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**Self-Assembly with Fluorescence Readout in a Free Base
Dipyrrin-Polymer Triggered by Metal Ion Binding in
Aqueous Solution**

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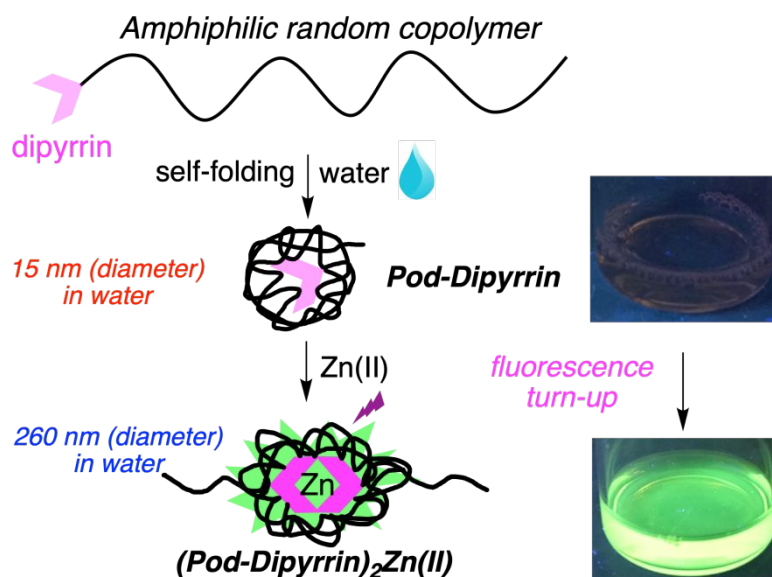
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26 **TOC graphic**



Profound morphological and fluorogenic changes ensue upon binding of zinc ion by two polymers, each of which bears a single dipyrrin at one terminus, forming the bis(dipyrrinato)Zn(II) complex.

Abstract

A hydrophobic dipyrin has been attached to a heterotelechelic amphiphilic random copolymer as part of a single-polymer–single-cargo strategy. The polymer–dipyrin (“Pod–dipyrin”) exhibits an extended conformation in the organic solvent *N,N*-dimethylformamide (DMF) but in aqueous solution forms a unimeric nanoparticle (i.e., a foldamer) as indicated by dynamic light-scattering (DLS) data (hydrodynamic diameter $D_h = 140$ nm versus 15 nm in the two solvents, respectively). Treatment of the Pod–dipyrin with Zn(II) in aqueous solution causes formation of a bis(Pod–dipyrinato)Zn(II) complex as evidenced by (1) a slight change in absorption spectrum, (2) ~50-fold increase in fluorescence quantum yield (Φ_f from 0.0035 to 0.17), and (3) profound increase in D_h (from 15 nm to 260 nm). The D_h value for the bis(Pod–dipyrinato)Zn(II) complex is only slightly smaller in water (260 nm) than in DMF (310 nm), where a more extended conformation is expected. The increase in fluorescence of the bis(Pod–dipyrinato)Zn(II) complex is readily observed by visual inspection (i.e., naked-eye readout). The conversion of a compact folded unimer (15 nm) to a more extended dimer (260 nm) triggered by metal ion addition denotes a conformationally malleable, environmentally sensitive molecular architecture. The self-assembly process can be reversed upon treatment with a metal-ion sequestering agent such as EDTA. The facile synthesis of a fluorogenic architecture that undergoes profound spontaneous change in molecular morphology upon binding an exogenous agent in water under physiological conditions may open a number of opportunities for sensory-mechanical studies in the life sciences.

Introduction

The absorption and fluorescence spectra of organic chromophores make for diverse applications in the life sciences yet the tailoring of chromophores for solubility in aqueous media often presents significant challenges. Absorption and fluorescence in the visible region inevitably entails a sizeable π -chromophore, at least for intense π - π^* transitions. The vast majority of dyes contain a chromophore that bears intrinsic charge,¹ which also facilitates solubilization in aqueous media. For chromophores that lack intrinsic charge (e.g., carotenoids, perylenes, quinones, tetrapyrroles),¹ the challenge of aqueous solubilization is acute. While solubilizing substituents can often be incorporated, in many cases the size of the π -system requires construction of a daunting superstructure to impart adequate aqueous solubility.

We recently reported a new and simpler approach for aqueous solubilization of neutral or moderately polar chromophores.² The approach – extending extensive work by the groups of Sawamoto^{3,4} and Zimmerman^{5,6} – entails covalent attachment of the chromophore to one terminus of an amphiphilic random copolymer, thereby affording a single-polymer–single-chromophore construct. The polymer “folds” around the unadorned chromophore and enables the resulting “pod-chromophore” to dissolve in aqueous solution. This approach separates the chromophore design and aqueous solubilization into two separate spheres – chromophore cargo and polymer package – with covalent joining of the two entities in the final synthetic step. To date, the polymer that we have employed (**F-Ph**)² is a polyacrylate/polyacrylamide containing three types of pendant groups and distinct functional groups at the two backbone termini (i.e., heterotelechelic). The three types of pendant groups: a methoxy-terminated oligoethylene glycol (PEG9; polar but nonionic), a sulfonate-terminated short alkyl chain (ionic), and a lauryl (hydrophobic) group are incorporated from three corresponding monomers via living radical polymerization. The chain-transfer agent in the polymerization ultimately provides the two

terminal functional groups, which in this case are carboxylic acid and dithiobenzoate groups (Chart 1). The success relied on identification of the appropriate ratios of the three monomers (m:n:p = 1:1:5 ratio of PEG9, lauryl and sulfonate-terminated units), affording polymer **F-Ph**² of size 40 kDa where m, n ~20 and p ~100.

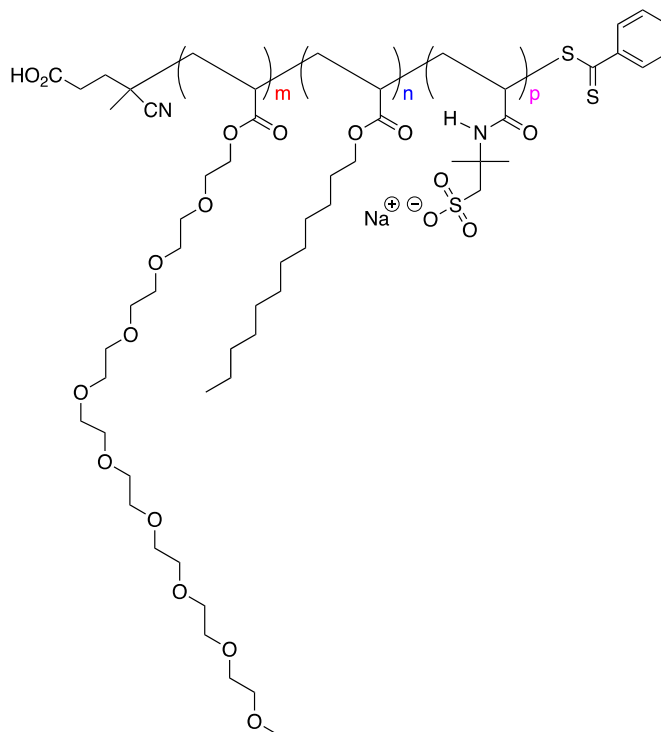


Chart 1. General design of the heterotelechelic amphiphilic random copolymer for encapsulation of a covalently attached hydrophobic fluorophore in aqueous solution. For **F-Ph**, m:n:p = 1:1:5 ratio with overall molecular weight of 40 kDa (m, n ~20 and p ~100).

This foldamer approach proved viable with 8 classes of chromophores including those that are neutral and hydrophobic (bacteriochlorin, BODIPY, chlorin, coumarin, perylene, phthalocyanine) or intrinsically charged (cyanine, rhodamine).² The presence of one and only one chromophore per pod resulted in absorption and fluorescence features of the pod-chromophore in aqueous solution that were essentially identical to those of the hydrophobic chromophore in an organic solvent (e.g., toluene; *N,N*-dimethylformamide, DMF). In addition, the fluorescence quantum yield (Φ_f) was little changed for the pod-chromophore in water versus

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3 the chromophore alone in organic media, thus exhibiting none of the insidious diminution of
4 fluorescence brightness⁷ so often encountered upon placing an organic chromophore in aqueous
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6 media.
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10 One objective in developing the single-polymer–single-chromophore foldamer approach
11 was to embed the chromophore in the inner confines of the polymer, insulated from the
12 surrounding environment. One chromophore examined was a fluorogenic rhodamine–lactam
13 that upon exposure to metal ions is known to undergo ring-opening, thereby eliciting
14 fluorescence. Upon treatment of the **Pod-Rhodamine** with various metal ions, the fluorescence
15 turn-on phenomenon was observed.² This result implies that the fluorophore is not irrevocably
16 incarcerated in the inner space of the folded polymer but does have exposure or at least
17 excursions to the outside milieu.
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21 Here, we report our results concerning the metal binding of a single dipyrin in the
22 polymer. We first describe the synthesis of a maleimide-containing free base dipyrin, then
23 attach the dipyrin to the thiol group in the heterotelechelic amphiphilic random copolymer
24 described above. The resulting single-polymer–single-dipyrin architecture (**Pod-Dipyrin**) was
25 examined for binding of divalent zinc and copper ions. The formation of the
26 bis(dipyrinato)metal(II) complex would entail intermolecular interaction of two pods rather than
27 interaction with a single pod as in the **Pod-Rhodamine** study. The fluorogenic response here
28 also is completely distinct from the common process where amine-based photoinduced electron
29 transfer is suppressed upon metal binding, thereby turning on fluorescence. The studies have led
30 to new insights concerning fluorescence of the free base dipyrin, intrinsic properties of the pod
31 architecture, and morphological changes of the polymer–dipyrin entity upon binding metal ions.
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Results and Discussion

1. Dipyrin Molecular Design. Free base dipyrins bearing diverse substituents react readily with various metal ions (such as Zn^{2+} , Mg^{2+} , Ni^{2+} , Co^{2+} , Cu^{2+} , Pd^{2+} , In^{3+} , Ga^{3+} , etc.) to form the corresponding bis(dipyrinato)metal(II) (or tris(dipyrinato)metal(III)) complexes. The bis(dipyrinato)metal(II) complexes absorb strongly in the blue-green region ($\epsilon \sim 50,000\text{--}100,000 \text{ M}^{-1}\text{cm}^{-1}$). Although first reported by Fischer in 1924,⁸ the complexes were long considered to be non-fluorescent and were little investigated.⁹ Replacement of the phenyl group at the dipyrin 5-position with the sterically encumbering mesityl group increased the fluorescence quantum yield of the corresponding bis(dipyrinato)Zn(II) complexes by a factor of 60, from $\Phi_f = 0.006$ of the 5,5'-phenyl substituted complex **I** to 0.36 of the 5,5'-mesityl substituted complex **II** (Chart 2).¹⁰ The very large increase in fluorescence intensity makes the resulting complexes viable for use as legitimate fluorescent dyes. Accordingly, for the free base dipyrin here, we chose an analogous system wherein the 5-aryl group is equipped with 2,6-dimethyl groups and a tether at the 4-position. The presence of an amino or alkoxy group at the *p*-position of the 5-phenyl group is known to cause quenching of the BODIPY analogues¹¹ (presumably via internal charge transfer), hence we chose to join the tether to the aryl *p*-position via a carbonyl group.

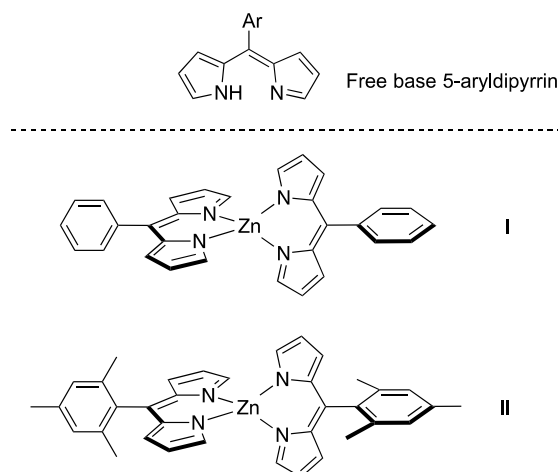
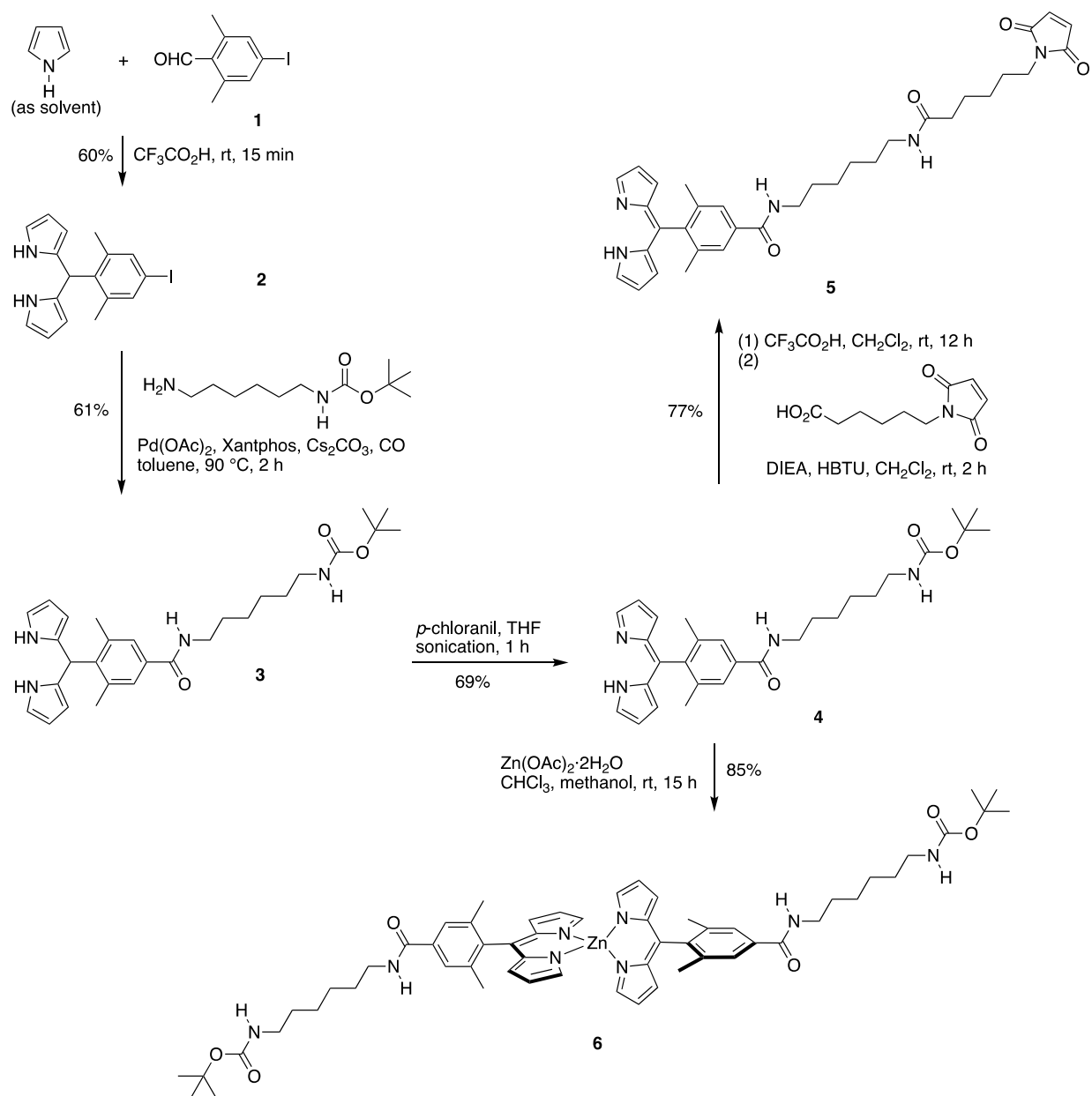


Chart 2. Free base dipyrin and two bis(dipyrinato)Zn(II) complexes.

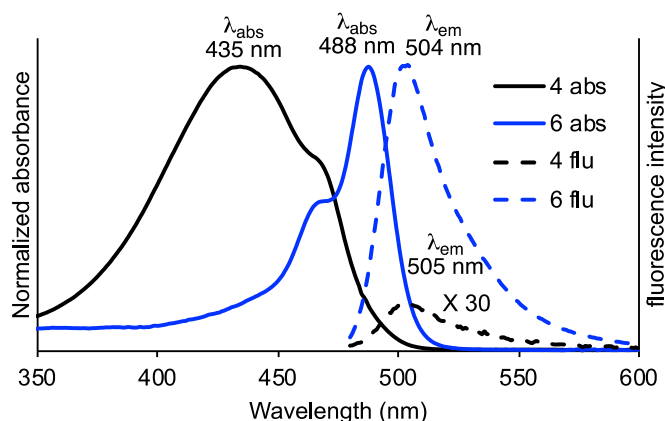
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3 **2. Synthesis of the Bioconjugatable Dipyrin.** The known aryl aldehyde 4-iodo-2,6-
4 dimethylbenzaldehyde (**1**)^{12,13} was dissolved in pyrrole (serving as reactant in excess and solvent)
5 and treated with trifluoroacetic acid at room temperature in a one-flask synthesis^{14,15} (Scheme 1).
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7 The corresponding dipyrromethane **2** was obtained in 60% yield without aqueous/organic
8 extraction. The aminocarbonylation of **2** with *N*-(*tert*-butoxycarbonyl)-1,6-diaminohexane was
9 attempted using Mo(CO)₆ as a solid source of carbon monoxide,¹⁶⁻²⁰ but the mono- and di-
10 carbonylated products were obtained in yields of 11% and 10%, respectively. Upon use of
11 Pd(OAc)₂/Xantphos/Cs₂CO₃ in an atmosphere of CO,^{21,22} only the desired product
12 dipyrromethane **3** was obtained in 61% yield. Treatment of **3** with *p*-chloranil in THF at room
13 temperature for 24 h afforded **4** in 69% yield. The oxidation could be completed within 1 h by
14 sonicating the reaction mixture instead of stirring at room temperature for 24 h. The removal of
15 the *tert*-butoxycarbonyl group of **4** and introduction of the maleimido moiety were carried out in
16 a single flask process. Thus, treatment of **4** with trifluoroacetic acid in CH₂Cl₂ for 12 h caused
17 removal of the *tert*-butoxycarbonyl group as confirmed by TLC analysis. The resulting residue
18 after the removal of the volatile substances was treated with 6-maleimidohexanoic acid and *O*-
19 (benzotriazol-1-yl)-*N,N,N',N'*-tetramethyluronium hexafluorophosphate (HBTU) in CH₂Cl₂
20 containing *N,N*-diisopropylethylamine (DIEA) for 2 h. Workup and chromatography afforded
21 dipyrin **5** in 77% yield. Finally, treatment of the dipyrin **4** with zinc acetate afforded
22 bis(dipyrinato)Zn(II) complex **6**. Each new compound was characterized by ¹H NMR and ¹³C
23 NMR spectroscopy as well as accurate mass determination by electrospray ionization mass
24 spectrometry (ESI-MS). The ¹H NMR spectrum of the free base dipyrins (**4**, **5**) showed peaks
25 due to all protons, with the exception of that of the pyrrole N-H in the dipyrin moiety. This
26 feature has been observed previously with other free base dipyrins.⁹
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Scheme 1. Synthesis of dipyrromethanes **2** and **3**; dipyrins **4** and **5**; and bis(dipyrinato)Zn(II) complex **6**.

The absorption and fluorescence properties of the bis(dipyrinato)Zn(II) complex **6** were examined in toluene (Figure 1). Coordination complex **6** has sharp absorption (fwhm 35 nm) and fluorescence (fwhm 32 nm) bands compared to those of free base dipyrin **4** in toluene. The bis(dipyrinato)Zn(II) complex **6** exhibits $\Phi_f = 0.16$, to be compared with 0.0011 of the free base dipyrin **4**. For these and all subsequent measurements, the fluorescence standard is a BODIPY-

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3 hydrazide ($\Phi_f = 0.97$).² Self-assembly of the two free base dipyrins into a bis(dipyrinato)Zn(II)
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5 complex equipped with 2,6-dimethylaryl moieties affords a substantially brighter chromophore.
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7 The fluorescence spectra of matched samples of **6** and **II** in toluene were found to be essentially
8
9 identical, although the Φ_f value of **6** (0.18) was exactly half that of **II** (0.36)¹⁰ using **II** as the
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12 standard. The complexes differ only in the nature of the *p*-aryl substituent (amido versus methyl);
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14 hence the amido substituent causes diminution of the fluorescence intensity.
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32 **Figure 1.** Electronic spectra in toluene at room temperature. Absorption (blue solid line, $\lambda_{\max} =$
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34 488 nm) and fluorescence ($\lambda_{\text{exc}} 460$ nm, blue dashed line, $\lambda_{\max} = 504$ nm) of bis(dipyrinato)Zn(II)
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36 complex **6**, and absorption (black solid line, $\lambda_{\max} = 435$ nm) and fluorescence ($\lambda_{\text{exc}} 390$ nm, black
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38 dashed line, $\lambda_{\max} = 505$ nm) of free base dipyrin **4**.
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42 The absorption and fluorescence spectral properties of the bis(dipyrinato)Zn(II) complex
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44 **6** also were examined in organic solvents of a range of polarity. The solvents range from the
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46 nonpolar toluene to the quite polar DMF. The spectra are displayed in Figure 2. The absorption
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48 maximum varies little (from 482–488 nm) although slightly greater variation occurs in the
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50 position of the fluorescence maximum (493–503 nm), giving a Stokes shift range of 9–15 nm
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52 (Table 1). The Φ_f values decline with increasing polarity, from $\Phi_f = 0.18$ in toluene to 0.041 in
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54 DMF, although the spectra shapes are little changed with solvent polarity.
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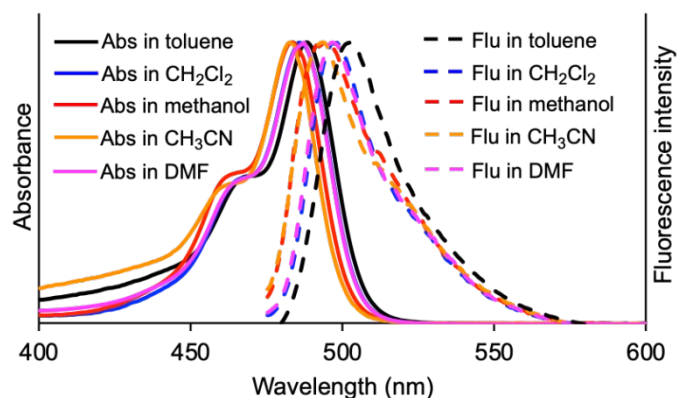


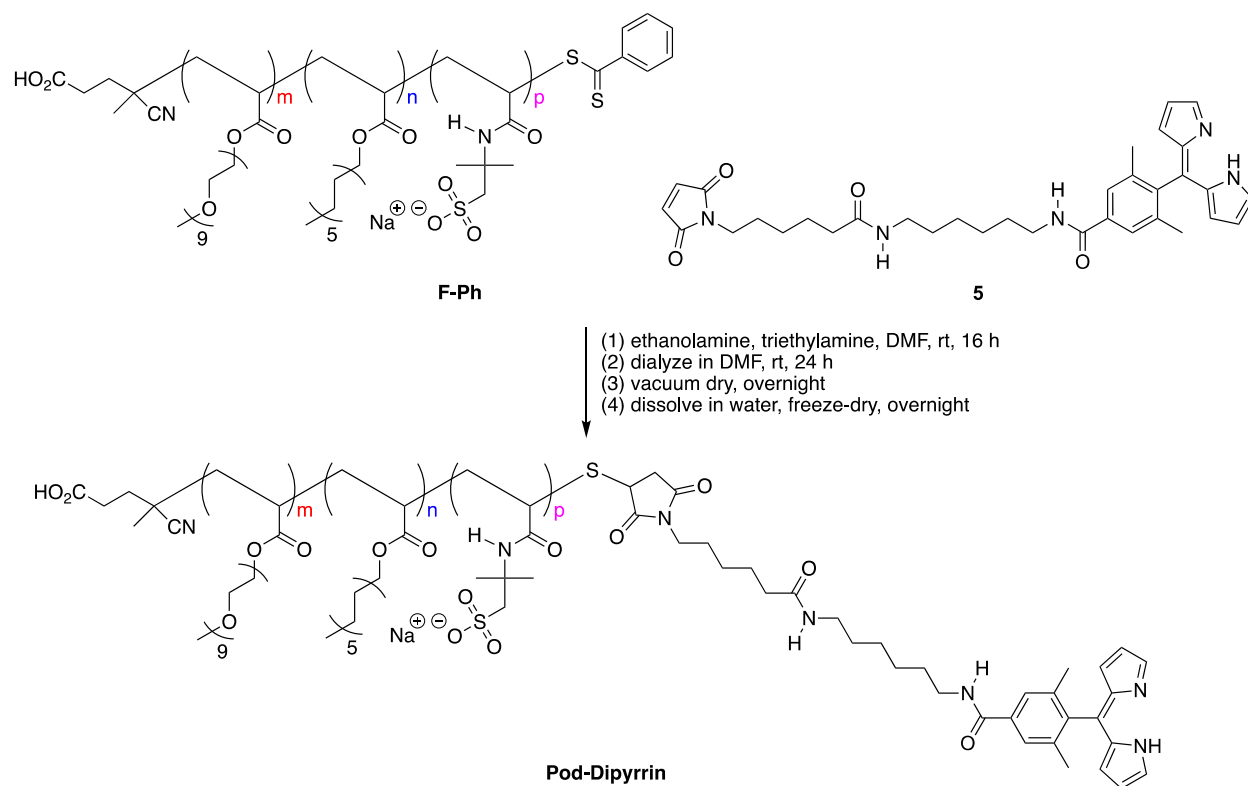
Figure 2. Absorption and fluorescence spectra (normalized) of **6** in toluene (black line), dichloromethane (blue line), methanol (red line), acetonitrile (orange line), and DMF (magenta line).

Table 1. Spectral properties of **6** in organic solvents.

Entry	Solvent	Dielectric constant	λ_{abs} (nm)	λ_{em} (nm)	Stokes shift (nm)	fwhm λ_{em} (nm)	Φ_f^a
1	toluene	2.4	488	503	15	30	0.18
2	CH ₂ Cl ₂	9.1	486	498	12	27	0.079
3	methanol	32.6	485	494	9	35	0.032
4	acetonitrile	36.6	482	493	11	34	0.050
5	DMF	38.3	487	496	9	27	0.041

^aAll yield data were obtained with compound **II** as a reference ($\Phi_f = 0.36$).¹⁰

3. Synthesis of the Polymer–Dipyrrin Conjugate. Synthesis of **Pod-Dipyrrin** follows the procedure employed previously² (Scheme 2). Treatment of polymer **F-Ph** in DMF containing triethylamine and ethanolamine liberated the free thiol, which was reacted *in situ* with dipyrrin-maleimide **5**. The crude reaction mixture in DMF was dialyzed to remove any free fluorophore, which has been readily achieved previously with diverse chromophores.² The conjugation and purification process required ~4 days, whereupon **Pod-Dipyrrin** was obtained as a yellow friable powder.



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Scheme 2. Synthesis of **Pod-Dipyrrin**.

4. Characterization of the Pod-Dipyrrin. The **Pod-Dipyrrin** was examined by ^1H NMR spectroscopy in D_2O at room temperature. Characteristic peaks were observed for the polymer constituents but not of the dipyrrin moiety, which is not surprising given that the latter is present on one terminus of the polymer and represents only $\sim 1.5\%$ of the mass of the construct. While the extent of conjugation cannot be readily established by NMR analysis, evidence presented below supports the interpretation that the conjugation has gone to completion. We turned to studies of the photophysical properties of **Pod-Dipyrrin** in water. The absorption spectrum of **Pod-Dipyrrin** (Figure 3A, black solid line) in aqueous solution shows a narrower absorption peak (full-width-at-half-maximum, fwhm 39 nm) compared to that of the hydrophobic benchmark **5** (fwhm 80 nm) in toluene (Figure 3B, red solid line). The narrower absorption feature may stem from restricted conformational motion of the dipyrrin unit in **Pod-**

Dipyrrin due to the folded state of the amphiphilic polymer in aqueous solution, whereas the dipyrrin chromophore of hydrophobic benchmark **5** has a fuller range of motion in toluene. The motions include butterfly flexing^{10,23} of the dihedral planes of the pyrroles with respect to each other, and the rotation of the aryl group about its *p*-substituted axis. **Pod-Dipyrrin** in water has a similar weak and broad fluorescence peak compared to that of **5** in toluene.

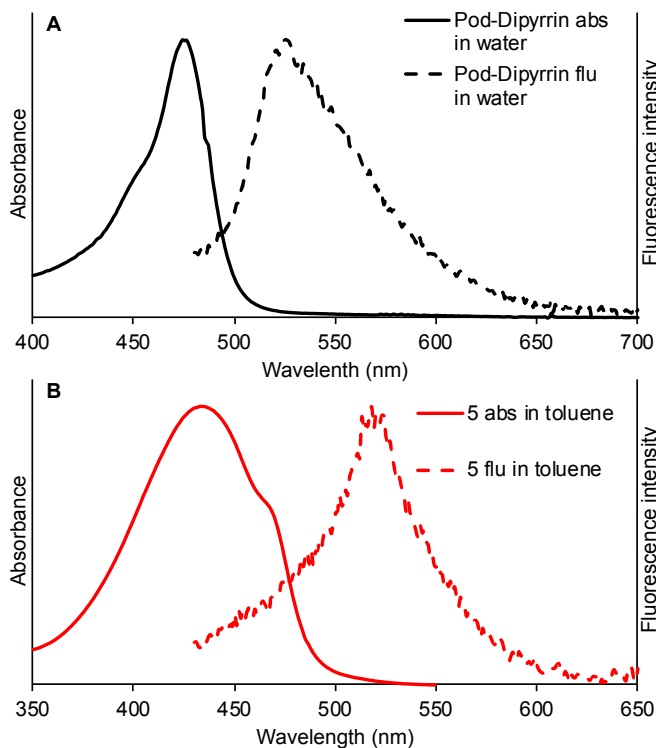


Figure 3. (A) Absorption (solid line) and fluorescence (λ_{exc} 460 nm, dashed line) spectra of **Pod-Dipyrrin** in water; and (B) absorption (red solid line) and fluorescence (λ_{exc} 390 nm, red dashed line) spectra of free base dipyrrin **5** in toluene.

The **Pod-Dipyrrin** was examined by dynamic light-scattering (DLS) in aqueous solution at concentrations ranging from 10–1 mg/mL, the approximate lower limit of the DLS instrument. Essentially identical size distributions were observed over this concentration range indicating the integrity of the folded architecture (Figure 4A). On the other hand, the **Pod-Dipyrrin** showed a profound difference in molecular size in DMF versus water (Figure 4B). In DMF the size

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3 distribution is peaked at a hydrodynamic diameter $D_h = 140$ nm, whereas the distribution peak is
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5 15 nm in water. The results are consistent with folding to form a unimer in aqueous solution
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7 from a more extended and unstructured set of conformations in the organic solvent DMF. The
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9 absorption spectrum of **Pod-Dipyrrin** (Figure 4C) also is much narrower in water (fwhm 39 nm)
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11 than in DMF (fwhm 79 nm). These absorption spectral changes are similar to those for **Pod-**
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13 **Dipyrrin** in water versus **5** in toluene (Figure 3B). The data are fully consistent with
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15 expectations concerning folding in aqueous solution on the basis of prior data of the same
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17 polymer bearing other families of chromophores.²
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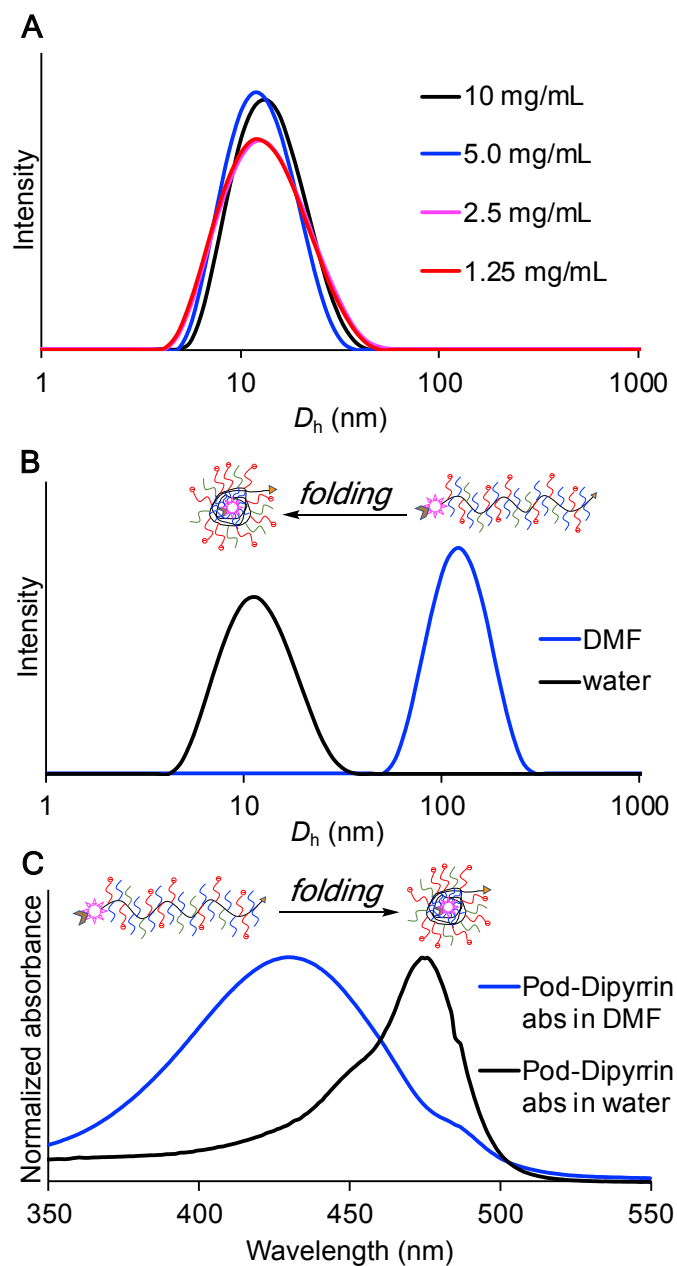


Figure 4. Properties of **Pod-Dipyrin** at room temperature. (A) DLS data in 1 M aqueous NaCl at various concentrations: (A) 10 mg/mL, average diameter 14.8 nm; 5.0 mg/mL, 13.4 nm; 2.5 mg/mL, 14.8 nm; and 1.25 mg/mL, 14.5 nm. (B) DLS data in DMF (blue line) and water (black line) at 1.0 mg/mL, and (C) absorption in DMF (blue line) and water (black line) at 1.0 mg/mL.

5. Self-assembly of the Pod-Dipyrin via Metal Ion Chelation in Water. We have demonstrated that a related polymer–fluorophore is able to bind metals in aqueous solution.² Here, we tested the ability of **Pod-Dipyrin** to form dimeric pod assemblies owing to formation

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3 of the dipyrinato)metal(II) complexes upon the introduction of divalent metal ions (Zn(II) and
4 Cu(II)). A solution of **Pod-Dipyrrin** in aqueous solution was treated with excess Zn(OAc)₂ (440
5 equiv) at room temperature. Examination by absorption spectroscopy after 5 minutes revealed a
6 bathochromic shift (12 nm) compared to the monomeric **Pod-Dipyrrin** in water (Figure 5A).
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8 The fluorescence quantum yield ($\Phi_f = 0.17$) also increased profoundly (~50-fold) compared to
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10 that of **Pod-Dipyrrin** ($\Phi_f = 0.0035$ in water) (Figure 5B). Such changes in absorption and
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12 fluorescence are attributed to the formation of the bis(dipyrinato)Zn(II) complex, which must
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14 then join two pod architectures; the resulting assembly is termed **(Pod-Dipyrrin)₂Zn(II)**. The
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16 aforementioned spectral changes are visibly evident: (1) the free base dipyrin in solution is
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18 yellow whereas the bis(dipyrinato)Zn(II) complex is light yellow; and (2) upon illumination, no
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20 fluorescence is visible from the free base dipyrin whereas the bis(dipyrinato)Zn(II) complex
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22 exhibits a green glow. The fluorescence properties of the **Pod-Dipyrrin** species in water are
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24 shown in Table 2 compared with **Pod-Dipyrrin** (in DMF) and **6** in toluene. The spectral
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26 properties of the **(Pod-Dipyrrin)₂Zn(II)** are described further in the Discussion section.
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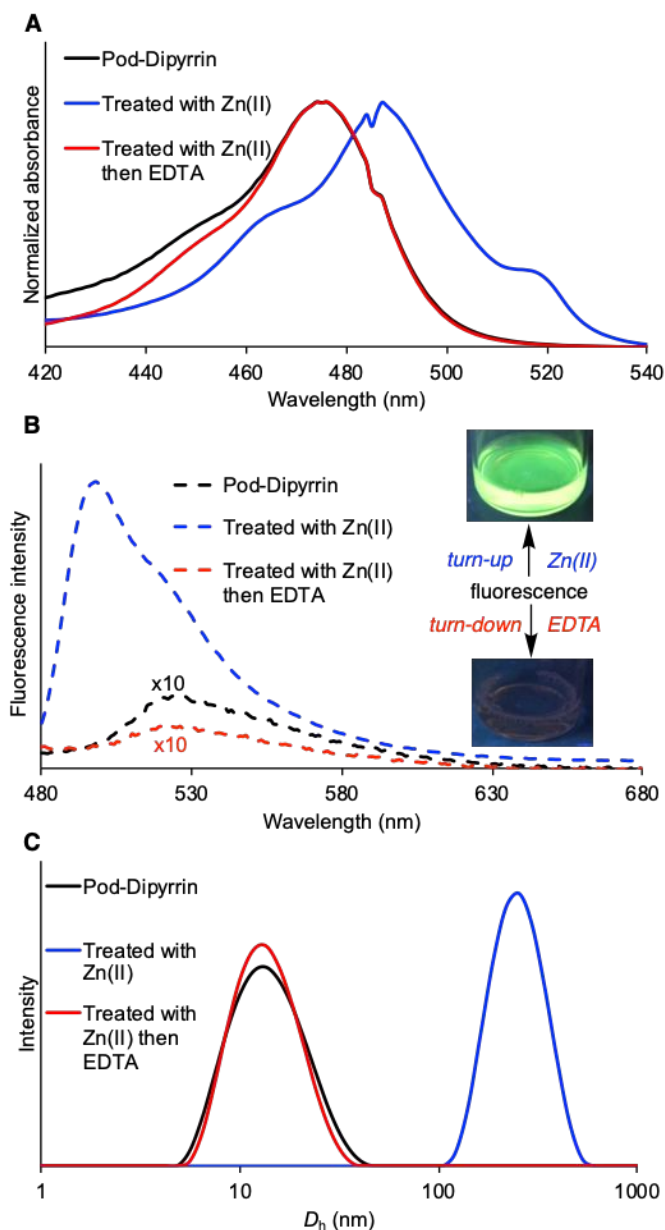


Figure 5. Spectral and DLS data in water at room temperature. (A) Absorption spectra of **Pod-Dipyrrin** at 1 mg/mL (black line), **Pod-Dipyrrin** treated with Zn(II) (blue line), and **Pod-Dipyrrin** treated with Zn(II) then EDTA (red line); (B) fluorescence spectra of **Pod-Dipyrrin** (black dashed line), **Pod-Dipyrrin** treated with Zn(II) (blue dashed line), and **Pod-Dipyrrin** treated with Zn(II) then EDTA (red dashed line); (C) DLS data of **Pod-Dipyrrin** at 1 mg/mL (black line), **Pod-Dipyrrin** treated with Zn(II) (blue line), and **Pod-Dipyrrin** treated with Zn(II) then EDTA (red line). In all treatments, Zn(II) refers to $Zn(OAc)_2$ (440 equiv).

Table 2. Photophysical properties of diverse constructs in organic or aqueous media.

Entry	Compound	Solvent	λ_{abs} (nm)	λ_{exc} (nm)	λ_{em} (nm)	fwhm λ_{em} (nm)	Φ_{f}^a
1	4	toluene	435	390	505	34	0.0011
2	5	toluene	434	390	518	54	0.0006
3	6	toluene	488	460	504	32	0.16
4	Pod-Dipyrrin	water	474	460	526	61	0.0035
5	Pod-Dipyrrin	DMF	430	390	530	80	0.0021
6	(Pod-Dipyrrin)₂Zn(II)	DMF	486	460	496	27	0.11
7	(Pod-Dipyrrin)₂Zn(II)	water	487	470	498	43	0.17

^aAll yield data obtained with **BDPY-hydrazone** as a reference ($\Phi_{\text{f}} = 0.97$).^{2,24}

The free base **Pod-Dipyrrin** and the assembled **(Pod-Dipyrrin)₂Zn(II)** were examined by DLS. The free base **Pod-Dipyrrin** is unimeric in water with a peak in the DLS distribution at 15 nm (Figure 5C). On the other hand, the assembled **(Pod-Dipyrrin)₂Zn(II)** exhibits large very assemblies of 260 nm in dimension; such large assemblies are unexpected because the simplest architecture would be simply twice the size of a single **Pod-Dipyrrin**. The data imply the **(Pod-Dipyrrin)₂Zn(II)** must unfold upon formation of the bis(dipyrrinato)Zn(II) motif, causing extensive aggregation.

Upon treatment of the **(Pod-Dipyrrin)₂Zn(II)** assembly in aqueous solution with excess EDTA (3.2 equiv per the total quantity of Zn(II)), several distinct changes occurred: (1) the absorption features reverted to those of the unimeric free base **Pod-Dipyrrin** (Figure 5A), reflecting a change in color from light green to yellow; (2) the fluorescence diminished markedly and visibly, to a slightly lower level than for the **Pod-Dipyrrin** alone (Figure 5B); and (3) DLS data revealed the reversion from the large assembly to give the unimeric folded architecture characteristic of **Pod-Dipyrrin** (Figure 5C). The combined evidence of absorption shifts, fluorescence brightness and size changes reveal a reversible conversion of **Pod-Dipyrrin** to

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3 **(Pod-Dipyrrin)₂Zn(II)** in water accompanied by a profound mechanical change and
4 fluorescence readout: fluorescence turn-up upon addition of Zn(II) and turn-down upon removal
5 of Zn(II).
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10 The self-assembly experiment with **Pod-Dipyrrin** was also carried out with Cu(II) in
11 water. Upon treatment with 600 equiv of CuCl₂, the absorption spectrum shows a bathochromic
12 shift (17 nm) (Figure 6A), the fluorescence was strongly quenched (Figure 6B), and the DLS
13 data showed conversion of the unimer ($D_h = 15$ nm) to a large assembly ($D_h = 230$ nm) (Figure
14 6C), all of which signal the formation of **(Pod-Dipyrrin)₂Cu(II)**. Treatment with excess EDTA
15 (2.3 equiv per the total quantity of Cu(II)) caused reversion to the absorption spectrum and
16 unimeric size of the **Pod-Dipyrrin**. The fluorescence increases but only slightly, without full
17 recovery of the fluorescence expected of the **Pod-Dipyrrin**; one interpretation is that the copper–
18 EDTA complex (green) is associated with the **Pod-Dipyrrin** and causes quenching of the free
19 base dipyrin. The combined evidence of absorption, fluorescence brightness and size changes
20 reveal a reversible self-assembly of **Pod-Dipyrrin** in water with fluorescence turn-off upon
21 addition of Cu(II) and turn-on upon removal of Cu(II).
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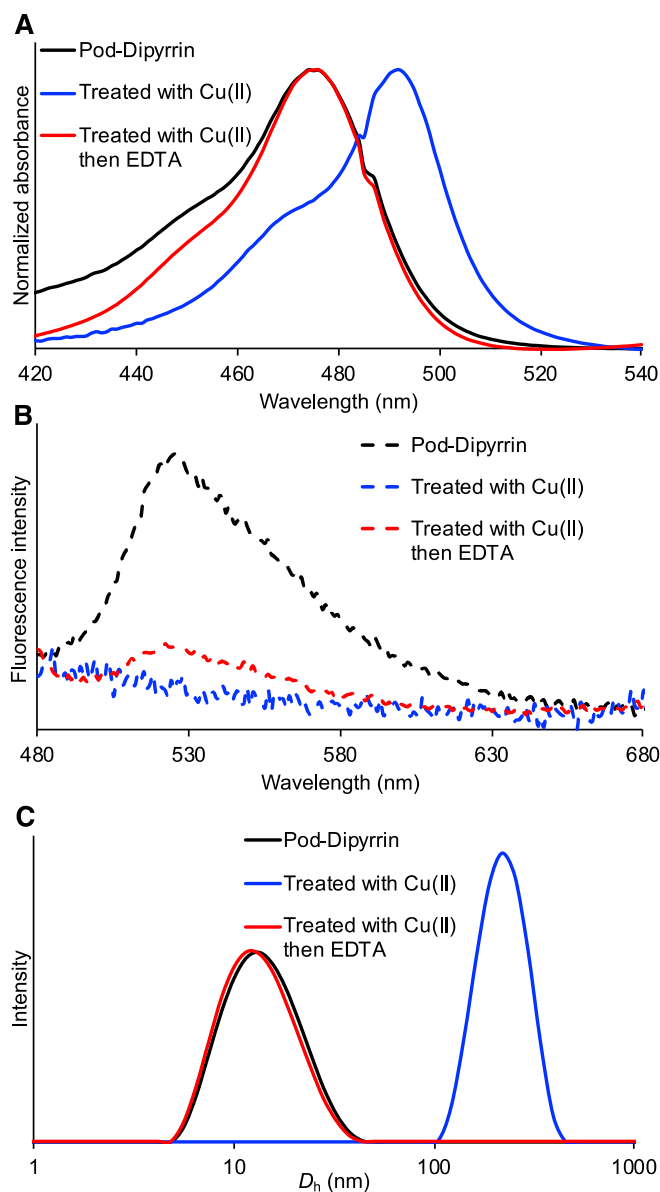


Figure 6. All data were collected in water at room temperature. (A) Absorption spectra of **Pod-Dipyrrin** at 1 mg/mL (black solid line), **Pod-Dipyrrin** treated with Cu(II) (red solid line), and **Pod-Dipyrrin** treated with Cu(II) then EDTA (red solid line); (B) fluorescence spectra of **Pod-Dipyrrin** (black dashed line), **Pod-Dipyrrin** treated with Cu(II) (red dashed line), and **Pod-Dipyrrin** treated with Cu(II) then EDTA (red dashed line); (C) DLS data of **Pod-Dipyrrin** at 1mg/mL (black solid line), **Pod-Dipyrrin** treated with Cu(II) (red solid line), and **Pod-Dipyrrin** treated with Cu(II) then EDTA (red solid line). In all treatments, Cu(II) refers to CuCl₂ (600 equiv).

The observed size distribution of **(Pod-Dipyrrin)₂Zn(II)** (260 nm) in water matches that of two unfolded **F-Ph** (130 nm * 2) polymers in DMF, but is far larger than expected for the simple dimerization of two intact unimers (15 nm * 2) in water. To explore such large size distributions, the assembly of **Pod-Dipyrrin** was carried out in DMF. Treatment of **Pod-Dipyrrin** in DMF with Zn(OAc)₂ result in formation of the corresponding **(Pod-Dipyrrin)₂Zn(II)** as seen by the absorption spectrum (Figure 7). Examination by DLS shows that the free base **Pod-Dipyrrin** in DMF exhibits $D_h = 140$ nm (Figure 8A) whereas the **(Pod-Dipyrrin)₂Zn(II)** exhibits a much larger $D_h = 310$ nm (Figure 8B). Finally, treatment of polymer **F-Ph** with Zn(OAc)₂ in water resulted in no noticeable size change (Figure 8C), as expected for a polymer that lacks a dipyrrin unit altogether.

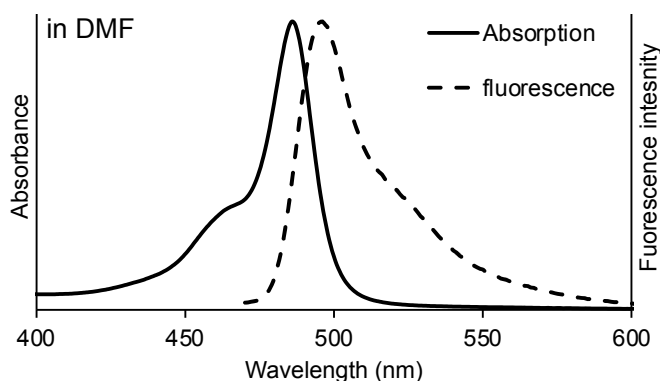


Figure 7. Absorption (solid line) and fluorescence (dashed line) spectra of **(Pod-Dipyrrin)₂Zn(II)** in DMF at room temperature.

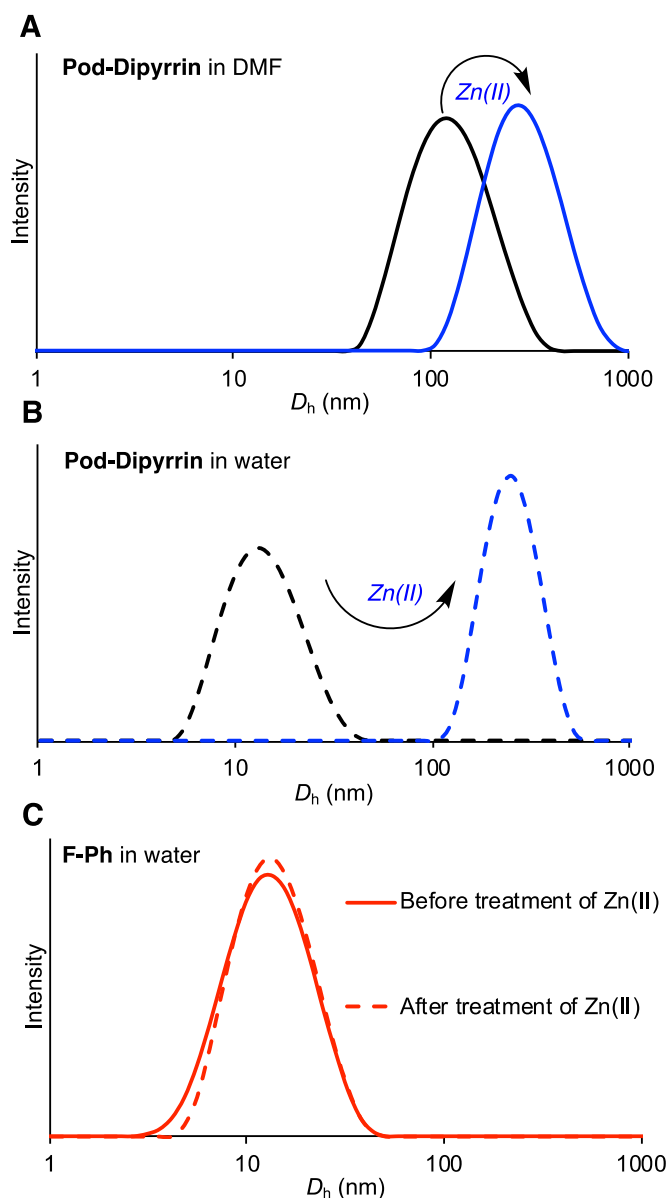


Figure 8. DLS data at room temperature. (A) **Pod-Dipyrin** in DMF before (black solid line) and after (blue solid line) treatment of Zn(II); (B) **Pod-Dipyrin** in water before (black dashed line) and after (blue dashed line) treatment of Zn(II); (C) **F-Ph** in water before and after treatment with Zn(II). In all treatments, Zn(II) refers to Zn(OAc)₂ (440 equiv).

To probe the self-assembly process, **Pod-Dipyrin** in aqueous solution was titrated with zinc acetate. The addition of 1 equiv gave two peaks upon DLS examination, including that of the uncomplexed unimer ($D_h = 13$ nm) and that for the putative dimer at $D_h = 200$ nm (Figure 9). Treatment with 10 equiv caused loss of the unimer peak and only the putative dimer peak was

observed. No further change was observed with 100 equiv. In this titration, no peaks intermediate between that of unimer and dimer were observed.

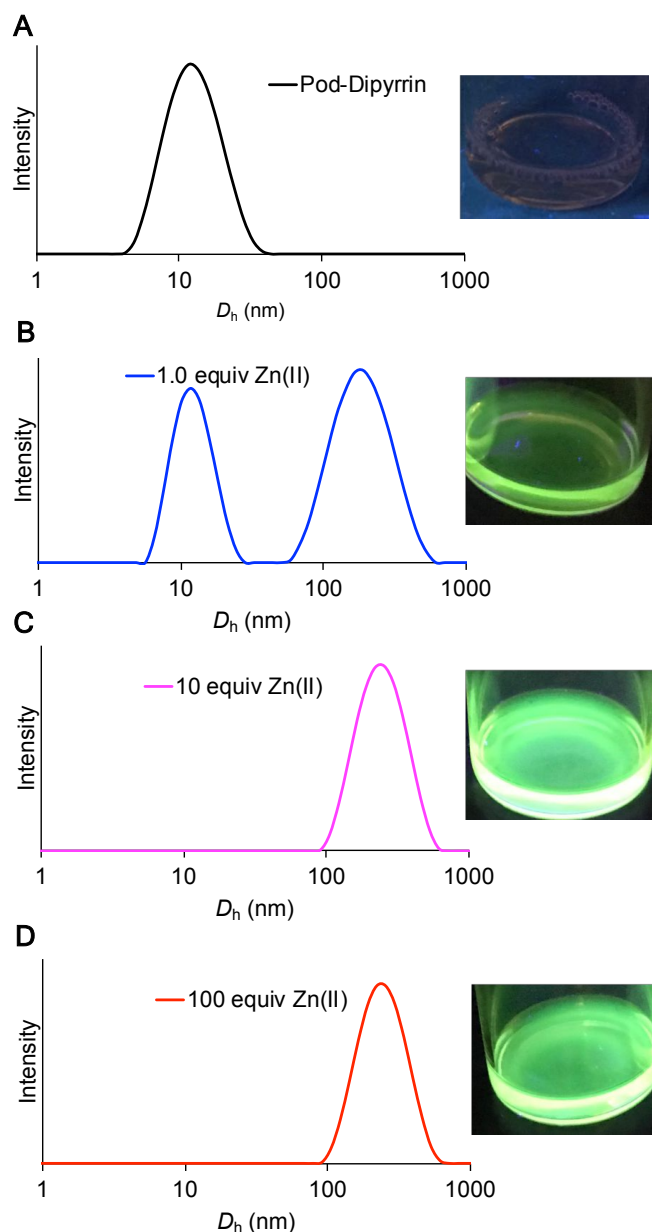


Figure 9. All data were collected in water at room temperature. DLS data (left panel) and photographs of the **Pod-Dipyrin** solution under ultraviolet illumination (right panel). (A) **Pod-Dipyrin**; (B) **Pod-Dipyrin** treated with 1.0 equivalent of Zn(II); (C) **Pod-Dipyrin** treated with 10 equivalents of Zn(II); and (D) **Pod-Dipyrin** treated with 100 equivalents of Zn(II). In all treatments, Zn(II) refers to Zn(OAc)₂.

Discussion

The development of foldamers enables diverse molecular entities to be packaged in a relatively hydrophobic enclosure for use in aqueous solution. The distinct feature of the strategy we are developing enables packaging of a single item of molecular cargo per polymer chain. The idea behind individual packaging of a single molecular item was developed to preclude quenching of fluorophores and deleterious contact with water while achieving use in water. A limiting view is that the fluorophore or other hydrophobic cargo item is ensconced in the interior of the self-assembled foldamer. Yet the results herein show that the cargo item – the dipyrin – can bind with metal ions in aqueous solution, and in so doing cause substantial structural change of the foldamer package. Hence the foldamer is not like a rigid marble but is highly malleable and poised for morphological alteration. In the following sections, we first discuss the spectra and photophysics of dipyrins. We then discuss experiments that bear on the environmental polarity of the encapsulated bis(dipyrinato)Zn(II) complex. We finish by considering the mechanical change associated with the self-assembly process leading to the bis(dipyrinato)metal complexes from the foldamer–dipyrin.

1. Spectra and photophysics of dipyrins. The chemistry of dipyrins^{25,26} is rapidly growing particularly because of the facile synthesis and versatile coordination features that can be exploited to create supramolecular structures.²⁷ The dipyrinatoboron complexes (i.e., BODIPYs)^{28,29} are the best known but the bis(dipyrinato)metal complexes have assembly features that the former lack. The fluorescence properties of dipyrinato–metal complexes have been comprehensively reviewed,^{11,27,30-32} including the turn-on fluorescence that accrues upon exposure of a free base dipyrin to a suitable metal salt, termed chelation-enhanced fluorescence (CHEF). Free base dipyrins are at best only very weakly fluorescent,³⁰⁻³² although distinctions between one extreme of “complete lack of fluorescence”³³ and no fluorescence even at 1.2 K,³⁴

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3 the common report of no detectable fluorescence ($\Phi_f < 5 \times 10^{-4}$;³⁵ ≤ 0.001 ;^{11,32,36-38} $\leq 10^{-4}$;³⁹), and
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5 the other extreme of a measurable albeit tiny fluorescence ($\Phi_f = 8.1 \times 10^{-4}$;⁴⁰ 2×10^{-3} ;⁴¹ 0.01–
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7 0.02;⁴²) may stem in part from different structural features of the dipyrin in question. Some
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9 studies concerned the HBr salt of the free base dipyrin, which may remain partially complexed
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11 in solution and contribute to the observed properties. Regardless, the photodynamics of free base
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13 dipyrins were recognized early on to be dominated by internal conversion rather than by
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15 intersystem crossing.^{34,41} Obvious mechanisms for internal conversion include rotation of the
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17 pyrrolic planes from coplanarity with respect to each other, and proton tunneling from the
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19 pyrrolic unit to the neighboring imino nitrogen.³⁴ A very recent study points instead, however, to
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21 the presence of a conical intersection of the S_1 and S_0 potential energy surfaces; such an
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23 intersection is less accessible energetically upon coordination of the nitrogen atoms. If the latter
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25 view holds, the diminished non-radiative decay upon metal coordination to the dipyrin is less
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27 about structural rigidification and absence of proton transfer than shifting of molecular orbital
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29 energies.⁴³
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36 Here, three observations concerning the fluorescence of the **Pod–Dipyrin** system
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38 warrant discussion. First, the **Pod–Dipyrin** exhibits very low fluorescence in DMF ($\Phi_f =$
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40 0.0021) but ~ 1.7 -fold increase upon unimer assembly in water, albeit to a still very low value (Φ_f
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42 = 0.0035) (Figures 1 and 3). The dipyrin in the foldamer undergoes a striking spectral change in
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44 the unfolded state in DMF (broad, unstructured absorption) to the folded unimer in water (more
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46 sharp absorption) (Figure 4C). The origin of such spectral change is unclear, as the dipyrin in
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48 the foldamer in aqueous solution (versus that in the extended conformation in DMF) is in a more
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50 hydrophobic environment, may be somewhat constrained conformationally, and perhaps is
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52 hydrogen-bonded where (1) the backbone N–H serves as the hydrogen-bond donor, (2) the amide
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54 carbonyl serves as a hydrogen-bond acceptor, and/or (3) the lone carboxylic acid at the polymer
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3 terminus serves as a proton donor. Such hydrogen bonds to and from the dipyrin would likely
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5 supplant the known intramolecular hydrogen bond of the dipyrin⁴⁴ and hence are only
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7 conjectural; a general rigidification of the dipyrin owing to the constrained environment of the
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9 assembly may also contribute. Evidence for the unimer assembly stems not only from the DLS
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11 data but also, *inter alia*, from 2D ¹H NMR NOESY analysis of polymer **F-Ph** in DMSO-*d*₆ or
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13 D₂O.² The Φ_f value of a cyanine dye also increased (by 2.5-fold) upon encapsulation in the pod
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15 architecture.²
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20 Second, complexation with Zn(II) to form **(Pod-Dipyrin)₂Zn(II)** causes the
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22 fluorescence intensity to be *turned up* markedly in DMF ($\Phi_f = 0.11$) and for the unimer in water
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24 ($\Phi_f = 0.17$) (Figure 5B), resembling that of the dipyrin lacking the attached foldamer (Figure 1,
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26 Table 1); on the other hand, the already low fluorescence quantum yield of the free base dipyrin
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28 is further *turned down* (to the level of noise) by complexation with Cu(II) to form **(Pod-**
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30 **Dipyrin)₂Cu(II)** (Figure 6A,B). The absorption spectral features of the **(Pod-Dipyrin)₂Zn(II)**
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32 differ slightly in water versus DMF (compare Figures 5A and 7); both the significant spectral
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34 changes of the dipyrin and slight spectral changes of the bis(dipyrinato)Zn(II) unit in DMF and
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36 water will require further investigation.
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41 Third, treatment of **(Pod-Dipyrin)₂M(II)**, where M = Cu or Zn, with EDTA in each
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43 case failed to give full reversion to the expected (limited) fluorescence of the free base dipyrin
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45 (Figures 5B and 6B). A tentative interpretation is that the M(II)-EDTA complex may associate
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47 with the **Pod-Dipyrin** and cause quenching of the free base dipyrin. Exploring the
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49 photophysics of the dipyrins, particularly upon metal complexation,⁴⁵⁻⁴⁹ remains an area of
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51 active investigation, and gaining a deep understanding will require further experimental and
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53 theoretical work.
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3 **2. Polarity-dependent fluorescence probe in the self-assembled polymer.** The
4 fluorescence quantum yields of bis(dipyrrinato)Zn(II) complexes bearing α - and/or β -alkyl
5 substituents are known to be dependent on solvent polarity.^{50,51} Such complexes are fluorescent
6 in a non-polar solvent such as toluene, but weakly or essentially non-fluorescent in modestly
7 polar organic solvents (e.g., dichloromethane). The diminution in fluorescence is attributed to an
8 energetically accessible and non-emissive excited-state charge-transfer process.^{50,51} The
9 benchmark bis(dipyrrinato)Zn(II) complex **6** also exhibited a decrease in Φ_f value with
10 increasing solvent polarity, but the effect was of lesser magnitude over a greater range of polarity:
11 the decline was ~ 4.5 -fold in total over the increase in polarity from toluene to DMF (Figure 2
12 and Table 1). The benchmark bis(dipyrrinato)Zn(II) complex **6** lacks α - and β -alkyl substituents;
13 alkyl substituents can profoundly alter the energetics of pyrrole molecules⁵² and are similarly
14 expected to influence the energetics of dipyrrin analogues.

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31 The solvent-dependence of the fluorescence of bis(dipyrrinato)Zn(II) complexes can be
32 exploited as an initial gauge of the polarity of the “Pod”. The Φ_f values of (**Pod-**
33 **Dipyrrin**)₂Zn(II) in water (0.17) and DMF (0.11) compared with **6** in toluene (0.18) and DMF
34 (0.041) indicate that in water (Tables 1 and 2), the bis(dipyrrinato)Zn(II) complex of (**Pod-**
35 **Dipyrrin**)₂Zn(II) is in a nonpolar environment comparable to that of toluene. The hydrophobic
36 environment is attributed to the pendant lauryl groups and the polymer backbone in the folded
37 assembly. This agrees with our previously reported several “Pod-fluorophores”, which have Φ_f
38 values in aqueous solution similar to those of their hydrophobic benchmarks in toluene.² On the
39 other hand, in DMF the bis(dipyrrinato)Zn(II) complex of (**Pod-Dipyrrin**)₂Zn(II) exhibits
40 fluorescence intensity that reflects a modestly polar environment, but still less polar than that
41 upon full exposure to bulk DMF. More in-depth analysis of the local environment will require
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3 examination of solvatochromic chromophores that exhibit changes in spectra alone or in
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5 conjunction with changes in fluorescence intensity.
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8 **3. Self-assembly and mechanical change.** Self-assembly processes afford larger
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10 architectures the structures of which are manifested by the spontaneous interactions of the
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12 individual constituents.⁵³⁻⁵⁸ While self-assembling architectures offer numerous attractive
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14 features, some limitations remained to be addressed. Common methods to characterize
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16 supramolecular assemblies include NMR spectroscopy, mass spectrometry, and microscopy (if
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18 size permits), all of which can pose difficulties in interpretation, which is rendered more complex
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20 when a distribution of species is present in an equilibrium mixture. Moreover, many self-
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22 assembly processes need to be carried out in organic solution given that most of the materials are
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24 hydrophobic, shutting off many applications in life sciences.
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28 The **Pod-Dipyrrin** in aqueous solution is folded, compact and unimeric as evidenced by
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30 DLS interrogation ($D_h = 15$ nm) over a range of concentrations but more extended in the organic
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32 solvent DMF ($D_h = 140$ nm) (Figure 4A, 4B). In DMF, the dimeric **(Pod-Dipyrrin)₂Zn(II)**
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34 remains intact as evidenced by absorption and fluorescence spectroscopy (Figure 7) yet exhibits
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36 a size approximately twice that of the monomer ($D_h = 310$ versus 140 nm). While we cannot
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38 rule out larger assemblies, the cleanliness of the DLS data (Figures 4A, 4B, 5C, 8 and 9) upon
39
40 adding zinc acetate is consistent with (1) complete reaction of **F-Ph** with **5** to form the **Pod-**
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42 **Dipyrrin** construct (no signal corresponding to a monomer remained upon adding excess zinc
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44 acetate), and (2) a monomer – dimer system (no signals corresponding to larger assemblies were
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46 observed). In DMF, both **Pod-Dipyrrin** and **(Pod-Dipyrrin)₂Zn(II)** architectures are expected
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48 to be extended with exploration of diverse conformational landscapes. On the other hand, in
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50 water, treatment with Zn(II) causes the folded, compact unimeric **Pod-Dipyrrin** to partially
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52 unfold upon metal-ion binding and join with another unfolded **Pod-Dipyrrin** to make a dimer.
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3 The change in size – from $D_h = 15$ nm to 260 nm indicates the unimer has largely unfolded. The
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5 **(Pod-Dipyrrin)₂Zn(II)** has rather similar size in DMF ($D_h \sim 310$ nm) and water ($D_h \sim 260$ nm)
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7 consistent with a largely extended conformation in both solvents, in contrast to the distinct sizes
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9 for the free base **Pod-dipyrrin** ($D_h = 140$ nm versus 15 nm) in the same solvents in the absence
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11 of Zn(II) complexation.
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15 Toward the end of this study we returned to examine a minor feature of the absorption
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17 spectrum of the **(Pod-Dipyrrin)₂Zn(II)**. The absorption spectrum shown in Figure 5A exhibits a
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19 small shoulder (~ 517 nm) on the long-wavelength side of the main peak, which is not
20
21 characteristic of typical bis(dipyrrinato)Zn(II) complexes as illustrated by **6** in diverse solvents
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23 (Figures 1 and 2) and the **(Pod-Dipyrrin)₂Zn(II)** in DMF (Figure 7). Illumination into the long-
24
25 wavelength shoulder did not result in any distinguishable fluorescence apart from that of the
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27 **(Pod-Dipyrrin)₂Zn(II)**. Complexation of **Pod-Dipyrrin** with Zn(II) containing other
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29 counterions (chloride, triflate, sulfate) at 10 equiv or 440 equiv gave the **(Pod-Dipyrrin)₂Zn(II)**
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31 with a clean absorption spectrum lacking the long-wavelength shoulder and also the expected
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33 unimeric behavior as assessed by DLS (Figure S1, see Electronic Supplementary Information).
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35 The presence of the long-wavelength shoulder is tentatively attributed to the effect of the acetate
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37 counterion (although the molecular origin remains unknown) and does not alter the conclusions
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39 concerning assembly and disassembly of the fluorogenic bis(dipyrrinato)Zn(II) complex. During
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41 the course of these ZnX₂ complexation experiments, two observations were made concerning the
42
43 size of the **(Pod-Dipyrrin)₂Zn(II)** complexes in water (Figures S1 and S2): (1) the size
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45 decreased by 9–30 nm in going from 10 to 440 equiv of ZnX₂, and (2) the use of 440 versus 10
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47 equiv of ZnCl₂ or Zn(OAc)₂ resulted in slight broadening of the peak observed upon DLS
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49 examination. The change in size distribution may reflect specific effects of counterions, and
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51 warrants further examination.
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3 The size of the assembled complex **(Pod-Dipyrin)₂Zn(II)** in water or DMF is
4 comparable to or larger than the discrete assemblies derived from metals and organic ligands
5 bearing two or three free base dipyrin ligands.⁵⁹ In limited studies where the **Pod-Dipyrin** was
6 titrated with Zn(II), no intermediates were observed; in other words, the assembly to give the
7 **(Pod-Dipyrin)₂Zn(II)** was an all-or-nothing phenomena giving the homoleptic product. In
8 contrast, a dibenzo-annulated 1,9-dicarboethoxydipyrin (L-H) embedded in a dendrimer for
9 aqueous solubilization underwent complexation with zinc acetate to give the heteroleptic
10 complex MLX where M = Zn(II) and X = acetate; the architecture was proposed as a biosensor
11 for detection of Zn(II).⁶⁰
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26 Outlook

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28 In summary, the single-polymer–single cargo strategy with a dipyrin as the cargo has
29 revealed fundamental features concerning the conformational pliability of the amphiphilic
30 polymer. The presence of a divalent ion (Zn(II), Cu(II)) triggers profound conformational
31 change of the polymer–dipyrin architecture. The molecular architecture of **Pod-Dipyrin** may
32 be unusual – and with fecund possibilities in supramolecular chemistry – in that mechanical
33 change prompted by Zn(II) is accompanied by a striking ~50-fold increase in fluorescence. The
34 change in fluorescence is visible by the naked eye although the change in absorption, while
35 measurable by absorption spectroscopy, is far less evident by visual inspection. A common and
36 versatile approach for metal ion sensing relies on binding to the lone pair of electrons on an
37 amino group, thereby suppressing photoinduced electron transfer to an adjacent chromophore,
38 thereby unleashing fluorescence.⁶¹ The fluorogenic mechanism here, based on dipyrin
39 dimerization, is complementary. The bis(dipyrinato)metal assembly process is robust and can
40 be reversed by metal ion sequestration. Mechanosensory receptors, which sense mechanical
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changes caused by pressure, vibration or electric fields, are well known in biology.⁶² Given the importance of mobile Zn(II) as a signaling molecule in neurophysiology,⁶³⁻⁶⁵ the ability to chelate Zn(II), detect the chelation fluorogenically, and also exert a mechanical change may enable the design of novel sensory-mechanical constructs for fundamental manipulations in neurobiology.

Experimental Procedures and Characterization Data

General Methods. All chemicals obtained commercially were used as received unless otherwise noted. Reagent-grade solvents (CH₂Cl₂, hexanes, methanol, toluene, ethyl acetate) and HPLC-grade solvents (toluene, CH₂Cl₂, hexanes) were used as received. THF was freshly distilled from sodium/benzophenone ketyl and used immediately. Electrospray ionization mass spectrometry (ESI-MS) data are reported for the cationized molecular ion.

Synthesis

5-(4-Iodo-2,6-dimethylphenyl)dipyrromethane (2). Following an established procedure,^{21,22} a solution of 4-iodo-2,6-dimethylbenzaldehyde (**1**,^{12,13} 1.82 g, 7.00 mmol) in pyrrole (50.3 g, 52.0 mL, 750 mmol) was degassed with a stream of Ar for 20 min. Trifluoroacetic acid (82.0 mg, 54.0 μ L, 700 μ mol) was then added, and the solution was stirred under Ar at room temperature for 15 min. The solution was concentrated and chromatographed [silica, hexanes/CH₂Cl₂/ethyl acetate (75:20:5)] to obtain a pale yellow solid (1.58 g, 60% yield): mp 162–163 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.06 (s, 6H), 5.90 (s, 1H), 5.96–5.98 (m, 2H), 6.17–6.20 (m, 2H), 6.68–6.70 (m, 2H), 7.43 (s, 2H), 7.95 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 20.5, 38.7, 93.2, 107.1, 109.0, 116.8, 130.5, 137.8, 138.4, 140.3; ESI-MS obsd 377.0495, calcd 377.0509 [(M + H)⁺, M = C₁₇H₁₇IN₂]; Anal. Calcd for C₁₇H₁₇IN₂: C, 54.27; H, 4.55; N, 7.45. Found: C, 54.09; H, 4.44; N, 7.27.

5-[4-(6-(*tert*-Butoxycarbonylamino)hexylaminocarbonyl)-2,6-dimethylphenyl]dipyrromethane (3). Following an established carbonylation procedure,^{21,22} a Schlenk flask containing samples of **2** (188 mg, 500 μ mol), Pd(OAc)₂ (11.0 mg, 50.0 μ mol), Xantphos (29.0 mg, 50.0 μ mol) and cesium carbonate (490 mg, 1.50 mmol) was evacuated and purged with carbon monoxide. A solution of *N*-(*tert*-butoxycarbonyl)-1,6-diaminohexane (216 mg, 1.00 mmol) in toluene (5.00 mL) was purged with Ar (30 min) and subsequently with carbon monoxide (30 min). The resulting solution was transferred into the Schlenk flask containing the solid materials under an atmosphere of CO. The reaction flask was placed in a preheated oil bath and stirred for 2 h at 90 °C. The resulting mixture was allowed to cool to room temperature. The solid material were filtered off and washed with toluene. The filtrate was concentrated, and the resulting residue was chromatographed [silica, hexanes/ethyl acetate (2:1)] to obtain a pale yellow solid (150 mg, 61% yield): mp 55–58 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.35–1.52 (m, 13H), 1.56–1.66 (m, 4H), 2.13 (s, 6H), 3.11 (q, *J* = 6.6 Hz, 2H), 3.43 (q, *J* = 6.6 Hz, 2H), 4.54 (br s, 1H), 5.93–5.95 (m, 2H), 5.97 (s, 1H), 6.17 (q, *J* = 3.0 Hz, 2H), 6.29 (br s, 1H), 6.68 (dd, *J*₁ = 2.4 Hz, *J*₂ = 4.2 Hz, 2H), 7.43 (s, 2H), 7.98 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 20.9, 26.1, 26.3, 28.5, 29.6, 30.1, 38.9, 39.7, 40.2, 79.2, 106.8, 108.6, 116.8, 128.0,

130.5, 133.1, 138.2, 141.5, 156.3, 167.6; ESI-MS obsd 515.2991, calcd 515.2993 [(M + Na)⁺, M = C₂₉H₄₀N₄O₃]; λ_{abs} (CH₂Cl₂) 406 nm.

5-[4-(6-(*tert*-Butoxycarbonylamino)hexylaminocarbonyl)-2,6-dimethylphenyl]dipyrrin (4). Following an established procedure,⁹ a solution of **3** (98.0 mg, 200 μmol) and *p*-chloranil (62.0 g, 250 μmol) in THF (5.00 mL) was sonicated in a laboratory sonication bath for 1 h. The solution was concentrated and chromatographed [silica, hexanes/ethyl acetate (2:1)] to obtain a yellow solid (68.0 mg, 69% yield): mp 58–60 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.36–1.55 (m, 15H), 1.66 (qt, *J* = 7.2 Hz, 2H), 2.18 (s, 6H), 3.16 (q, *J* = 6.4 Hz, 2H), 3.48 (q, *J* = 6.4 Hz, 2H), 4.58 (m, 1H), 6.32 (dq, *J*₁ = 1.2 Hz, *J*₂ = 4.8 Hz, 4H), 6.43 (br s, 1H), 7.53 (s, 2H), 7.63 (t, *J* = 1.2 Hz, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 20.2, 26.2, 26.4, 28.6, 29.7, 30.2, 39.8, 40.3, 79.2, 118.2, 125.8, 127.1, 134.6, 137.7, 139.6, 140.0, 144.0, 156.3, 167.8; ESI-MS obsd 491.3013, calcd 41.3017 [(M + H)⁺, M = C₂₉H₃₈N₄O₃]; λ_{abs} (CH₂Cl₂) 429 nm.

5-[4-(6-(6-Maleimidohexanoylamino)hexylaminocarbonyl)-2,6-dimethylphenyl]dipyrrin (5). A solution of **4** (49.0 mg, 100 μmol) in CH₂Cl₂ (1.50 mL) was purged with Ar (for 5 min). Trifluoroacetic acid (1.50 mL) was added, and the mixture was stirred for 12 h at room temperature. The volatile components (solvent and trifluoroacetic acid) were removed by purging with Ar. The resulting residue was treated with HBTU (76.0 mg, 200 μmol), 6-maleimidohexanoic acid (32.0 mg, 150 μmol), CH₂Cl₂ (5.00 mL) and *N,N*-diisopropylethylamine (500 μL) under Ar. The resulting solution was stirred for 2 h at room temperature. The mixture was concentrated and chromatographed [silica, ethyl acetate (2:1)] to obtain a brown solid (45.0 mg, 77% yield): mp 44–46 °C; ¹H NMR (300 MHz, CDCl₃) δ 1.26–1.69 (m, 14H), 2.15–2.20 (m, 8H), 3.26 (q, *J* = 6.6 Hz, 2H), 3.46–3.53 (m, 4H), 5.69 (m, 1H), 6.31–6.35 (m, 4H), 6.45 (br s, 1H), 6.68 (s, 2H), 7.52 (s, 2H), 7.63 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 20.3, 25.4, 26.2, 26.3, 26.5, 28.5, 29.7, 29.8, 36.7, 37.8, 39.2, 39.8, 118.3, 125.8, 127.2, 134.3, 134.6, 137.7, 139.7, 140.0, 144.0, 167.9, 171.0, 173.0; ESI-MS obsd 584.3231, calcd 584.3231 [(M + H)⁺, M = C₃₄H₄₁N₅O₄]; λ_{abs} (CH₂Cl₂) 468 nm.

Bis[5-(4-(6-(*tert*-butoxycarbonylamino)hexylaminocarbonyl)-2,6-dimethylphenyl)dipyrrinato]zinc(II) (6). A solution of **4** (15 mg, 30 μmol) in CHCl₃ (2.0 mL) was treated with Zn(OAc)₂·2H₂O (150 μmol, 10 equiv) in methanol (0.5 mL). After stirring for 15 h at room temperature, the mixture was diluted with CHCl₃, washed with water, dried with anhydrous sodium sulfate and chromatographed [silica, hexanes/ethyl acetate, (1:1)]. Decomplexation occurred during chromatography, and the starting material **4** was recovered. The reaction was repeated in identical fashion with the recovered material. The mixture was washed with water. The organic layer was concentrated to dryness to obtain a red solid (13.6 mg, 85% yield): mp 205 °C (discoloration); ¹H NMR (400 MHz, CDCl₃) δ 1.40–1.54 (m, 30H), 1.68 (qt, *J* = 6.8 Hz, 4H), 2.27 (s, 12H), 3.16 (q, *J* = 6.4 Hz, 4H), 3.50 (q, *J* = 6.4 Hz, 4H), 4.58 (br s, 2H), 6.37 (d, *J* = 4.4 Hz, 4H), 6.43 (br t, *J* = 4.8 Hz, 2H), 6.52 (d, *J* = 4 Hz, 4H), 7.49 (br s, 4H), 7.57 (br s, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 20.3, 26.2, 26.4, 28.7, 29.9, 30.3, 39.9, 40.4, 79.3, 117.8, 125.7, 131.1, 134.5, 137.6, 139.5, 141.5, 146.9, 149.8, 156.4, 167.9; ESI-MS obsd 1043.5079, calcd 1043.5096 [(M + H)⁺, M = C₅₈H₇₄N₈O₆Zn]; λ_{abs} (toluene) 487 nm, ε_{488 nm} = 52,900 M⁻¹cm⁻¹.

Pod-Dipyrrin. A solution of **F-Ph**² (30 mg), **5** (0.53 mg, 0.90 μmol), and ethanolamine (0.24 μL) in DMF (400 μL) was treated with triethylamine (1 drop). The resulting mixture was stirred at room temperature for 16 h. Then the mixture diluted with 1.0 mL DMF. The resulting

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3 solution was transferred into a dialysis membrane as described in a prior supporting information
4 section² in detail. The solution was then dialyzed in DMF to remove the excess fluorophore.
5 The dialysis reservoir volume was replaced with fresh DMF four times over the course of ~24 h.
6 The resulting solution was dried under high vacuum at 50 °C, and the resulting solid was
7 dissolved in DI water. The aqueous solution was then freeze-dried to give a pink solid (26 mg).
8 The characterization and properties of the construct are described in the Results section.
9

10 **Absorption Spectroscopy.** Absorption spectra were recorded at room temperature as
11 described previously.² Unless stated otherwise, samples were examined with **Pod-Dipyrrin** at
12 ~20 μM ($A \sim 1$ in a 1-cm pathlength cuvette).
13

14 **Fluorescence Spectroscopy.** Fluorescence spectra were recorded as described
15 previously² at room temperature using BDPY-hydrazide in methanol ($\Phi_f = 0.97$)^{2,24} and **II** in
16 toluene ($\Phi_f = 0.36$)¹⁰ as quantitative standards with correction for instrument sensitivity but
17 without correction for solvent refractive indices. Unless stated otherwise, samples were
18 examined with **Pod-Dipyrrin** at ~2 μM in a 1-cm pathlength cuvette. The typical wavelength of
19 excitation of the **(Pod-Dipyrrin)₂Zn(II)** was into the short-wavelength shoulder at ~460 nm.
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21

22 **Dynamic Light Scattering (DLS) Measurements.** DLS analysis of the **Pod-Dipyrrin**
23 was performed in cuvettes with a Zetasizer Nano ZS instrument. A typical analysis was as
24 follows: a sample was dissolved in HPLC-grade water containing 1.0 M NaCl (filtered with a
25 220-nm membrane) for analysis, in some cases with a preceding step of filtration through a 220-
26 nm membrane. Illumination was performed at 632.8 nm at room temperature. Unless stated
27 otherwise, samples were examined with **Pod-Dipyrrin** at ~20 μM .
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30 **Nuclear Magnetic Resonance (NMR) Spectroscopy.** ¹H NMR spectra were measured
31 at room temperature.
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33 **Metal Coordination Reactions.**

34 The number of equivalents of a metal salt in a coordination reaction of a dipyrin is
35 described relative to the bis(dipyrinato)metal complex; e.g., 5.5 μmol of $\text{Zn}(\text{OAc})_2$ and 0.025
36 μmol of **Pod-dipyrrin** corresponds to 440 equiv of $\text{Zn}(\text{OAc})_2$.
37

38 **General Method for Metal Coordination Reactions of Pod-Dipyrrin, Illustrated with**
39 **Zn(II) in Water.** A solution of **Pod-Dipyrrin** in water (1.0 mg/mL) was measured directly,
40 without dilution, by DLS and absorption spectroscopy in a 1-cm pathlength cuvette. Then a part
41 of the solution (~ 200 μL) was diluted (by ~ 6-fold) by water and transferred into another 1-cm
42 pathlength cuvette. The resulting solution measured by absorption and fluorescence
43 spectroscopy. A solution of **Pod-Dipyrrin** (1.0 mL, 1.0 mg/mL) was treated $\text{Zn}(\text{OAc})_2$ (1.0 mg,
44 5.5 μmol , ~440 equivalent of **Pod-Dipyrrin**). The resulting solution was allowed to sit for 5 min
45 and measured directly, without dilution, by DLS and absorption spectroscopy in a 1-cm
46 pathlength cuvette. Then a part of the Zn(II)-containing **Pod-Dipyrrin** solution (~ 200 μL) was
47 diluted (by ~6-fold) by water and transferred into another 1-cm pathlength cuvette. The resulting
48 solution was measured by absorption and fluorescence spectroscopy. The Zn(II) containing
49 **Pod-Dipyrrin** solution (0.8 mL, 1.0 mg/mL) was treated EDTA (4.0 mg, 14 μmol , ~3.2
50 equivalent per total Zn(II)). The resulting solution was allowed to sit for 5 min and measured
51 directly, without dilution, by DLS and absorption spectroscopy in a 1-cm pathlength cuvette.
52 Then a part of the EDTA treated **Pod-Dipyrrin** solution (~ 200 μL) was diluted (by ~ 6-fold) by
53 water and transferred into another 1-cm pathlength cuvette. The resulting solution was measured
54 by absorption and fluorescence spectroscopy.
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4 **Metal Binding Reactions of Pod-Dipyrrin with Cu(II) in water.** Following the
5 method describe for the reaction of **Pod-Dipyrrin with Zn(II)**, a solution of **Pod-Dipyrrin** in
6 water (1.0 mL, 1.0 mg/mL) was treated with CuCl₂ (1.0 mg, 7.5 μmol, ~600 equivalent). The
7 resulting solution was measured by DLS, absorption and fluorescence spectroscopy. Then the
8 resulting solution was treated with EDTA (4.0 mg, 14 μmol, 2.3 equivalent per total Cu(II)) and
9 measured by DLS, absorption and fluorescence spectroscopy.

10
11 **Metal Binding Reactions of Pod-Dipyrrin with F-Ph in water.** Following the method
12 describe for the reaction of **Pod-Dipyrrin** with Zn(II) in water, a solution of **F-Ph** in water (1.0
13 mL, 1.0 mg/mL) was measured by DLS and treated with Zn(OAc)₂ (1.0 mg, 5.5 μmol). The
14 resulting solution was then measured by DLS.

15
16 **Metal Binding Reactions of Pod-Dipyrrin with Zn(II) in DMF.** Following the method
17 describe for the reaction of **Pod-Dipyrrin** with Zn(II) in water, a solution of **Pod-Dipyrrin** in
18 DMF (1.0 mL, 1.0 mg/mL) was measured by DLS, absorption and fluorescence spectroscopy.
19 Then the mixture was treated with Zn(OAc)₂ (1.0 mg, 5.5 μmol, ~440 equivalent). The resulting
20 solution was measured by DLS, absorption and fluorescence spectroscopy.
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24 **Conflict of Interest Statement:** J.S.L. is cofounder of NIRvana Sciences, which develops
25 fluorophores for use in clinical diagnostics.
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30
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36 were carried out in the Molecular Education, Technology, and Research Innovation Center
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45 **Electronic supplementary information (ESI) available:** Results upon complexation of **Pod-**
46 **Dipyrrin** with various ZnX₂; and NMR spectra for all new compounds.
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