TOP with PdCl₂ in pyridine-d₅ at TOP : Pd ratio of 1 and 2 showed peaks at 29.0 ppm and 11.8 ppm corresponding to trans-PdCl₂(TOP)(py) and trans-PdCl₂(TOP)₂, respectively. Therefore, we assign the peak at 22.9 ppm in pyridine-d₅-hexanol to the trans-Pd(OAc)₂(TOP)(py) complex and the peak at 23.5 ppm in toluene-d₈-hexanol tentatively to Pd(OAc)₂(TOP)(Tol). Our EXAFS modeling results of the coordination of Pd(OAc)₂ to TOP in pyridine and toluene (Fig. S14-S16 and Table S2) are consistent with the ³¹P NMR results. In pyridine, Pd was found to be coordinated, on average, with 1 and 2 TOP molecules at TOP : Pd molar ratios of 1 and 2 respectively. In toluene, Pd was coordinated to 2 TOP molecules (on average) at TOP : Pd molar ratio of 2.

Our ³¹P NMR results indicate that the major Pd complex in pyridine changes from trans-Pd(OAc)₂(TOP)(py) to trans-Pd(OAc)₂(TOP)₂ as the TOP : Pd ratio increases from 1 to 2. On the other hand, the major complex in toluene is trans-Pd(OAc)₂(TOP)₂ at both TOP : Pd ratios. The difference in the relative amounts of the complexes formed in toluene compared with pyridine is because Pd acetate is present as monomers in pyridine while it is present mostly as trimers in toluene as shown by our EXAFS results (see Fig. S12 and Table S1) and previous work⁵⁷. In toluene each Pd₃(OAc)₆ trimer requires 6 TOP molecules per trimer (2 TOP per Pd center) to break into monomers at room temperature as we previously reported⁵⁷ (see also Tables S1-2). However, dissociation of a TOP ligand from trans-Pd(OAc)₂(TOP)₂ is energetically unfavorable⁵⁷ which is why trans-Pd(OAc)₂(TOP)₂ is the major complex in toluene-hexanol at TOP : Pd ratio = 1. The results indicate that at TOP : Pd molar ratio of 1, TOP would bind to ca. half the Pd acetate forming trans-Pd(OAc)₂(TOP)₂ while the remaining Pd acetate trimers are not bound to TOP and do not appear as phosphine bonded complexes in the ³¹P NMR spectrum. However, we note that once the temperature increases during the nanoparticle synthesis experiments, the remaining trimers in toluene would dissociate into monomers and all the Pd acetate would be available for TOP binding.

In summary, the NMR and EXAFS results indicate that in toluene, pyridine and by extension the other solvents, a mixture of complexes are present in solution and their relative amounts depend on the thermodynamic equilibrium (dictated by concentrations and temperature in each solvent). Based on the EXAFS and NMR results, we anticipate that at the nanoparticles synthesis conditions, the major pre-reduction Pd complexes present in solution are Pd(OAc)₂(TOP)(solv) and Pd(OAc)₂(TOP)₂ at TOP : Pd molar ratio of 1 and 2 respectively, while Pd(OAc)₂(solv)₂ is a minority species (solv represents a solvent molecule).

Solvent effect on speciation and reducibility of Pd complexes using DFT

The NMR and EXAFS results show that N-coordinating solvents act as ligands and bind to Pd acetate. The coordination of the solvents with Pd acetate could modify the reactivity of the resulting complexes depending on the solvent electron

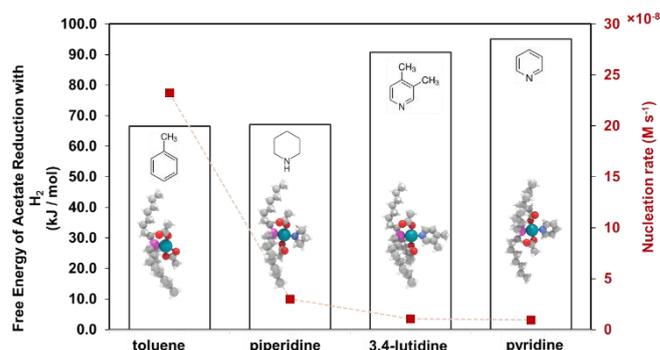
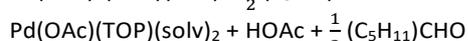
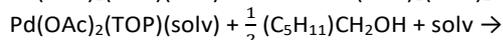
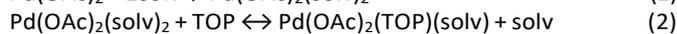


Figure 4 DFT calculated Gibbs free energy of reduction (with H₂) of Pd(OAc)₂(TOP)(solv) (and Pd(OAc)₂(TOP) in toluene), and nucleation rate at 10% conversion of Pd complex in each solvent. Color code of atoms: Pd – teal, N – light blue, H – white, O – red, P – magenta, and C – gray.

donating ability⁶¹ and binding stoichiometry.^{58, 62, 63} Such complexes, which are discrete molecular precursors different from both the initial reagents and final nanoparticles, can affect the nanoparticle formation mechanisms⁶⁴⁻⁶⁶. Therefore, we performed density functional theory calculations to provide details on the Pd complexes formed in the different solvents, their concentrations at equilibrium, as well as reducibility. Below we list the possible initial binding, solvent-TOP exchange and reduction reactions, where solv represents the solvent molecule:



The equations represent: Eq. 1, solvent binding to Pd(OAc)₂ (i.e. Pd(OAc)₂ dissolving in each solvent); Eq. 2, first TOP ligand substituting solv in Pd(OAc)₂(solv)₂; Eq.3, second (subsequent) TOP substituting solv in Pd(OAc)₂(solv)(TOP); Eq. 4, potential initial reduction reaction by hexanol of the Pd Pd(OAc)₂(solv)(TOP) complexes through acetate dissociation. First, to investigate the complex stability and speciation, we calculated the Gibbs free energy for the formation of all the possible unreduced Pd complexes before the addition of hexanol (Fig. 3). Compared with Pd(OAc)₂, the Gibbs free energies of Pd complexes are in the following order (lowest to highest free energy): Pd(OAc)₂(TOP)₂ < Pd(OAc)₂(solv)(TOP) < Pd(OAc)₂(solv)₂ < Pd(OAc)₂(solv) < Pd(OAc)₂. The complex is readily bound by the solvents (reacts exothermically) except for toluene (consistent with the stability of Pd acetate trimers in toluene)⁵⁷ and the solvents are favorably displaced with the addition of TOP. Although the Gibbs free energies reported in Fig. 3 are all calculated from gas phase calculations, we note that when implicit solvent is included the trends in binding strength do not change as seen in Fig. S17. Further, a recent benchmark study on Pd-phosphine complexes found ideal gas rigid rotator harmonic oscillator approach with solvent effects yielded accurate free energies of reaction.⁶⁷ Overall, this shows our gas-phase free energy calculations should at least trend with the equivalent solvent-phase reactions.

Table 1 Mole fraction of different precursor complexes (speciation) existing in different solvents based on equilibrium thermodynamics calculations from DFT at TOP : Pd = 1, T = 293.15 K.

Solvent	mol % Pd(OAc) ₂ (TOP)(solv)	mol % Pd(OAc) ₂ (TOP) ₂	mol % Pd(OAc) ₂ (solv) ₂
Toluene	96.00	2.00	2.00
Pyridine	98.70	0.65	0.65
3,4-Lutidine	99.70	0.15	0.15
Piperidine	47.31	26.35	26.35

The nucleation is dependent on the reduction rate which is affected by the reducibility of the Pd pre-reduction complexes in solution. Thus, to estimate the major pre-reduction complex, we performed DFT calculations on the free energies of solvent coordination of Eq. 1, as well as equilibrium thermodynamics of Eq. 2 and 3. From the DFT-based thermodynamic equilibrium results presented in Table 1 for TOP : Pd = 1 (see Table S3 for equilibrium calculation details and results for TOP : Pd = 2), the most abundant complex in all solvents is trans-Pd(OAc)₂(TOP)(solv) (and cis-Pd(OAc)₂(TOP) in toluene).

Therefore, our next step was to address the reducibility of the most abundant complexes by calculating their reduction Gibbs free energies, Pd(OAc)₂(TOP)(solv) $\xrightarrow{+\frac{1}{2}H_2, -HOAc}$ Pd(OAc)(TOP)(solv). Our analysis ignores hexoxy formation or hexanol displacement of ligands, since hexoxy displacement of acetate and hexanol displacement of ligands were both found to be endothermic (see Table S4 and Table S5). We used H₂ as the reference reducing agent as the exact reduction byproducts (potential reduction reaction in Eq. 4) are unknown and using H₂ as a common reference allows us to calculate consistent trends in reduction affinity. The results in Fig. 4 show that without the solvent coordination in toluene, the Gibbs free energy of Pd(OAc)₂(TOP) reduction is the most favorable.

In addition, as Fig. 4 demonstrates, the binding of more basic N-coordinating solvents makes the reduction affinity of the complexes more endothermic (piperidine < 3,4-lutidine < pyridine). For reactions in the gas phase, liquid phase and on surfaces, the activation energies for a series of reactions have been shown to be linearly correlated to the reaction enthalpies (or free energies) through Brønsted-Evans-Polanyi (BEP) relationship.⁶⁸⁻⁷² Therefore, an increase in the reduction free energy (more endothermic) is expected to increase the activation energy of reduction and consequently result in lower reduction and nucleation rates. Consistent with this argument, the DFT calculated trend of thermodynamic reducibility presented in Fig. 4, toluene (lowest reduction free energy, highest reducibility) > piperidine > 3,4-lutidine > pyridine (highest reduction free energy, lowest reducibility), appears to be correlated with the trend in nucleation rates measured experimentally (Fig. 1 and also shown in Fig. 4). Furthermore, we observed that the same trends in reduction free energy with different solvents also hold for the partially-reduced precursors (i.e. further reduction of Pd(OAc)(TOP)(solv)₂, Fig. S19). Beyond the reducibility preference of the predominant precursor complex in Fig. 4, the weak binding (non-coordination) of toluene to the precursor complexes likely contributes to faster (less-inhibited) nucleation rates. Thus, the weak binding of toluene to the precursor complexes may have a dual impact on

the nucleation rates of Pd nanoparticles, resulting in the especially high nucleation rate observed experimentally.

Solvent effect on Pd nanoparticle surface growth

The growth rate is affected by the binding and coverage of ligands on the nanoparticle surface. As the ligand coverage increases, the number of available sites for growth becomes limited, and the growth rate on the nanoparticle would decrease. The relative coverage of the solvent and TOP on the surface of nanoparticles is dynamic throughout a nanoparticle synthesis and depends on several factors including their respective concentrations and binding energies. Thus, periodic DFT calculations were conducted to compare binding energies of the solvent molecules to Pd(111) surface partially covered by triethylphosphine (TEP, as an approximation of TOP to reduce computational cost). The Pd(111) facet was selected as it is one of the most abundant facets in previous calculations⁷³ (even for

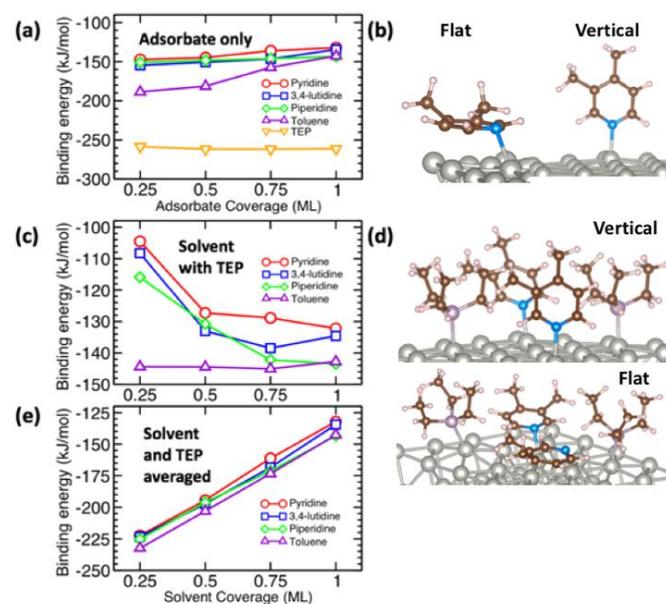


Figure 5 Binding energy (BE) per mole of solvent and/or TEP with coverage ranging from 0.25 monolayer (ML) to 1 ML. BE is averaged over vertical and flat binding configurations for pyridine, 3,4-lutidine, and piperidine (toluene only binds flat). (a) BE of solvents and TEP as a function of coverage on clean Pd(111). (b) Examples of vertical and flat binding of 3,4-lutidine on clean Pd. (c) BE of solvents as a function of coverage where TEP occupies remaining sites. (d) Examples of vertical and flat binding of 3,4-lutidine where TEP occupies remaining sites. (e) Combined BE of solvents and TEP as a function of solvent coverage, where TEP occupies remaining sites.

small nanoparticle sizes, 1.8-3.0 nm)⁷⁴ and experiments⁷⁵ as also seen from our high resolution TEM images in Fig. S20.

We calculated the binding energies of all solvents in both vertical and flat configurations with and without TEP competing for binding sites as a function of solvent monolayer fraction (Fig. 5, S21, S22, S24, S25). Flat binding (Fig. S24) is in general more exothermic than vertical binding (Fig. S25), though vertical-bound solvents better protect the nanoparticle surface by allowing more adsorbates per unit surface area whilst exhibiting comparable binding energies. However, considering either vertical or flat configurations in isolation does not capture the full physical picture. Dynamic fluctuation between both configurations is possible on the nanoparticle surface. To account for the collective solvent configurational variation, we present in Fig. 5a,c the average binding energy of both vertical and flat configurations at each solvent coverage fraction. The results in Fig. 5a show that TEP has the most negative binding energy, indicating the strongest binding affinity and consequently the tendency toward high surface coverage at equilibrium, see discussion below. Similarly, if we consider the combined molar binding energy of all adsorbates (averaged over TEP and solvent as a function of solvent coverage), we observe a monotonic increase in binding strength (more negative binding energy) as TEP replaces solvent on the Pd surface as shown in Fig. 5e. What is remarkable is that the configurationally-averaged solvent binding energies when co-adsorbed with TEP (Fig. 5c,e) follow the same order as the experimental growth rate: toluene (strongest binding, slowest growth) > piperidine > 3,4-lutidine > pyridine (weakest binding, fastest growth) as shown in Fig. 6 (with one exception for the case of 0.5 ML solvent, 0.5 ML TEP where the order of piperidine and 3,4-lutidine is reversed). We note in Fig. S23 that overall similar binding trends between the ligand/solvents on Pd(111) appear to hold on the surface of a small 13-atom Pd nanoparticle, which indicates similar ligand-dependent binding trends may occur on different facets of the Pd nanoparticles present during growth.

Our results show that in the presence of solvents, despite their less exothermic binding energy compared with TOP (Fig. 5), the nanoparticle surface will be covered by both the solvents and TOP. This is due to the much higher concentration of the solvents and their less bulky structures compared with TOP allowing co-adsorption, especially for N-coordinating solvents through vertical binding. Therefore, regardless of whether the coverage of TOP on the surface of the nanoparticles is at equilibrium (high coverage), or if the coverage of TOP is low at early times and increases with time (as shown from our previous in-situ EXAFS and kinetic modeling^{37, 48} studies and reports by other groups⁷⁶⁻⁷⁸), the solvent will act as a second ligand that helps passivate the surface. Since the solvents bind significantly weaker than TOP (more labile than TOP) as shown in Fig. 5, S21-22, the sites occupied by the solvent will be less effectively passivated and allow adsorption of Pd adatoms and growth of the nanoparticle (the weaker the binding of the solvent, the less effective passivation and faster growth). This can be also explained in terms of adsorption/desorption rates of the solvents. Assuming a Brønsted-Evans-Polanyi (BEP) relationship

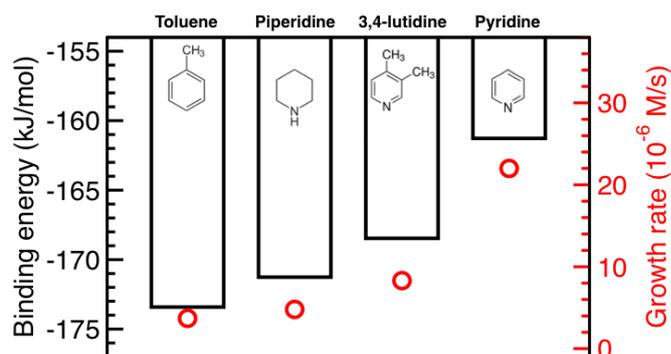


Figure 6 Combined binding energies of 0.25 ML TEP with 0.75 ML of toluene, piperidine, 3,4-lutidine, or pyridine on Pd(111) as also shown in Fig. 5e (black bars) and the growth rates at 10 % Pd conversion (red circles).

between the solvent binding energy and its desorption/adsorption rates at the nanoparticle surface, weaker binding solvents (e.g. pyridine) would show higher rates of desorption, which creates open sites at the nanoparticle surface leading to faster nanoparticle growth. This is consistent with the increase in the growth rate in the same order as the decrease in solvent binding energy shown in Fig. 6 (weaker binding energy, i.e. less negative, of solvent or combined solvent-TEP results in less effective passivation and faster growth).

Insights on deconvoluting the effects of solvents on nucleation and growth rates

The final size of the nanoparticles is controlled by the relative nucleation and growth rates which are affected by the precursor reactivity and ligand capping on the nanoparticle surface, respectively. For example, introducing stronger binding ligands was used to stabilize the ligated-metal complex and inhibit the nucleation, leading to larger nanoparticle size.^{8, 12, 13, 16} Other studies showed that a stronger capping ligand or higher ligand to metal ratio lead to smaller nanoparticle size which was attributed to slower surface growth (e.g. Au,⁴⁹ Pd,^{25, 37, 48} Pt,^{79, 80} and quantum dots⁵¹). However, the reported trends were mostly qualitative and relied on correlations with the final nanoparticle size. Additionally, since the ligand and/or solvent can play more than one role⁸¹ by binding to both, the metal precursor and nanoparticle surface, the deconvolution of these two factors is necessary to understand the effects on nucleation and growth kinetics. In this study, we used solvents of different coordinating abilities to investigate the effects on the reducibility of Pd precursors, ligand capping on the nanoparticle surface and provide mechanistic details on the synthesis kinetics of Pd colloidal nanoparticles. We directly correlate the ligated-metal complex reduction and ligand-surface binding thermodynamics (free energy of reduction and binding energies, respectively) calculated using DFT to the nucleation and growth rates measured using in-situ SAXS. The results show that solvents (and ligands in general) can be used to tune the reactivity of the metal-ligand pre-reduction complexes. More importantly, the reduction free energy of the metal-ligand pre-

reduction complex in each solvent was found to correlate with the measured nucleation rates (Fig. 4). Furthermore, the solvent is shown to affect the effective capping on the nanoparticle surface (and possibly the reactivity of surface atoms) which was reflected in a correlation between the solvent/ligand surface binding energy and the measured growth rates (Fig. 6). The results show the importance of understanding the distinct roles ligands and solvents play in affecting the nucleation and growth rates during the synthesis of colloidal nanoparticles. Additionally, the results suggest that general correlations, i.e. Brønsted-Evans-Polanyi relationships, likely exist between thermodynamics of metal-ligand binding and the kinetics of nucleation and growth. We note that although microkinetic models have been attempted in other works studying nanoparticle growth,^{82, 83} such studies necessarily require heavy assumptions due to the complex chemical environment present during growth. Rather than focusing effort on addressing the complex chemical environment during growth and get accurate nanoparticle growth kinetics from our theoretical predictions, here, we identify simple theory-derived quantities that correlate with the experimental kinetics as a means of understanding factors dictating nanoparticle nucleation and growth. Our future efforts will focus on investigating other ligands and metals to determine if they follow similar correlation between the metal-ligand thermodynamic interactions and nucleation and growth rates. Such correlations would enable the selection of ligands and/or solvents, *a priori*, to synthesize nanoparticles with specific sizes.

Conclusions

We investigated the effect of solvent coordinating ability on the final size of colloidal Pd nanoparticles and provide a direct correlation between the metal-ligand-solvent interactions and the synthesis kinetics using in-situ SAXS, XAFS, NMR and DFT calculations. The nanoparticle size can be tuned from 1.4-5.0 nm with narrow size distribution solely through changing the solvent coordinating ability (toluene vs. N-coordinating solvents). Our in-situ kinetic measurements indicate slower nucleation and faster growth in N-coordinating solvents compared with toluene. Through ³¹P NMR characterization and theoretical calculations of the Pd pre-reduction complexes, we show that the reduction free energy of the most abundant complexes increases with the solvent electron donating ability. The trend in the DFT calculated free energy of reduction is shown to correlate with the nucleation rates measured experimentally (increase in reduction free energy corresponds to lower nucleation rate), which is consistent with a Brønsted-Evans-Polanyi relationship. Similarly, the experimental growth rates are correlated with the solvent-surface binding energies (weaker binding corresponds to faster growth). Our study demonstrates the importance of solvents in colloidal synthesis of nanoparticles and their role in affecting the synthesis kinetics. The results provide insights on the microscopic relationship between the metal-ligand-solvent interactions and nucleation and growth rates and introduces a promising method for tuning the size of colloidal metal nanoparticles.

Conflicts of interest

The authors have no conflict of interest to report.

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Electronic supplementary information (ESI) available

Estimations of the number of nanoparticles and extent of reaction, extracting the rates of nucleation and growth from experimental data, precursor speciation calculations from density functional theory (DFT), representative SAXS spectra, TEM images of nanoparticles, in-situ SAXS measurement fitting results, kinetics extracted from in-situ SAXS, XANES and XAFS results, ³¹P NMR spectra, mole fraction of different precursor complexes from DFT calculations, rendered images of all precursors, Gibbs free energy of H₂ reduction of Pd(OAc)(TOP)(solv)₂, DFT calculation results and images for solvent-nanoparticle / Pd(111) surface binding.

Notes and references

- K. M. Bratlie, H. Lee, K. Komvopoulos, P. Yang and G. A. Somorjai, *Nano Lett.*, 2007, **7**, 3097-3101.
- K. Sugawa, H. Tahara, A. Yamashita, J. Otsuki, T. Sagara, T. Harumoto and S. Yanagida, *ACS Nano*, 2015, **9**, 1895-1904.
- R. Narayanan and M. A. El-Sayed, *The Journal of Physical Chemistry B*, 2005, **109**, 12663-12676.
- L. Zhang, H. Jing, G. Boisvert, J. Z. He and H. Wang, *ACS Nano*, 2012, **6**, 3514-3527.
- J. D. Aiken and R. G. Finke, *J. Mol. Catal. A: Chem.*, 1999, **145**, 1-44.
- Z. Zhou, X. Liu, L. Yue and I. Willner, *ACS Nano*, 2018, **12**, 10725-10735.
- Y. Wang, S. Xie, J. Liu, J. Park, C. Z. Huang and Y. Xia, *Nano Lett.*, 2013, **13**, 2276-2281.
- L. Lian, Y. Xia, C. Zhang, B. Xu, L. Yang, H. Liu, D. Zhang, K. Wang, J. Gao and J. Zhang, *Chem. Mater.*, 2018, **30**, 982-989.
- L. Wu, H. Lian, J. J. Willis, E. D. Goodman, I. S. McKay, J. Qin, C. J. Tassone and M. Cargnello, *Chem. Mater.*, 2018, **30**, 1127-1135.
- N. Zheng, J. Fan and G. D. Stucky, *J. Am. Chem. Soc.*, 2006, **128**, 6550-6551.
- A. J. Bicchi and R. E. Schaak, *ACS Nano*, 2015, **9**, 1707-1720.
- N. Ortiz and S. E. Skrabalak, *Angewandte Chemie-International Edition*, 2012, **51**, 11757-11761.
- W. W. Yu, Y. A. Wang and X. Peng, *Chem. Mater.*, 2003, **15**, 4300-4308.
- B. Hu, K. Ding, T. Wu, X. Zhou, H. Fan, T. Jiang, Q. Wang and B. Han, *Chem. Commun.*, 2010, **46**, 8552-8554.
- H. Qian, Y. Zhu and R. Jin, *ACS Nano*, 2009, **3**, 3795-3803.
- Z. Q. Yang and K. J. Klabunde, *J. Organomet. Chem.*, 2009, **694**, 1016-1021.
- Y. Xia, X. Xia and H. C. Peng, *J. Am. Chem. Soc.*, 2015, **137**, 7947-7966.
- A. J. Bicchi and R. E. Schaak, *ACS Nano*, 2011, **5**, 8089-8099.
- J. Song, D. Kim and D. Lee, *Langmuir.*, 2011, **27**, 13854-13860.
- Y. Jiang, Y. Y. Huang, H. Cheng, Q. H. Liu, Z. Xie, T. Yao, Z. Jiang, Y. Y. Huang, Q. Bian, G. Q. Pan, Z. H. Sun and S. Q. Wei, *Journal of Physical Chemistry C*, 2014, **118**, 714-719.
- W. Yu, W. Tu and H. Liu, *Langmuir.*, 1999, **15**, 6-9.
- W. Yu, M. Liu, H. Liu and J. Zheng, *J. Colloid Interface Sci.*, 1999, **210**, 218-221.
- C. B. Williamson, D. R. Nevers, T. Hanrath and R. D. Robinson, *J. Am. Chem. Soc.*, 2015, **137**, 15843-15851.
- S.-W. Kim, J. Park, Y. Jang, Y. Chung, S. Hwang, T. Hyeon and Y. W. Kim, *Nano Lett.*, 2003, **3**, 1289-1291.
- T. Teranishi and M. Miyake, *Chem. Mater.*, 1998, **10**, 594-600.
- A. P. LaGrow, B. Ingham, M. F. Toney and R. D. Tilley, *J. Phys. Chem. C*, 2013, **117**, 16709-16718.
- A. Henglein and M. Giersig, *The Journal of Physical Chemistry B*, 1999, **103**, 9533-9539.
- D. D. Vaughn, S.-I. In and R. E. Schaak, *ACS Nano*, 2011, **5**, 8852-8860.
- M. J. Hostetler, J. E. Wingate, C.-J. Zhong, J. E. Harris, R. W. Vachet, M. R. Clark, J. D. Londono, S. J. Green, J. J. Stokes, G. D. Wignall, G. L. Glish, M. D. Porter, N. D. Evans and R. W. Murray, *Langmuir*, 1998, **14**, 17-30.
- C. Evangelisti, N. Panziera, A. D'Alessio, L. Bertinetti, M. Botavina and G. Vitulli, *J. Catal.*, 2010, **272**, 246-252.
- H. P. Choo, K. Y. Liew and H. Liu, *J. Mater. Chem.*, 2002, **12**, 934-937.
- N. Sakamoto, M. Harada and T. Hashimoto, *Macromolecules*, 2006, **39**, 1116-1124.
- J. Wang, R. E. Winans, S. L. Anderson, S. Seifert, B. Lee, P. J. Chupas, Y. Ren, S. Lee and Y. Liu, *J. Phys. Chem. C*, 2013, **117**, 22627-22635.
- J. Park, K. An, Y. Hwang, J. G. Park, H. J. Noh, J. Y. Kim, J. H. Park, N. M. Hwang and T. Hyeon, *Nat Mater*, 2004, **3**, 891-895.
- X. Yin, M. Shi, J. Wu, Y.-T. Pan, D. L. Gray, J. A. Bertke and H. Yang, *Nano Lett.*, 2017, **17**, 6146-6150.
- L. Scarabelli, M. Coronado-Puchau, J. J. Giner-Casares, J. Langer and L. M. Liz-Marzán, *ACS Nano*, 2014, **8**, 5833-5842.
- A. M. Karim, N. Al Hasan, S. Ivanov, S. Seifert, R. T. Kelly, N. G. Hallfors, A. Benavidez, L. Kovarik, A. Jenkins, R. E. Winans and A. K. Datye, *J. Phys. Chem. C*, 2015, **119**, 13257-13267.
- P. Abellan, L. R. Parent, N. Al Hasan, C. Park, I. Arslan, A. M. Karim, J. E. Evans and N. D. Browning, *Langmuir.*, 2016, **32**, 1468-1477.
- S. Mozaffari, W. Li, C. Thompson, S. Ivanov, S. Seifert, B. Lee, L. Kovarik and A. M. Karim, *Nanoscale*, 2017, **9**, 13772-13785.
- S. Mozaffari, W. Li, C. Thompson, S. Ivanov, S. Seifert, B. Lee, L. Kovarik and A. M. Karim, *JoVE*, 2018, DOI: doi:10.3791/57667, e57667.
- J. Polte, T. T. Ahner, F. Delissen, S. Sokolov, F. Emmerling, A. F. Thünemann and R. Kraehnert, *J. Am. Chem. Soc.*, 2010, **132**, 1296-1301.
- M. Harada, N. Tamura and M. Takenaka, *J. Phys. Chem. C*, 2011, **115**, 14081-14092.
- M. A. Watzky and R. G. Finke, *J. Am. Chem. Soc.*, 1997, **119**, 10382-10400.
- V. K. LaMer and R. H. Dinegar, *J. Am. Chem. Soc.*, 1950, **72**, 4847-4854.
- C. Besson, E. E. Finney and R. G. Finke, *Chem. Mater.*, 2005, **17**, 4925-4938.
- S. R. K. Perala and S. Kumar, *Langmuir.*, 2013, **29**, 9863-9873.
- M. Harada and R. Ikegami, *Crystal Growth & Design*, 2016, **16**, 2860-2873.
- S. Mozaffari, W. Li, C. Thompson, S. Ivanov, S. Seifert, B. Lee, L. Kovarik and A. M. Karim, *Nanoscale*, 2017, **9**, 13772-13785.
- A. E. Saunders, M. B. Sigman and B. A. Korgel, *The Journal of Physical Chemistry B*, 2004, **108**, 193-199.
- C. R. Bullen and P. Mulvaney, *Nano Lett.*, 2004, **4**, 2303-2307.
- J. van Embden and P. Mulvaney, *Langmuir.*, 2005, **21**, 10226-10233.
- B. H. Wu, H. Y. Yang, H. Q. Huang, G. X. Chen and N. F. Zheng, *Chin. Chem. Lett.*, 2013, **24**, 457-462.
- E. Redel, P. Mirtchev, C. Huai, S. Petrov and G. A. Ozin, *ACS Nano*, 2011, **5**, 2861-2869.
- M. L. Personick and C. A. Mirkin, *J. Am. Chem. Soc.*, 2013, **135**, 18238-18247.
- S. Mozaffari, W. Li, C. Thompson, S. Ivanov, S. Seifert, B. Lee, L. Kovarik and A. M. Karim, *Journal of visualized experiments: JoVE*, 2018.
- S. Mozaffari, W. Li, M. Dixit, S. Seifert, B. Lee, L. Kovarik, G. Mpourmpakis and A. M. Karim, *Nanoscale Advances*, 2019, **1**, 4052-4066.
- W. Li, S. Ivanov, S. Mozaffari, N. Shanaiah and A. M. Karim, *Organometallics*, 2019, **38**, 451-460.
- C. Amatore, A. Jutand and M. A. M'Barki, *Organometallics.*, 1992, **11**, 3009-3013.
- J. A. Goodfellow, T. A. Stephenson and M. C. Cornock, *J. Chem. Soc., Dalton Trans.*, 1978, DOI: 10.1039/DT9780001195, 1195-1200.

60. E. G. Moschetta, K. M. Gans and R. M. Rioux, *J. Catal.*, 2013, **302**, 1-9.
61. P. R. Vashi and H. M. Marques, *J. Inorg. Biochem.*, 2004, **98**, 1471-1482.
62. C. Amatore and A. Jutand, *Acc. Chem. Res.*, 2000, **33**, 314-321.
63. C. Amatore, A. Jutand and A. Thuilliez, *Organometallics.*, 2001, **20**, 3241-3249.
64. L. E. Marbella, D. M. Chevrier, P. D. Tancini, O. Shobayo, A. M. Smith, K. A. Johnston, C. M. Andolina, P. Zhang, G. Mpourmpakis and J. E. Millstone, *J. Am. Chem. Soc.*, 2015, **137**, 15852-15858.
65. G. Mpourmpakis, S. Caratzoulas and D. G. Vlachos, *Nano Lett.*, 2010, **10**, 3408-3413.
66. X. Yin, X. Liu, Y.-T. Pan, K. A. Walsh and H. Yang, *Nano Lett.*, 2014, **14**, 7188-7194.
67. M. Besora, P. Vidossich, A. Lledós, G. Ujaque and F. Maseras, *The Journal of Physical Chemistry A*, 2018, **122**, 1392-1399.
68. M. G. Evans and M. Polanyi, *Trans. Faraday Society*, 1936, **32**, 1333-1360.
69. T. Bligaard, J. K. Nørskov, S. Dahl, J. Matthiesen, C. H. Christensen and J. Sehested, *J. Catal.*, 2004, **224**, 206-217.
70. J. K. Nørskov, F. Abild-Pedersen, F. Studt and T. Bligaard, *Proceedings of the National Academy of Sciences*, 2011, **108**, 937.
71. F. Calle-Vallejo, D. Loffreda, M. T. M. Koper and P. Sautet, *Nature Chemistry*, 2015, **7**, 403.
72. M. Dixit, X. Peng, M. D. Porosoff, H. D. Willauer and G. Mpourmpakis, *Catalysis Science & Technology*, 2017, **7**, 5521-5529.
73. M. Crespo-Quesada, A. Yarulin, M. Jin, Y. Xia and L. Kiwi-Minsker, *J. Am. Chem. Soc.*, 2011, **133**, 12787-12794.
74. H. Ramezani-Dakheel, P. A. Mirau, R. R. Naik, M. R. Knecht and H. Heinz, *Phys. Chem. Chem. Phys.*, 2013, **15**, 5488-5492.
75. J. M. Petroski, Z. L. Wang, T. C. Green and M. A. El-Sayed, *The Journal of Physical Chemistry B*, 1998, **102**, 3316-3320.
76. S. R. K. Perala and S. Kumar, *Langmuir.*, 2013, **29**, 9863-9873.
77. S. R. K. Perala and S. Kumar, *Langmuir.*, 2014, **30**, 12703-12711.
78. S. Lazzari, P. M. Theiler, Y. Shen, C. W. Coley, A. Stemmer and K. F. Jensen, *Langmuir.*, 2018, **34**, 3307-3315.
79. K. Wikander, C. Petit, K. Holmberg and M.-P. Pileni, *Langmuir*, 2006, **22**, 4863-4868.
80. M. Osmić, J. Kolny-Olesiak and K. Al-Shamery, *CrystEngComm*, 2014, **16**, 9907-9914.
81. N. Ortiz and S. E. Skrabalak, *Langmuir*, 2014, **30**, 6649-6659.
82. C. H. Turner, Y. Lei and Y. Bao, *Nanoscale*, 2016, **8**, 9354-9365.
83. X.-Y. Li, B. Zhu, R. Qi and Y. Gao, *Advanced Theory and Simulations*, 2019, **2**, 1800127.

