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The effect of size-scale on the kinematics of elastic energy release^{\dagger}

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Elastically-driven motion has been used as a strategy to achieve high speeds in small organisms and engineered micro-robotic devices. We examine the size-scaling relations determining the limit of elastic energy release from elastomer bands that efficiently cycle mechanical energy with minimal loss. The maximum center-of-mass velocity of the elastomer bands was found to be size-scale independent, while smaller bands demonstrated larger accelerations and shorter durations of elastic energy release. Scaling relationships determined from these measurements are consistent with the performance of small organisms and engineered devices which utilize elastic elements to power motion.

Organisms can use elastically-driven motion to bypass the forcevelocity trade-off of an alternate method of actuation. This includes organisms that load internal spring-like elements to circumvent both the power limitations of muscle^{1,2} and other system-level constraints³. For example, mantis shrimp store elastic bending energy in the exoskeleton of their raptorial appendages. When released, this energy can drive their appendages at speeds greater than those achievable with muscle alone, reaching velocities up to 30 m/s and rapidly delivering high forces to prey^{4–7}. Although large organisms make some use of elastic structures (e.g. tendon), elastic energy storage and release can be crucial for small organisms (typically < 10 cm in length) to achieve rapid movement^{1,8}. These small organisms - such as mantis-shrimp, trap-jaw ants, locusts and fleas - use a latch to separate the phase of elastic energy storage (via muscle contractions) from that of energy release⁹. Disentangling energy storage from release enables these organisms to achieve astonishing kinematic performance (high velocities, large accelerations, and short durations of movement), and perhaps most remarkably, to perform these motions in a repeatable manner sustained by their metabolic processes.

Organisms that store and release elastic energy have served as inspiration for recent robotics research $^{10-17}$. In an attempt to match (or exceed) biological performance using engineered de-

vices, several research groups have taken a biomimetic or bioinspired approach. This approach has led to new techniques for robotic manipulation^{11,14,17}, the ability to move robots on difficult terrain^{10,15,16}, and has been used to test scientific hypotheses about locomotion^{12,14,15}. As of yet, none of these devices can match the kinematic performance of the fastest organisms⁹. However, these engineered devices are typically larger than their biological counterparts, and it is unclear to what extent size might account for the differences in kinematic performance.

We seek to explain the gap in performance between biological and synthetic elastic systems by examining how kinematic performance depends on size-scale and properties of system components. In addition to an elastic element (i.e. spring), these elastically-driven systems contain three other major components³: (i) a motor (in many animals, muscle) that generates sufficient work to load the elastic element, (ii) an energy-efficient latch to hold and release the elastic element without significant dissipation, and (iii) a load mass that is moved by the elastic element and not actively involved in elastic energy release. The coupling between these components leads to trade-offs and scalings that have been explored in our previous work^{9,18}.

In this work, we focus exclusively on the elastic element to determine the limits of elastic energy release due to only spring properties. We take a reductionist approach by examining the dynamics of a freely-retracting spring in isolation - externalizing the motor and latch. This externalization decouples the motor and latch from the fast movement of the spring, which is similar to the way some fast elastically-driven organisms operate¹⁹. Isolating the spring removes size-dependent force limitations of the loading motor, which enables the stored elastic energy to be ultimately limited by the failure properties (maximum strain) of the isolated spring material. This isolation can be taken one step further by measuring the dynamics of an unloading spring that carries no additional load mass. Traditionally, spring properties have

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been measured while controlling the strain rate of the material. These types of analyses are sufficient to quantify spring properties in systems where the mass of the spring is much smaller than the load mass. At very small load masses (or zero load mass, as we explore here), the elastic wave propagation of the spring material becomes a limiting factor. In this study, we examine recoil kinematics of springs driving no mass to quantify the limits of an unloading elastic element.

The unloading of a spring is constrained by its material properties and geometry⁹. Springs in engineered devices are often optimized to maximize elastic energy storage^{20,21}. While metals have significantly higher elastic wavespeeds²², the large strain to failure exhibited by many elastomers can often lead to enhanced performance in elastically-driven motion. However, intrinsic dissipative properties of many typical elastomers can also limit their effectiveness in these applications. In biological systems, spring components are often composite materials^{23–25}, comprised of both highly resilient* soft protein (e.g. elastin or resilin^{26–28}) matrices and stiffer components, such as chitin. This diversity of materials in engineered and biological examples, in light of their varied performance, suggests that material properties play an important role; however, a general framework for this necessary understanding has not yet been established.

To understand the upper limit of elastic energy release in a resilient material system, we perform experiments measuring the high speed free retraction of a polyurethane elastomer composite with similar resilience and wavespeed to resilin. Building upon recent work²⁹⁻³⁴, we release long thin bands of the elastomer from an initially uniform uniaxial extension, and track its displacement field on unloading. The displacement field is used to obtain the center-of-mass motion of the band, which allows for a functional determination of the scaling relations that define the limits of impulsive elastic performance. We focus on the sizescale and materials properties of a spring and how these factors affect its elastically-driven performance by examining three key parameters often used to assess kinematic performance in biology and micro-robotics 1,2,4,10,35-38: maximum center-of-mass velocity (v_{max}) , maximum center-of-mass acceleration (a_{max}) , and duration of elastic energy release (Δt). Utilizing this experimental approach we ask two guiding questions: Does kinematic performance depend on the size of an elastic element? How does the kinematic performance of elastically-driven biological organisms and engineered devices compare to the isolated recoil of a resilient elastomer composite?

Expected scaling relations for the center-of-mass kinematic performance of a recoiling elastomer band can be rationalized based on physical principles. First, the center-of-mass acceleration of the band is given by the ratio of the net force acting on the band divided by its mass. Just after the release of the band from one end, if the only external force acting on the band is from the clamp at the other (fixed) end, then center-of-mass acceleration is

$$a_{\max} = \frac{\sigma_{in}}{\rho L_0}$$

where σ_{in} is the initial stress from which the band is released, while L_0 and ρ are the equilibrium length of the band and its density, respectively. To separate the role of materials and loading strain, we can rewrite this equation as

$$a_{\max} = \frac{c_{\text{sec}}^2 \varepsilon_{\text{in}}}{L_0},\tag{1}$$

with the secant elastic wavespeed (c_{sec}) from an initial strain (ε_{in}) defined as

$$c_{\rm sec} = \sqrt{\frac{\sigma_{\rm in}}{\rho \varepsilon_{\rm in}}}.$$
 (2)

During the unloading of a uniform, long thin strip of elastic material stretched to an initial strain of ε_{in} , the center-of-mass travels a displacement $\varepsilon_{in}L_0/2$. Using this displacement and assuming a constant center-of-mass acceleration given by Eq. (1), leads to the duration of elastic energy release

$$\Delta t = \frac{L_0}{c_{\text{sec}}}.$$
(3)

Finally, with those same assumptions, the maximum center-ofmass velocity is determined by the product of acceleration and duration, $v_{\text{max}} = a_{\text{max}}\Delta t$, yielding an expression consistent with the maximum velocity found in previous work for a linear elastic material³²

$$v_{\max} = c_{\sec} \varepsilon_{\sin}.$$
 (4)

Notably, these scaling relations depend on the loading of the material before release (ε_{in}). In elastically-driven systems more generally, ε_{in} might be imposed by other constraints on the system. For example, for a system with a maximum motor force, ε_{in} would be set by a coupling between the motor maximum force, spring modulus, and spring geometry⁹. In this work, however, we focus on the spring to first test the kinematic scaling relations of Eqs. (1-4). Then, by setting the loading of the material to the maximum allowable by its material failure properties, we explore the upper kinematic limits of a resilient elastomer.

1 Experimental

1.1 Polyurethane sample preparation and mechanical characterization.

To experimentally verify the kinematic scaling relations, a 1.6 mm thick commercially available pre-fabricated polyurethane elastomer sheet (McMaster-Carr, 8716K61, durometer 40A) was sectioned into long, thin bands using a razor blade. For the narrowest bands (width, $w_0 < 2$ mm), a laser cutter (Universal Laser Systems) was first used to create shallow grooves to guide the razor blade, reducing variation in the band width. Samples were cut to ensure a uniaxial geometry ($L_0 >> w_0$), with $1.6 \text{ mm} \le w_0 \le 27 \text{ mm}$ and $17 \text{ mm} \le L_0 \le 267 \text{ mm}$. The average density of all samples was $\rho = 1125 \text{ kg/m}^3$, with the mass of each sample measured using an analytical balance.

The mechanical properties of the material were character-

^{*} Resilience is a measure of energy recovery, and is defined by the ratio of energy recovered upon unloading divided by the energy expended during loading a material

ized by performing cyclic loading/unloading of the bands at low strain-rate ($\dot{\varepsilon} < 0.01 \, \text{s}^{-1}$), using a tensile testing apparatus (Instron 5564). The polyurethane elastomer has a similar modulus as resilin (Fig. 1A-B) at low strain-rates, and a resilience r > 97%at up to 300% strain. Beyond 300% strain, the material would typically fail due to stress concentrations at the clamped ends of the band. While resilin can strain up to 300% reliably³⁹, it is not generally observed to stretch to this extent in vivo. The highest suggested in vivo strain for a recoiling insect spring is in the flea pleural arch, where the resilinous portion is hypothesized to strain 100%⁴⁰ - thus our experimental elastomer strain covers the whole range of the strains observed in vivo. Over the full range of the polyurethane elastomer, the secant wavespeed of the polyurethane depends on strain and varies between $\sim 24 - 40 \,\mathrm{m/s}$ (Fig. 1C), as calculated from Eq. 2 using the stress-strain relationship in Fig. 1A and the density of the material.

1.2 High-speed kinematics.

Free retraction measurements were performed by initially loading a band clamped between two pneumatic grips to a given initial strain (ε_{in}) using the tensile testing apparatus, and then releasing one of the grips (Fig. 2A). Upon release, the band rapidly contracts, and the motion was recorded using a high speed camera (Photron Fastcam SA3) at a frame rate of 20-75 kfps. A macro zoom lens (Nikon AF Nikkor 24-85mm) was used to maximize the image of the band to cover the full 1024 pixel CCD of the camera along the direction of motion (x-axis), giving a pixel resolution of $33 - 420 \mu m$ depending on the band length and initial strain. Markings were drawn along the band using a metallic silver Sharpie[®] marker were then digitized from the high speed videography to determine the position (x) of each point of the band as a function of time (t) (Fig. 2B). To generate velocity, acceleration, and higher order derivatives of the position with respect to time, the digitized position data was fit to free knot splines^{42,43}. Combining the motion of each section of the band, the center-of-mass kinematics were then deduced, allowing for the determination of v_{max} and a_{max} . The duration was defined as the time between the onset of the propagating elastic wave (determined by a minimum onset threshold of jerk) until the kinetic energy of the band reached its maximum (which occurs at $v = v_{max}$). Jerk was used to determine the onset time because over a large range of jerk threshold (between 20-80 % of the maximum jerk), the measured duration was approximately constant (<10% difference for even the smallest measured duration).

2 Results & discussion

The kinematic performance of 13 different bands with varying geometry (varying L_0 and w_0) was measured (Fig. 2 shows an example measurement) as a function of the strain energy loaded into the band (between 1-8 values of ε_{in} for each band, for a total of 57 unique measurements). The center-of-mass kinematic performance does not depend on w_0 for the uniaxial geometry used in these experiments. The maximum center-of-mass velocity, acceleration, and duration all increase with increasing initial strain (Fig. 3A-C). The center-of-mass velocity is independent of the



Fig. 1 Low strain-rate characterization of a resilin-like polyurethane elastomer. A The polyurethane elastomer used in this study has a similar modulus (slope of stress-strain response at small strain) to resilin (from ref.³⁹) at low strain-rate ($\dot{\varepsilon} < 0.01$). The small difference in stress between loading and unloading the polyurethane (inset) can be quantified by the material's resilience. B The polyurethane elastomer has a high resilience (r > 0.97) for all samples measured in this study, up to $\varepsilon = 3$. Its resilience is similar to natural resilin (r = 0.96 - 0.97)⁴¹, and slightly higher than recombinant resilin from ref.²⁸. C The secant wavespeed ($c_{\rm sec}$) depends on strain for the polyurethane elastomer, as determined from the stress-strain response and Eq. (2). As will be shown, $c_{\rm sec}$ is a characteristic velocity that governs the recoil dynamics of the elastomer.

band length (Fig. 3A), however, the maximum center-of-mass acceleration and duration both depend on band length (Fig. 3B-C); the acceleration is inversely proportional to band length (Fig. 3B, bottom panel) and the duration scales with band length (Fig. 3C, bottom panel), as demonstrated by the data collapse after appropriately normalizing a_{max} and Δt with L_0 .



Fig. 2 The center-of-mass kinematic performance (velocity, acceleration, and duration) is measured for a retracting elastomer band. A Five images of a retracting elastomer band ($L_0 = 140 \,\mathrm{mm}, w_0 = 8.5 \,\mathrm{mm}$) from a high speed image sequence. To visualize the motion of the band, silver markings are placed along the band and on the clamps at the top and bottom of the image (colored points to the left of the images were added in postprocessing to uniquely label the points of the band, and correspond to the colors used in B-D). The last two images in the sequence show the band undergoing compressive buckling, and occur after the center-of-mass has reached its maximum velocity. B After the bottom clamp releases, motion propagates up through the band in a spatially non-uniform release of strain energy. The center-of-mass motion (black points) is determined from a weighted average of the individual segments of the band (colored points). The inset shows a zoom in of the center-of-mass position along with a free knot spline fit (solid, black curve) in close agreement with the data. The maximum difference between the data and spline fit is on the order of a single pixel ($\sim 0.2\%$ of the total displacement of the centerof-mass). t=0 is set by the propagation of the elastic wave unloading, and determined by a minimum threshold in the derivate of the acceleration (jerk). C,D Derivatives of the free knot splines give the velocity (C), and acceleration (D) of each segment of the band (colored curves), along with the center-of-mass (black curve). From the center-of-mass velocity and acceleration, the kinematic performance is determined (here $v_{\text{max}} = 23 \text{ m/s}, \ a_{\text{max}} = 6.2 \times 10^3 \text{ m/s}^2, \ \Delta t = 4.6 \text{ ms}).$

The scaling relations predicted by Eqs. (1-4) are comparable to the observed recoil kinematics (dashed curves in Fig. 3A-C), using c_{sec} measured from the tensile test (Fig. 1C). The scalings agree with the data for acceleration and duration with no free parameters (Fig. 3B-C). However, the scaling relationship for velocity of Eq. (4) systematically exceeds the observed recoil velocity (Fig. 3A).

To understand the systematic difference in predicted and measured recoil velocity, it is helpful to examine the predicted velocity scaling of Eq. (4) in the context of the kinematic data in Fig. 2D. The equation assumes a constant acceleration over the entire duration recoil. Although this is a reasonable approximation, the measured duration also includes the ramp-up time to reach a_{max} ($\sim 1\,\mathrm{ms}$ in Fig. 1D) and the ramp-down to zero acceleration (also \sim 1 ms in Fig. 1D). During this ramp-up and ramp-down period the acceleration is less than a_{max} , which leads to a breakdown in the predicted scaling of Eq. (4). Factors that could affect the ramp-up/ramp-down time include frictional losses from interaction of the band with the pneumatic clamp³², inertia of elastomer material inside the clamp, dispersion of the elastic wave due to losses within the material or to the environment²⁹, and residual strain left in the band at the point of buckling ³⁰. These losses depend on both material properties of the band and external factors. Since these factors are challenging to accurately model, as a first approximation we assume these losses are constant for all the bands measured, and introduce an effective resilience of the recoiling elastomer $r_{\rm eff}$ through the scaling

$$v_{\rm max} = \sqrt{r_{\rm eff}} c_{\rm sec} \varepsilon_{\rm in}.$$
 (5)

This effective resilience accounts for both energy loss within the band and external dissipation (such as friction of the clamp), and $r_{\rm eff}$ is defined by the ratio of output kinetic energy to input elastic energy (i.e. $r_{\rm eff} = \rho v_{\rm max}^2/2u_{\rm in}$, where $u_{\rm in}$ is the stored elastic energy density). The energy loss occurs primarily during the ramp-up and ramp-down periods of the recoil, which accounts for why $a_{\rm max}$ and Δt do not depend on $r_{\rm eff}$. Using $r_{\rm eff}$ as a free parameter to fit the measured recoil velocity (Fig. 3A, solid curve) results in $r_{\rm eff} = 0.5 \pm 0.1$. This effective resilience is significantly lower than that measured at low strain-rate (recall r > 0.97 from Fig. 1B).

Armed with scaling relations that agree with the observed recoil kinematics, we now return to answering our first guiding question: Does kinematic performance depend on the size of an elastic element? The size-scaling limits of the resilinlike polyurethane elastomer for repeatable, elastic energy release (Fig. 4, dashed lines) are determined by setting the initial strain to $\epsilon_{in}=3$ in the scaling relations from Fig. 3 (recall for $\epsilon_{in}>3$ failure of the polyurethane was often observed). The maximum velocity of the polyurethane elastomer recoil is size-scale independent (Fig. 4A), while the maximum acceleration and duration of movement depend on size (Fig. 4B-C). The dashed lines in Fig. 4 represent the kinematic performance of this particular material choice of polyurethane elastomer, under a specific loading geometry (uniaxial extension), and driving zero added load mass. In the next two paragraphs we justify two specific claims about the recoil scaling limits shown in Fig. 4: (1) the overall scaling of kinematic performance with size does not depend on the specific choice material, geometry, or load mass, and (2) the dashed lines in Fig. 4 are an approximate upper bound for the particular material choice used in this study, independent of geometry and load mass.



Fig. 3 Scaling relations from Eqs. (1-4) are tested by recoil experiments. A The maximum velocity from 57 recoil measurements increases with the initial strain before release (ε_{in}), and is independent of the band length. B The maximum acceleration increases with initial strain, and varies inversely with the band length (bottom panel shows data collapsed in terms of $L_0 a_{max}$). C Recoil duration has a weak (convex downward) dependence on initial strain, and varies inversely with band length (bottom panel shows collapsed data). The dashed curves are the predicted scaling relations from Eqs. (1-4), while the solid curve in panel A is from Eq. (5).

First, the size-scaling of kinematic performance (summarized in the first column of Table 1) should theoretically be independent of the specific choice of materials, geometry, and load mass. Changing the elastic material would alter the pre-factors in the scaling limits through changing csec, failure properties, and resilience, without altering the fundamental trade-offs with sizescale²¹. A different geometry (e.g. using a cantilevered beam as a spring) or adding load mass to the system would alter the absolute kinematic performance of the system. However, if the relative size of elements all change with system size, then changing geometry or mass simply introduces a lengthscale-independent pre-factor to the scaling relations. As a specific example, for a cantilevered beam driving a heavy load mass the scaling relations shown in Fig. 3 still hold, but with added coefficients that depend on two dimensionless parameters: the aspect ratio of the beam (length to thickness), and the ratio of the spring mass to load mass (see Supplementary Information). Since these are independent of size-scale when relative size proportions are held constant, the scaling of kinematic performance with characteristic length shown in Fig. 4 are robust descriptions of the size-scale dependence of elastically-driven motion.

The second claim above — that for the specific polyurethane used in this study the dashed lines in Fig. 4 are an approximate upper bound to elastically-driven performance — is also related to the geometry and load mass. In both of these cases, changing geometry or adding load mass, the net effect is a decrease in the system's kinematic performance and does not change the scaling argument in Fig. 4. Intuitively, adding load mass to the system would decrease the kinematic performance compared to the unloaded elastomer bands used here. The uniaxial geometry used in this work ensures a nearly uniform strain energy density in the material. Other geometries (such as bending) result in a non-uniform strain energy density, and material failure will likely occur at a lower average strain energy density than for uniaxial extension (see Supplementary Information). And although geometries which introduce a mechanical advantage in the system through a lever arm can amplify displacement, they also increase inertial load. As a result, a longer lever arm does not improve performance of the three kinematic parameters in Fig. 4. Therefore, changing geometry or load mass would shift the polyurethane scaling to lower performance (lowering the intercepts in the plots of Fig. 4), without altering the size-scaling relationship (the slopes in Fig. 4).

Putting these results in a larger context, we return to our second guiding question: How does the kinematic performance of elastically-driven biological organisms and engineered devices compare to the isolated recoil of a resilin-like elastomer? We interpret our results by comparing the size-scale dependence of the



Fig. 4 Scaling relations from recoil experiments have a similar size scaling as organisms which use elastically-driven motion and show similar performance limits. The kinematic performance for organisms and engineered devices (from refs. $^{7,10-15,36,37,44-60}$, summarized in Table 1 of ref.⁹) are compared with the limits of the recoil measurements (this work) in terms of A velocity, B acceleration, and C duration. The dashed line is taken using the scaling relations found in Fig. 3 (using $\varepsilon_{in}=3$).

kinematic performance of the model elastomer with the performance of organisms and engineered devices that incorporate elastic elements (Fig. 4). The limits of kinematic performance for the polyurethane elastomer shows a similar size-scaling to elasticallydriven organisms and engineered devices (Table 1). We are cautious in the interpretation of this result because each organism or engineered device in this dataset represents a unique embodiment of material properties and geometry of elastic energy release. Moreover, there are fewer engineered devices in the dataset and they span a narrower range of lengthscales than the organisms. However, the connection between size-scale dependence of the recoil performance and elastically-driven organisms suggests a possible universality to the size-scaling limits of elastic energy release.

Another notable feature that emerges from Fig. 4 is the ability for examples from biology to match the performance of the synthetic elastomer system. The scaling limits of kinematic performance for the elastomer recoil is similar to the performance of hydra, trap-jaw ants, and mantis shrimp. This is impressive for three reasons. First, compared to our isolated polyurethane elastomer, we would expect a diminished performance for organisms because they have load mass that does not contribute to elastic energy storage. For example, in the raptorial appendage of mantis shrimp, the two regions that move furthest (the propodus and dactyl) do not store significant elastic energy⁶, and the added mass of these regions slows the release of elastic energy. Second, dissipation is likely much more significant at the lengthscales of these organisms⁸, and remarkably, both the hydra and mantis shrimp achieve their kinematic performance under water in a viscous environment. Finally, since performance of organisms in the lab is often inferior to that in nature^{38,61}, the kinematic performance of these organisms could potentially be higher in a natural setting. The remarkable performance of hydra, trap-jaw ants, and mantis shrimp despite these hindering factors, suggests that the materials properties of the biological springs are likely critical to their kinematics. While resilin is often discussed as an energy store (going back to refs. ³⁹ and ⁴⁰), many arthropods also use the much harder chitin as a primary material to store energy, as is the case for chitinous springs in locusts ^{24,62}, froghoppers ^{23,63}, plan-thoppers ⁶³, mantis shrimp ⁶⁴, and trap-jaw ants ⁶⁵. Chitin, having an elastic modulus orders of magnitude larger than resilin ⁶⁶, may account for the ability of arthropod systems to surpass the maximums observed in our experiments which use a resilin-like elastomer as the primary energy store. This difference in modulus is consistent with observed elastic mechanisms in the highest performing organisms, including the chitinous exoskeletal elastic materials in mantis shrimp ⁴ and trap-jaw ants ⁶⁷, along with mini-collagen fibrils in hydra ⁶⁰.

The diminished performance of the engineered devices compared to biological organisms demonstrates that there are opportunities for improved design. From our experimental scaling relations, we find that kinematic performance is constrained by a trade-off between resilience, elastic wavespeed, and maximum strain in materials. Recent evidence indicates that similar tradeoffs persist in biological systems. While resilin and elastin are highly resilient materials, their capacity for elastic energy storage is low. This suggests that the coupling of resilient and stiff materials commonly found in biological systems may offset these inherent trade-offs²³. For small-scale devices, performance enhancements could be developed from a bioinspired approach, utilizing composite elastic materials with both resilient and stiff components. Depending on the desired function of the device, this work also suggests some advantage to engineering devices at smaller size-scales to maximize performance (e.g. maximizing acceleration or minimizing duration might be a goal).

The impact of size-scale on kinematic performance is complicated by the choice of using either absolute performance, or scaling the performance relative to body size (relative performance). Biologists examine kinematics of organisms both in an absolute sense (a cheetah runs more quickly than an ant) and in a relative sense (relative to body size, some ants are faster than cheetahs). Relative performance of running (body lengths per

second) and jumping (jump height per body length) have been used to characterize both biological organisms^{36,68–70} and engineered devices ^{10,37,71,72}. In biology, relative size has been used to standardize for size differences between animals in the same species⁶⁸ or across several species⁶⁹, and it has been suggested that relative performance is more ecologically relevant as it correlates well with the ability to evade predators⁷³. Relative performance can also be used to normalize for drag effects, which become significant at small size-scales⁸. However, in contrast to the prevalent use of relative performance, we find that absolute velocity is a size-scale independent quantification of elastic performance for the lengthscales probed in the current work. Higher relative velocities (along with higher accelerations or shorter durations) can be achieved simply by reducing the size of an elastic element. Therefore, comparing the performance of systems that are orders of magnitude different in size-scale requires caution.

3 Conclusions

In summary, we have measured the kinematic performance of elastic energy release for a resilient elastomer. In agreement with expected scaling relations, the maximum center-of-mass velocity of a freely retracting band is independent of length, and depends only on the initial strain at which the band was released and the elastic wavespeed of the material. The maximum center-of-mass acceleration and duration of elastic energy release were found to depend on the length of the elastomer band, with an improved performance at smaller size-scales. Previously reported measurements of kinematic performance for elastically-driven organisms and engineered micro-robotic devices show similar size-scaling to the elastomer studied here. The current results, which probe the upper bound of elastically-driven kinematics of a resilin-like material, show a similar limit as some of the fastest biological systems. Future work which seeks to delineate the role of elastic wavespeed, maximum strain, and resilience in elastic biological systems could lead to a foundational understanding for improved engineering design. Specifically, the mechanical properties of resilin, chitin, and resilin/chitin composites would be of great importance to compare to engineered systems.

4 Conflicts of interest

The are no conflicts to declare.

Table 1 Dependence of velocity, acceleration, and duration on the characteristic lengthscale (L_c) for recoil measurements along with two parameter power law fits to the organisms and engineered devices in Fig. 4. Here we report the power law exponent α and 95% confidence intervals obtained by fitting to AL_c^{α} (using a linear regression on a log-transform of the data), where both A and α are adjustable fitting parameters.

	Recoil α	Organisms α	Devices α
Velocity	0	$-0.1{\pm}0.5$	$0.2{\pm}0.4$
Acceleration	-1	$-0.9{\pm}0.7$	$-0.5{\pm}0.8$
Duration	1	$1.1{\pm}0.4$	$0.9{\pm}0.7$

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Notes and references

- 1 R. S. James, C. A. Navas and A. Herrel, *Journal of Experimental Biology*, 2007, **210**, 923–933.
- 2 S. N. Patek, D. M. Dudek and M. V. Rosario, *Journal of Experimental Biology*, 2011, **214**, 1973–1980.
- 3 S. J. Longo, S. M. Cox, E. Azizi, M. Ilton, J. P. Olberding, R. St Pierre and S. N. Patek, *Journal of Experimental Biology*, 2019, **222**, year.
- 4 S. N. Patek, W. L. Korff and R. L. Caldwell, *Nature*, 2004, **428**, 819–820.
- 5 T. I. Zack, T. Claverie and S. N. Patek, *Journal of Experimental Biology*, 2009, **212**, 4002–4009.
- 6 M. V. Rosario and S. N. Patek, *Journal of Morphology*, 2015, **276**, 1123–1135.
- 7 M. J. McHenry, P. S. Anderson, S. Van Wassenbergh, D. G. Matthews, A. P. Summers and S. N. Patek, *The Journal of Experimental Biology*, 2016, **219**, 3399–3411.
- 8 H. C. Bennet-Clark, *Scale Effects in Animal Locomotion*, Academic Press Ltd., London, 1977, pp. 185–201.
- 9 M. Ilton, M. S. Bhamla, X. Ma, S. M. Cox, L. L. Fitchett, Y. Kim, J.-S. Koh, D. Krishnamurthy, C.-Y. Kuo, F. Z. Temel, A. J. Crosby, M. Prakash, G. P. Sutton, R. J. Wood, E. Azizi, S. Bergbreiter and S. N. Patek, *Science*, 2018, **360**, eaa01082.
- 10 M. Kovač, M. Fuchs, A. Guignard, J. C. Zufferey and D. Floreano, Proceedings - IEEE International Conference on Robotics and Automation, 2008, pp. 373–378.
- 11 A. Debray, Bioinspiration & Biomimetics, 2011, 6, 026002.
- 12 J. S. Koh, S. P. Jung, M. Noh, S. W. Kim and K. J. Cho, Proceedings IEEE International Conference on Robotics and Automation, 2013, pp. 26–31.
- 13 J. Zhao, J. Xu, B. Gao, N. Xi, F. J. Cintron, M. W. Mutka and L. Xiao, *IEEE Transactions on Robotics*, 2013, **29**, 602–614.
- 14 S.-W. Kim, J.-S. Koh, J.-G. Lee, J. Ryu, M. Cho and K.-J. Cho, *Bioinspiration & Biomimetics*, 2014, **9**, 036004.
- 15 J.-S. Koh, E. Yang, G.-P. Jung, S.-P. Jung, J. H. Son, S.-I. Lee, P. G. Jablonski, R. J. Wood, H.-Y. Kim and K.-J. Cho, *Science*, 2015, **349**, 517–521.
- 16 V. Zaitsev, O. Gvirsman, U. Ben Hanan, A. Weiss, A. Ayali and G. Kosa, IEEE International Conference on Intelligent Robots and Systems, 2015, p. 066012.
- 17 D. W. Haldane, M. M. Plecnik, J. K. Yim and R. S. Fearing, *Science Robotics*, 2016, 1, eaag2048.
- 18 G. P. Sutton, E. Mendoza, E. Azizi, S. J. Longo, J. P. Olberding, M. Ilton and S. N. Patek, *Integrative and Comparative Biology*, 2019.
- 19 S. M. Cox, D. Schmidt, Y. Modarres-Sadeghi and S. N. Patek, *Bioinspiration and Biomimetics*, 2014, **9**, 16014.

- 20 W. Lee, *M.Sc. Thesis*, Massachusetts Institute of Technology, 1990.
- 21 M. F. Ashby, *Materials Selection in Mechanical Design*, Butterworth-Heinemann, Oxford, 4th edn, 2011.
- 22 K. Maier, Product Engineering, 1957, 28, 167-174.
- 23 M. Burrows, S. R. Shaw and G. P. Sutton, *BMC Biology*, 2008, 6, 41.
- 24 M. Burrows and G. P. Sutton, *Journal of Experimental Biology*, 2012, **215**, 3501–3512.
- 25 M. Burrows, The Journal of Experimental Biology, 2016, **219**, 2449–2457.
- 26 R. J. King, *M.Sc. Thesis*, Virginia Polytechnic Institute and State University, 2010.
- 27 J. Gosline, M. Lillie, E. Carrington, P. Guerette, C. Ortlepp and K. Savage, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 2002, **357**, 121–132.
- 28 C. M. Elvin, A. G. Carr, M. G. Huson, J. M. Maxwell, R. D. Pearson, T. Vuocolo, N. E. Liyou, D. C. C. Wong, D. J. Merritt and N. E. Dixon, *Nature*, 2005, 437, 999–1002.
- 29 R. Vermorel, N. Vandenberghe, E. Villermaux, N. V. R. Vermorel and E. Villermaux, Proceedings of the Royal Society A: Mathematical, Physical and Engineering Sciences, 2007, 463, 641–658.
- 30 R. B. Bogoslovov and C. M. Roland, *Journal of Applied Physics*, 2007, **102**, 1–6.
- 31 J. Niemczura and K. Ravi-Chandar, *Journal of the Mechanics* and *Physics of Solids*, 2011, **59**, 457–472.
- 32 L. B. Tunnicliffe, A. G. Thomas and J. J. Busfield, *Polymer Testing*, 2015, **47**, 36–41.
- 33 S. H. Yoon and C. R. Siviour, Journal of Dynamic Behavior of Materials, 2017, 3, 12–22.
- 34 A. T. Oratis and J. C. Bird, Phys. Rev. Lett., 2019, 122, 014102.
- 35 S. M. Rogers, J. Riley, C. Brighton, G. P. Sutton, D. A. Cullen and M. Burrows, *Journal of Experimental Biology*, 2016, 219, 635–648.
- 36 M. Burrows, Journal of Experimental Biology, 2006, **209**, 4607–4621.
- 37 A. P. Gerratt and S. Bergbreiter, *Smart Materials and Structures*, 2013, **22**, 014010.
- 38 D. J. Irschick, Integrative and Comparative Biology, 2003, 43, 396–407.
- 39 T. Weis-Fogh, Journal of Molecular Biology, 1961, 3, 520–531.
- 40 H. C. Bennet-Clark and E. C. Lucey, *The Journal of Experimental Biology*, 1967, **47**, 59–67.
- 41 M. Jensen and T. Weis-Fogh, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 1962, **245**, 137–169.
- 42 H. Schwetlick and T. Schütze, *BIT Numerical Mathematics*, 1995, **35**, 361–384.
- 43 T. Schütze and H. Schwetlick, *BIT Numerical Mathematics*, 1997, **37**, 105–137.
- 44 V. Zaitsev, O. Gvirsman, U. Ben Hanan, A. Weiss, A. Ayali and G. Kosa, *Bioinspiration & Biomimetics*, 2015, 10, 066012.
- 45 R. Niiyama, A. Nagakubo and Y. Kuniyoshi, Proceedings IEEE

International Conference on Robotics and Automation, 2007, pp. 2546–2551.

- 46 S. Kajita, F. Kanehiro, K. Kaneko, K. Fujiwara, K. Harada, K. Yokoi and H. Hirukawa, Proceedings 2003 IEEE/RSJ International Conference on Intelligent Robots and Systems (IROS 2003) (Cat. No.03CH37453), 2003, pp. 1644–1650.
- 47 W. A. Churaman, L. J. Currano, C. J. Morris, J. E. Rajkowski and S. Bergbreiter, *Journal of Microelectromechanical Systems*, 2012, **21**, 198–205.
- 48 Y. Forterre, J. M. Skotheim, J. Dumals and L. Mahadevan, *Nature*, 2005, **433**, 421–425.
- 49 O. Vincent, C. Weisskopf, S. Poppinga, T. Masselter, T. Speck,
 M. Joyeux, C. Quilliet and P. Marmottant, *Proceedings of the Royal Society B: Biological Sciences*, 2011, 278, 2909–2914.
- 50 H. C. Bennet-Clark, *Journal of Experimental Biology*, 1975, **63**, 53–83.
- 51 C. V. Anderson, Scientific Reports, 2016, 6, 18625.
- 52 C. V. Anderson, T. Sheridan and S. M. Deban, *Journal of Morphology*, 2012, **273**, 1214–1226.
- 53 R. S. James and R. S. Wilson, Physiological and Biochemical Zoology, 2008, 81, 176–185.
- 54 M. Burrows, Journal of Experimental Biology, 2012, 215, 3612–3621.
- 55 S. N. Patek, J. E. Baio, B. L. Fisher and A. V. Suarez, Proceedings of the National Academy of Sciences, 2006, 103, 12787– 12792.
- 56 D. L. Whitaker, L. A. Webster and J. Edwards, *Functional Ecology*, 2007, **21**, 219–225.
- 57 J. Edwards, D. Whitaker, S. Klionsky and M. J. Laskowski, *Nature*, 2005, **435**, 164.
- 58 A. Pringle, S. N. Patek, M. Fischer, J. Stolze and N. P. Money, *Mycologia*, 2005, **97**, 866–871.
- 59 A. W. Koch, T. W. Holstein, C. Mala, E. Kurz, J. Engel and C. N. David, *Journal of Cell Science*, 1998, **111**, 1545–1554.
- 60 T. Nüchter, M. Benoit, U. Engel, S. Özbek and T. W. Holstein, *Current Biology*, 2006, **16**, R316–R319.
- 61 H. C. Astley, E. M. Abbott, E. Azizi, R. L. Marsh and T. J. Roberts, *Journal of Experimental Biology*, 2013, **216**, 3947– 3953.
- 62 C. Wan, Z. Hao and X. Feng, Scientific reports, 2016, 6, 35219.
- 63 I. Siwanowicz and M. Burrows, *eLife*, 2017, 6, e23824.
- 64 M. Tadayon, S. Amini, A. Masic and A. Miserez, *Advanced Functional Materials*, 2015, **25**, 6437–6447.
- 65 F. J. Larabee, W. Gronenberg and A. V. Suarez, *The Journal of Experimental Biology*, 2017, **220**, 3062–3071.
- 66 J. F. Vincent and U. G. Wegst, Arthropod Structure and Development, 2004, **33**, 187–199.
- 67 W. Gronenberg, Journal of Comparative Physiology A, 1996, 178, 727–734.
- 68 R. Avery and D. Bond, *Amphibia-Reptilia*, 1989, **10**, 77–84.
- 69 J. C. Spagna and A. M. Peattie, *Journal of Insect Physiology*, 2012, **58**, 599–606.
- 70 M. V. Rosario, G. P. Sutton, S. N. Patek and G. S. Sawicki, Proceedings of the Royal Society B, 2016, **283**, 20161561.

- 71 U. Saranli, M. Buehler and D. E. Koditschek, *The International Journal of Robotics Research*, 2001, **20**, 616–631.
- 72 W. Gao, S. Sattayasamitsathit and J. Wang, Chemical Record,

2012, **12**, 224–231.

73 R. Van Damme and T. J. M. Van Dooren, *Animal Behaviour*, 1999, **57**, 347–352.