



Quantitative Colorimetric Paper Analytical Devices Based on Radial Distance Measurements for Aqueous Metal Determination

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5 2 **Quantitative Colorimetric Paper Analytical Devices Based on Radial Distance**
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8 3 **Measurements for Aqueous Metal Determination**
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2
3 19 **Abstract**
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5 20 Here we report a new microfluidic paper-based analytical device (mPAD) for quantifying
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7 21 metals in water. Metals represent an important class of water contaminants that come
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10 22 from a variety of sources including mining, transportation, manufacturing, waste
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12 23 management, and energy production. Current technologies for quantifying aquatic
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14 24 metals in water are expensive, relatively slow, tedious, provide inadequate performance,
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17 25 and difficult to use in a field setting. As a result, a need exists for simple, portable,
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19 26 power-free measurement tools that enable rapid in-field quantification of aquatic metals.
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21 27 The reported metal test cards, referred to as the On-Target Water Chemistry test cards,
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23 28 represent a major improvement over previously reported linear distance-based
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26 29 detection systems comprised of paper. With the On-Target approach, the sample flows
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28 30 outwards radially and reacts with colorimetric complexing agents, significantly reducing
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31 31 assay time. The diameter of the resulting color formation is directly proportional to
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33 32 analyte concentration. The On-Target cards were used for detecting copper, iron, and
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35 33 zinc with detection limits as low as 0.1 ppm in ~ 3 min and single ppb in combination
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38 34 with a membrane pre-concentration system.
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35 **Introduction**

36 Aqueous metals, such as Fe, Ni, and Cu, are regulated by numerous State and Federal
37 agencies because they are recognized as significant threats to human and ecosystem
38 health.¹ Cobalt (Co), copper (Cu), chromium (Cr), iron (Fe), magnesium (Mg),
39 manganese (Mn), nickel (Ni), and zinc (Zn) are also essential nutrients for humans as
40 they participate in important biochemical and physiological functions.² However,
41 overexposure to these metals can be carcinogenic and cause cellular and tissue
42 damage thereby leading to a variety of adverse health effects and human diseases.^{2,3}
43 Other possible health effects from over exposure to metals include gastrointestinal
44 illness, reproductive problems, and neurological disorders.⁴ Many industries, such as
45 mining, energy production, waste management, and transportation contribute to
46 pollutant release. Greater than 60% of rivers, lakes, and estuaries in the US are
47 presently 'impaired' with levels of metals, pathogens, and organic matter above EPA
48 limits due to the release of pollutants.⁵ Operators in the environmental remediation
49 sector and those handling wastewater disposal are required to monitor and control the
50 release of metals.⁶ Many legacy mines and industrial sites continue to release metals
51 into the environment at levels significantly exceeding EPA limits; these sites frequently
52 perform analytical testing to verify remediation but the frequency of testing is limited by
53 the cost of current analytical methods. There are also approximately 36,000 abandoned
54 mines in the western United States that are in need of reclamation.⁷ The federal
55 Comprehensive Environmental Response, Compensation, and Liability Act (CERCLA) –
56 also known as 'Superfund' – was passed in 1980 to fund the cleanup of such
57 uncontrolled or abandoned hazardous-waste sites, accidents, spills, etc.⁸ As a result,

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3 58 tens of thousands of samples are analyzed for metals each day in the US alone.
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5 59 Dissolved aquatic metals are presently measured using two distinct methods: in-
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8 60 field or laboratory. Common laboratory methods include atomic absorption spectroscopy
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10 61 (AAS) and inductively coupled plasma (ICP) coupled with either optical emission
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12 62 spectroscopy (OES) or mass spectrometry (MS).^{9,10} These methods are sensitive and
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14 63 selective but require large complicated instruments that delay the time between sample
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17 64 and results. Consequently, in-field assays that are not as sensitive or selective are
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19 65 frequently used.
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21 66 The in-field analysis technologies can be broken down into three categories: non-
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24 67 instrumented analysis (i.e., dipstick style tests), portable instrument supported analysis,
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26 68 and indirect 'point and shoot' analysis. Non-instrumented tests include basic visual
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28 69 comparative colorimetric products such as the Hach Colorwheel™ or Color Discs,
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30 70 Millipore Mquant™ and MColortest™, and CHEMetrics Colorimetrics and Titrimetrics.^{11–}
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32 71 ¹⁴ These solution chemistry kits currently require up to 19 steps which can introduce
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35 72 human error, have a narrow dynamic range, and can be inherently subjective since the
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38 73 result is based on human observation of visual color comparison. Test subjectivity can
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40 74 be reduced through electronic 'reading' of the colorimetric reaction with a fluorimeter or
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42 75 spectrophotometer but these external instruments add hundreds to thousands of dollars
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44 76 in cost.^{15,16} The CHEMetrics Instrumental Colorimeter and Hach SL1000 Portable
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47 77 Parallel Analyzer can streamline analysis by removing the sample preparation and
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49 78 interference blocking steps via disposable Vacu-vial® and Chemkey® consumables,
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51 79 respectively, but require a relatively high upfront investment.^{17,18}
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54 80 Point and shoot methods, such as handheld X-Ray Fluorescence (XRF)
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3 81 instruments are an alternative for solution-phase measurements.¹⁹ XRF instruments are
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5 82 often mentioned synonymously with aquatic testing, but can actually only analyze
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7 83 solids/soil for total elemental components and cannot differentiate between dissolved
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10 84 contaminants (mobile or leachable) and sequestered immobile ones. As many firms
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12 85 now remediate metals via chemical sequestration (i.e., converting the metals to an
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14 86 immobile state and leaving them in the ground), there is a critical need to differentiate
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17 87 between mobile and immobile metals, a major drawback of the XRF method.^{20,21} The
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19 88 relatively low sample throughput from third-party analyses limits on-site decision making,
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21 89 slows the time-to-completion, and increases overall project costs. In the academic realm,
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23 90 researchers have also made advancements towards fieldable detection systems for
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26 91 detection of metals, including paper-based platforms.^{22–34}

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28 92 Here we report a microfluidic paper-based analytical device, named On-Target
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31 93 Chemistry cards, for rapid quantification of metals in water. The cards are a modified
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33 94 version of the previously reported linear distance-based paper analytical devices where
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35 95 the analyte concentration is read as the length of color formation along a test
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37 96 channel.^{24,33–35} The distance-based approach requires no external instrumentation and
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40 97 eliminates the need to differentiate hues and intensities common to other colorimetric
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42 98 paper-based analytical devices. Linear distance-based paper analytical devices have
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44 99 been demonstrated for a variety of analytes, including metals.^{24,33–35} While the distance-
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47 100 based detection has proven effective, the analysis can take up to 30 min based on the
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49 101 slow liquid transport down the channel. The On-Target Water Chemistry test cards still
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51 102 possess the advantages of the linear distance-based paper analytical devices (portable,
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54 103 low-cost, easy-to-use, and quantitative) with the additional benefit of results being
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3 104 generated in 2-3 min. The reduced analysis time is achieved using a circular flow path
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5 105 where the sample flows radially outward from the tests central inlet (**Figure 1**, label 5)
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8 106 and, in the presence of the target analyte, creates a circular color change whose
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10 107 diameter correlates with analyte concentration (**Figure 1**, label 7). The flow rate is faster
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12 108 because the increasing diameter results in higher capillary pressure to offset the
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14 109 increasing viscous drag. Without preconcentration, low ppm (mg/L) detection limits have
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17 110 been achieved. To address lower analyte levels, a membrane pre-concentration system
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19 111 is also reported as a system that integrates with the On-Target test cards. Using the
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21 112 combined systems, it was possible to achieve single ppb detection limits of metal ions
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23 113 within 9 min. The new radial detection technology will ultimately allow sampling to occur
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26 114 more frequently and in more settings to ultimately permit better monitoring and
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28 115 mitigation of environmental pollutants than can be achieved at present.
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32 33 117 **Experimental Section**

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35 118 **Chemicals and materials.** All solutions were prepared using deionized (DI) water.
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37 119 Ascorbic acid, bathophenanthroline, cadion, cetylpyridinium bromide hydrate, L-ascorbic
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39 120 acid, nitric acid, potassium acetate, sodium hydroxide, zincon, Whatman Grade 4
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42 121 Chromatography Paper, and Empore Extraction Disks (Model 2271) were all purchased
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44 122 from Sigma-Aldrich (St. Louis, MO). Laminationers (3 Mil, 9" x 11.5") were purchased
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46 123 from Amazon (Seattle, WA) and mylar bags (3" x 5") were purchased from Uline
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48 124 (Seattle, WA). The real-world water samples were collected from the Animas River at
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50 125 the Gold King Mine (Silverton, CO) and the Argo Tunnel Water Treatment Plant (Idaho
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53 126 Spring, CO).
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3 127 **On-Target device fabrication.** The On-Target test cards are comprised of three
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5 128 material layers. The central layer is made of Whatman Grade 4 filter paper. Wax
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8 129 designs are printed onto the central layer to contain lateral fluid movement (**Figure 1**)
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10 130 using a Xerox ColorQube 8570DN. The wax is melted using a Puhui T-962C infrared
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12 131 oven at 150 °C for 4 min. Inkjet printing (Epson ET-2550) was used to add a QR code
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14 132 and details on the specific assay. Finally, inkjet printing was used to deposit assay
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17 133 reagents that both generate colors in the presence of analyte and control selectivity in
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19 134 the 3.8 cm test zone. The top and bottom layers of the card are commercial lamination
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21 135 sheets, which are impermeable to fluid, and control evaporation. A sample introduction
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23 136 orifice (i.e., test inlet) is laser cut in the top lamination layer using a Trotec Speedy100™
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26 137 laser system for sample addition. The lamination and porous layers are assembled,
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28 138 precisely aligned with a custom fixture, and then laminated together at 145 °C (Akiles
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31 139 Prolam). The three combined layers are then laser cut around their perimeter to yield
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33 140 individual test cards, each having the dimensions of a standard business card (3.5" ×
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35 141 2.0"). The test cards are stored in Mylar bags at ambient conditions (22 ± 2 °C) until
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38 142 used. Adobe Illustrator CS6 (version 19.2.1) was used for the design of the On-Target
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40 143 cards printed features and laser cutting pattern. The diameter to concentration
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42 144 relationship was plotted using Origin Pro8 SR4 v8.0951 (Northampton, MA).

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146 **Results and Discussion**

147 **Diameter-based colorimetric detection method.** The On-Target test cards can be
148 used to test water from mine water runoff, storm water, pool/spa, industrial wastewater,
149 household water supplies, or recreational bodies of water (**Figure 2a**). To use an On-

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3 150 Target test card, 100 μL of sample is collected (**Figure 2b**) and added to a vial
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5 151 containing 10 μL of buffer (**Figure 2c**) that adjusts the sample pH to an acceptable
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7 152 range for the assay, ensures the analyte is in the correct oxidation state, masks
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10 153 interferences, and, depending on the test type, assists with immobilization or color
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12 154 change enhancement. After mixing by briefly shaking the vial, the contents of the vial
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14 155 are added to the sample inlet port of an On-Target card using a disposable pipet
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17 156 (**Figure 2d**). After 3 min, the metal concentration is determined by using a ruler to
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19 157 measure the diameter of the circular color change area (**Figure 2f**). The resulting
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21 158 diameter is compared to a calibration curve or conversion table to convert the diameter
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23 159 to analyte concentration. The test cards allow for quantitative readout without needing
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25 160 expensive external analysis equipment. This is accomplished using the unique
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27 161 diameter-to-concentration relationship that results from the target ion being trapped and
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29 162 depleted from the bulk sample upon reacting with the pre-printed colorimetric
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31 163 chemistries.

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35 164 On-Target Water Chemistry test cards have been developed and thoroughly
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37 165 tested for 3 metal ions: copper (Cu^{2+}), total iron (Fe^{2+} and Fe^{3+}), and zinc (Zn^{2+}). We
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39 166 adapted assays for the three metals using previously reported mPAD
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41 167 methods^{23,24,29,34,35} along with the following colorimetric reagents and their referenced
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43 168 procedures: zincon for Cu,^{36–39} bathophenanthroline for Fe,^{40,41} and dithizone for Zn.^{42,43}

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49 170 **On-Target device performance.** As part of the rigorous testing to understand the
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51 171 performance of each metal test, we completed calibration curves and interference
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53 172 testing for Cu, Fe, and Zn. The calibration curves were collected following the procedure

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3 173 outlined in **Figure 2** with standard metal solutions at concentrations ranging from 0 ppm
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5 174 to 100 ppm. The calibration curves in **Figure 3** show the relationship between test
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7 175 diameter and metal concentration for each test along with representative test cards
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10 176 images for 1, 10, 25, 50, and 100 ppm. The working range of the test cards is 0.1 to 20
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12 177 ppm and the full range calibration curves follow a second order polynomial. The
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14 178 associated fit equations for the Cu, Fe, and Zn curves are $y=-0.26x^2+4.85x+1.99$
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17 179 ($R^2=0.98$), $y=-0.02x^2+1.39x+2.01$ ($R^2=0.99$), and $y=-0.01x^2+1.26x+2.6$ ($R^2=0.99$),
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19 180 respectively. The calibration curves are non-linear due to evaporation through the inlet
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22 181 port and the device being mass transfer limited. In other words, the metal ions within the
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24 182 sample front are depleted (i.e., complexed with the colorimetric reagent) before
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26 183 reaching the outer edges of the device along with evaporation preventing the unbound
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28 184 metal ions in the last plug of fluid from flowing outward. The lowest concentration
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31 185 discernible by eye for the Cu, Fe, and Zn test cards is 100 ppb. The average percent
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33 186 coefficient of variation is 1.4%, 2.0%, and 2.1% (n=8) for Cu, Fe, and Zn, respectively.
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35 187 The test cards are stable for at least 8 months at room temperature (22 ± 2 °C) in Mylar
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38 188 bags. To determine the stability of the test cards we have an ongoing stability study
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40 189 where the test cards are tested in quadruplicate every month. After 10 months of testing,
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42 190 there has been no change (results are all within error) in signal diameter.

44 191 The specificities of all tests were determined through a series of interference
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47 192 screening experiments.⁴⁴ Thirteen different potential interferents were tested both
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49 193 individually and combined with the target metal using the On-Target format. All results
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51 194 were compared to a positive (standard solution of target metal) and negative control
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54 195 (blank of deionized water) to determine whether a non-specific interaction occurred. For
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3 196 example, an aqueous mixture of 10 ppm Zn and 10 ppm Cd was tested on a Zn On-
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5 197 Target card and compared to that of an aqueous solution of 10 ppm Zn. The Zn and Cd
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7 198 mixture produced the expected pink color change but with a diameter of 25 mm, which
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10 199 is 11 mm larger than the Zn solution alone (14 mm). The tolerance ratio, defined as the
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12 200 ratio of interferent to analyte that causes a change of 10% or more,²⁴ is equal to 1 for
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14 201 Cd in the Zn test. This means that the cadmium interferes with the chemistry of the Zn
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16 202 On-Target card and is therefore classified as an interfering species at or above 10 ppm.
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18 203 Tests that included the interfering species alone (i.e., no target metal present) and
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20 204 yielded a color change not matching the expected color (**Table 1**) were considered to
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22 205 not interfere so long as in the presence of the target metal, the color formation was of
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24 206 the expected diameter and color. By following this approach, the interferents in **Table 1**
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26 207 were identified and their tolerance ratio was determined.

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30 208 **Membrane pre-concentration system.** While ppm detection limits are sufficient for
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32 209 some metals, lower levels are needed to assess environmentally relevant levels of
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34 210 many metals. We expanded the range of detectable concentrations on the On-Target
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36 211 Water Chemistry test cards down to single ppb levels through the development of a new
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38 212 membrane pre-concentration system. The pre-concentrator (depicted in **Figure 4**)
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40 213 housing was designed as an assembly of 2-dimensional acrylic pieces. The base pieces
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42 214 are 1/4 inch acrylic to increase rigidity and the upper housing is 1/8 inch acrylic. These
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44 215 pieces were cut on a laser cutter to allow for rapid design iterations. The reported pre-
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46 216 concentrator takes 1 L of aqueous sample and concentrates 95% of the contained
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48 217 metals onto an EmporeTM Chelation Disk (160.0 ± 8.0 mL/min or 5.7 ± 0.3 min, 18.8
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50 218 Watts, n=24) located in the filter cassette on the pre-concentrator outlet. The metals are
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3 219 then eluted off the disk using 500 μ L of nitric acid (pre-loaded and sealed in a syringe)
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5 220 that's pushed through the filter cassette and into a buffer vial specific to the metal test of
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7 221 interest. The buffer vial is pre-loaded with 3 M potassium hydroxide to neutralize the
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10 222 acid and a concentrated buffer to set the pH for the specific test. The resulting solution
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12 223 is 1000X more concentrated and can be analyzed directly on an On-Target test card.
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14 224 **Figure 5** is a plot of the diameter of color change vs. the concentration of aqueous Zn
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16 225 (**Figure 5a**) and Cu (**Figure 5b**) detected on the On-Target cards with and without use
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18 226 of the pre-concentration system. When the pre-concentration system is used in
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20 227 combination with On-Target test cards, the limit of detection is 2.5 ppb for Zn and 1 ppb
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22 228 for Cu and the percent coefficient of variation is 18.5% and 6.5% for Zn and Cu,
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24 229 respectively. While these values are higher than some traditional assays, the short
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26 230 assay time combined with the simplicity of the system still make these systems of
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28 231 interest, particularly in screening experiments.

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33 232 **Real-world detection of the *On-Target System*.** The Cu, Fe, and Zn On-Target Water
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35 233 Chemistry test cards were tested with three water samples: Gold King Mine water
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37 234 before and after remediation and untreated water from the Argo Tunnel. The samples
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39 235 were also analyzed by Flame Atomic Adsorption Spectroscopy (FAAS) analysis for
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41 236 validation. The performance of the On-Target cards relative to the traditional FAAS
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43 237 method are shown in **Figure 6**. The concentration of metal ions in the three water
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45 238 samples ranged from 0.1 ppm to 34.8 ppm. At concentrations between 1 ppm and 30
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47 239 ppm, the two methods agree at the 95% confidence interval. At concentrations above
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49 240 30 ppm the results deviate for the two methods due to the On-Target cards calibration
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51 241 curves plateauing from saturation of the color formation diameter as explained above.
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3 242 The EPA levels range from 0.1 to 500 ppm,⁴⁵ depending on the metal, which is within
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5 243 the working range of the reported system. Concentrations detected above the EPA
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7 244 cutoff are considered 'impaired' and of serious health concern. The On-Target test
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9 245 cards allow for quantitative aqueous metal analytics to be generated on-site for
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11 246 emergency response and on-going monitoring and remediation. Looking to the future,
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13 247 the test card will be expanded for more analytes and sample types.
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19 249 **Conclusions**

21 250 In conclusion, a new form of distance-based detection using radius or diameter instead
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23 251 of length is reported. The approach achieves similar or better detection limits in one-
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25 252 tenth the time of linear devices enabling results in 2-3 minutes and represents a
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27 253 significant step forward in this field. To demonstrate the utility of the approach, Cu, Zn,
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29 254 and Fe assays were developed and used to measure levels in mining water samples.
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31 255 To improve detection limits and enable measurement of ppb concentrations, a simple
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33 256 pre-concentration system was developed that uses a portable pump to concentrate
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35 257 metals onto an ion-exchange membrane. After elution and neutralization, the final
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37 258 system could achieve 1000-fold sample enrichment. When the distance-based test
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39 259 cards were used for real samples, the method agreed with the results from AAS from
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41 260 0.1 to 30 ppm but did deviate at higher concentrations due to color saturation.
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49 262 **Acknowledgments**

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Figure Captions

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3 353 **Figure 1.** Photograph of an On-Target test card for Fe with labels on the different
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5 354 device features: inkjet printed labeling, hydrophobic wax barrier defining test zone,
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7 355 hydrophobic wax barrier around the perimeter of the card, bare hydrophilic
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9 356 chromatography paper, laser cut test inlet, printed colorimetric reagent for aqueous Fe,
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11 357 and color change resulting from 100 ppm of Fe.
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17 359 **Figure 2.** Photographs of an Fe On-Target test card being used to test a Colorado River,
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19 360 including (a) the sampling location, (b) water being collected from river, (c) 100 μL
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21 361 aliquot of collected sample being added and mixed with buffer in a vial, (d) addition of
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23 362 the buffered sample to the test card inlet, (e) sample flowing radially outward with color
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25 363 change occurring from Fe complexation, and (f) measuring of the resulting color change
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27 364 diameter to determine concentration.
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33 366 **Figure 3.** Calibration curves for Fe, Zn, and Cu On-Target cards with representative
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35 367 images of test zones for 1, 10, 25, 50, and 100 ppm on each test type. Each data point
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37 368 represents the average of four independent test card results and the error bars are the
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39 369 standard deviations of each concentrations average diameter.
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44 371 **Table 1.** The expected color changes and known interfering species for each test type.
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49 373 **Figure 4.** Photograph of the pre-concentration system with labels on the key
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51 374 components.
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3 376 **Figure 5.** On-Target calibration curves for (a) Zn and (b) Cu with and without the pre-
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5 377 concentration system. The calibration curves are plotted on a logarithmic scale. Each
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7 378 data point represents the average diameter measured from four independent samples
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9 379 and test cards, and the error bars correspond to the standard deviation.
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14 381 **Figure 6.** Performance of the On-Target test cards for Cu, Fe, and Zn relative to
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16 382 traditional flame atomic absorption spectroscopy laboratory-based testing. The solid line
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18 383 represents perfect agreement between the two methods.
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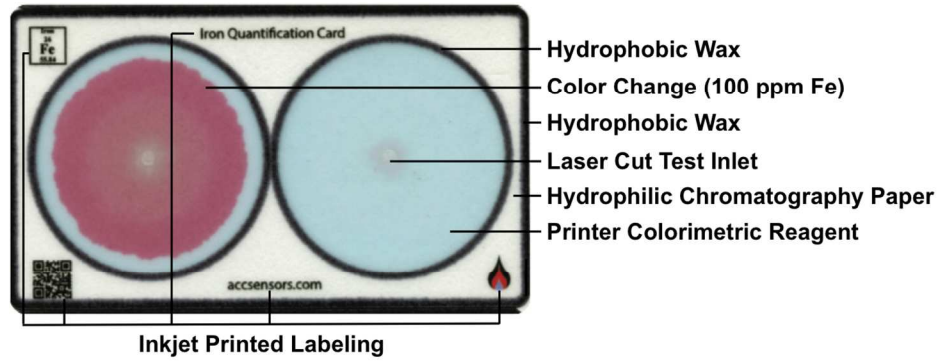
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23 385 **TOC Graphic.** The On-Target test cards are a rapid testing method for untrained users
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25 386 to quantitatively assess contaminants in aqueous samples.
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Figures



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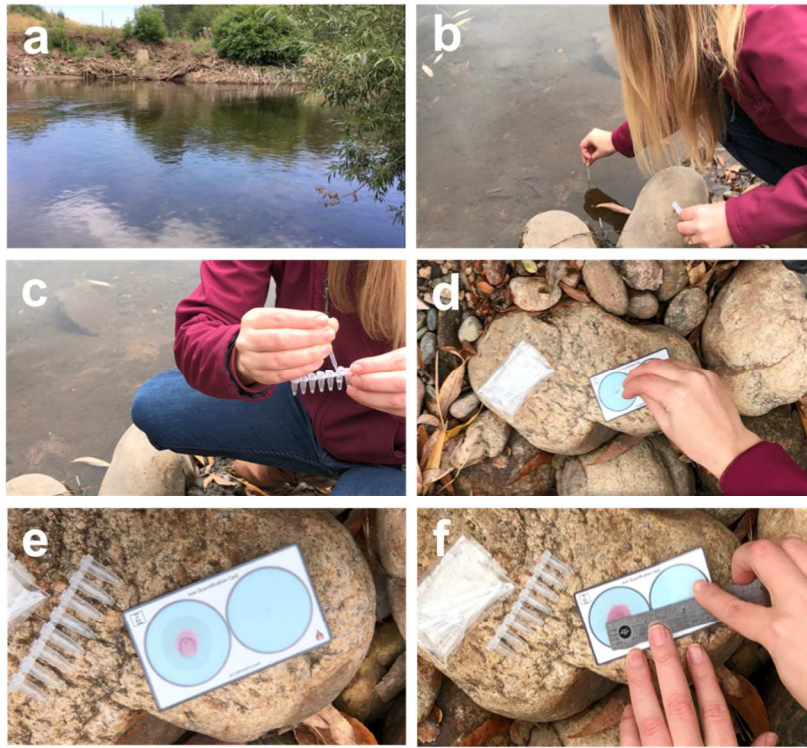
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Figure 1 / Hofstetter et al.

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Figure 2 / Hofstetter et al.

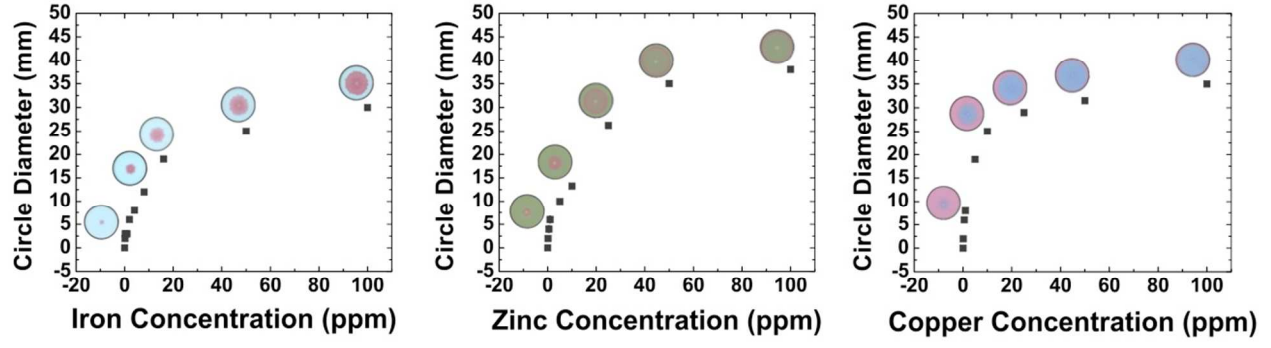


Figure 3 / Hofstetter et al.

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Test Metal (M ⁺)	Test Color Change	Known Interferent (Tolerance Ratio)
Zinc (Zn ²⁺)	Green to Pink	Cu ²⁺ (1), Fe ²⁺ (1), Cd ²⁺ (1), and Ni ²⁺ (1)
Iron (total Fe)	Blue to Pink	Cu ²⁺ (1)
Copper (Cu ²⁺)	Pink to Blue	None

Table 1 / Hofstetter et al.

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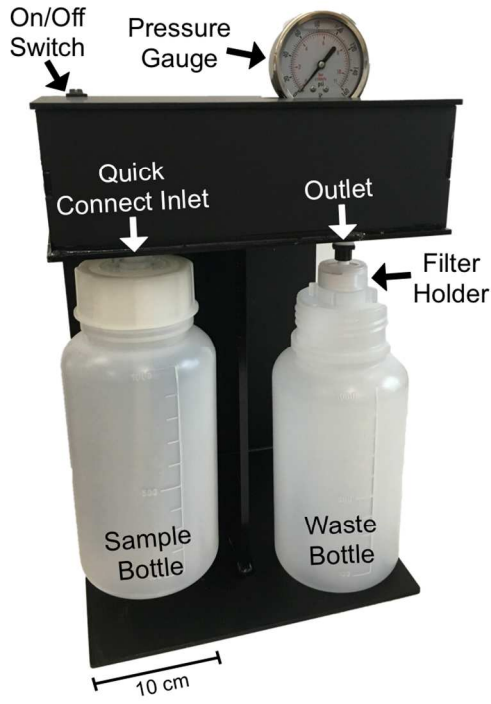


Figure 4 / Hofstetter et al.

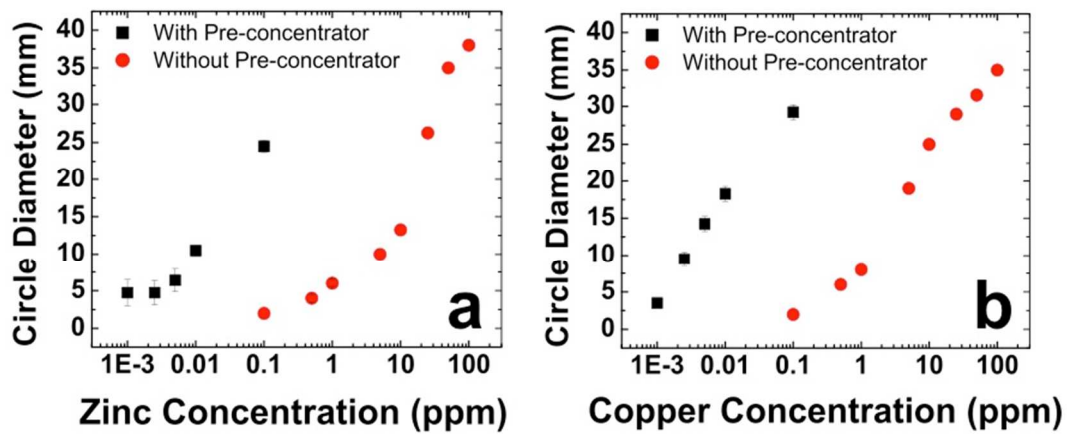


Figure 5 / Hofstetter et al.

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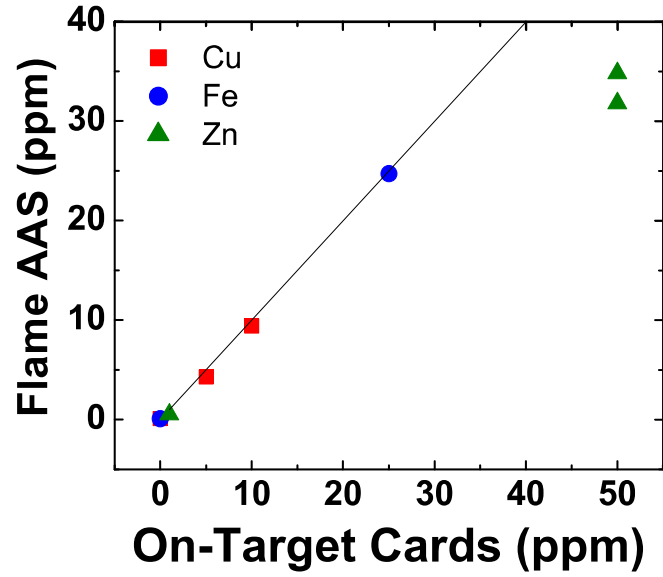


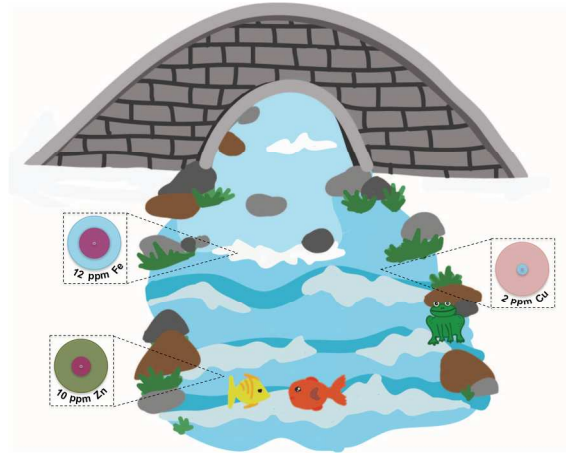
Figure 6 / Hofstetter et al.

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