



A mechanical model reveals that non-axisymmetric buckling lowers the energy barrier associated with membrane neck constriction

Journal:	<i>Soft Matter</i>
Manuscript ID	SM-ART-07-2019-001494.R1
Article Type:	Paper
Date Submitted by the Author:	31-Oct-2019
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1 **A mechanical model reveals that non-axisymmetric buckling lowers** 2 **the energy barrier associated with membrane neck constriction**

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4 November 27, 2019

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13 **Abstract**

14 Membrane neck formation is essential for scission, which, as recent experiments on tubules have demon-
15 strated, can be location dependent. The diversity of biological machinery that can constrict a neck such as
16 dynamin, actin, ESCRTs and BAR proteins, and the range of forces and deflection over which they operate,
17 suggest that the constriction process is functionally mechanical and robust to changes in biological environment.
18 In this study, we used a mechanical model of the lipid bilayer to systematically investigate the influence of lo-
19 cation, symmetry constraints, and helical forces on membrane neck constriction. Simulations from our model
20 demonstrated that the energy barriers associated with constriction of a membrane neck are location-dependent.
21 Importantly, if symmetry restrictions are relaxed, then the energy barrier for constriction is dramatically lowered
22 and the membrane buckles at lower values of forcing parameters. Our simulations also show that constriction
23 due to helical proteins further reduces the energy barrier for neck formation compared to cylindrical proteins.
24 These studies establish that despite different molecular mechanisms of neck formation in cells, the mechanics
25 of constriction naturally leads to a loss of symmetry that can lower the energy barrier to constriction.

Significance statement

26 Membrane tubule constriction is a critical step of cellular membrane trafficking processes and is thought to be mechanically regulated. Mechanical modeling techniques employing the Helfrich Hamiltonian and axisymmetric continuum frameworks have previously described energy barriers to constriction as a function of location along a membrane tubule. Recent advances in numerical modeling using spline basis functions (Isogeometric Analysis) enable us to conduct our analyses of membrane mechanics in a generalized 3D framework. Here, we implement a novel 3D Isogeometric Analysis framework and juxtapose it against an axisymmetric model to study the influence of location, symmetry constraints and helical collars on the constriction pathway. We show that an unsymmetric, “crushed soda can” neck consistently displays a lower energy barrier than a symmetric neck.

27 **Introduction**

28 Many cellular transport processes involving the plasma membrane including different forms of endocytosis [1, 2, 3],
29 exocytosis [4, 5], and vesicle budding from intracellular organelles [6, 7] require mechanical deformation of the
30 cellular membranes. The generation of membrane curvature is essential to trafficking, and the morphology of
31 membranes has often been characterized as distinct shapes including U- and Ω -shaped bud profiles [8, 9, 10] and
32 tubulovesicular structures [11, 12]. The molecular mechanisms of these processes can be attributed to biochemical
33 components of the protein machinery involved [13, 14]. For example, in the case of clathrin-mediated endocytosis
34 (CME), more than 50 proteins are involved in regulating the different steps of membrane invagination such as
35 nucleation, cargo selection, coat assembly, neck formation and scission [3, 15, 16] and contribute to the robustness
36 and progression of endocytosis.

37 The formation of a membrane neck and scission are the last steps during many trafficking processes preceding
38 vesicle formation. This neck formation is mediated by multiple biochemical mechanisms including mechanoen-
39 zymes belonging to the dynamin family [17], helix insertion due to BAR domain proteins [18] and ESCRT proteins
40 [19, 20]. A common organizational feature of these different proteins is that they form helical assemblies at the
41 membrane neck through oligomerization [21, 22].

42 Studies using reconstituted systems of lipid tubules decorated with protein assemblies have identified certain ge-
43 ometric and mechanical features of scission. Notably, studies of dynamin-mediated scission [23, 24, 25], the
44 most investigated scission mechanism, have shown that the location of neck formation along a membrane tube,
45 membrane tension, and bending rigidity play important roles in membrane tube constriction and scission [26]. Col-
46 lectively, these studies support an emerging view that fundamental physical laws and geometric bounds confer a
47 universality on membrane constriction phenomena and scission.

48 Crucially, neck formation occurs at a length scale of < 10 nm, which is challenging to image even with high res-
49 olution electron tomography (ET) as radiation damage and low signal to noise ratio (SNR) can limit contrast [27].
50 Alternatively, equipped with extensive information from experiments such as those described above, mathemati-
51 cal and computational models can provide insight to the mechanics and energetics of membrane neck formation.
52 Almost all of these models are rooted in the Helfrich elastic energy framework [28]. The physical principles under-
53 lying the Helfrich model are simple enough – the elastic energy of membrane deformation depends primarily on the
54 curvatures of the membrane. Computational implementation of the governing equations resulting from this model,
55 however, remain extremely challenging (see [29] for a detailed review). Therefore, many studies have assumed an
56 axisymmetric configuration of the membrane for ease of computation [9, 10, 30, 31, 32, 33, 34].

57 In the most relevant of these studies to the present work, we and others have shown that a snap-through instability

58 governs the first energy barrier associated with the formation of a membrane neck during CME [9, 35, 36]. An
59 important limitation of the assumption of axisymmetry is that membrane deformation pathways associated with
60 neck constriction that may have lower symmetries are not accessible (Figure 1B) and helicoidal protein assemblies
61 [12, 37, 38] cannot be explicitly modeled.

62 In this study, we systematically investigate the energy barriers to constriction at different locations of a membrane
63 geometry with and without symmetry restrictions (Figure 1). Importantly, we tackle the challenging problem of
64 modeling non-axisymmetric membrane deformations with a benchmark comparison to axisymmetric modeling.
65 We use a minimal, but fundamental, model of collar pressure-mediated tube constriction to obtain insights from a
66 mechanical and energetic perspective. Using this model, we seek to answer the following foundational questions
67 for the broader field of membrane deformation processes: *First*, how does the local pre-existing curvature along
68 a tube influence the energy barrier associated with neck constriction? *Second*, how does relaxation of *a priori*
69 imposed symmetry restrictions impact the energy barriers associated with constriction of the neck? *And finally*,
70 how do cylindrical versus helical protein assemblies modulate this energy barrier?

71 To answer these questions, we have developed a computational framework for solving membrane mechanics prob-
72 lems on complex geometries using numerical techniques that exploit Galerkin methods, specifically Isogeometric
73 Analysis [39]. This framework draws upon recent far-reaching advances on the use of spline basis functions in com-
74 putational mechanics and brings them to the world of biological membranes, while building upon recent literature
75 on finite element modeling of liquid shells [40]. As a result, we can now investigate membrane deformation using
76 simulations of neck constrictions under conditions that are notably less restrictive than those adopted previously in
77 the literature (i.e. no enforced axis of symmetry). Importantly, this allows us to probe realistic helical constriction
78 pathways within a continuum framework, a different approach than recent efforts using coarse-grained modeling
79 [37]. Using this framework, we applied constriction pressures at three different locations along the membrane
80 tube (see Figure 2) – the ‘cap’ (positive mean and Gaussian curvature), ‘cylindrical tube’ (positive mean and zero
81 Gaussian curvature), and ‘base’ (Positive- negative mean and negative Gaussian curvature). Our simulations show
82 that the energy barriers associated with membrane neck constriction are indeed curvature-dependent, and therefore
83 location-dependent, regardless of symmetry restrictions. Most importantly, we show that access to less symmetric
84 shapes of membrane deformation lowers the energy barrier for scission considerably. These results suggest that
85 loss of symmetry of the membrane neck may be an important energetic feature of successful neck formation.

Table 1: Model parameter values

Parameter	Value	Reference
1. Boundary membrane tension (λ_0)	10^{-2} - 10^{-1} pN·nm ⁻¹	[51, 52]
2. Bending rigidity of bare membrane (κ)	320 pN·nm	[53]
3. Length scale for non-dimensionalization (R_0)	20 nm	[9]

86 Model development and simulations

87 Helfrich energy

88 The lipid bilayer is modeled as a thin elastic shell using the Helfrich energy [28] based on the assumption that the
 89 thickness of the bilayer is negligible compared to its radius of curvature [23, 41]. The Helfrich energy density is
 90 defined as

$$W = \kappa H^2 + \kappa_G K, \quad (1)$$

91 where κ is the bending rigidity, H is the mean curvature, K is the Gaussian curvature and κ_G is the Gaussian
 92 rigidity. Furthermore, we assume that the membrane is incompressible (*i.e* the membrane area is constant) [42]—a
 93 constraint that is implemented using a Lagrange multiplier field. Thus, while the Helfrich energy is defined entirely
 94 in terms of the geometry of the surface, the Lagrange multiplier, often interpreted as membrane tension [43, 44],
 95 is an important parameter that determines the minimum energy configuration. We ignore any fluid [45, 46] and
 96 friction [47, 48, 49] properties of the bilayer, guided by the dominance of unstable and stable equilibrium states
 97 over relaxation/rate processes. The augmented Helfrich Hamiltonian that is being minimized on the surface Ω ,
 98 including the Lagrange multiplier λ is given as [44, 45, 50]

$$E = \int_{\Omega} (\kappa H^2 + \kappa_G K + \lambda) dA. \quad (2)$$

99 **Simulations in axisymmetric coordinates**

100 In axisymmetry, the membrane is modeled using coordinates defined in Figure 1A. As the membrane tubule (Fig.
101 2E) has three distinct shape features (Fig. 2A - Cap, Tube and Base), local membrane geometries were modeled as
102 a hemispherical cap (Fig. 2B, Case 1), cylindrical tube (Fig. 2C, Case 2) and a curved base with negative Gaussian
103 curvature (Fig. 2D, Case 3). Cases 1 and 2 are constant mean curvature shapes and are solved as two-point
104 boundary value problems. Case 3 is a negative Gaussian curvature shape with an inflection point in mean curvature
105 with respect to the arc length and is solved as a three point boundary value problem. Case 4 includes local geometric
106 variations in both mean and Gaussian curvature and is solved as a three point boundary value problem. The third
107 point in these cases is an additional interface point enforced at the location of constriction, such that it satisfies
108 continuity requirements [54]. The resulting system of equations is solved using the partial differential equation
109 solution routines in Matlab, specifically bvp4c [54, 55]. Importantly, these equations are solved using both force
110 control (compute membrane shape for a certain applied force) and displacement control (compute applied force
111 for a certain membrane shape). These two approaches can lead to the same equilibrium membrane shape. In the
112 presence of membrane bending instabilities, displacement control can access regimes of the response curve that
113 force control cannot reach. However, this requires a precise prescription of the kinematic path. In order that a
114 system be free to find the lowest energy pathways through a region of instability in its energy landscape, it is
115 important that, while the force and displacement vary in a coupled manner, neither quantity be fully prescribed
116 [9, 56]. Parameters for the bending rigidity, membrane tension and non-dimensionalization length R_0 are specified
117 in Table 1. Details of the numerical methods are provided in the Supplementary Online Material (SOM).

118 **3D numerical model development and validation**

119 The membrane deformation problems considered in this paper can be modeled using classical thin shell theories
120 of mechanics. However, given the geometric complexity and the associated boundary conditions, analytical so-
121 lutions are inaccessible. Instead, we obtain three dimensional numerical solutions to the membrane deformation
122 problems using the framework of Isogeometric Analysis (IGA) [39]. An IGA method-based membrane mechanics
123 framework has been developed for this work, and is build on top of the PetIGA [57] open source library. In an IGA
124 approach, the membrane geometry is discretized using a spline mesh and the governing equations (Fig. 1B, see thin
125 shell formulation in the Supplementary Information) are converted to a nonlinear system of equations. This non-
126 linear system of equations is then solved to obtain the deformed membrane shape, and the related force and energy
127 metrics. Of importance to our central result is that this framework naturally admits both symmetric and asymmetric
128 deformation modes driven by the underlying physics. This framework has three key assumptions. First, a funda-
129 mental conjecture of the Helfrich model is that the characteristic length scales of the problem are much larger than

130 the thickness of the bilayer [28]. This assumption allows us to neglect the effect of transverse shear deformations
131 and consider the classical Kirchhoff–Love shell kinematics for thin shell geometries [58]. Second, numerical so-
132 lutions to the membrane shape equations (Equations (12) and (13)) in general coordinates are challenging because
133 of continuity requirements in the numerical scheme. We have overcome this challenge by adopting both B-Spline
134 basis functions, which allow high-order continuity, and the numerical framework of Isogeometric Analysis [39].
135 Finally, an inherent limitation of the Helfrich energy formulation in three dimensional simulations is the lack of
136 resistance to shear deformation modes. The zero energy modes corresponding to shear deformation are eliminated
137 in this framework by adding shear stabilization terms of smaller magnitude relative to the traditional bending terms
138 in the Helfrich energy [40], thus restoring stability to the numerical model. A companion manuscript (in prepara-
139 tion by the authors) describes the details of the mathematical methods and numerical formulation, and establishes
140 the validity of the computational framework by modeling a range of problems in membrane mechanics. Here, we
141 present a validation of the 3D computational framework by comparing the output from the simulation with a known
142 analytical solution of the classical tube pulling problem (Figure S1A, B). In addition to demonstrating good agree-
143 ment with the analytical solution, the 3D model also resolves the symmetric pathways of deformation if they are
144 indeed the energy minimizing modes (Figure S1A). Having validated the 3D numerical scheme, we then proceeded
145 to simulate the different cases shown in (Figure 2B-E) and compared them against axisymmetric pathways. We use
146 three key metrics to compare the two models – (1) the radial pinching load, represented by the collar pressure that
147 drives constriction, (2) structural stiffness of the membrane, defined as the slope of the load-displacement response,
148 and (3) membrane bending energy. We track these metrics for different pinching radii, which are defined as the
149 shortest distances between the membrane and the center of the necking region. For fully symmetric configurations
150 and those with lower symmetry, this distance is the radius of the smallest circle that can be fit in the necking region.

151 **Results**

152 We systematically investigated the role of preexisting curvature (varying with location on the membrane) in the
153 constriction process and the associated energy landscape using both the traditional axisymmetric calculations and
154 the 3D computational framework. The constriction process is modeled using a collar pressure (pN/nm^2) applied
155 onto a fixed membrane height (nm). In this study, we include the effect of the height of applied pressure by
156 reporting a force per unit length, or an effective surface pressure (pN/nm), as the product of the applied collar
157 pressure and the fixed height. Our main results can be summarized as follows – *first*, the energy landscape for
158 constriction depends on the preexisting curvature of the membrane; *second*, 3D modes of constriction with less
159 than full symmetry encounter lower energy barriers when compared to pathways of higher symmetry; and *finally*,
160 helical constriction modes can have the lowest energy barriers of all in 3D. We elaborate on these findings in detail

161 below.

162 **The energy barrier associated with constriction depends on preexisting membrane curvature**

163 We investigated the effect of local, preexisting curvature on the energy barrier associated with tubule constriction
164 in axisymmetry. We pulled out a membrane tube by applying an external axial force (f_{axial}) on a small patch of
165 the membrane to mimic a point load while maintaining a membrane tension of 0.2 pN/nm [59]. We then applied
166 a radial collar pressure at different locations on the tube (Figure 3A) while maintaining the membrane height, a
167 setup that can be generalized to *in-vitro* membrane tubules pulled by optical tweezers. In the absence of a fixed
168 height applied as a boundary condition, the membrane deforms freely in the axial direction at negligible collar
169 pressures (Figure S2). Results from our simulations show that pinching the tube at the cap (positive mean and
170 Gaussian curvature) and along the cylinder (positive mean and zero Gaussian curvature) results in similar force-
171 shape relationship (Figure 3D) and the cross section of the pinched profile remains circular by construction due to
172 the restriction of axisymmetry. (Figure 3B,C). Surprisingly, for the same range of collar pressure applied to the base
173 (positive-negative mean and negative Gaussian curvature), we observed the existence of a snap-through instability
174 as the membrane constriction progresses, as shown by the red line in Figure 3D. The dotted lines (Figure 3D, base)
175 are calculated using displacement control, i.e., compute the applied force given the membrane shape. However,
176 given our initial conditions and the mechanism of neck formation via increasing pressure, these shapes are not
177 accessible during constriction. As in all snap-through instabilities, this pinching instability arises from a reduced
178 energy barrier and associated reduction in neck radius, and has been reported in other membrane physical processes
179 as well [9, 35, 36]. Despite the existence of the snap-through instability at the base, the pressure needed for
180 further constriction becomes unbounded as the pinching radius approaches zero. This suggests that fully symmetric
181 membrane shapes are not favorable for constriction below a certain critical radius.

182 **Relaxation of symmetry constraints lowers the energy barrier associated with membrane constrict-** 183 **tion**

184 We next asked if relaxation of symmetry constraints alters the energy landscape of location-dependent constrict-
185 tion. To answer this question, we used our 3D model. For these simulations, we initialized the computation as a
186 pre-formed membrane tubule to limit computational complexity (see SOM). Strikingly, we observed that once the
187 symmetry constraints are relaxed, membrane constriction at all three locations requires a lower collar pressure by
188 more than an order of magnitude when compared to the axisymmetric deformation (compare Figure 4D and Fig-
189 ure 3D). To verify this result, we enforced axisymmetry constraints in the 3D model and repeated our calculations
190 for the “cap” (Figure 2B) and “tube” (Figure 2C). We observed that the collar pressures increased by an order of

191 magnitude (Figures S3 and S4) when symmetry is imposed, resulting in comparable pinching profiles between the
192 axisymmetric and 3D models. However, without the imposed axisymmetry, the collar pressures reduced signifi-
193 cantly (Figure S5). These results allow us to conclude that absence of enforced axisymmetry alone is responsible
194 for the significant decrease of collar pressure. We next analyzed the shapes of the membrane cross sections during
195 3D constrictions, which we found to be distinctly reminiscent of buckling phenomena that are observed in thin
196 walled elastic structures [60, 61]. The pinching profiles shown in Figure 4C correspond closely to the classical
197 result of the first buckling mode of a thin ring subjected to inward pressure on its walls. These deformation modes
198 of buckling/pinching in thin-walled elastic rings and tubes have been known since the early twentieth century in the
199 context of structural engineering applications [62, 63, 64, 65, 66, 67], and are also observed experimentally [68].

200 Finally, we observed that the base, with the preexisting negative Gaussian curvature, needed lower collar pressure
201 to undergo constriction. This result is consistent with the observation that membranes with a negative Gaussian
202 curvature are more amenable to constriction. Furthermore, contrary to the case of axisymmetric deformation,
203 the collar pressure associated with increasing constriction at the base does not continue to grow with constriction
204 (compare Figure 4D, red line and Figure 3D, red line). While the collar pressure increases sharply for initial
205 constriction (Figure 4D, inset), it only rises gradually for the tube and cap geometries as constriction continues
206 to increase several-fold. The initial increase in compressibility represents the natural stiffness of the membrane,
207 after which symmetry breaking occurs leading to near spontaneous constriction. This result suggests that without
208 the arbitrary restriction to axisymmetric deformation, near-spontaneous collapse of the neck is possible after a
209 critical collar pressure is reached. The sudden drop in stiffness observed for the base geometry (Figure 4D) is
210 associated with symmetry-breaking. As shown, this symmetry-breaking and loss of stiffness is not immediate, but
211 occurs after a small amount of constriction has occurred. Thus, we predict that the energy landscape of membrane
212 neck constriction is location dependent, but more importantly, predict that lower symmetry shapes attained by 3D
213 constriction can significantly lower the energy barrier at the base to promote easier constriction.

214 **Easier constriction at the base is accompanied by reduced membrane stiffness.**

215 We further investigated the energy landscape at the base of the tubule (Figure 2D) to identify the mechanisms
216 associated with easier constriction. For the axisymmetric pinching pathway, an increase in collar pressure results
217 in progressive transformation of the tubule into a half-catenoid-shaped membrane. Indeed, this is the shape that
218 is commonly seen in schematics of membrane pinching (Figure 5A). On the other hand, application of increasing
219 collar pressure in 3D demonstrates that the membrane base is quick to break symmetry, and assumes the iconic
220 shape of a soda can crushed by radial pinching (Figure 5B). We thus observe that the axisymmetric and 3D models
221 invoke different constriction pathways; the axisymmetric model yields uniform pinching (Figure 5A), but the 3D

222 model captures an asymmetric flattened tubule geometry (Figure 5B).

223 As before, the axisymmetric mode shows a snap-through instability (Figure 5C, Figure S6B). This instability can
224 be attributed to a build up of negative tangential stress or tension work (Figure S6E). The sharp increase of both
225 the bending energy (Figure S6D) and tension work (Figure S6E) at large constriction corresponds to a sharp in-
226 crease in the pressure required for constriction (Figure 5C, Figure S6B). However, in 3D, a narrow constriction
227 radius is accessible at a much lower pressure when compared with the axisymmetric mode. This result can be
228 understood by analyzing the relationship between stiffness and constriction of the membrane in different modes
229 of deformation (Figure 5D). The stiffness of the membrane is significantly reduced in the lower symmetry mode
230 attained in 3D when compared with the axisymmetric mode (compare black circle lines and red diamond lines in
231 Figure 5D). Similar comparisons for the tube (Figure S5) show a significantly reduced stiffness in lower symmetry
232 pathways of 3D constriction when compared with axisymmetric pathways. Comparison of the bending energy in
233 the axisymmetric and 3D modes of deformation shows that while the bending energies in both cases are similar
234 in magnitude, the energy landscape is different (Figure 5E). All intermediate energy states along the constriction
235 of the neck radius are accessible in the 3D pathway of deformation while in the axisymmetric pathway the energy
236 states associated with the snap-through regime are inaccessible (dashed grey region in Figure 5E). From these anal-
237 yses, we conclude that easier constriction at the base of the tube, revealed by full 3D computations, is accompanied
238 by a reduced membrane stiffness and accessibility to all intermediate energy states along the constriction path.

239 **Helical force collar further reduces the barrier to membrane neck constriction**

240 The 3D model of membrane deformation allows us to probe the response of the membrane to non-symmetric force
241 distributions such as those exerted by helical arrangements of proteins that cannot be modeled in the axisymmetric
242 framework. Membrane scaffolding proteins such as dynamin [33, 38] and ESCRT-III [69] self-assemble into helical
243 collars that can constrict the neck. Recent Cryo-EM maps of human dynamin-1 (dyn-1) polymer report detailed
244 structural and molecular information on its helical geometry [12]. With GTP hydrolysis, the helical polymer
245 actively constricts the membrane from a diameter of more than 20 nm to below 3.4 nm [12]. Disassembly of
246 dynamin is thought to promote scission via hemifission [37, 70, 71]. Despite the well-established mechanisms of
247 dynamin-mediated constriction, the response of the membrane to constriction and subsequent scission is not yet
248 fully understood. This led us to investigate the role of a helical collar pressure, which is a mimic of force generated
249 by helical protein assemblies, on membrane constriction. We explore the role of two geometric parameters of a
250 helical collar – the pitch, defined as the distance along the axis for a complete helical turn, and the number of rings.
251 We first consider a single helical ring exerting a collar pressure on a cylindrical tube (Figure 6A) with different
252 values of the pitch, non-dimensionalized by the height of the collar. Increasing helical pitch corresponds to an

253 increased span (with same collar area) over which the load is distributed on the geometry. Qualitatively, we observe
254 that the cross section of the neck is non-axisymmetric for different values of the pitch, confirming that the lower
255 symmetry modes of deformation are still preferred for neck constriction with helical rings of pressure (Figure 6B,
256 C). Quantitatively, the collar pressure associated with constriction decreases as the pitch increases (Figure 6D).
257 Correspondingly, the stiffness also decreases for increased pitch (Figure 6E), indicating that the bending energy
258 (Figure 6F) becomes slightly less steep. The pink and blue shaded regions in Figure 6E indicate regions of high
259 and low stiffness (easier constriction) respectively. From these observations, we conclude that helical collars have
260 the ability to further reduce the energy barrier to membrane constriction by a decrease in collar pressure and
261 stiffness associated with this process, and that the pitch of the helix is an important determinant of this barrier.

262 To our knowledge, this is the first numerical characterization of the effect of helical squeezing forces on membrane
263 constriction in a continuum framework. However, from a soft matter perspective, it is well-known that helical
264 structures are known to exert squeezing forces more effectively on their support; an excellent example of this
265 behavior can be found in the twining of plant vines [72, 73, 74] and other naturally occurring soft materials. Like
266 cylindrical forces, helices also exert tangential and radial forces. Additionally, because of the pitch of the helix,
267 they also exert axial forces.

268 Given our observation that the base of the tube is energetically favorable to constriction (Figure 5), we next added
269 a helical collar pressure to the base to investigate the effect of coupling the three key design elements – negative
270 Gaussian curvature, access to non-axisymmetric modes, and helical collar pressure (Figure 7A) – on membrane
271 neck constriction. We found that this combination also results in reduced collar pressure (Figure 7D) and energy
272 barrier (Figure 7F) with increasing constriction when compared to a ring of collar pressure (Figure 7D,F for zero
273 pitch). However, a ring of lower pitch was advantageous in terms of a lower stiffness; the stiffness for Pitch 4 is
274 greater than that associated with Pitch 0 (Figure 7E). This is possibly the result of recruiting the stiffer cylindrical
275 portion of the membrane tube with greater pitch.

276 Comparing the effect of helical pinching on the tube (Figure 6) versus the base (Figure 7), we arrive at the following
277 conclusions. For a single helical ring, the relationship between helix geometry and the membrane geometry is non-
278 trivial. It appears while both the tube and base geometry show a clear relationship between helical pitch and collar
279 pressure (Figure 6D, Figure 7D), the same is not true for the stiffness (Figure 6E, Figure 7E). This suggests a
280 complex interaction between the geometries of the membrane and the helical ring, possibly due to a mechanical
281 feedback between membrane curvature and the constricting action of the helices.

282 **Effects of multiple helical collars are also location-dependent**

283 Since a helical ring of collar pressure offers a lower energy barrier to constriction, we next asked if an increase
284 in the number of helical rings can further promote scission. This question is motivated by observations that an
285 increase in the number of dynamin rings is a predicted response to delayed scission and higher membrane tension
286 [24, 75, 76, 77]. To answer this question, we simulated 3 rings of helical collar pressure reminiscent of dynamin
287 rings assembled on a membrane tubule [12, 38, 76]. More than 3 rings of a dynamin collar are unlikely to exist
288 *in vivo* due to disassembly of the dynamin oligomer [78]. Collar pressure due to multiple helical rings not only
289 leads to membrane constriction as expected (Figure 8A) but also appears to stabilize the membrane tube against a
290 sideways wobble that is observed with one ring (compare Figure 6B with Figure 8B). We also observed that while
291 the values of the collar pressure are of the same order of magnitude for 3 rings as with 1 ring (compare Figure 6D
292 and Figure 8C), the stiffness profile is different (Figure 8D). During the early part of the constriction, the stiffness
293 values go from high to low, quickly leading to a region of easier constriction (Figure 8D, blue shaded region).
294 However as constriction increases, the stiffness increases again over a finite range of constriction and at a lower
295 value of constriction (Figure 8D, pink shaded region), unlike the very narrow range of stiffening in Figure 6E.
296 This may be due to the increase in the surface area that is constricted by three rings as compared to the surface
297 area constricted by one ring, increasing the structural resistance to constriction. Thus, multiple rings assist neck
298 formation on a tubule during an initial constriction region (Figure 8D, 4 - 15 nm of constriction), after which
299 disassembly and possibly additional proteins are required.

300 Interestingly, the presence of 3 rings at the base resulted in an increase in both the collar pressure (Figure 8G) and
301 stiffness (Figure 8H), such that the region of easier constriction (Figure 8H, blue shaded region) is much smaller
302 than for a single helical ring (Figure 7E, blue shaded region). The membrane then transitions into a region of high
303 stiffness at a smaller value of constriction (Figure 8H, pink shaded region) due to the larger surface area of the three
304 rings that recruits more of the cylindrical tube to resist constriction.

305 Furthermore, multiple helical rings achieve easier constriction at shorter constriction distances for a cylindrical ge-
306 ometry (Figure 8D, blue region) and at larger constriction distances for the base geometry (Figure 8H, blue region).
307 However, they resist further constriction at narrow radii independently of the pre-existing curvature (Figure 8D and
308 Figure 8H, pink region) . Given these observations, it is possible that helical polymers might preferentially undergo
309 conformational rearrangements such as a change in pitch or number of rings based on feedback with the underly-
310 ing membrane curvature so as to achieve a lower energy barrier to constriction. Such structural rearrangements in
311 dynamin have also been reported in experiments [12].

312 Discussion

313 Membrane constriction and subsequent scission are universal to membrane remodeling processes *in vitro* and *in*
314 *vivo*. While the molecular machineries may differ across systems, these deformation processes likely share the
315 same common physical principles. In this study, using computational modeling, we show that there are three
316 key design elements that play important roles in promoting membrane constriction – (1) location *i.e.* preexisting
317 curvature of the membrane being constricted, (2) access to lower-symmetry modes of deformation, and (3) access
318 to helical loading.

319 From a mechanical standpoint, membrane constriction can be interpreted as a deformation mechanism driven by
320 a radial collar pressure applied by the scission proteins in the vicinity of the necking region. For axisymmetric
321 constriction, the pinching pressure needed to cause membrane constriction increases with the narrowing of the neck
322 radius. This monotonic growth of the radial pressure results in a high energy barrier for pinching. Interestingly,
323 many elastic structures have inherent modes of instability that result in enhanced deformation or even collapse in
324 response to loading and are associated with lower energy barriers. Such modes are ubiquitous in thin elastic shells
325 and manifest as folding, wrinkling, creasing, and buckling deformations (e.g. wrinkling of thin membranes and
326 graphene sheets [79], surface tension induced buckling of liquid-lined elastic tubes [80], snap-through of elastic
327 columns [81], barrelling modes of thin cylinders [60, 61], etc.). Notably, they have lower symmetry than the
328 fully axisymmetric deformations. If such modes exist, and are accessible in cell membranes, their being triggered
329 would naturally lead to a reduction in the energy barrier to constriction and scission. Building on this conventional
330 understanding of buckling analysis of thin-walled structures, we predict the existence of lower energy modes of
331 constriction in membrane tubules. The conclusions from our simulations provide insight to a number of recent
332 experimental studies and suggest new experimental design as discussed below.

333 Dynamin and dynamin-related proteins (DRPs) have been shown to be essential for scission events during mito-
334 chondrial division [82] and during clathrin-mediated endocytosis via mechanical feedback with actin in both yeast
335 [83, 84] and mammalian cells [85]. In dynamin-mediated fission during endocytosis [86, 87], dynamin preferen-
336 tially interacts with curved membranes [88, 89], indicating a curvature dependence. Morlot *et al.* [23] showed
337 that the local energy barrier to constriction is lower at the edge of the dynamin helix (large curvature) in optical
338 tweezer experiments of dynamin-mediated fission. More recent experiments and models suggest that fission can
339 also occur in the middle of the dynamin-coated region [37, 90]. While our results cannot confirm where fission
340 will occur, we predict two important effects - (1) constriction is indeed curvature-dependent and (2) the membrane
341 shape at the center of a given helical pitch is highly curved in 3D. These predictions are consistent with observa-
342 tions [23, 90, 91]. For example, Dar *et al.* [90] showed that dynamin1 polymers cause membrane constriction with
343 high probability when the tubule radius approaches 16 nm or less, consistent with predictions from our model (see

344 Figure 2F of [90] and compare against Figure 6E, J).

345 A central conclusion from this study is that a crushed soda can shape of the neck is energetically favorable for
346 constriction over radially symmetric pinching. This prediction suggests that mechanisms such as those proposed
347 in Figure 6 of Dar *et al.* can be revised to include lower degrees of symmetry (compare Figure 6 of [90] with
348 Figure 6B). With advances in 3D imaging methods such as electron tomography, it should be possible to examine
349 the cross-sections of necks during the progression of constriction by different molecular machines and quantify the
350 relationship between membrane tubule symmetry and the particular protein assembly. Furthermore, determining
351 the curvature-dependent rate constants for these proteins binding to the membrane will be important to quantify
352 the relationship between the shape of the buckled membrane and the disassembly of monomers from polymerizing
353 helical filaments such as dynamin [88, 92]. We predict that this feedback between membrane curvature and ki-
354 netics of helix assembly-disassembly is particularly important for the membrane curvatures where our simulations
355 determine that it is energetically expensive for multiple rings to achieve the progression to scission.

356 Our results also apply to cases where dynamin is not involved in the scission process. In the absence of dynamin,
357 BAR domain proteins and actin are thought to work closely in the formation of long tubular necks [93]. Indeed, in
358 dynamin and clathrin-independent endocytosis, actin is the primary driver of scission of tubular invaginations via
359 a constriction force [94]. These observations suggest that while scission may be less efficient, it is still functional
360 in the absence of dynamin. Our results show that cylindrical collars, such as those enforced by actin, can promote
361 constriction in the absence of dynamin.

362 From a structural mechanics standpoint, the differences in the membrane responses to helical versus cylindrical col-
363 lars can also be understood by drawing analogies again with the buckling of thin cylindrical tubes. The distributed
364 radial pressure in a helical collar creates an lateral torque that induces a bending moment on the tube. Under these
365 conditions, the cylindrical tubes are now susceptible to both radial collapse (through pinching) and buckling under
366 bending moment, which can cause accelerated pinching. The soda can shape also locally reduces the area moment
367 of inertia and this can induce buckling through a process called buckling by ovalization [95]. In a completely dif-
368 ferent setting, the helical structures of twining vines are also known to exert squeezing forces on the support rods,
369 suggesting that helical structures as force generating mechanisms are quite common in nature at different scales
370 [73]. The helical collar mechanism opens up a wider parameter space (helical pitch, collar height, lateral bending,
371 squeeze induced by helical twist, etc.) to optimize for achieving effective pinching. Such analogies with common
372 engineering principles and with biological materials can help build our intuition on membrane-protein interactions;
373 however, we note that the results presented in this work are specific to elastic, incompressible membranes only.

374 Based on the insights derived from our simulations, future work should include further complexities such as the in-
375 fluence of the structure of the helical polymer, the compositional heterogeneity of cellular membranes and the effect

376 of contact constraints between the protein and tubule that can permit potential sliding of the protein on the tubule
377 during the scission process. While recent molecular dynamics (MD) simulations of dynamin-mediated fission also
378 reveal non axisymmetric pathways of constriction via the formation of transient pores [37], better connections be-
379 tween continuum descriptions of the lipid bilayer and membrane-protein interactions at the mesoscale need to be
380 developed to close this gap. This is an ongoing research effort in our group.

381 **Acknowledgements**

382 We would like to thank David Drubin, Jasmine Nirody, and Morgan Chabanon for their feedback on the study.
383 P.R. would like to acknowledge the Office of Naval Research N00014-17-1-2628. S.R. would like to acknowledge
384 the Wisconsin Alumni Research Foundation (WARF) and the Grainger Institute for Engineering at UW-Madison
385 for funding support, and thank Prof. Xiaoping Qian at UW-Madison for his advise on geometric modeling. M.A.
386 would like to acknowledge the Arnold O. Beckman Postdoctoral Fellowship. KG would like to acknowledge NSF
387 DMREF grant #1729166.

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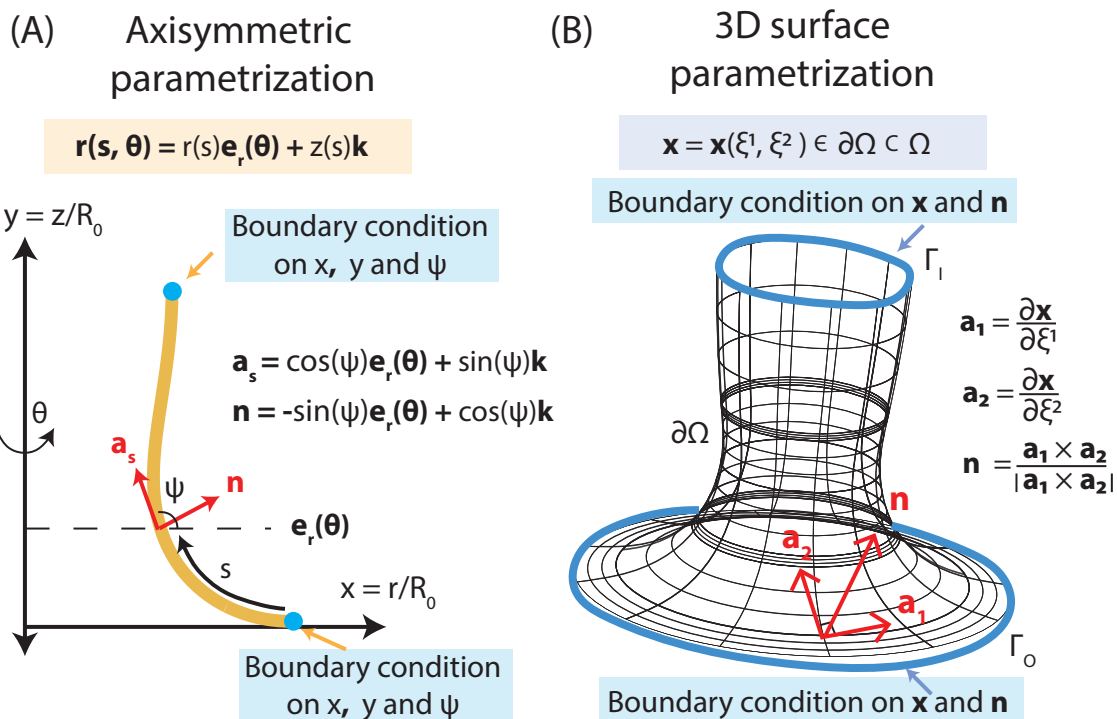


Figure 1: Schematics showing surface parametrization of the membrane geometry in the axisymmetric and 3D formulations. (A) The axisymmetric coordinate system is parametrized in terms of the unit tangent vector (\mathbf{a}_s), unit surface normal vector (\mathbf{n}) and arc length (s), where $\mathbf{r}(s, \theta)$ is the position vector, s is the arc length along the axisymmetric curve, θ is the out-of-plane rotation angle, r is the radius, z is the height, \mathbf{e}_r is the unit radial vector and \mathbf{k} is the unit axial vector. $(\mathbf{e}_r, \mathbf{e}_\theta, \mathbf{k})$ forms the coordinate basis (see SOM for more details). (B) Parametrization of a surface ($\partial\Omega$) embedded in a 3D volume (Ω). Here, \mathbf{x} is the position vector of a point on the surface parametrized in terms of the surface coordinates (ξ^1, ξ^2) which are associated with a flat 2D domain that is then mapped to $\partial\Omega$ by $\mathbf{x} = \mathbf{x}(\xi^1, \xi^2)$. \mathbf{a}_1 and \mathbf{a}_2 are the local tangent vectors to the surface at \mathbf{x} , and \mathbf{n} is the corresponding surface normal. $(\mathbf{a}_1, \mathbf{a}_2, \mathbf{n})$ forms the local coordinate basis. The axisymmetric coordinate system in (A) is a specialization of the general curvilinear coordinate system depicted in (B).

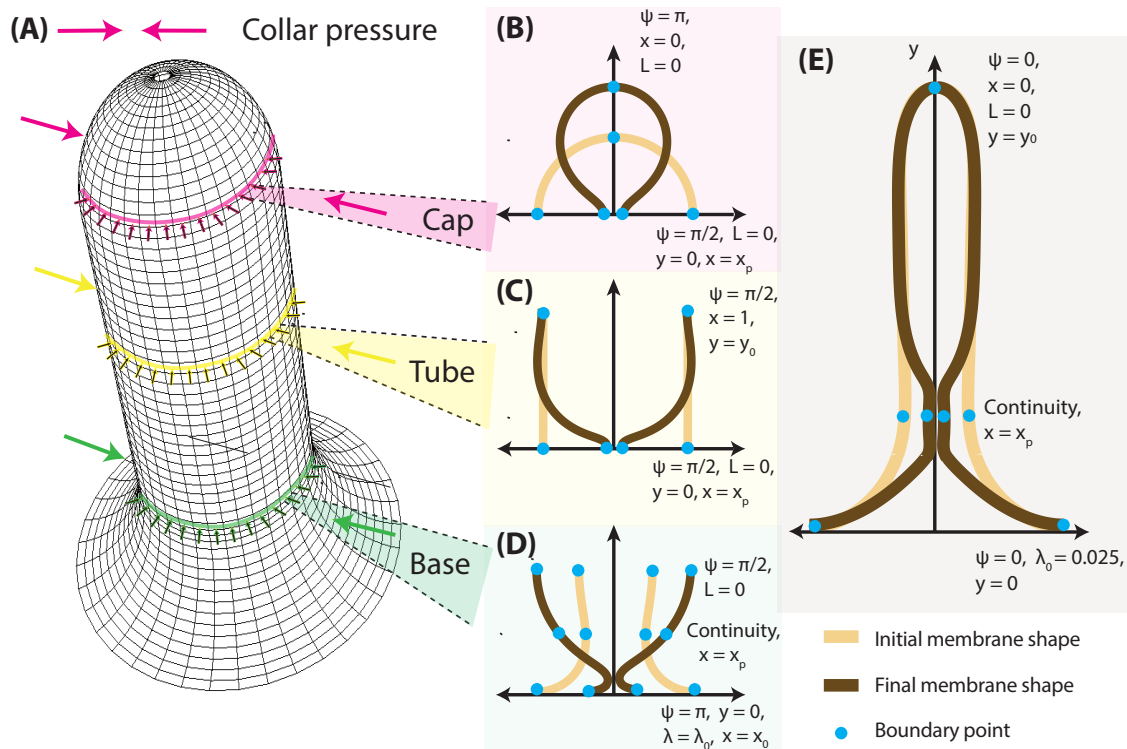


Figure 2: Schematic depicting the modeling framework and simulation set up. Localized forces acting on the membrane were simulated as a collar pressure (A). Three different pinching locations are considered along the membrane tubule. (B) Case 1: Collar pressure applied at a circumference near the cap of the tube, where the mean and Gaussian curvatures are positive. (C) Case 2: Collar pressure applied at the center of the tube, where the mean curvature is positive and Gaussian curvature is zero. (D) Case 3: Collar pressure applied at a circumference at the base of the tube, where the mean curvature is positive along the cylindrical region and negative along the boundary, and Gaussian curvature is negative. (E) Case 4: Collar pressure applied along a tubule of fixed length pulled from an initially flat membrane. Shown are the initial membrane shape (light brown), final membrane shape (dark brown) and boundary/interface points (blue dots). $x = x_p$ is an interface condition enforced in axisymmetry to solve for the collar pressure as an unknown parameter.

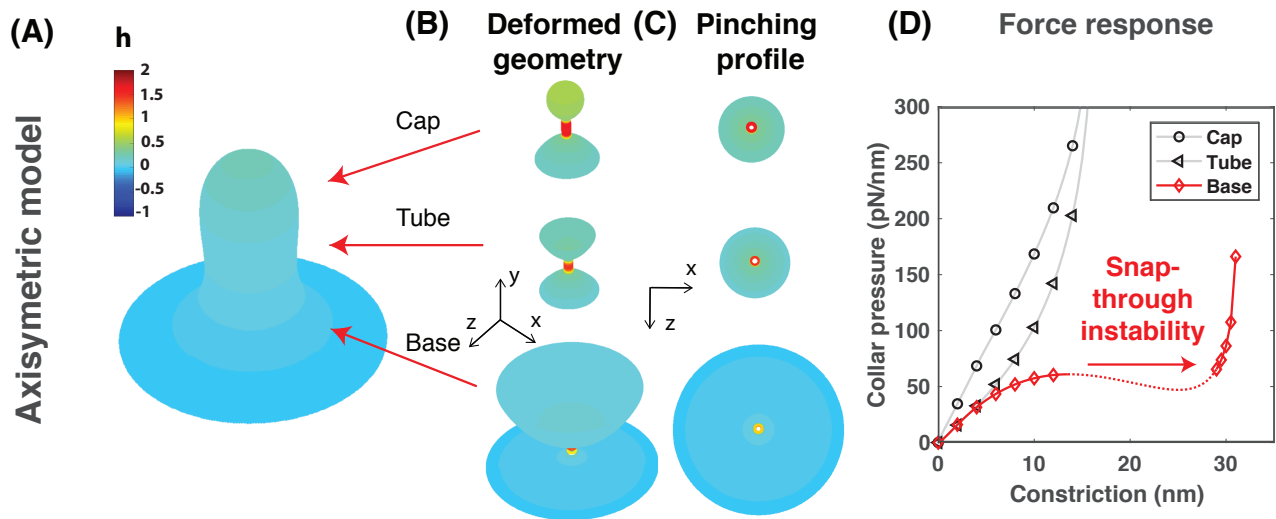


Figure 3: Location dependence of membrane tube constriction in axisymmetry. Shown are the different locations of constriction, cap, tube and base (A), the corresponding membrane shapes (pinched configurations) obtained (B, C) and the evolution of the collar pressure as a function of constriction (force response, D). The bending rigidity κ is 320 pN-nm, membrane tension λ is 0.2 pN/nm, radius of the tube is 20 nm, the radially inward-directed collar pressure is applied over a strip of height 1 nm, and the height of the membrane tubule is 100 nm. The cap and the tube locations deform smoothly, while a snap-through instability is observed at the base (D). The dotted solution path is never realized during the loading phase, leading to a transition to a wider tube morphology that is markedly different from the other cases. The colorbar in (A) shows the non-dimensional mean curvature.

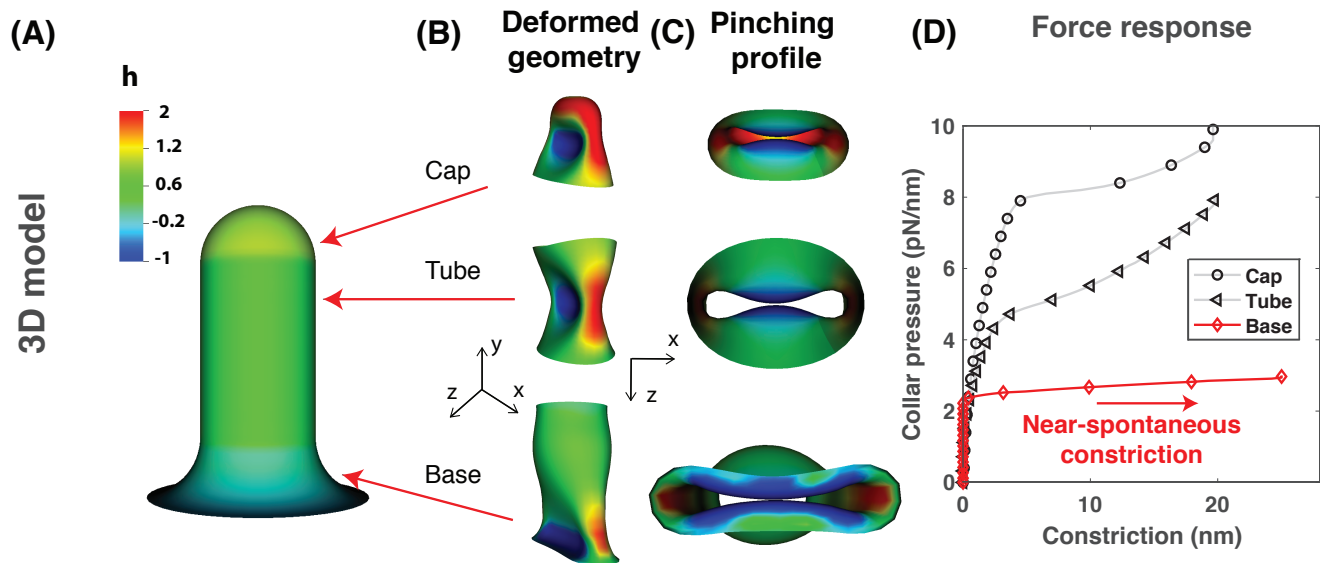


Figure 4: Location dependence of membrane tube constriction in 3D. Shown are the different locations of constriction: cap, tube and base locations (A), the corresponding membrane shapes (pinched configurations) obtained (B, C) and the evolution of the collar pressure as a function of constriction (force response, D). Bending rigidity κ is 320 pNnm, membrane tension λ is 0.2 pN/nm, radius of the tube is 20 nm, height of the applied force is 1 nm, height of the membrane tubule is 100 nm. A near-spontaneous collapse is observed for the base, and a relatively stable constriction evolution for the cap and the tube locations (D). The colorbar in (A) indicates non-dimensional mean curvature. See Movies M1-M3 in the supplementary information for the evolution of the constriction process for the Cap, Tube and Base locations.

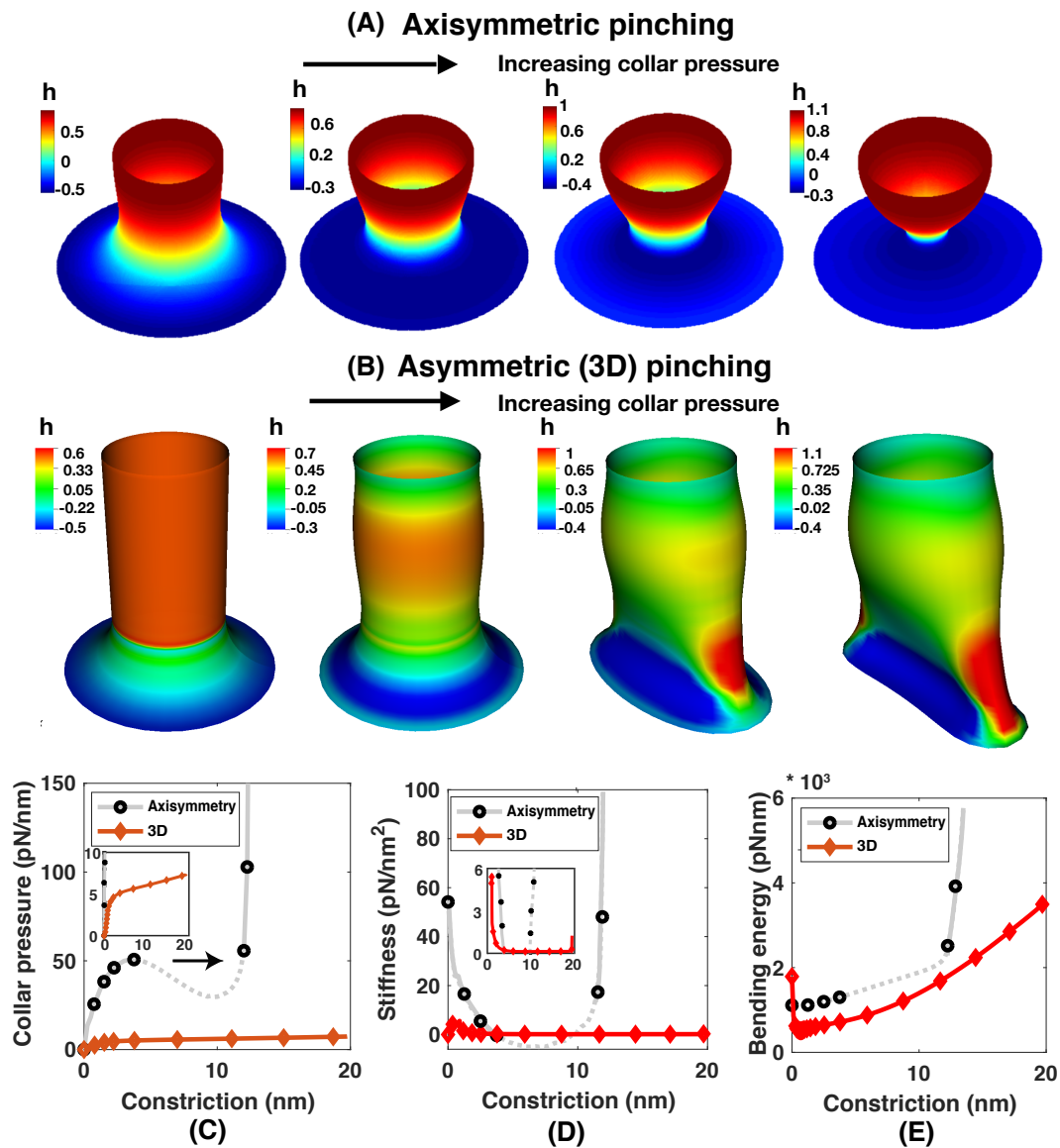


Figure 5: Pinching response of the base geometry and the corresponding evolution of pressure, stiffness and bending energy obtained using the axisymmetric and 3D models. Boundary conditions are shown in Fig. 1, Case 3. Bending rigidity κ is 320 pN·nm, membrane tension is 0.2 pN/nm, the collar pressure is applied over a height of 1 nm, and the length scale is set by the initial radius of 20 nm. Shown are the membrane shape evolution obtained from the axisymmetric (A) and 3D models (B), and the corresponding variation of the collar pressure (C, with inset), stiffness (D, with inset), and bending energy (E). Colorbars in (A) and (B) indicate non-dimensional mean curvature. See Movie M3 in the supplementary information for the evolution of the constriction process for the base geometry.

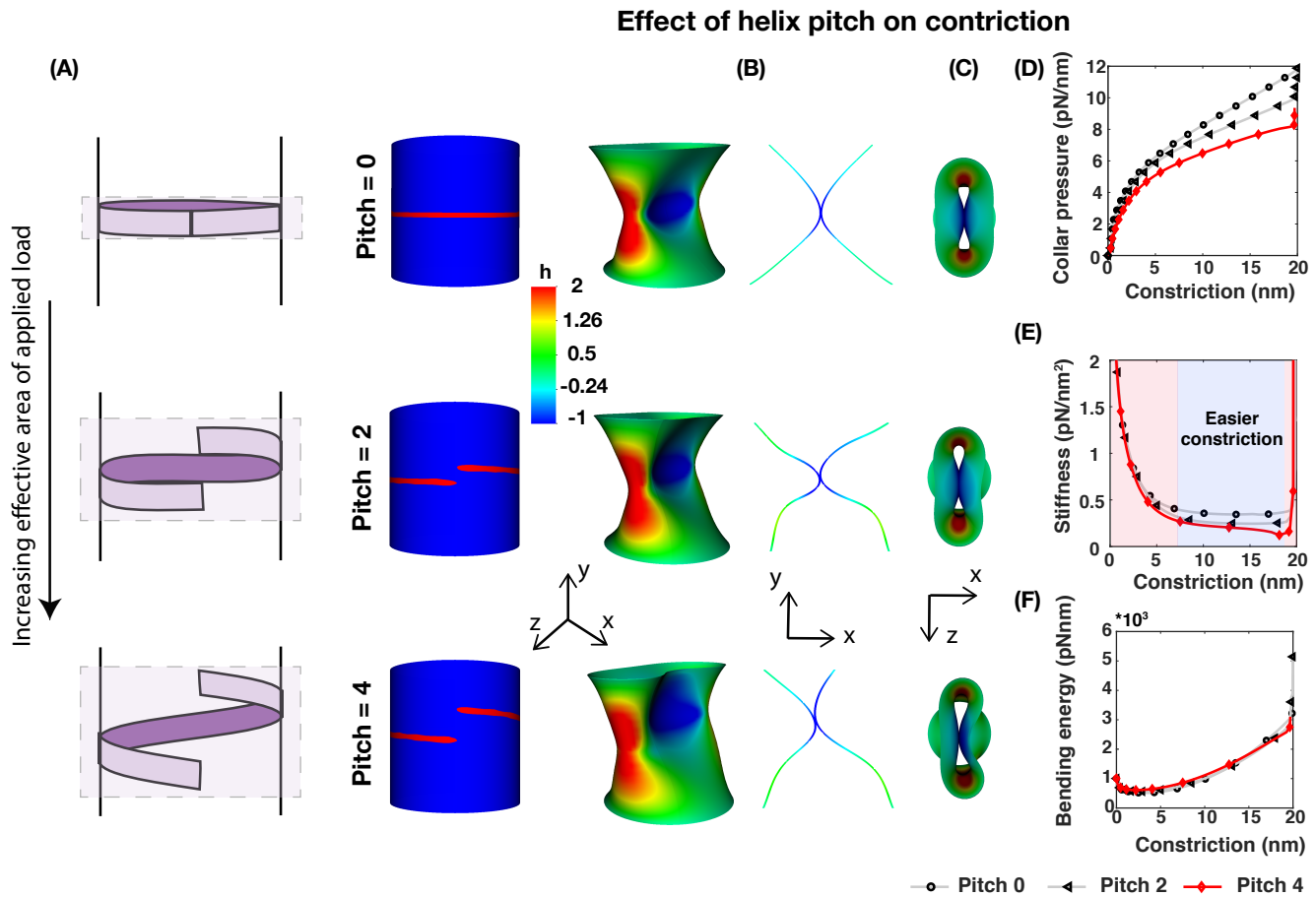


Figure 6: A helical force collar further reduces the barrier to constriction. Considering force collars with a normalized pitch of zero, two and four, shown are the shape of the helical collar (A) with a schematic depicting increased span of load distribution area with increasing helical pitch, deformed shape and corresponding pinching shape (B), the pinching profile (C), and evolution of the collar pressure (D), stiffness (E), and membrane bending energy (F). Shaded regions of pink and blue in (E) represents a region of high stiffness ($> \sim 0.25$ pN/nm²) and low stiffness ($< \sim 0.25$ pN/nm²) respectively. The colorbar under (B) indicates non-dimensional mean curvature. See Movies M4-M6 in the supplementary information for the evolution of the constriction process due to a helical force collar at the tube location with a non-dimensional pitch of zero, two and four, respectively.

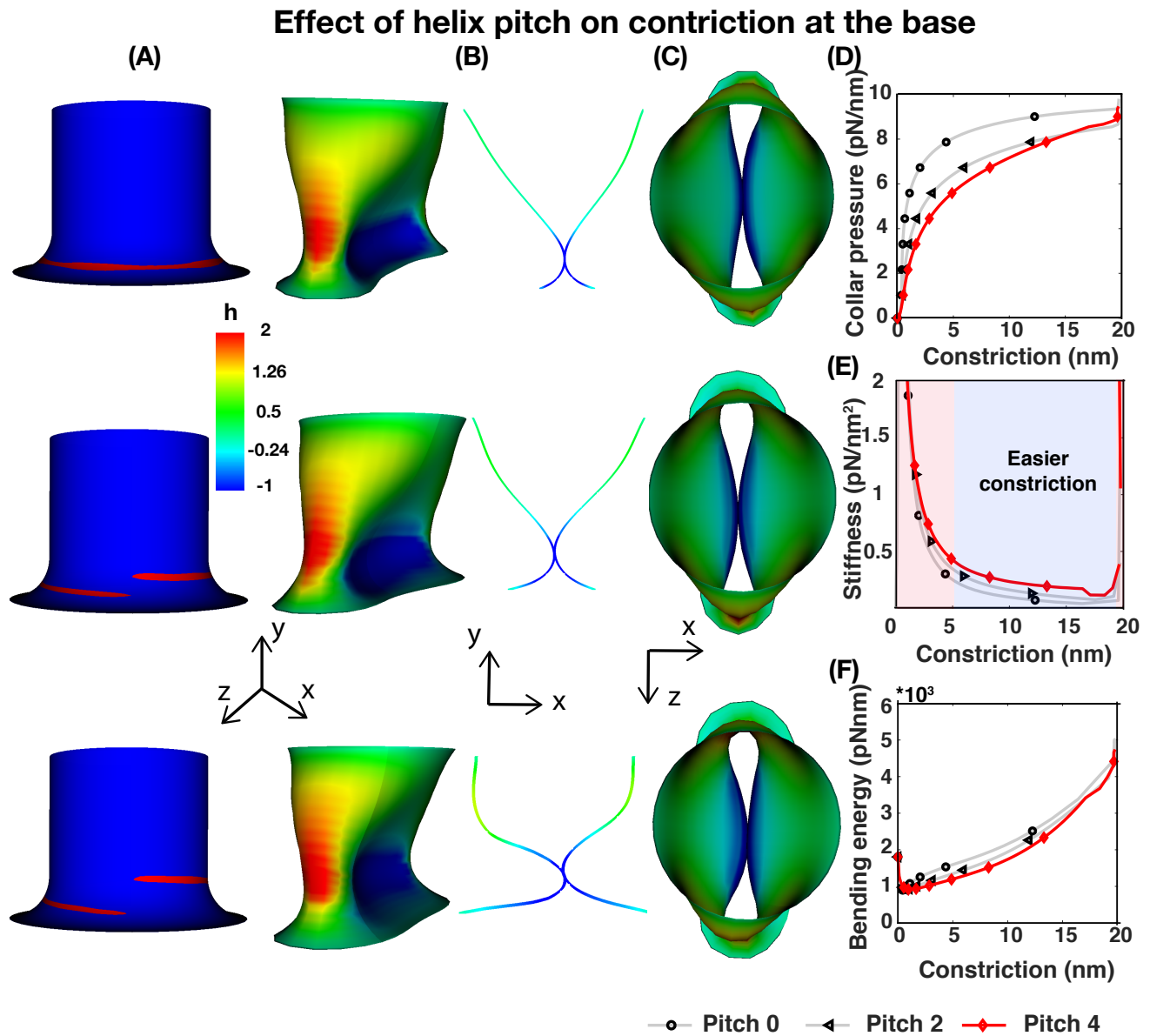


Figure 7: A helical force collar increases stiffness to constriction at the base. Considering force collars with a normalized pitch of zero, two and four, shown are the shape of the helical collar and deformed shape for pitch two and four (A-C), evolution of the collar pressure (D), stiffness (E), and membrane bending energy (F). Shaded regions of pink and blue in (E) represents a region of high stiffness ($> \sim 0.25$ pN/nm²) and low stiffness ($< \sim 0.25$ pN/nm²) respectively. The colorbar under (B) indicates non-dimensional mean curvature. See Movies M8-M10 in the supplementary information for the evolution of the constriction process due to a helical force collar at the base location with a non-dimensional pitch of zero, two and four, respectively.

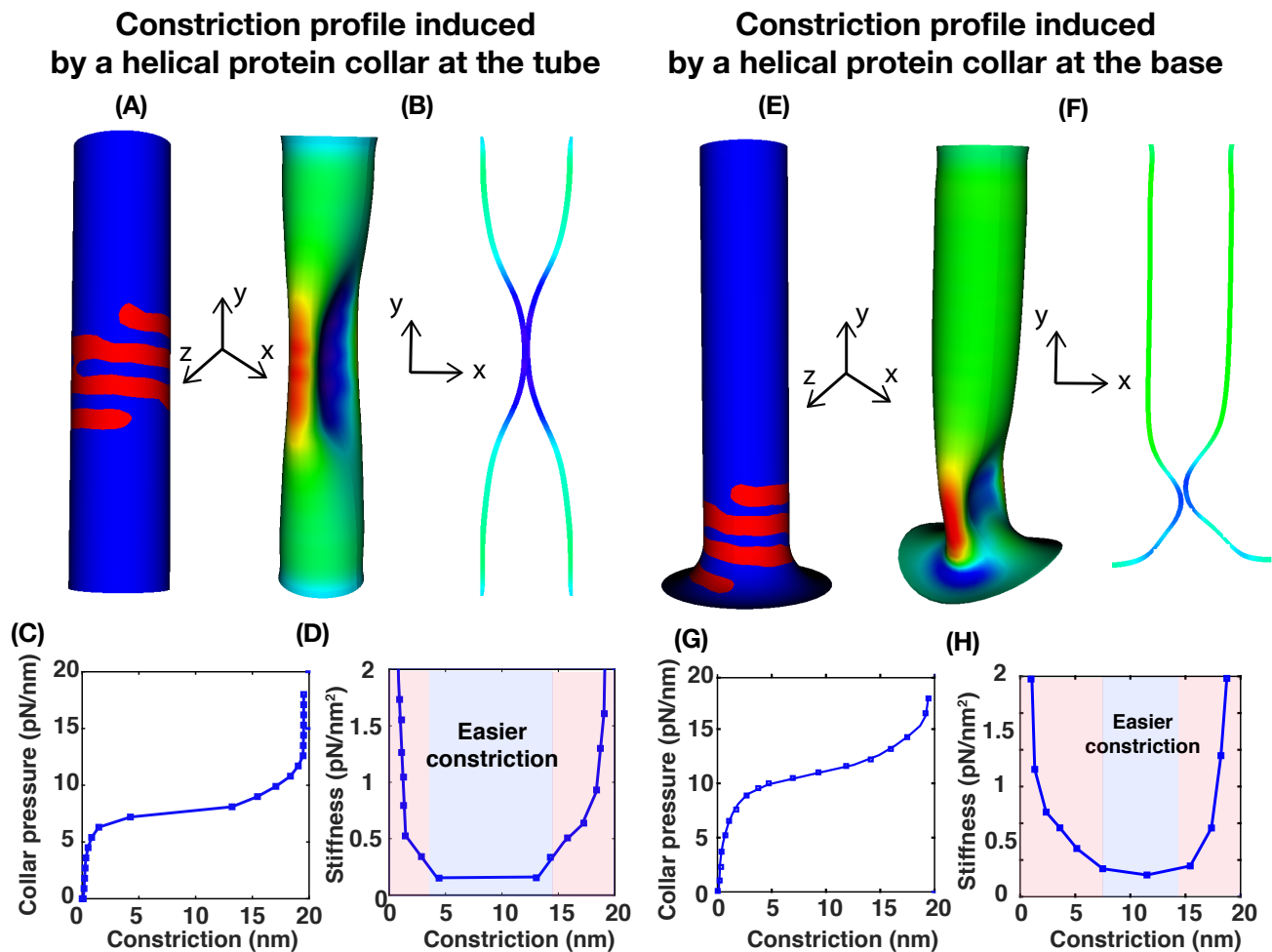


Figure 8: Constriction by multiple helical rings are also location dependent. For both the tubule and base geometry, shown are the initial geometry and location of the force collar with three helical rings (A, E), the deformed shape and corresponding pinching profile (B, F) and the corresponding variation of collar pressure (C, G) and stiffness (D, H). See movies M7 and M11 for the corresponding evolution of the constriction process due to a force collar with three helical rings.