# Environmental Science Water Research & Technology

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#### Water Impact Statement

We present for first time the concept **METIand** that merges Microbial Electrochemical Technologies (**MET**) with constructed wet**lands**. METIands are based on electroconductive biofilters for treating urban wastewater in decentralized systems in a sustainable way with no energy cost. Our strategy was the seed for an innovative European H2020 project devoted to construct full scale applications of METlands (www.imetland.eu).

**Technology Accepted Manuscript** 

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**Environmental Science: Water Research** 

1	Microbial Electrochemical Systems outperform fixed-bed biofilters for
2	cleaning-up urban wastewater
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17	In this work we present for first time the concept of integrating Microbial Electrochemical
18	Technologies (MET) with the natural wastewater treatments biofilters used in constructed
19	wetlands (CW) to form METlands. In order to validate this technology, three lab-scale
20	horizontal subsurface flow (HSSF) biofilters, two hosting electroconductive material and one
21	gravel biofilter (control) were operated for 525 days to define the best design and operational
22	conditions to maximize removal of wastewater pollutants. Organic loading rates tested ranged
23	from 2 to 24 g BOD <sub>5</sub> m <sup>-2</sup> d <sup>-1</sup> at hydraulic retention times (HRT) from 4 to as low as 0.5 days,
24	respectively. The electroconductive biofilter showed the best COD and BOD removal rates per
25	volume of bed, achieving mean values of 213 g COD $m^{-3}d^{-1}$ and 119 g BOD $m^{-3}d^{-1}$ at the lowest
26	HRT (0.5 d). Ammonia and total nitrogen maximum removal efficiency at 3.4 days of HRT were
27	97 and 69 %, respectively, in the electroconductive biofilter. Bacterial communities were
28	studied by 16S rDNA Illumina sequencing with the aim of understanding the role of the
29	electrically conductive material in selecting microbial populations. Deltaproteobacteria (a
30	known electroactive taxon) were enriched in presence of electrically conductive bed.
31	Geobacter and Geothrix were the dominant genera in the deeper zone of the electrically
32	conductive bed where oxidation of organic matter occured. The results suggest that the
33	enhancement in biodegradation rate will significantly reduce the area requirements of classical
34	CW.

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Introduction

36 Conventional wastewater treatments require high energy, operation and maintenance 37 costs. In addition, due to population growth and urban expansion, the volume of sewage sludge produced by wastewater treatment is constantly increasing <sup>1</sup>. Thus, a 38 39 different water-energy nexus is required to cope with the future global water demand. 40 Since the discovery of electroactive microorganisms, Microbial Fuel Cells (MFC) 41 were proposed to play an important role in wastewater treatment for converting the waste into clean energy, by oxidizing organic and inorganic matter to generate electrical 42 current <sup>2,3</sup>. In these devices, electrons produced by the microbial metabolism are first 43 transferred to an electrode (anode), and then to a second electrode (cathode) via a 44 45 conductive material containing a resistor <sup>3</sup>. In this configuration, the anode act as terminal electron acceptor as any other natural acceptor like oxygen, nitrate or Fe(III). 46 47 The clear advantage of exploiting electro-stimulated communities is that electrodes can 48 boost microbial metabolism in anaerobic systems that are typically electron acceptor 49 limited. Electroconductive material may represent an inexhaustible source of electron 50 acceptors, hosting the additional advantage of providing a more easily modulated redox potential compared to standard, low-reducing redox species that generally drive these 51 systems<sup>4</sup>. The redox potential of the anode depends on the chemistry and 52 53 bioelectrochemistry around the electrode. Moreover, the electrochemical 54 characteristics of those microbial-assisted devices can be simply controlled by altering 55 their configuration. Thus, they can be operated in different configurations, such as i) short-circuit, no resistors between electrodes <sup>5</sup>; ii) MFC, able to harvest energy in 56 presence of a resistor <sup>6</sup>; and iii) Microbial Electrolysis Cell (MEC) by poising a certain 57 potential through a potentiostat or a power source  $^{7,8}$ . 58 59 A suitable scenario for testing this emergent technology is the Constructed Wetlands 60 (CW) since they are a good alternative for wastewater treatment in small communities and are used worldwide<sup>9</sup>. Low cost operation and maintenance, low energy requirements, low 61 production of sewage sludge (just in primary treatment) and good landscape integration are 62

- 63 some of the most attractive advantages of CW compared to conventional treatment systems
- <sup>10</sup>. However CW treatment is constrained by limitations such as large land requirements (3-10
- 65 m<sup>2</sup> PE<sup>-1</sup>\* depending on design) (Vymazal and Kropfelova, 2008, Tilley et al., 2008) and clogging

<sup>\*</sup> PE Population equivalent is the number expressing the ratio of the sum of the BOD load produced during 24 hours by industrial facilities and services to the individual BOD load in household sewage produced by one person in the same time. For practical calculations it is assumed that one unit equals to 60 g of BOD per 24 hours.

by the accumulation of solids <sup>13,14</sup>. Recommended surface organic inlet load for HSSF CW is 66 reported as 6.0 g BOD<sub>5</sub> m<sup>-2</sup> d<sup>-1</sup> in order to achieve a value under 30 mg BOD<sub>5</sub> L<sup>-1</sup> in the effluent 67 and avoid clogging <sup>15,16</sup>. HSSF CW were initially presented as environments that could take 68 advantage of depth-depending redox potential gradients <sup>17,18</sup>. Previous reports argued that 69 redox conditions in CW could be controlled by altering the organic loading rate, the hydraulic 70 design and the mode of operation <sup>19</sup>. Following this strategy, different groups have integrated 71 MFC elements to lab-scale CW with the purpose of harvesting electricity  $2^{20-22}$ . In spite of using 72 wastewater as organic fuel, the power densities reported were as low as 1.84 - 44.63 mW m<sup>-2</sup> 73 <sup>23</sup>, which is a range typical for sMFC operating in soil or sediments, but still far from 10 W m<sup>-2</sup> 74 values obtained using filter press bioelectrochemical reactors<sup>8</sup>. This is mainly due to the fact 75 76 that redox gradients are not broad enough in this kind of environments and in situ 77 implementation of power-harvesting devices is indeed limited. 78 However, we still believe that CW are a suitable environment for implementing 79 microbial electrochemical systems. Our aim was not to harvest energy but to enhance the rate 80 of pollutant removal by converting the classical inert biofilter into an electroconductive

- 81 biofilter where its redox state could be tuned or controlled by electrochemical tools. Our
- 82 results revealed how the integration of METs in wetlands resulted in a powerful hybrid
- 83 technology so-called METland <sup>24</sup>, that strongly outperforms the treatment of urban
- 84 wastewater through the stimulation of different microbial populations.
- 85

# 86 Experimental

# 87 Design and construction of electroconductive biofilters

88 In this study, four laboratory-scale HSSF biofilters were constructed for determining the best 89 design and operational conditions to maximize wastewater pollutants removal. A control unit 90 used standard siliceous gravel (Ø 6-12 mm) as biofiltering bed (Fig.1, A). An electroconductive 91 bed configuration (Fig.1, B) was constructed with a single material, acting as a whole electrode. 92 This configuration did not allow the conversion of microbial metabolism into electrical current 93 to be monitored, since anode and cathode were not differentiated. In order to harvest 94 electrochemical information about the process, a three electrodes system was additionally 95 constructed by using a hybrid unit made of inert gravel and polarized coke bed (Fig.1, C). An 96 additional hybrid unit operating under short-circuit (Fig.1, D) was constructed as control. In 97 these hybrid biofilters, conductive material was vertically inserted into the gravel. The short-98 circuit hybrid unit acted as a single electrode without differentiated anode and cathode.



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Fig.1 Simplified design of the four systems A) Gravel biofilter (control), B) Coke biofilter, C)
Hybrid biofilter, D) Hybrid polarized biofilter.

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The conductive material in the bed was coke granules ( $\emptyset$  5-10 mm). The dimension of the biofilters were 0.52 m long, 0.34 m wide and 0.30 m high, and material layer was 0.20 m deep, with a total bed volume of 0.034 m<sup>3</sup> and a water volume of 0.011 m<sup>3</sup>. Each biofilter had a drainage pipe, located on the flat bottom, for effluent discharge and the water level was kept below the surface.

109 The hybrid polarized biofilter hosted a coke anode of 0.006 m<sup>3</sup> as schemed in Fig.1. A 110 plate of graphite (3 cm x 3 cm x 0.5 cm, Sofacel) buried into the coke anode acted as electron collector. The cathode was made of carbon cloth (0.34 m x 0.15 m, Resinas Castro, 420 g m<sup>-2</sup>). 111 112 Anode and cathode were connected by a copper wire to a potentiostat unit (Nanoelectra S.L., 113 Spain). A third electrode (Ag/AgCl) buried in the anodic bed acted as reference to polarize the 114 anode at 0.3 V (vs. Ag/AgCl). The anode potential and the current were periodically measured 115 using a digital multimeter (Model 2700, Keithley Instruments, USA). Data was recorded every 116 10 s on a spreadsheet using ExceLINX (Keithley) via an interface card (GPIB Interface Boards, 117 Keithley) linked to a personal computer. The performance of the polarized biofilter was

evaluated in terms of coulombic efficiency (CE, %) comparing the total electrons harvested by the anode to the electrons possibly generated by the microbial oxidation of the substrate. For continuous flow through the system, we calculate CE based on the COD change, and the flow rate, q<sup>25</sup>, as

$$CE = \frac{8I}{Fq \ \Delta COD}$$

122

where 8 is a constant used for COD, based on conversion from  $gO_2$  (MW = 32 g mol<sup>-1</sup>) to mol e<sup>-1</sup> (4 mol e<sup>-</sup>/mol O<sub>2</sub>), I is the current obtained over time and F is the Faraday's constant.

125 The systems were operated in parallel and fed with real urban wastewater from the 126 municipality of Carrión de los Céspedes (Sevilla, Spain) (2500 inhabitants) under discontinuous 127 flow regime during 525 days (75 weeks). Wastewater was pretreated in an Imhoff tank in order 128 to remove solids and prevent a potential clogging of the systems. The feeding from the Imhoff 129was made by programmed pumping, by means of 12 daily periods, simulating the130production of wastewater in small populations  $^{26}$ . Several organic loading rates were tested131 $(2.0 \pm 1.0; 4.2 \pm 0.7, 9.2 \pm 2.8, 13.8 \pm 9.5 \text{ and } 24.0 \pm 12.7 \text{ g BOD}_5 \text{ m}^{-2} \text{ d}^{-1}$  in average) at the132following hydraulic retention times (HRT): 4.0, 3.4, 1.7, 0.8 and 0.5 days, respectively.

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## 134 Physical, chemical and statistical analysis

135  $BOD_5$ , total suspended solids (TSS), total nitrogen (TN), ammonia (NH<sub>4</sub>) and nitrate (NO<sub>3</sub>) were 136 analysed weekly; COD was analysed twice a week, following the standard methods (APHA-137 AWWA-WEF, 2005). Temperature (T), pH, electrical conductivity (EC), dissolved oxygen (DO), 138 and redox potential (ORP) were measured weekly with a handheld multiparameter (YSI 556 139 MPS). Samples were taken at the inlet and the outlet of the systems and water flow was daily 140 measured. Moreover, hybrid systems were also sampled through sampling tubes buried in the bed, before and after the electroconductive barrier (anode), in order to calculate the 141 142 coulombic efficiency. Inlet wastewater analyses is shown in Table 1S. Removal rates were 143 calculated as grams per cubic meter of bed material per day. Removal efficiencies were 144 calculated as percentage.

Statistical procedures to evaluate the effect of HRT for every water quality parameter
were conducted using the Statgraphics Centurion XVII statistical software package. T-test or
Wilcoxon tests were used to determine the differences of every water quality parameter
among the effluents, depending on the type of data (95 % confidence).

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#### 150 Microbial communities

151 Sampling, DNA extraction and 16S rDNA sequencing. Samples were taken from lab-scale 152 biofilters and inlet wastewater to determine the composition of their microbial community at 153 four different spots: anode in the hybrid polarized biofilter (B1), upper area of the coke 154 biofilter (B5), upper area of the gravel biofilter (B6) and inlet wastewater (B7). Either granules 155 of coke (B1, B5) or gravel pebbles (B6) were sampled with tweezers and loosely attached 156 bacteria were removed by dipping them in 3 consecutive sterile saline solutions (50ml, NaCl 7 157 g/l). Coke and gravel pebbles were then frozen for 1 week until performing DNA extraction. 158 Around 10 granules/pebbles were extracted per spot. DNA was extracted with PowerSoil spin 159 columns (MO BIO Laboratories), suspended in 60 µl of sterile MilliQ water and quantified with 160 PicoGreen (Invitrogen). A total of 3 ng of DNA were amplified with primers 515F-CS1 161 (ACACTGACGACATGGTTCTACAGTGCCA GCMGCCGCGGTAA) and 806R-CS2 162 (TACGGTAGCAGAGACTTGG TCTGGACTACHVGGGTWTCTAAT). The polymerase used was Q5 163 Hot Start High-Fidelity (New England Biolabs) and the PCR conditions were: initial denaturation at 98°C for 30" followed by 30 cycles of 98°C x 10", 60°C x 20" and 72°C x 20", and a final
elongation step of 72°C for 2'. A 1/100 dilutions of PCR products were then re-amplified (15
cycles) with Illumina's primers. Finally, products were run on a Bioanalyzer (Agilent) and the
successful generation of equimolar pools was confirmed by qPCR. Sequencing was performed
in a MiSeq equipment using the 2x250 bp format and following Illumina's protocol.

169 The Illumina Miseq sequence reads have been deposited in the European Nucleotide170 Archive (ENA) database under accession Nr. PRJEB10685.

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Bioinformatic analysis. The total sequence reads were analysed with the QIIME 1.7 pipeline <sup>28</sup> 172 with few stitches along the way. Briefly, complementary reads were merged using fastq-join <sup>29</sup>. 173 174 Subsequently, our quality filtering strategy removed complemented sequences that had one of 175 the following characteristics: (i) deviated more than 10 bp from the expected length (292); (ii) 176 contained primers with more than 1 mismatch; or (iii) contained nucleotides with Phred score 177 <20. Filtered seqs were organised in OTUs by de novo picking using Usearch <sup>30</sup> and one 178 representative sequence per OTU was chosen. Taxonomy was assigned using the GreenGenes 179 database <sup>31</sup> version 10 12 at the 97% identity rate. Furthermore, sequences were aligned and a tree generated using FastTree 2.1.3<sup>32</sup>. Finally, in order to investigate alpha diversity with 180 181 QIIME, OTUs containing less than 0.005% of the total sample reads were removed according to 182 Bokulich <sup>33</sup>. The results have been represented as relative abundance of a specific sequence in 183 every sample. Taking into account the possible effect of deviation introduced by the 184 implemented protocol and that not all the bacterial species have the same number of copies of 16S rRNA gen in their genomes <sup>34</sup>, the values can be related to percentage of cells of every 185 186 species that were part of the sampled communities.

187

### 188 Results and discussion

189 HSSF CW are biofilter setups that exploit biofilm-based natural process by means of inert 190 material like gravel with the purpose of treating urban wastewater. Plants are typically 191 integrated in CW for oxygenating the root zone and for providing aerobic microorganisms an habitat within the anoxic environment <sup>35</sup>. Our approach consists in substituting inert material 192 193 for an electroconductive material in order to stimulate electroactive microorganism and 194 consequently biodegradation rates. Due to the oxygen supply role by plants we did not 195 include vegetal species in our experimental set up in order to achieve a better control of the 196 redox interaction between bacteria and bed.

198 Urban wastewater treatment by horizontal subsurface flow (HSSF) biofilters:

- 199 electroconductive versus non-electroconductive biofilters
- 200

201 Influence of the material on the wastewater pollutants removal. In order to quantify the 202 influence of the material, we tested two independent HSSF biofilters fully constructed with 203 electroconductive and inert material (Fig.1A,B). The organic matter removal rates, in terms of 204 COD and BOD<sub>5</sub>, were similar under a low organic loading rate regardless of the material (Table 2S). However, significant differences among both systems appeared when the organic loading 205 206 rate was increased. The coke biofilter performed removal efficiencies close to 100% despite 207 increasing the organic loading rate, while the gravel biofilter efficiency decreased as the organic loading rate increased (Fig.2). Indeed, the coke biofilter showed the best COD and 208 BOD<sub>5</sub> removal rates, achieving mean values of 213 g COD m<sup>-3</sup>d<sup>-1</sup> and 119 g BOD<sub>5</sub> m<sup>-3</sup>d<sup>-1</sup> (Table 209 210 2S). Furthermore, the gravel biofilter showed a more variable performance.

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Fig.2 A) Relation between normalized COD removed and COD inlet loading of the coke and the
gravel biofilters, B) Relation between normalized NH<sub>4</sub>-N removed and NH<sub>4</sub>-N inlet loading of
the coke and the gravel biofilters.

216

217 Statistical tests revealed that there were significant differences (p < 0.05) in the

effluent's concentration of COD and BOD<sub>5</sub> at every HRT (Table 3S) when the coke and gravel

219 biofilters where compared. The coke biofilter biodegradation rates led to effluents with 220 residual values up to 3-fold lower for COD and 4.5-fold lower for  $BOD_5$  (Fig. 3). COD and  $BOD_5$ coke biofilter effluent values never exceeded the limits of discharge, which are 125 mg COD L<sup>-1</sup> 221 (or > 75 % removal) and 25 mg BOD<sub>5</sub> L<sup>-1</sup> (or 70-90 % removal) (Dir. 91/271/EEC of 21 May 222 1991)<sup>36</sup>, in contrast with the gravel biofilter performance from 3.4 days of HRT onwards, which 223 average effluent concentration exceeded 25 mg BOD<sub>5</sub> L<sup>-1</sup> (Fig. 3). Even at the lowest HRT the 224 performance of the coke biofilter fulfilled the COD and BOD<sub>5</sub> discharge requirements in 225 226 percentage (91 % and 96 %, respectively), compared to hardly 73% and 86 % for the gravel biofilter (Table 2S). Caselles-Osorio and García<sup>37</sup> reported COD removal efficiencies of 71-85 % 227 in intermittent fed HSSF CW experimental systems with a nominal HRT of 3.4 days, which is 228 229 comparable to removal efficiencies of our control system at the same HRT (83 %). Coke 230 biofilter achieved mean BOD<sub>5</sub> removal rates as high as 99 % at high HRT (3.4 days).



- 232
- Fig.3 COD (A) and BOD<sub>5</sub> (B) influent and effluent average values of the coke and gravel filters.
  Error bars represent 95 % confidence interval.
- 235
- 236 The BOD<sub>5</sub> surface inlet loads applied at 1.7, 0.8 and 0.5 days of HRT (Table 2S) were
- 237 1.5, 2.3 and 4-fold, respectively, the recommended load (6.0 g  $BOD_5 m^{-2}d^{-1}$ ) and  $BOD_5$  average
- values of the coke biofilter effluent were always under 10 mg  $L^{-1}$  (Fig. 3). Even at very high inlet

239 organic loads, the coke biofilter had a great capacity to remove organic matter, without any evidence of clogging during the long course (525 days) of the experiment. A remarkable 240 241 conclusion is that just the coke biofilter fulfilled the Directive for COD and BOD<sub>5</sub> at a HRT as 242 low as 0.5 days. In contrast, for standard gravel biofilter a HRT as high as 3.5 days was required 243 for fulfilling the limits. Moreover, there were not significant TSS differences in the effluents of 244 the two biofilters, and both fulfilled the limit values of discharge (35 mg  $L^{-1}$ ) (Table 1S). 245 Nitrogen removal was also analysed under both electroconductive and inert materials 246 and a very similar result was found. Statistical analysis revealed significant differences (p < 247 0.05) among TN and NH<sub>4</sub>-N effluent concentrations at every HRT. The coke biofilter exhibited 248 the highest removal rates at every HRT (Table 4S). Interestingly, differences with gravel 249 biofilter were more noticeable than those found for organic matter removal. In the coke 250 biofilter, the maximum amount of nitrogen was removed at 0.5 days of HRT (TN 11.9 gN m<sup>-3</sup>d<sup>-1</sup>; NH₄ 11.2 gN m<sup>-3</sup>d<sup>-1</sup>) although the removal efficiency (%) decreased with decreasing HRT. This 251 trend has been reported in other studies <sup>16,38</sup>. The coke biofilter showed maximum average 252 253 removal efficiency values at 3.4 days, 97% of ammonia and 69% of total nitrogen compared to 254 71% and 51%, respectively, in the gravel biofilter. The minimum values were reached at 0.5 255 days, 39 % of NH<sub>4</sub>-N and 37 % of TN compared to 16 % and 19 %, respectively, in the gravel 256 biofilter (Table 4S). Fig. 2B shows that the coke biofilter had a trend to maintain higher 257 removal rates than gravel biofilter. The higher biodegradation rates generated effluents with 258 residual TN and NH<sub>4</sub>-N significantly lower (Fig. 1S). The results demonstrate that the coke 259 biofilter removed at least 2-fold the amount of TN and 2.5-fold the amount of NH<sub>4</sub> than 260 removed by the gravel biofilter (HRT 0.5 days). Therefore, at HRT shorter than 4 days 261 nitrification was higher in the coke biofilter compared to the gravel biofilter. Moreover, at 262 lower HRT, ammonia concentration in the effluent increased while nitrate was decreased 263 (Fig.1S). The improvement of the conversion of ammonia to nitrate and nitrogen removal 264 suggests the enhancement of other metabolic pathways in the electroconductive bed. 265

Electrochemical analysis using hybrid electroconductive setups. In order to quantify the role
of the electroconductive bed for accepting charge from microbial metabolism we constructed
a hybrid polarized biofilter (figure 1D). In contrast with the sole-coke biofilter, this setup allows
an accurate control of the electrical current by polarizing the system at 0.3 V (vs. Ag/AgCl).

The electrical current monitored throughout the assay revealed an expected profile, a stable value around 100 mA was measured (Fig. 4). Interestingly, an increase in the organic loading rate did not result in a clear increase in electrical current, suggesting that the electroactive biofilm was not limited in electron donor. In contrast, the increase in the organic

274 loading rates showed very good correlation with the organic removal rates only in the 275 presence of electroconductive material so we concluded that some other biodegradation 276 pathways, although not contributing to current production, are definitively being enhanced. As 277 the electron donor is not a limiting factor, other degradation routes must have a major 278 influence on the performance. In that sense, coulombic efficiency (CE) ranged from 37 % at 279 low organic loading rate to 9 % at maximum organic loading rate, which indicates that low 280 organic loading rates enhance the CE. The bacteria can biodegrade part of the COD through fermentation or the use of alternative electron acceptors <sup>39</sup> such sulphate or nitrate. This is 281 282 consistent with previous reports that showed how, under higher organic loading rates, electron flow is channelled towards methanogenesis or sulphate reduction so CE is reduced <sup>39</sup>. 283 284 Methane emissions are common in HSSF CW because these systems present appropriate 285 environmental conditions for methanogens and sulphate-reducing bacteria. These Archaea 286 and Eubacteria require environments with similar redox potentials and use the same types of electron donors (i.e., hydrogen, methanol, and acetic acid)<sup>9</sup>. Methane emission rates are very 287 288 variable and they are usually greater at the inlet than the outlet, given that methanogens activity is directly dependent of the organic load <sup>40</sup>. Further research about this topic should be 289 290 carried out to evaluate the contribution of METlands to methane emissions.



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Fig.4. Profile of electrical current, COD loading rate (g  $m^3d^{-1}$ ) and COD removal rate (g  $m^3d^{-1}$ ) during long term operation of the hybrid biofilter polarized at 0.3 V (vs. Ag/AgCl).

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296Together with the hybrid polarized system, a non-polarized hybrid biofilter was also297constructed (Fig. 1C) to evaluate the influence of the polarization versus the mere effect of the298coke. Interestingly, despite polarizing the anode our assays did not reveal significant299differences (p>0.05) in terms of COD and BOD removal among the two hybrid configurations300(Fig. 2S). This fact strongly suggests that the electroconductivity of the material exert a positive

301	influence on the microbial metabolism regardless of the existence of an electron flow among
302	the different electrodes. Our Hybrid biofilter is a single electrode configuration, a simplified
303	design of a short-circuited system that cannot provide current but optimizes the pollutants
304	removal. In that sense, our results are consistent with previous studies that reported how
305	compact short-circuited system provided higher biodegradation performance than MFCs
306	operating at maximum power <sup>41</sup> .
307	Redox potential was measured in both the electroconductive and the gravel biofilters.
308	There was a noticeable redox potential gradient with depth and distance from the inlet in the
309	systems which corresponded to COD and BOD. This gradient was greater in the
310	electroconductive biofilter (Fig. 3S). This gradient suggests the presence of an electron flow
311	from the deep bed to the more oxidized top layer of the coke bed.
312	In the hybrid systems the differences between materials were also remarkable. COD
313	removal rates in the electroconductive bed (Table 1) were ca. 5-fold higher than in the gravel
314	bed of the same hybrid device. Regarding nitrogen removal, both hybrid systems removed
315	similar amounts of total nitrogen and ammonia at high and medium HRT (Table 4S).

316

317 **Table 1** Urban wastewater treated by hybrid biofilter setups. COD overall averages ± SD, at

318 HRT = 3.4 d. Removal efficiencies in the conductive bed (%) were referred to the COD before 319 conductive bed.

COD levels (mg L <sup>-1</sup> )	Hybrid biofilter	Hybrid polarized biofilter
Influent	231 ± 58	231 ± 58
Before conductive bed	188 ± 55	182 ± 59
After conductive bed	89 ± 49	78 ± 31
Effluent	37 ± 20	35 ± 14
COD removal		
Removed in conductive bed (g m <sup>-3</sup> d <sup>-1</sup> )	50.9 ± 24.8	55.5 ± 26.0
Removal efficiency in conductive bed (%)	52 ± 18	56 ± 14
Removed in gravel before conductive bed (g m <sup>-3</sup> d <sup>-1</sup> )	12.8 ± 7.8	15.8 ± 13.4
Removed in gravel after conductive bed (g m <sup>-3</sup> d <sup>-1</sup> )	$10.4 \pm 7.0$	8.1 ± 4.8

320

## 321 Microbial communities

322 The analysis of four microbial communities revealed 696,288 raw reads that yielded a total of

689,911 high quality sequences with an average length of 292 bp (Table 5S). This volume of

324 sequences is around one order of magnitude greater than previously reported studies of

diversity in bioelectrochemical systems <sup>42</sup>, as result of improved sequencing technologies.

326 Clustering these sequences generated 16,582 OTUs evenly distributed between the four

327 samples. 2.33% of the sequence reads were not classified.

The classifiable sequences included members of 48 phyla of which an average of 64 % were *Proteobacteria*, ranging between 52% (anode of the hybrid polarized biofilter) and 74% (gravel biofilter).

331 Rarefaction curves showed saturation, indicating that a reasonable number of 332 sequence reads per sample were collected to reveal diversity at the sites (Fig. 4S). Rarefaction 333 curves indicate that predicted diversity was much less in the inlet wastewater than in the rest of the niches (around 70% of the number of identified taxa). Diversity estimators such as 334 335 observed OTUs, Chao1 and Shannon-Wienner, were significantly higher for coke granules 336 samples when compared to the gravel samples (Table 5S). The Good's coverage estimator 337 denoted that the sizes of the libraries were enough to cover almost 100% of the bacterial 338 communities. Shannon diversity indexes (H), which includes the information of both richness 339 (the number of species present) and evenness (how the abundance of each species is 340 distributed) were obtained for our system. They were distinctly higher (between 6.27 and 341 7.38) than those in other studies on electrochemical CW treating urban wastewater (4.36-5.5 342  $^{43}$ , 5.6 – 6.3  $^{44}$ ) and similar to the results of Lu et *al*.  $^{45}$  (*H*: 7.33-7.47). These results, together with the high number of taxa found in the samples, indicated a very high diversity. 343

344 Weighted Fast UniFrac analysis and Correspondence analysis (CA) were used to 345 identify the differences of the bacterial community structures based on their phylogenetic 346 lineages. CA showed that the four communities separated distinctly from one another despite 347 the same origin (Fig. 5S). CA plot revealed that coke and hybrid polarized biofilters are closely 348 related and that electroactive bacteria (Deltaproteobacteria) had the higher component 349 weight in both systems. Another closely related taxa to these biofilters were the classes 350 Holophagae (with the genus Geothrix, an electroactive bacteria of the phylum Acidobacteria), 351 and Brocadiae (phylum Planctomycetes). The class Brocadiae, involved in annamox processes, 352 only appeared in the anode of the polarized biofilter (table 6S). Alpha, Beta and 353 Gammaproteobacteria had the higher component weight in the inlet wastewater and the 354 gravel biofilter.

355

Presence of *Deltaproteobacteria* as indicator of microbial electroactivity. Our analysis of microbial communities revealed the presence of similar taxonomic groups with the exception of some remarkable ones. An interesting finding was the high presence of *Deltaproteobacteria* (Fig. 5) when the electroconductive material was the substrate (27.2 % in the coke biofilter and 23.4% in the hybrid polarized biofilter) in comparison with the gravel biofilter (8.1 %). Bacteria belonging to this group have been reported associated to electroactive biofilm from the very beginning <sup>46</sup> as they share the capacity for generating ATP from very low thermodynamic value

reactions <sup>47,48</sup>. In the anaerobic treatment of wastewater, *Deltaproteobacteria* assures the 363 364 removal of fatty acids of low energetic value as acetate which is typically the metabolic bottleneck of these systems <sup>49</sup>. In addition, *Deltaproteobacteria* can compete with 365 366 methanogenic microorganisms and their preponderance may reduce methane emissions. 367 However, we cannot confirm any outcompeting effect on methanogenic populations because, 368 apparently, some of the taxa were not amplified with the primer sets 515F/806R utilised for the sequencing <sup>50</sup>. In fact, only 0.1 % of OTUs correspond to the Kingdom Archaea, which 369 370 contains the main methanogenic groups. It must also be noted that community members with 371 multiple 16S copies may be over-represented. Nevertheless, our main purpose was to estimate 372 those genera associated with degradation processes and electroactive bacteria, groups that 373 were adequately represented.

374



375

376 Fig.5 Relative abundances of OTUs at class level (larger than 5% in average).

377

378 Some Deltaproteobacteria, like bacteria from the genus Geobacter, are able to transfer electrons to conductive materials <sup>51</sup>. Indeed, the largest presence of *Geobacter* was found in 379 380 the coke biofilter (2.9%) (Table 7S). Surprisingly, although at lower levels it was also found in 381 the inlet wastewater (0.45 %) and in the gravel biofilter (0.3%). Some studies have previously 382 reported the presence of Geobacter species in anaerobic digesters suggesting a role in performing direct interspecies electron transfer (DIET)<sup>52–55</sup> with a direct impact on methane 383 384 production. Interestingly, inlet wastewater for our assays was generated in an Imhoff tank, 385 which host environmental conditions similar to those found in an anaerobic digester. It seems 386 reasonable to expect the presence of Geobacter associated with other biofilm species in our gravel biofilter. In the Deltaproteobacteria, it is remarkable the dominance of some genera of 387 388 the family Desulfobulbaceae (Table 7S) in both the anode of the hybrid polarized biofilter 389 (20.8%) and also in the coke biofilter (16.8%), in contrast with low presence in the gravel

390 biofilter (1.6%). Moreover, other studies also reported *Desulfobulbus* species colonizing anodes 44,56–58 and, for instance, *D. propionicus* was previously reported to use the electrode 391 392 surface as an electron acceptor when pyruvate, lactate, propionate or hydrogen was provided as electron donor <sup>59</sup>. The presence of *Desulfobulbus* is especially relevant due to its fascinating 393 capacity for generating electrically conductive-microbial filaments <sup>60,61</sup>. These microbial 394 395 filaments transport electrons from the bottom of sediment, rich in hydrogen sulphide, up to 396 the oxygen-rich sediment that is in contact with the water. Interestingly, this situation is 397 similar to the one found in our METlands where a redox gradient is generated among bottom 398 and upper layers of the electroconductive bed. So, our results have revealed that specific 399 microbial consortia previously related to electrical current production were selected for by our 400 electroconductive biofilters from our inlet wastewater.

401 On top of that, other electroactive microorganism like *Geothrix*, an *Acidobacteria*<sup>62</sup>, 402 were also found in all the systems (table 7S), with a significant presence in the anode of the 403 hybrid polarized biofilter (3.2 %) and in the coke biofilter (2.2 %). Interestingly, *Geothrix* was 404 almost absent in the inlet wastewater and scarce in the gravel biofilter (0.2%).

405

406 Nitrogen cycle bacteria: nitrification and denitrification. Nitrogen removal is typically poorly
407 achieved under anaerobic conditions, showing a bottleneck in the ammonium oxidizing
408 process. Apparently this is not the case when electroconductive material is supporting the
409 biofilm growth (Fig. 1S) since this material outperforms gravel to remove nitrogen by 2-fold
410 (table 4S).

A deep analysis into the microbial communities' distribution may helps us to understand what different nitrogen metabolisms are active in our systems. The detection of ammonium oxidizers, like *Nitrosomonadaceae*, associated to the electroconductive material is remarkable if we consider that this family was absent in both the gravel and the inlet wastewater. Even more interesting was the presence of bacteria from the *Brocadiaceae* family (1.7%) in the anode of the polarized biofilter. This family of bacteria include several genera involved in the anaerobic oxidation of ammonia to dinitrogen via ANNAMOX<sup>63</sup>.

Another nitrogen pathway that could be enhanced by the presence of the electroconductive material is based on direct interspecies electron transfer <sup>64</sup>. Focusing on nitrogen removal, it has been reported that *Geobacter* bacteria can transfer electrons directly to *Thiobacillus* which in turn may reduce nitrate <sup>65</sup>. Interestingly, both microbial genera are colonizing our electroconductive biofilters although further research is required to find out if these redox syntrophic relationships are the ones after nitrogen removal in our systems.

Environmental Science: Water Research & Technology Accepted Manuscript

425	Conc	lusions		
426	Problems with wastewater treatment in small communities are different that in large cities			
427	owing to the scarcity of economical and technical resources. It is necessary to find solutions			
428	that generate minimum energy cost, simple maintenance and functional robustness. With this			
429	aim, the successful integration of microbial electrochemical technologies into well tested			
430	treat	treatments, such as constructed wetlands, represents a substantial advance since the new		
431	system can be operated a surface inlet load 4-fold higher than the standard systems. Indeed,			
432	our la	our lab scale METland design for treating urban wastewater was able to fulfil the Directive		
433	91/271/EEC and produced water with BOD $_5$ levels as low as 6 mg/L. Our research suggests that			
434	surface area requirements of classical Constructed Wetlands (CW) can be significantly reduced.			
435				
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442				
443	Refe	rences		
444	1	M. R. Ghazy, T. Dockhorn and N. Dichtl, Fifteenth Int. Water Technol. Conf. IWTC-15		
445		2011, Alexandria, Egypt, 2011.		
446	2	H. Liu, R. Ramnarayanan and B. E. Logan, <i>Environ. Sci. Technol.</i> , 2004, <b>38</b> , 2281–5.		
447	3	B. E. Logan, B. Hamelers, R. Rozendal, U. Schröder, J. Keller, S. Freguia, P. Aelterman, W.		
448		Verstraete and K. Rabaey, Environ. Sci. Technol., 2006, 40, 5181–5192.		
449	4	A. Kato Marcus, C. I. Torres and B. E. Rittmann, Biotechnol. Bioeng., 2007, 98, 1171-		
450		1182.		
451	5	B. Wu, C. Feng, L. Huang, Z. Lv, D. Xie and C. Wei, Bioresour. Technol., 2014, 157, 305-		
452		309.		
453	6	K. Rabaey and W. Verstraete, Trends Biotechnol., 2005, 23, 291–298.		
454	7	P. D. Kiely, R. Cusick, D. F. Call, P. a. Selembo, J. M. Regan and B. E. Logan, Bioresour.		
455		Technol., 2011, <b>102</b> , 388–394.		
456	8	Z. Borjas, J. Ortiz, A. Aldaz, J. Feliu and A. Esteve-Núñez, Energies, 2015, 8, 14064-		
457		14077.		
458	9	J. García, D. P. L. Rousseau, J. Morató, E. Lesage, V. Matamoros and J. M. Bayona, Crit.		

459 *Rev. Environ. Sci. Technol.*, 2010, **40**, 561–661.

460	10	J. García, J. Vivar, M. Aromir and R. Mujeriego, Water Res., 2003, <b>37</b> , 2645–2653.
461	11	J. Vymazal and L. Kropfelova, Wastewater Treatment in CWs with HSF, Environmental
462		Pollution 14, 2008.
463	12	E. Tilley, C. Lüthi, A. Morel, C. Zurbrügg and R. Schertenleib, Compendium of Sanitation
464		Systems and Technologies, 2008.
465	13	C. C. Tanner and J. P. Sukias, Water Sci. Technol., 1995, <b>32</b> , 229–239.
466	14	D. P. L. Rousseau, P. a. Vanrolleghem and N. De Pauw, <i>Ecol. Eng.</i> , 2004, <b>23</b> , 151–163.
467	15	US EPA, Manual. Constructed Wetlands Treatment of Municipal Wastewaters, US
468		Environmental Protection Agency, Cincinnati, Ohio, National R., 2000.
469	16	R. Kadlec and S. Wallace, Treatment wetlands, Taylor & Francis Group, Boca Raton, FL,
470		2009.
471	17	C. Corbella, M. Garfí and J. Puigagut, <i>Sci. Total Environ.</i> , 2014, <b>470–471</b> , 754–758.
472	18	J. Villaseñor, P. Capilla, M. a Rodrigo, P. Cañizares and F. J. Fernández, Water Res., 2013,
473		<b>47</b> , 6731–8.
474	19	J. L. Faulwetter, V. Gagnon, C. Sundberg, F. Chazarenc, M. D. Burr, J. Brisson, A. K.
475		Camper and O. R. Stein, <i>Ecol. Eng.</i> , 2009, <b>35</b> , 987–1004.
476	20	Z. Fang, H. Song, N. Cang and X. Li, <i>Biosens. Bioelectron.</i> , 2015, <b>68</b> , 135–41.
477	21	S. Liu, H. Song, S. Wei, F. Yang and X. Li, <i>Bioresour. Technol.</i> , 2014, <b>166</b> , 575–83.
478	22	Y. Zhao, S. Collum, M. Phelan, T. Goodbody, L. Doherty and Y. Hu, Chem. Eng. J., 2013,
479		<b>229</b> , 364–370.
480	23	L. Doherty, Y. Zhao, X. Zhao, Y. Hu, X. Hao, L. Xu and R. Liu, <i>Water Res.</i> , 2015, <b>85</b> , 38–45.
481	24	A. Esteve-Núñez, A. Berná Galiano, A. Reija Maqueda, C. Aragón, A. Aguirre-Sierra, T.
482		Bachetti de Gregoris, R. Esteve-Núñez, B. Barroeta García, J. R. Pidre, J. Fernández
483		Ontivero and J. J. Salas, in MFC4-4th International Microbial Fuel Cell Conference,
484		Cairns, Australia, 2013, pp. 130–131.
485	25	B. E. B. Logan, Microbial fuel cells, 2008.
486	26	E. Ortega, Y. Ferrer, J. J. Salas, C. Aragón and Á. Real, Manual para la implantación de
487		sistemas de depuración en pequeñas poblaciones, Ministerio de Medio Ambiente y
488		Medio Rural y Marino, Madrid, 2010.
489	27	American Public Health Asociation, Standard methods for the examination of water and
490		wastewater, American Public Health Association/American Water Works
491		Association/Water Environmental Federation, Washington DC, 21st edn., 2005.tandard
492		methods for the, American Public Health Association/American Water Works
493		Association/Water Environmental Federation, Washington DC, 21st edn., 2005.
494	28	J. G. Caporaso, J. Kuczynski, J. Stombaugh, K. Bittinger, F. D. Bushman, E. K. Costello, N.

495 Fierer, A. G. Peña, J. K. Goodrich, J. I. Gordon, G. A. Huttley, S. T. Kelley, D. Knights, J. E. 496 Koenig, R. E. Ley, C. A. Lozupone, D. McDonald, B. D. Muegge, M. Pirrung, J. Reeder, J. 497 R. Sevinsky, P. J. Turnbaugh, W. A. Walters, J. Widmann, T. Yatsunenko, J. Zaneveld and 498 R. Knight, Nat. Methods, 2010, 7, 335-6. 499 29 E. Aronesty, Ea-Utils Command. Tools Process. Biol. Seq. Data, 2011. 500 30 R. C. Edgar, *Bioinformatics*, 2010, 26, 2460-1. 501 T. Z. DeSantis, P. Hugenholtz, N. Larsen, M. Rojas, E. L. Brodie, K. Keller, T. Huber, D. 31 502 Dalevi, P. Hu and G. L. Andersen, Appl. Environ. Microbiol., 2006, 72, 5069–72. 503 32 M. N. Price, P. S. Dehal and A. P. Arkin, *PLoS One*, 2010, 5, e9490. 504 N. A. Bokulich, S. Subramanian, J. J. Faith, D. Gevers, J. I. Gordon, R. Knight, D. A. Mills 33 505 and J. G. Caporaso, Nat. Methods, 2013, 10, 57-9. 506 34 J. A. Klappenbach, Nucleic Acids Res., 2001, 29, 181–184. 507 H. Brix, Water Sci. Technol., 1994, 29, 71-78. 35 508 36 European Union, Directive 91/271/EEC of 21 May 1991, concerning urban waste-water 509 treatment. Official Journal L 135, 30/05/1991 P. 0040 - 0052 510 37 A. Caselles-Osorio and J. García, Sci. Total Environ., 2007, 378, 253–262. 511 38 C. C. Tanner, J. P. S. Sukias and M. P. Upsdell, J. Environ. Qual., 1998, 27, 448. 512 39 K. Rabaey, P. Clauwaert, P. Aelterman and K. Verstraete, W., Environ. Sci. Technol., 513 2005, **39**, 8077–8082. 514 40 S. Teiter and Ü. Mander, Ecol. Eng., 2005, 25, 528–541. B. Erable, L. Etcheverry and A. Bergel, *Biofouling*, 2011, 27, 319–326. 515 41 516 42 J. F. Miceli, P. Parameswaran, D.-W. Kang, R. Krajmalnik-Brown and C. I. Torres, Environ. 517 Sci. Technol., 2012, 120828164231002. 518 43 C. Corbella, M. Guivernau, M. Viñas and J. Puigagut, Water Res., 2015, 84, 232-242. 519 44 J.H. Ahn, W.S. Jeong, M.Y. Choi, B.Y. Kim, J. Song and H.Y. Weon, J. Microbiol. 520 Biotechnol., 2014, 24, 1707–1718. L. Lu, D. Xing and Z. J. Ren, *Bioresour. Technol.*, 2015. 521 45 522 S. Bicciato, M. Pandin, G. Didone and C. Di Bello, *Biotechnol Bioeng*, 2003, **81**, 594–606. 46 523 47 M. J. McInerney, L. Rohlin, H. Mouttaki, U. Kim, R. S. Krupp, L. Rios-Hernandez, J. Sieber, 524 C. G. Struchtemeyer, A. Bhattacharyya, J. W. Campbell and R. P. Gunsalus, Proc. Natl. 525 Acad. Sci. U. S. A., 2007, 104, 7600–7605. 526 48 D. Lovley, *Prokaryotes SE - 69*, 2013, 287–308. 527 49 Z. Zhao, Y. Zhang, S. Chen, X. Quan and Q. Yu, Sci. Rep., 2014, 4, 6658. 528 50 A. Parada, D. M. Needham and J. a. Fuhrman, *Environ. Microbiol.*, 2015, n/a-n/a. 529 51 D. R. Bond, D. E. Holmes, L. M. Tender and D. R. Lovley, *Science*, 2002, **295**, 483–485.

530	52	M. Morita, N. S. Malvankar, A. E. Franks, Z. M. Summers, L. Giloteaux, A. E. Rotaru, C.
531		Rotaru and D. R. Lovley, <i>mBio</i> , 2011, <b>2</b> , 5–7.
532	53	AE. Rotaru, P. M. Shrestha, F. Liu, M. Shrestha, D. Shrestha, M. Embree, K. Zengler, C.
533		Wardman, K. P. Nevin and D. R. Lovley, Energy Environ. Sci., 2014, 7, 408–415.
534	54	P. M. Shrestha, N. S. Malvankar, J. J. Werner, A. E. Franks, A. Elena-Rotaru, M. Shrestha,
535		F. Liu, K. P. Nevin, L. T. Angenent and D. R. Lovley, Bioresour. Technol., 2014, 174, 306-
536		10.
537	55	Z. Zhao, Y. Zhang, T. L. Woodard, K. P. Nevin and D. R. Lovley, Bioresour. Technol., 2015,
538		<b>191</b> , 140–5.
539	56	L. De Schamphelaire, A. Cabezas, M. Marzorati, M. W. Friedrich, N. Boon and W.
540		Verstraete, Appl. Environ. Microbiol., 2010, 76, 2002–2008.
541	57	Y. Sun, J. Zuo, L. Cui, Q. Deng and Y. Dang, <i>J. Gen. Appl. Microbiol.</i> , 2010, <b>56</b> , 19–29.
542	58	Z. Wang, T. Lee, B. Lim, C. Choi and J. Park, <i>Biotechnol. Biofuels</i> , 2014, <b>7</b> , 9.
543	59	D. E. Holmes, D. R. Bond and D. R. Lovley, Appl. Environ. Microbiol., 2004, 70, 1234-
544		1237.
545	60	C. Pfeffer, S. Larsen, J. Song, M. Dong, F. Besenbacher, R. L. Meyer, K. U. Kjeldsen, L.
546		Schreiber, Y. A. Gorby, M. Y. El-Naggar, K. M. Leung, A. Schramm, N. Risgaard-Petersen
547		and L. P. Nielsen, <i>Nature</i> , 2012, <b>491</b> , 218–221.
548	61	S. Larsen, L. P. Nielsen and A. Schramm, Environ. Microbiol. Rep., 2015, 7, 175–179.
549	62	D. R. Bond and D. R. Lovley, Appl. Environ. Microbiol., 2005, <b>71</b> , 2186–2189.
550	63	B. Kartal, L. van Niftrik, J. T. Keltjens, H. J. M. Op den Camp and M. S. M. Jetten, Adv.
551		Microb. Physiol., 2012, <b>60</b> , 211–62.
552	64	AE. Rotaru, P. M. Shrestha, F. Liu, B. Markovaite, S. Chen, K. P. Nevin and D. R. Lovley,
553		Appl. Environ. Microbiol., 2014, <b>80</b> , 4599–4605.
554	65	S. Kato, K. Hashimoto and K. Watanabe, Proc. Natl. Acad. Sci., 2012, 109, 10042–10046.
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# **Table of contents**

Water impact: MET biofilters outperform gravel biofilters for wastewater treatment and will reduce the surface for CW, selecting certain genera of bacteria reported to be electroactive.

# **GRAPHICAL ABSTRACT**

