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COMMUNICATION

Photoactivation of the Ni-SI_r state to Ni-SI_a state in [NiFe] hydrogenase: FT-IR study on the light reactivity of the ready Ni-SI_r state and as-isolated enzyme revisited†

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Hulin Tai,^{a,b} Liyang Xu,^a Seiya Inoue,^c Koji Nishikawa,^c Yoshiki Higuchi,^{b,c} and Shun Hirota^{a,b,*}

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The Ni-SI_r state of [NiFe] hydrogenase from *Desulfovibrio vulgaris* Miyazaki F was photoactivated to its Ni-SI_a state by Ar⁺ laser irradiation at 514.5 nm, whereas the Ni-SI state was light induced from a newly identified state, which was less active than any other identified state and existed in the “as-isolated” enzyme.

Introduction

Hydrogenase is a metalloenzyme which catalyzes the reversible H₂ oxidation reaction, H₂ ⇌ 2H⁺ + 2e⁻.¹⁻⁴ According to the active site metal composition, hydrogenases are classified into three types: [NiFe], [FeFe], and [Fe].⁴ [NiFe] hydrogenase from *Desulfovibrio vulgaris* Miyazaki F (*Dv*MF) is a membrane-attached enzyme comprising two subunits, one large and one small.⁵⁻⁷ The large subunit contains the Ni-Fe active site, where the Ni and Fe ions are bridged with two cysteinyl thiolates (Fig. 1). Another two cysteine residues are terminally bound to the Ni ion, whereas one CO and two CN⁻ ligands are coordinated to the Fe ion.⁷⁻¹⁰ The small subunit contains three Fe-S clusters which mediate the electron transfer between the Ni-Fe active site and cytochrome *c*₃.¹¹

Aerobically isolated [NiFe] hydrogenase, herein referred to as “as-isolated”, is a mixture of mainly two paramagnetic Ni-A (Ni³⁺) and Ni-B (Ni²⁺) states with some other EPR-silent states.^{10,12,13} The Ni-B state is readily activated in the presence of H₂ or under electrochemically reducing conditions, while the Ni-A state requires longer time for activation.^{14,15} A bridging hydroxo (OH⁻) ligand between the Ni and Fe ions has been identified for the Ni-B state (Fig.1).^{6,16} For the Ni-A state, the nature of an oxygenic bridging ligand remains contentious,^{6,16-21} however, bridging OH⁻ and cysteine-sulfenate ligands between the Ni and Fe ions have been indicated recently.²² One electron reduction of the Ni-A and Ni-B states produces EPR-silent unready Ni-SU and ready Ni-SI_r states (Ni²⁺), respectively.^{10,16} The Ni-SI_r state is activated into another EPR-silent Ni-SI_a state (Ni²⁺) by protonation at the Ni-Fe active site

through an acid–base equilibrium, where the Ni-SI_r and Ni-SI_a states represent the deprotonated and protonated states, respectively.^{10,12,23} Several mechanisms have been proposed to explain the acid–base equilibrium. In one of them, the bridging OH⁻ ligand is present in the Ni-SI_r state, and a proton is transferred to the OH⁻ ligand, which then leaves the active site as a H₂O molecule.^{2,12,24,25} In the other proposals, a bridging OH⁻ ligand may be present, absent, or replaced by a hydride (H⁻) or a H₂O molecule in the Ni-SI_r state, and the proton is transferred to one of the terminal Ni-coordinating cysteine-thiolate^{26,27} or cysteine-sulfenate²⁸ ligand that acts as a proton accepting base in the Ni-SI_r state. The acid–base equilibrium between the Ni-SI_r and Ni-SI_a states is a common feature among [NiFe] hydrogenases, and thus the Ni-SI_r state has been identified as a key intermediate for the enzyme activation.⁴ Further reduction of the Ni-SI_a state produces a paramagnetic state (Ni-C, Ni³⁺) and a fully reduced EPR-silent state (Ni-R, Ni²⁺), where the Ni-SI_a, Ni-C, and Ni-R states form a catalytic cycle.^{3,4,10}

Light sensitivity of [NiFe] hydrogenase has been reported for various states and utilized to elucidate its catalytic reaction.²⁹⁻³⁶ For example, we have reported photo-conversion of the Ni-C state to the Ni-L and Ni-SI_a states for *Dv*MF [NiFe] hydrogenase, and proposed the Ni-L state as an intermediate between the transition of the Ni-C and Ni-SI_a states.³¹ The Ni-L state has also been shown to be a catalytic intermediate for [NiFe] hydrogenases from *Pyrococcus furiosus* and *Escherichia coli* by chemical potential jump kinetic and direct electrochemical studies.³⁷⁻⁴⁰ We have also simultaneously detected two Ni-L states (Ni-L2 and Ni-L3) by FT-IR, and proposed that Ni-coordinating Cys546 is deprotonated during the conversion from the Ni-L2 to Ni-L3 state.³² Furthermore, it has been proposed that the Ni-SI_r state is light sensitive, reversibly forming an EPR-



Fig. 1 Active site structure of *Dv*MF [NiFe] hydrogenase in the Ni-B state (PDB: 1WUJ). One CO and two CN⁻ ligands are assigned as Fe ligands.^{7,16,23} Carbon, nitrogen, oxygen, sulphur, nickel, and iron atoms are shown in grey, blue, red, yellow, green, and pink spheres, respectively.

^a Graduate School of Materials Science, Nara Institute of Science and Technology, 8916-5 Takayama-cho, Ikoma-shi, Nara 630-0192, Japan
E-mail: hirota@ms.naist.jp; FAX: +81-743-72-6119; Tel: +81-743-72-6110

^b CREST, Japan Science and Technology Agency, 4-1-8 Honcho, Kawaguchi, Saitama 332-0012, Japan

^c Graduate School of Life Science, University of Hyogo, 3-2-1 Koto kamigori-cho, Ako-gun, Hyogo 678-1297, Japan

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silent Ni-SL state (Ni²⁺) at 90–110 K.³⁶ However, in this work, we found that the Ni-SL state is not light induced from the Ni-SL_r state, but rather the Ni-SL_r state is photo-induced to the Ni-SL_a state.

Experimental

Preparation of [NiFe] hydrogenase

[NiFe] hydrogenase was isolated from sulfate reducing bacterium *DvMF*, and purified as described previously.⁵ The concentration of [NiFe] hydrogenase was adjusted with its absorption at 400 nm using its absorption coefficient ($\epsilon = 47 \text{ mM}^{-1}\text{cm}^{-1}$).¹¹

FT-IR measurements

[NiFe] hydrogenase (concentration 1.0–2.0 mM) in 25 mM Tris-HCl buffer (pH 7.4 at 298 K) was degassed with a vacuum line, purged with 1 bar of H₂, and incubated at 310 K for 5.5 h (if not mentioned) to obtain the H₂-activated sample. The sample solution was further degassed with the vacuum line and purged with 1 bar of N₂. The Ni-SL_r state was obtained by partial oxidation of the H₂-activated enzyme with an anaerobic addition of 5 equivalents of phenosafranin (Sigma-Aldrich) using a glove box (YSD-800L, UNICO, Tsukuba). The sample solution was transferred anaerobically into an infrared cell with CaF₂ windows in the glove box. FT-IR spectra were measured before, during, and after light irradiation at 103–238 K with a FT-IR spectrometer (FT-IR 6100V, JASCO, Tokyo) equipped with an MCT detector. A cryostat system (CoolSpeK IR USP-203IR-A, Unisoku, Hirakata) was used to control the temperature of the cell. The light irradiation spectra were measured 5–22 min after light-irradiation was started. Light irradiation of the sample was performed at 514.5 nm with an Ar⁺ laser (Model 2017, Spectra-Physics, Santa Clara). The laser power was adjusted to 0.5–3.3 W/cm² at the sample point. The corresponding buffer spectrum was collected as a reference spectrum and subtracted from the sample spectra. Spectral data were collected at 2-cm⁻¹ resolution and averaged from 1024 scans.

Results and discussion

Observation of the light-induced states at low temperatures

It has been reported that the midpoint potential (E_m) for the redox transition between the Ni-B and Ni-SL (Ni-SL_r and Ni-SL_a) states of *DvMF* [NiFe] hydrogenase is -151 mV at pH 7.4, whereas between the Ni-SL and Ni-C states it is -375 mV .²³ Under N₂ atmosphere, the H₂-activated enzyme contained the Ni-C and Ni-R states for $\sim 70\%$ and $\sim 30\%$, respectively (See S1, ESI[†]), with $\sim 90\%$ of the proximal Fe-S cluster reduced.²⁹ The Ni-SL_r state was obtained by partial oxidation of the H₂-activated enzyme with an anaerobic addition of 5 equivalents of phenosafranin under N₂ atmosphere, since phenosafranin exhibits its redox potential at $E_m = -252 \text{ mV}$ between -375 and -151 mV .

The CO stretching (ν_{CO}) and CN⁻ stretching (ν_{CN}) frequencies of the Fe site are reliable sensors for the changes in the electron density of the Fe ion in [NiFe] hydrogenase.⁴¹ Negative IR bands at 1924, 2056, and 2071 cm⁻¹ and positive bands at 1943, 2077, and 2089 cm⁻¹ were observed in the difference (light-minus-before) FT-IR spectra between the spectra during and before light irradiation by Ar⁺ laser (514.5 nm) for phenosafranin-oxidized [NiFe] hydrogenase at 178–238 K under N₂ atmosphere at pH 8.0 (Fig. 2A). The negative and

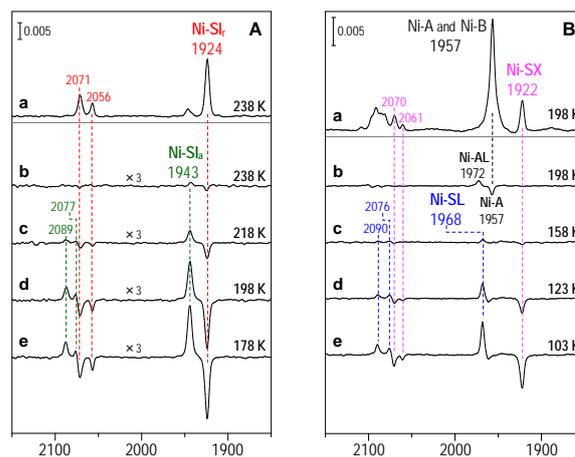


Fig. 2 FT-IR spectra of (A) phenosafranin-oxidized and (B) as-isolated *DvMF* [NiFe] hydrogenase at 178–238 and 103–198 K, respectively, under N₂ atmosphere at pH 8.0. (a) FT-IR spectra before light irradiation and (b–e) light-minus-before difference spectra between the spectra during and before light irradiation are shown. Phenosafranin-oxidized enzyme was obtained by partial oxidation of the H₂-activated enzyme with an anaerobic addition of 5 equivalents of phenosafranin. The difference spectra of phenosafranin-oxidized enzyme are expanded by three. The laser power was adjusted to 2.5 W/cm² at the sample point. The pH value was measured at 274 K.

positive bands were related to the light-sensitive reactant and light-induced product, respectively. The frequency of the negative band at 1924 cm⁻¹ corresponded to that of the ν_{CO} band of the Ni-SL_r state of *DvMF* [NiFe] hydrogenase, whereas 2056 and 2071 cm⁻¹ corresponded well to the frequencies of its conjugated ν_{CN} bands.²³ The positive frequencies at 1943, 2077, and 2089 cm⁻¹ corresponded well to those of the ν_{CO} and two conjugated ν_{CN} bands of the Ni-SL_a state of the H₂-activated enzyme.^{23,31,32} These results showed that the Ni-SL_r state was converted to the Ni-SL_a state by the light irradiation (Fig. 3). Ciaccavava *et al.* have reported that electrochemical activation of an O₂-tolerant [NiFe] hydrogenase from *Aquifex aeolicus* is promoted by UV-vis light irradiation, but the detailed activation mechanism was unspecified.³⁰ Although the Ni-SL_r state has not been observed by electrochemical FT-IR measurements for O₂-tolerant [NiFe] hydrogenases, the Ni-SL_r state may be highly reactive leading to the fast transition of the Ni-B state to Ni-SL_a state by the light irradiation.⁴² Judging from the intensities of the ν_{CO} bands of the Ni-SL_r state in the light-minus-before difference FT-IR spectra, approximately 3% of the Ni-SL_r state was converted to the Ni-SL_a state by the light irradiation at 238 K. The intensities of the ν_{CO} bands of the Ni-SL_r and Ni-SL_a states increased in the light-minus-before difference spectra with a decrease in the temperature, and approximately 34% of the Ni-SL_r state was converted to the Ni-SL_a state at 178 K. Notably, the light-induced conversion of the Ni-SL_r state decreased significantly at pH 9.6 (See S2, ESI[†]), indicating that protonation occurred in the photo-activation process. The reported photochemical reactions in various [NiFe] hydrogenases are usually associated with dissociation of non-protein ligands bound to the metal ions at the Ni–Fe active site.^{13,29–36,43} Stronger laser power was required for photo-activation of the Ni-SL_r state to the Ni-SL_a state compared to photo-activation of the Ni-C state to the Ni-L state associated with dissociation of the bridging H⁻.³¹ Considering these results, we propose that the protonation of the Ni-SL_r state is related to dissociation of the putative bridging OH⁻ ligand as a H₂O molecule by the light irradiation, although the possibility of the Ni-

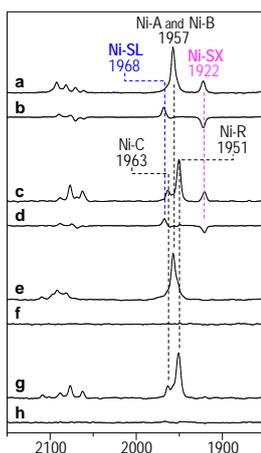


Fig. 5 FT-IR spectra of *DvMF* [NiFe] hydrogenase under N_2 atmosphere at pH 8.0 and 103 K: (a,c,e,g) Spectra before light irradiation and (b,d,f,h) light-minus-after difference spectra. The spectra of the as-isolated enzyme in the (a,b) absence and (c,d) presence of 10 equivalents of dithionite and the spectra of the air-oxidized enzyme (obtained by exposure of the H_2 -activated enzyme to air) in the (e,f) absence and (g,h) presence of 10 equivalents of dithionite are shown. The “after” spectra were measured 30–47 min after light irradiation was stopped. The laser power was adjusted to 2.5 W/cm^2 at the sample point. The pH value was measured at 274 K.

was unreactive toward CO (Fig. 4B), in agreement with the results that an acid–base equilibrium did not exist for the Ni-SX state (See S6B, ESI[†]).

Reaction of the Ni-SX state with dithionite

To gain information on the activation of the Ni-SX state, light-minus-after difference FT-IR spectra were measured for as-isolated and dithionite-reduced *DvMF* [NiFe] hydrogenase at 103 K (Fig. 5). The photo-conversion of the Ni-SX state to the Ni-SL state was successfully observed in the light-minus-after difference spectra, because the Ni-SL state converted back to the Ni-SX state (See S5, ESI[†]) but the light-induced Ni-SL_a and Ni-L states were trapped at 103 K when the light irradiation was stopped (See S7, ESI[†]).³¹ By assuming equal absorption coefficients for the ν_{CO} bands, approximately 16% was in the Ni-SX state for the as-isolated enzyme (Fig. 5, curve a), and almost (~95%) all the enzymes in the Ni-SX state converted to the Ni-SL state by the light irradiation at 103K (Fig. 5, curve b). However, approximately 10% of the enzyme was still in the Ni-SX state after reduction with dithionite (Fig. 5, curves c and d), indicating that the Ni-SX state was very inactive. For [NiFe] hydrogenases from the sulphur-metabolizing bacterium *Allochroamatium vinosum* and *Desulfovibrio fructosorans* (*Df*), an inactive state (Ni-‘S_{ox}’) similar to the Ni-SX state has been reported.^{12,22} The ν_{CO} and ν_{CN} frequencies of the Ni-‘S_{ox}’ state for *Df* [NiFe] hydrogenase (ν_{CO} : 1911, ν_{CN} : 2059 and 2068 cm^{-1}) were 1–5 cm^{-1} shifted from those of its Ni-SL_a state (ν_{CO} : 1913 cm^{-1} , ν_{CN} : 2054 and 2069 cm^{-1}),^{22,46} which was very similar to those of the Ni-SX state detected in the present study (Fig. 2B). The Ni-‘S_{ox}’ state has been proposed by X-ray crystallographic analysis to possess a cysteine-persulfide terminal Ni-coordinating ligand at the active site.²² The required reduction of the persulfide bond may explain the observed slow activation of the Ni-SX state.

As-isolated *DvMF* [NiFe] hydrogenase contained the Ni-SX state exhibiting a ν_{CO} band at 1922 cm^{-1} and two ν_{CN} bands at 2061 and 2070 cm^{-1} .^{13,36} The intensities of these bands decreased

exponentially with a time constant of ~50 min by incubation under H_2 atmosphere at 310 K (See Figs. S8 and S9, ESI[†]), revealing that the Ni-SX state was activated very slowly under H_2 atmosphere. However, the Ni-SX state was not observed for the air-oxidized enzyme, where the H_2 -activated enzyme was exposed to air (Fig. 5, curves e and f). Additionally, the Ni-SX state was not observed after further dithionite reduction of the air-oxidized enzyme (Fig. 5, curves g and h). These results reveal that although the as-isolated enzyme contained the Ni-SX state, the Ni-SX state was not formed during the generation or activation of the Ni-A and Ni-B states in vitro.

Conclusions

We have shown for the first time that the ready Ni-SL_a state of *DvMF* [NiFe] hydrogenase is converted to the active Ni-SL_a state by laser light irradiation at 514.5 nm (Fig. 3). From the pH-dependent light-reactivity of the Ni-SL_a state, we propose that the bridging OH-ligand dissociates as a H_2O molecule from the Ni–Fe active site by light irradiation at low pH. We have identified a light-sensitive Ni-SX state (ν_{CO} , 1922 cm^{-1} ; ν_{CN} , 2061 and 2070 cm^{-1}), which was photo-converted to the Ni-SL state. A certain amount of the enzyme was still in the Ni-SX state after treatment of the as-isolated enzyme with dithionite, although the enzyme was activated slowly by H_2 , revealing that the Ni-SX state was highly inactive. These findings provide new insights into the activation mechanism of [NiFe] hydrogenase.

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References

- P. M. Vignais and B. Billoud, *Chem. Rev.*, 2007, **107**, 4206–4272.
- J. C. Fontecilla-Camps, A. Volbeda, C. Cavazza and Y. Nicolet, *Chem. Rev.*, 2007, **107**, 4273–4303.
- A. L. De Lacey, V. M. Fernández, M. Rousset and R. Cammack, *Chem. Rev.*, 2007, **107**, 4304–4330.
- W. Lubitz, H. Ogata, O. Rüdiger and E. Reijerse, *Chem. Rev.*, 2014, **114**, 4081–4148.
- Y. Higuchi, H. Ogata, K. Miki, N. Yasuoka and T. Yagi, *Structure*, 1999, **7**, 549–556.
- H. Ogata, S. Hirota, A. Nakahara, H. Komori, N. Shibata, T. Kato, K. Kano and Y. Higuchi, *Structure*, 2005, **13**, 1635–1642.
- H. Ogata, K. Nishikawa and W. Lubitz, *Nature*, 2015, **520**, 571–574.
- R. P. Happe, W. Roseboom, A. J. Pierik, S. P. J. Albracht and K. A. Bagley, *Nature*, 1997, **385**, 126.
- A. J. Pierik, W. Roseboom, R. P. Happe, K. A. Bagley and S. P. Albracht, *J. Biol. Chem.*, 1999, **274**, 3331–3337.
- A. L. de Lacey, E. C. Hatchikian, A. Volbeda, M. Frey, J. C. Fontecilla-Camps and V. M. Fernandez, *J. Am. Chem. Soc.*, 1997, **119**, 7181–7189.
- T. Yagi, K. Kimura, H. Daidoji, F. Sakai and S. Tamura, *J. Biochem.*, 1976, **79**, 661–671.

12. B. Bleijlevens, F. A. van Broekhuizen, A. L. De Lacey, W. Roseboom, V. M. Fernandez and S. P. J. Albracht, *J. Biol. Inorg. Chem.*, 2004, **9**, 743-752.
13. M. E. Pandelia, H. Ogata, L. J. Currell, M. Flores and W. Lubitz, *Biochim. Biophys. Acta*, 2010, **1797**, 304-313.
14. V. M. Fernandez, E. C. Hatchikian and R. Cammack, *Biochim. Biophys. Acta*, 1985, **832**, 69-79.
15. S. E. Lamle, S. P. Albracht and F. A. Armstrong, *J. Am. Chem. Soc.*, 2004, **126**, 14899-14909.
16. A. Volbeda, L. Martin, C. Cavazza, M. Matho, B. W. Faber, W. Roseboom, S. P. J. Albracht, E. Garcin, M. Rousset and J. C. Fontecilla-Camps, *J. Biol. Inorg. Chem.*, 2005, **10**, 239-249.
17. H. Ogata, P. Kellers and W. Lubitz, *J. Mol. Biol.*, 2010, **402**, 428-444.
18. P. Amara, A. Volbeda, J. C. Fontecilla-Camps and M. J. Field, *J. Am. Chem. Soc.*, 1999, **121**, 4468-4477.
19. M. Stein, E. van Lenthe, E. J. Baerends and W. Lubitz, *J. Am. Chem. Soc.*, 2001, **123**, 5839-5840.
20. M. van Gastel, M. Stein, M. Brecht, O. Schröder, F. Lenzian, R. Bittl, H. Ogata, Y. Higuchi and W. Lubitz, *J. Biol. Inorg. Chem.*, 2006, **11**, 41-51.
21. J. L. Barilone, H. Ogata, W. Lubitz and M. van Gastel, *Phys. Chem. Chem. Phys.*, 2015, **17**, 16204-16212.
22. A. Volbeda, L. Martin, E. Barbier, O. Gutierrez-Sanz, A. L. De Lacey, P. P. Liebgott, S. Dementin, M. Rousset and J. Fontecilla-Camps, *J. Biol. Inorg. Chem.*, 2015, **20**, 11-22.
23. C. Fichtner, C. Laurich, E. Bothe and W. Lubitz, *Biochemistry*, 2006, **45**, 9706-9716.
24. S. Kurkin, S. J. George, R. N. F. Thorneley and S. P. J. Albracht, *Biochemistry*, 2004, **43**, 6820-6831.
25. S. J. George, S. Kurkin, R. N. F. Thorneley and S. P. J. Albracht, *Biochemistry*, 2004, **43**, 6808-6819.
26. A. L. De Lacey, A. Pardo, V. M. Fernandez, S. Dementin, G. Adryanczyk-Perrier, E. C. Hatchikian and M. Rousset, *J. Biol. Inorg. Chem.*, 2004, **9**, 636-642.
27. T. Krämer, M. Kamp, W. Lubitz, M. van Gastel and F. Neese, *ChemBioChem*, 2013, **14**, 1898-1905.
28. A. Volbeda, L. Martin, P. P. Liebgott, A. L. De Lacey and J. C. Fontecilla-Camps, *Metallomics*, 2015, **7**, 710-718.
29. H. Osuka, Y. Shomura, H. Komori, N. Shibata, S. Nagao, Y. Higuchi and S. Hirota, *Biochem. Biophys. Res. Commun.*, 2013, **430**, 284-288.
30. A. Ciaccafava, C. Hamon, P. Infossi, V. Marchi, M. T. Giudici-Orticoni and E. Lojou, *Phys. Chem. Chem. Phys.*, 2013, **15**, 16463-16467.
31. H. Tai, K. Nishikawa, M. Suzuki, Y. Higuchi and S. Hirota, *Angew. Chem. Int. Ed.*, 2014, **53**, 13817-13820.
32. H. Tai, K. Nishikawa, S. Inoue, Y. Higuchi and S. Hirota, *J. Phys. Chem. B*, 2015, **119**, 13668-13674.
33. P. Kellers, M.-E. Pandelia, L. J. Currell, H. Gerner and W. Lubitz, *Phys. Chem. Chem. Phys.*, 2009, **11**, 8680-8683.
34. E. Siebert, M. Horch, Y. Rippers, J. Fritsch, S. Frielingsdorf, O. Lenz, F. Velazquez Escobar, F. Siebert, L. Paasche, U. Kuhlmann, F. Lenzian, M.-A. Mroginski, I. Zebger and P. Hildebrandt, *Angew. Chem. Int. Ed.*, 2013, **52**, 5162-5165.
35. M. Horch, J. Schoknecht, M. A. Mroginski, O. Lenz, P. Hildebrandt and I. Zebger, *J. Am. Chem. Soc.*, 2014, **136**, 9870-9873.
36. M.-E. Pandelia, H. Ogata, L. J. Currell, M. Flores and W. Lubitz, *J. Biol. Inorg. Chem.*, 2009, **14**, 1227-1241.
37. R. Hidalgo, P. A. Ash, A. J. Healy and K. A. Vincent, *Angew. Chem. Int. Ed.*, 2015, **54**, 7110-7113.
38. B. J. Murphy, R. Hidalgo, M. M. Roessler, R. M. Evans, P. A. Ash, W. K. Myers, K. A. Vincent and F. A. Armstrong, *J. Am. Chem. Soc.*, 2015, **137**, 8484-8489.
39. B. L. Greene, C. H. Wu, P. M. McTernan, M. W. Adams and R. B. Dyer, *J. Am. Chem. Soc.*, 2015, **137**, 4558-4566.
40. B. L. Greene, C. H. Wu, G. E. Vansuch, M. W. Adams and R. B. Dyer, *Biochemistry*, 2016, **55**, 1813-1825.
41. M. Y. Darensbourg, E. J. Lyon and J. J. Smee, *Coord. Chem. Rev.*, 2000, **206**, 533-561.
42. M. E. Pandelia, V. Fourmond, P. Tron-Infossi, E. Lojou, P. Bertrand, C. Leger, M. T. Giudici-Orticoni and W. Lubitz, *J. Am. Chem. Soc.*, 2010, **132**, 6991-7004.
43. H. Ogata, Y. Mizoguchi, N. Mizuno, K. Miki, S.-I. Adachi, N. Yasuoka, T. Yagi, O. Yamauchi, S. Hirota and Y. Higuchi, *J. Am. Chem. Soc.*, 2002, **124**, 11628-11635.
44. J. W. Vanderzwaan, S. P. J. Albracht, R. D. Fontijn and Y. B. M. Roelofs, *Biochim. Biophys. Acta*, 1986, **872**, 208-215.
45. K. A. Bagley, C. J. Van Garderen, M. Chen, E. C. Duin, S. P. Albracht and W. H. Woodruff, *Biochemistry*, 1994, **33**, 9229-9236.
46. A. L. De Lacey, C. Stadler, V. M. Fernandez, E. C. Hatchikian, H. J. Fan, S. Li and M. B. Hall, *J. Biol. Inorg. Chem.*, 2002, **7**, 318-326.