

Analytical Methods

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4 1 **Simultaneous Determination of Tetrabutyl Ammonium and**
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6 2 **Tetrabutyl Phosphonium in Environmental Water Sample by Solid**
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9 3 **Phase Extraction and Ion Chromatography**
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14 5 Feng Liu, Hong Yu *

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16 6 College of Chemistry and Chemical Engineering, Harbin Normal University, Harbin
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18 7 150025, China; *E-mail address*: yuhonghsd@126.com; Fax: 86-451-88060571.
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23 9 **Abstract** Ion Chromatography method by solid-phase extraction which was used
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26 10 for simultaneous determination of tetrabutyl ammonium ([TBA]⁺) and tetrabutyl
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28 11 phosphonium ([TBP]⁺) in environmental water sample was developed. The samples
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31 12 were first concentrated and purified through a SCX solid-phase extraction column,
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34 13 and eluted with 0.02 mol L⁻¹ hydrochloric acid - 98% methanol (v/v), then analyzed
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37 14 by a carboxylic acid base cation exchange column. The optimized chromatographic
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40 15 conditions were 3 mmol L⁻¹ methanesulfonic acid-20% acetonitrile as mobile phase,
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43 16 flow rate of 1.2 mL min⁻¹ and column temperature of room temperature. Under these
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46 17 conditions, [TBA]⁺ and [TBP]⁺ achieved baseline separation. The retention times of
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49 18 the two cations were 11 min and 14 min, the resolution between peaks was 2.059. The
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52 19 recovery of solid-phase extraction reached 97.1%, the linear regression equations
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55 20 were $y = 0.0142 x - 0.0132$ and $y = 0.0195 x - 0.0265$, the correlation
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58 21 coefficients were $r = 0.9996$ and $r = 0.9997$, the detection limits (S/N = 3) were 0.502
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61 22 mg/L and 0.448 mg/L, the relative standard deviations ($n = 5$) of retention times of
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64 23 [TBA]⁺ and [TBP]⁺ were 1.22%, 1.23% and of peak areas were 0.24%, 0.25%,

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4 24 respectively. Recoveries were 94.8% to 103.2%. The method is accurate, rapid,
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7 25 sensitive and efficient to meet the requirements for quantitative analysis.
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10 26
11 27 **Keywords** tetrabutyl ammonium, tetrabutyl phosphonium, solid-phase extraction,
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13 28 ion chromatography, conductivity detection
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16 17 18 30 **1. Introduction** 19 20

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24 32 Quaternary ammonium salt disinfectant and quaternary phosphonium salt
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26 33 disinfectant are all used as cationic surfactant.¹⁻⁴ Nitrogen and phosphorus atoms
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28 34 linked to the alkyl formation of the cation is an effective part of sterilization and
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30 35 disinfection effects.⁵⁻⁸ Based on many researches of the quaternary ammonium salt
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32 36 and quaternary phosphonium salt physical and chemical properties, their bactericidal
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34 37 mechanism, influence factor, advantages of the low side effects, have been widely
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36 38 acknowledged, thus quaternary ammonium salt disinfectant and quaternary
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38 39 phosphonium salt disinfectant have been widely used in daily routine, healthcare, food
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40 40 and many other fields.⁹⁻¹² Additionally, it was found that with long chain alkyl
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42 41 quaternary ammonium salt and quaternary phosphonium salt groups, the antimicrobial
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44 42 properties of them were enhanced. There were reports of quaternary ammonium salts
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46 43 detection by ion chromatography method,¹³⁻¹⁶ mass spectrometry method¹⁷⁻²⁰.
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48 44 Analysis methods of quaternary phosphonium salt are rarely reported, only the
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50 45 infrared spectrum and nuclear magnetic resonance (NMR) were studied, it might be
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52 46 due to the structure and chemical properties of the quaternary phosphonium salt is
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54 47 similar to quaternary ammonium salt. However, the report of long chain quaternary
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3 48 ammonium salt and quaternary phosphonium salt analysis was scarce. Refer to the
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5 49 detection method of quaternary ammonium salt, quaternary phosphonium salt was
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8 50 studied simultaneously.
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11 51 Solid phase extraction is a kind of sample pretreatment technology developed in
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13 52 recent years. It is mainly used for sample separation, purification and concentration, it
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15 53 allows the separation of interfering component, thus provides the more effective
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17 54 analysis and reduces the sample pretreatment process. It was widely used in medicine,
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19 55 food, environment, commodity inspection, chemical and other fields.²¹⁻²⁵ For example,
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21 56 quaternary ammonium herbicides (e.g. paraquat) and its structural modifications used
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23 57 solid phase extraction to extract and enrich the sample.²⁶
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32 59 This study aims to develop an ion chromatography method by solid-phase
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34 60 extraction which was used for the simultaneous determination of tetrabutyl
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36 61 ammonium and tetrabutyl phosphonium. This method could be applied to determine
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38 62 quaternary ammonium salt and quaternary phosphonium salt in antiseptic and
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40 63 disinfectant agent to provide the corresponding reference.
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45 64 46 65 **2. Experimental** 47 48 49

50 66 51 67 **2.1. Instrumentation and Reagents** 52 53 54

55 68 56 69 **2.1.1. Instrumentation** 57 58 59

60 71 Sample preparation was carried out on an ASE-24 solid-phase extraction apparatus,

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4 72 which was equipped with a Model AP-9950 oil-free vacuum pump (Tianjin Automatic
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6 73 Science Instrument Co. Ltd). The whole experimentation was carried out on a 886
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8 74 Basic plus ion chromatograph, which was equipped with a Model 863 Compact
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10 75 Autosampler. The chromatographic system control and data acquisition were
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12 76 performed using the MagIC.net 3.1 workstation (Metrohm, Switzerland). Model
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14 77 BSA124S electronic analytical balance (Sartorius Scientific Instrument Corporation,
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16 78 China) was used. Model Milli-Q Reference water purification system (Millipore, USA)
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18 79 was used. To check the experiment pH, a Model PHSF-3F pH meter (Shanghai
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20 80 Precision Scientific Instrument Corporation, China) was used. Model EMS-9A
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22 81 Heating Magnetic Stirrer (Tianjin Honour Instrument Co. Ltd) was used to
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24 82 homogenize solutions. The SPE procedure used the UF-SCX solid phase extraction
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26 83 column (200 mg/ 3 mL, Dalian Zhongpu Technology Co. Ltd) . A Model
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28 84 DHG-9070A Electro-thermostatic blast oven (Shanghai Yiheng Instrument Co. Ltd)
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30 85 was used. Other instruments are conformed to the requirements of the
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32 86 chromatographic analysis.
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41 88 **2.1.2. Reagents**

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47 90 Tetrabutyl ammonium bromide, tetrabutyl phosphonium bromide and tetraethyl
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49 91 ammonium bromide (purity 99%) were purchased from Shanghai Cheng Jie Chemical
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51 92 Co. Ltd (Shanghai, China). Methanol and acetonitrile (chromatographically pure)
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53 93 were obtained from Sigma-Aldrich Co. (USA). Methanesulfonic acid, quinolinic acid
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55 94 and orthoboric acid (analytically pure) were purchased from Tianjin Guangfu Fine
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60 95 Chemical Research Institute (Tianjin, China), Tartaric acid, hydrochloric acid,

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4 96 phosphoric acid and sodium dihydrogen phosphate (analytically pure) were purchased
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7 97 from Beijing Bailingwei Technology Co. Ltd (Beijing, China).
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10 11 99 **2.2. Preparation of solutions**

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16 101 All the solutions in experiment were prepared with 18.2 MΩ cm deionized water.
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18 102 Stock standard solutions of concentration 1 g L⁻¹ were prepared. The solutions were
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20 103 diluted to the required concentration of this experiment, and then filtered using a 0.22
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22 104 μm membrane filter before injection. Buffer solutions of 10 mmol L⁻¹ phosphoric
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24 105 acid-sodium dihydrogen phosphate were prepared with 18.2 MΩ cm deionized water
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27 106 in 250 mL volumetric flask. The mobile phases were filtered through a 0.22 μm filter,
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30 107 and then degassed for 15 min prior to use.
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34 35 109 **2.3. Environmental water sample**

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39 111 The water sample was taken from river in Harbin for optimization and
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41 112 quantitative analysis. The pH of environmental water sample was 7.39-7.42. The
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43 113 water sample was taken from the upstream, which is the end of chemical plant and
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45 114 households sewage, therefore the freshwater sample contains many contaminants. The
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47 115 freshwater was passed through a quantitative filter paper of 9 cm diameter, and finally
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49 116 stored in the dark at ambient temperature.
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53 54 55 56 118 **2.4. Sample pretreatment**

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120 Prior using, the SCX solid-phase extraction column were conditioned with 5 mL

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4 121 of methanol followed by 5 mL of deionized water. After decompression, the loading
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7 122 was controlled in drain velocity of 1 mL/min, then 10 mL of 20% acetonitrile
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10 123 (prepared with deionized water) was added to buffering. After the buffering process
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13 124 was done, dry vacuum was applied for 10 min to remove the residual moisture.
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15 125 Sample was loaded into column, and the 5 mL of 0.24 mol L⁻¹ hydrochloric acid -
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18 126 98% methanol (v/v) was used to elute all the neutral and acidic impurities from
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21 127 sample. The chromatographic analysis was conducted on 5 mL of collected sample.
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23 128 The final eluent composition is discussed in the “Results and discussion” section.
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26 129 The solid-phase extraction protocol applied is shown in Fig. 1.
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32 131 **2.5. Chromatographic conditions**

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35 133 The separations were performed on a carboxylic acid base cation exchange column
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37 134 (Shodex IC YK-421, 4.6 mm I.D. × 125 mm, Showa Denko, Japan). The mobile
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40 135 phase was 3 mmol L⁻¹ methanesulfonic acid-20% acetonitrile (v/v, pH=2.51). The flow
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43 136 rate was 1.2 mL min⁻¹. Column temperature was room temperature. The injection
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46 137 volume was 20 μL. The detection mode was direct conductivity detection. The
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49 138 chromatographic system control and data acquisition were performed using the
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52 139 MagIC.net 3.1 workstation (Metrohm, Switzerland).
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141 **3. Results and discussion**

143 **3.1. Selection of solid-phase extraction conditions**

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3 145 **3.1.1. Selection of eluent**
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8 147 The elution capacity of eluent on solid-phase extraction was investigated using
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10 148 acetonitrile, 3 mmol L⁻¹ methanesulfonic acid - 20% acetonitrile (v/v), 10 mmol L⁻¹
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12 149 phosphoric acid - sodium dihydrogen phosphate and 0.24 mol L⁻¹ hydrochloric acid -
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14 150 98% methanol (v/v), respectively. Using 10 mmol L⁻¹ phosphoric acid-sodium
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16 151 dihydrogen phosphate and 0.24 mol L⁻¹ hydrochloric acid - 98% methanol (v/v) as
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18 152 eluents, the existence of analytes were able to be detected. The SCX solid-phase
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20 153 extraction has a strong acid property, it was because of the column consists of a
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22 154 polymerically bonded benzene sulfonic acid functional group with a proton
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24 155 counterion on a silica support. The solid phase, thus, be able to absorb or exchange
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26 156 the cations in solution, under all pH range, as long as the pH of solution also be able
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28 157 to guarantee that the analytes are in charged state. The SCX can be used to separate
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30 158 compounds of strong cation and weak cation. For the analysis of cationic compounds,
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32 159 pH of solution should be two units less than the pK_a of analytes. This is to ensure that
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34 160 the analytes are in electric charge, because when the elution was conducted in a pH
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36 161 two units larger than the target objects, the analytes could be joined with another
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38 162 cation instead. Hydrochloric acid - methanol was chosen as eluent and the
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40 163 concentration of HCl was further studied. This optimization study was conducted by
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42 164 varying HCl concentrations of 0.36, 0.24, 0.12, 0.09, 0.07, 0.05 and 0.02 mol L⁻¹.
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44 165 The result showed that 0.02 mol L⁻¹ hydrochloric acid - 98% methanol (v/v) as eluent
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46 166 recovery is the highest. The results of the study are shown in Table. 1.
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58 168 **3.1.2. Selection of buffer solutions**
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3 170 The sample washing capacity of buffer on solid-phase extraction was investigated
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5 171 using acetonitrile - water, 10 mmol L⁻¹ phosphoric acid - sodium dihydrogen
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7 172 phosphate and hydrochloric acid - methanol, respectively. The collected buffer
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9 173 solution was investigated by chromatographic analysis. It was found that the buffer
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11 174 consists of 10 mmol L⁻¹ of phosphoric acid - sodium dihydrogen phosphate and
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13 175 hydrochloric acid – methanol have washing effect. They destroyed the retention of
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15 176 sample, thus the retained samples were washed off. The acetonitrile – water had
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17 177 shown desirable effect and was capable of maintaining the equilibrium of samples to
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19 178 the stationary phase. To optimize the washing eluent, the variations of acetonitrile
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21 179 concentrations studied were 100, 80, 60, 40, 20%, together with water were studied.
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23 180 By considering that the enrichments of the environmental water sample are
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25 181 maintained on the solid phase extraction column, 20% acetonitrile - water as rinsing
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27 182 environmental water sample eluent was selected.
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184 **3.1.3. Breakthrough volume**

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186 In order to obtain a perfect balance between the maximum recoveries of the target
187 compound and the minimum consumption of loading eluent, various volumes of
188 loading eluent in the range of 5.0–35.0 mL were investigated. When the volume was
189 greater than 10 mL, the environmental water sample was detected in the loading
190 eluent. This indicated that the volume was beyond the solid phase extraction capacity,
191 which is in line with the standard calculation of adsorbent capacity. In the process of
192 enrichment and samples purification experiments, the weight of the sample was
193 controlled to be less than 5 mg, to reduce the effect of exceeding weight to recovery.

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3.2. Selection of chromatographic conditions

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3.2.1. Selection of eluent

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199 Using 5 mmol L⁻¹ tartaric acid - 50% acetonitrile (v/v) as mobile phase, the
200 determination and separation of [TBA]⁺ and [TBP]⁺ were investigated. The results
201 were peak shape was excellent and peaks appeared in 6 min, however it cannot
202 achieve the baseline separation. This result indicated that tartaric acid's elution ability
203 to these two kinds of cations was equal, as shown in Fig. 2. Another investigation was
204 conducted by using 3 mmol L⁻¹ methane sulfonic acid - 30% acetonitrile (v/v) as
205 eluent, the result showed that [TBA]⁺ and [TBP]⁺ were overlapping, the resolution
206 factor was less than 1. Thus, it was needed to further choose the different ratio of
207 methanesulfonic acid – acetonitrile, as shown in Fig. 2.

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3.2.2. Selection of methanesulfonic acid concentration

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211 Using methanesulfonic acid-30% acetonitrile as eluent, the effect of methanesulfonic
212 acid concentrations of 1, 2, 3, 4 and 5 mmol L⁻¹ were investigated. The
213 chromatographic analysis found that with the increase of methanesulfonic acid
214 concentration, the retention times were shortened. This complied with the principle of
215 ion exchange chromatography, however the resolution of two peaks was not obvious
216 improved. The value greater than 4 mmol L⁻¹ methanesulfonic acid (pH < 2.1) is
217 approaching the limit of the pH that column allows, affecting the system conductivity
218 and baseline instability. While if the concentration of methanesulfonic acid is less

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4 219 than 2 mmol L⁻¹, the retention time was at about 16 min as shown in Fig. 3. Thus,
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6 220 the 3 mmol L⁻¹ methanesulfonic acid - 30% acetonitrile eluent was chosen for further
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8 221 investigation.
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12 13 223 **3.2.3. Selection of acetonitrile concentration**

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15 224 Using 3 mmol L⁻¹ methanesulfonic acid and acetonitrile as the eluent, the effect of
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17 225 acetonitrile concentrations of 10, 20, 30, 40 and 50% were investigated. The
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19 226 chromatographic analysis found that as the concentration of acetonitrile was reduced,
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21 227 the retention time of both ions was increased. The reason is the increasing of organic
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23 228 modifier acetonitrile had enhanced the interaction between hydrophobic ions and the
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25 229 surface of the stationary phase. Result showed that there is an improved in separating
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27 230 degree by the decreasing the concentration of organic modifier, as shown in Fig. 4.
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29 231 However, when the concentration of acetonitrile was 10%, the tailing phenomenon
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31 232 was obvious. The concentration of 20% acetonitrile was chosen because the degree of
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33 233 separation reached 2.059 and the peak shape was excellent. The optimum
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35 234 compositions of the final eluent are for 3 mmol L⁻¹ methanesulfonic acid - 20%
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37 235 acetonitrile.
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45 46 237 **3.2.4. Selection of flow rate**

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51 239 Using 3 mmol L⁻¹ methanesulfonic acid - 20% acetonitrile as eluent, at room
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53 240 temperature, the effects of flow rate of 0.4, 0.6, 0.8, 1.0, and 1.2 mL/min were
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55 241 investigated. The influence of flow rates to the degree of separation, column pressure
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57 242 and column efficiency were studied. The experimental results showed that with the
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59 243 increase of flow rate, both ions' retention times were shortened, without impacting
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3 244 the degree separation, as shown in Fig. 5. The flow rate of 1.2 mL/min was chosen as
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5 245 the optimum flow rate used in the entire experiment, because it permitted the
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8 246 relatively short retention time and the stable baseline.
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11 248 **3.3. Quantitative parameter**

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17 250 Through the above discussions, the optimal chromatographic conditions for
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19 251 determination of [TBA]⁺ and [TBP]⁺ were as follow: the samples were first enriched
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21 252 and purified through a SCX solid-phase extraction column, and eluted with 0.02 mol
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23 253 L⁻¹ hydrochloric acid - 98% methanol (v/v), then analyzed by a carboxylic acid base
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25 254 cation exchange column, 3 mmol L⁻¹ methanesulfonic acid-20% acetonitrile(v/v,
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27 255 pH=2.51) as mobile phase, flow rate of 1.2 mL min⁻¹ and column temperature of
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29 256 room temperature was applied. Under these conditions, the chromatogram of two
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31 257 cations is shown in Fig. 6.
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37 258 The calibration curve, detection limits and precision of the method were
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39 259 determined by analyzing a series of standard solutions of [TBA]⁺ and [TBP]⁺ under
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41 260 the selected chromatographic conditions. Linear regression equations were obtained
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43 261 from the relationship between peak area (integral value) and ionic concentration (mg
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45 262 L⁻¹). Detection limits were calculated by a triple signal-to-noise ratio (S/N = 3), and
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47 263 noise value of experiment was 0.0029 μS/cm. Relative standard deviations (RSD)
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49 264 were obtained by five repeated measurement of a standard mixture solution of [TBA]
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51 265 ⁺ and [TBP]⁺ under the optimal chromatographic conditions. The results are shown in
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53 266 **Table 2**. It showed that the reproducibility and linearity of the method meet the
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60 267 requirements of quantitative analysis.

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3.4. Analysis of sample

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10 271 This method was applied to the simultaneous determination of quaternary ammonium
11 salts and quaternary phosphonium salts in environmental water samples. The samples
12 272 were left standing overnight, and then filtered through filter paper. The 5 mL sample
13 273 solutions were first enriched and purified through a SCX solid-phase extraction
14 274 column, and filtered through a 0.22 μm membrane filter. The final solutions were
15 275 used for the determination of $[\text{TBA}]^+$ and $[\text{TBP}]^+$ under optimized conditions.
16 276 Recoveries were determined by the standard addition method. Analytical results and
17 277 recoveries of $[\text{TBA}]^+$ and $[\text{TBP}]^+$ in the environmental water samples are listed in
18 278 **Table 3**. The data in Table 3 were the average values ($n = 5$), and the RSD were less
19 279 than 3%. Data from Table 3 proved that the method are accurate and reproducible,
20 280 and it meets the requirements of quantitative analysis of $[\text{TBA}]^+$ and $[\text{TBP}]^+$.
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4. Conclusion

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43 285 This paper reported a novel approach using solid phase extraction to concentrate and
44 286 purify tetrabutyl ammonium and tetrabutyl phosphonium in environmental water. The
45 287 method was using methanesulfonic acid and acetonitrile as the eluent and direct
46 288 conductivity for determination of tetrabutyl ammonium and tetrabutyl phosphonium
47 289 in environmental water. The study showed that in solid phase extraction, the change
48 290 of hydrochloric acid concentration could greatly affect the sample recovery. In ion
49 291 chromatography detection, reducing the content of acetonitrile in mobile phase had
50 292 significantly improved the tetrabutyl ammonium and tetrabutyl phosphonium

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3 293 separation. The appropriate adjustment of methanesulfonic acid and acetonitrile
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5 294 concentration was able to improve the retention time. This method has been
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8 295 successfully applied to determine the tetrabutyl ammonium and tetrabutyl
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10 296 phosphonium in environmental water. By using laboratory generally-available ion
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12 297 chromatography and conductivity detection, which has high practical value, the
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15 298 method was accurate, reliable, and relatively simple to be applied.
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301
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305 **References**

- 306
307 1. H. J. Xu, B. N. Liu, P Kang and X. Y. Bao. *J. Surfactants. Deterg.*, 2015, 18,
308 297.
309 2. S. Mondal, S. Das and S. Ghosh. *J. Surfactants. Deterg.*, 2015, 18, 471.
310 3. M. S. Kiakhani and K. Gharanjig. *J. Surfactants. Deterg.*, 2015, 18, 47.
311 4. B. Liu, W. Guo and J. F. Song. *Chin. J. Anal. Chem.*, 2002, 30, 1210.
312 5. E. R. Kenawy, F. I. Abdel-Hay1, A. R. El-Shanshoury and M. H. El-Newehy1. *J.*
313 *Polym. Sci., Polym. Chem.*, 2002, 40, 2384.
314 6. A. Kanazawa, T. Ikeda and T. Endo. *J. Appl. Polym. Sci.*, 1994, 53, 1237.
315 7. A. Kanazawa, T. Ikeda and T. Endo. *Antimicrob. Agents. Chemother.*, 1994, 38,
316 945.
317 8. A. Kanazawa, T. Ikeda and T. Endo. *J. Polym. Sci., Polym. Chem.*, 1993, 31,

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2
3 318 335.
4
5
6 319 9. A. Popa, G. Iliu, S. Iliescu, G. Dehelean, A. Pascariu, A. Bora and C. M.
7
8 320 Davidescu. *Mol. Cryst. Liq. Cryst. Sci. Technol., Sect. A*, 2004, 418, 923.
9
10 321 10. Y. Xue, H. Xiao and Y. Zhang. *Int. J. Mol. Sci.*, 2015, 16, 3626.
11
12 322 11. V. V. Egorov, E. M. Rakhman'ko, E. V. Pomelenok and E. B. Okaev. *Russ. J.*
13
14 323 *Phys. Chem.*, 2006, 80, 969.
15
16 324 12. Y. I. Kuznetsov, L. V. Frolova and E. V. Tomina. *Prot. Met.*, 2006, 42, 215.
17
18 325 13. E. N. Shapovalova, M. N. Ofitserova, E. V. Savost'yanova and O. A. Shpigun.
19
20 326 *J. Anal. Chem.*, 2001, 56, 160.
21
22 327 14. C. M. Zou, H. Yu and M. Y. Wang. *Chin. Chem. Lett.*, 2014, 25, 201.
23
24 328 15. N. K. Jagota, A. J. Chetram and J. B. Nair. *J. Chromatogr. A*, 1996, 739, 343.
25
26 329 16. Y. Zhang, H. Yu and M. Y. Wang. *Anal. Methods*, 2015, 7, 5654-5660.
27
28 330 17. U. Masaaki and Y. Miyuki. *Carbohydr. Res.*, 2005, 340, 1722.
29
30 331 18. K. C. Chimalakonda, C. Hailey, R. Black, A. Beekman, R. Carlisle, E.
31
32 332 Lowman-Smith, H. Singletary, S. M. Owens and H. Hendrickson. *Anal.*
33
34 333 *Methods*, 2010, 2, 1249.
35
36 334 19. G. J. Schad, M. R. Euerby, G. G. Skellern and J. N. A. Tettey. *Anal. Bioanal.*
37
38 335 *Chem.*, 2012, 404, 239.
39
40 336 20. M. J. Ruiz-Angel and A. Berthod. *J. Chromatogr. A*, 2006, 1113, 101.
41
42 337 21. E. Gemperline; K. Laha; C. O. Scarlett; R. A. Pearce; L. Li. *Anal. Methods*,
43
44 338 2014, 6, 6389.
45
46 339 22. P. Stepnowski and J. Nichthausser. *Anal. Sci.*, 2008, 24, 1255.
47
48 340 23. Z. Fahimeh, G. Mehrorang and D. Ali, *Anal. Methods*, 2015, 5, 70064.
49
50 341 24. S. H. Haa, N. L. Maib and Y.M. Koo. *J. Chromatogr. A*, 2010, 1217, 7638.
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342 25. C. X. Lu, Z. G. Tang, C. B. Liu, X. Chen, Y. Wang and F. M. Dang. Anal.

343 Methods, 2015, 7, 5924.

344 26. Y. Pico, G. Font, J. C. Molto, and J. Manes, J. Chromatogr. A, 2000, 885, 251.

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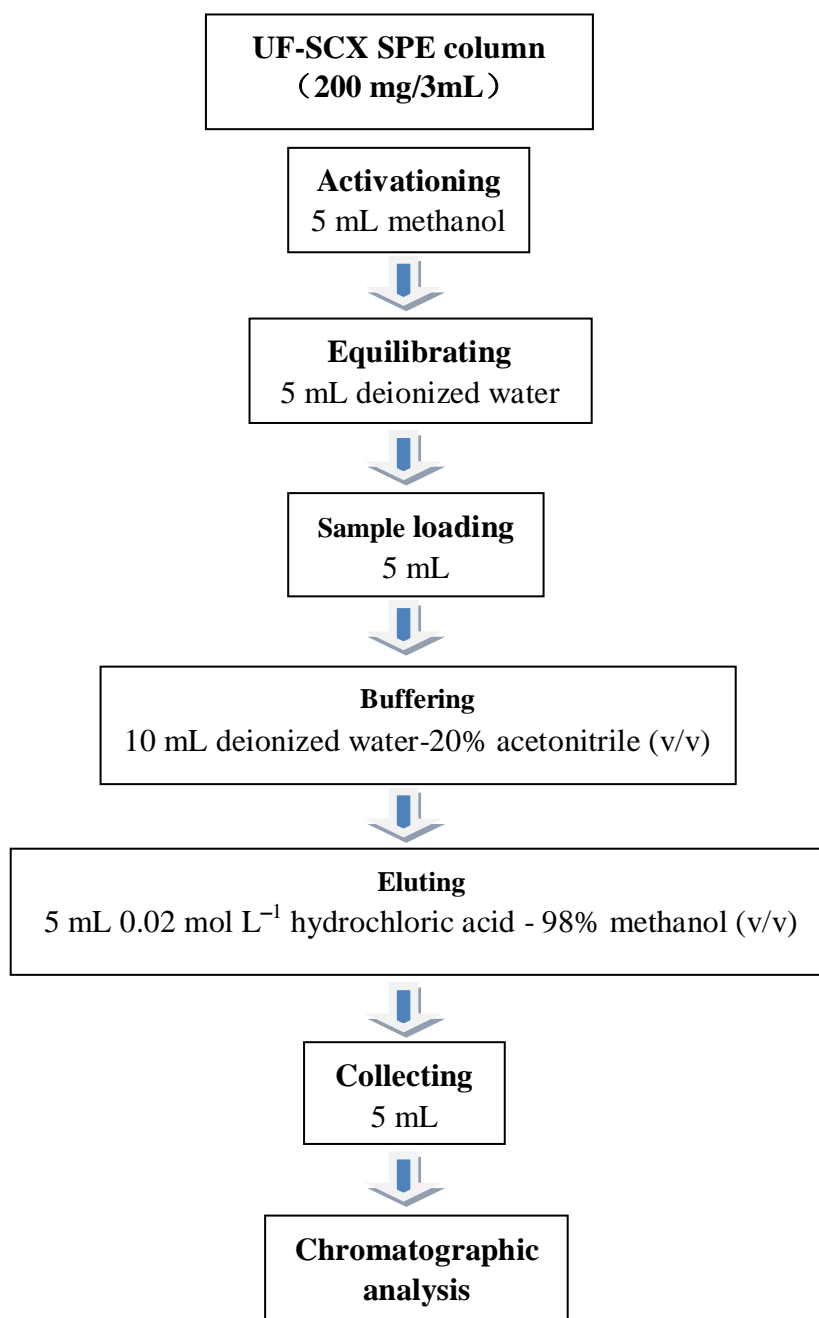
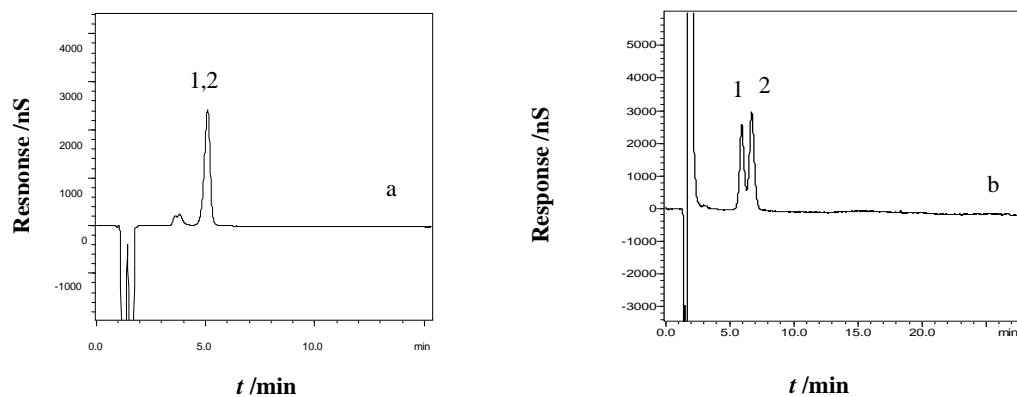


Fig. 1 Solid-phase extraction protocol of tetrabutyl ammonium cation and tetrabutyl phosphonium cation

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415 **Fig. 2** Chromatograms obtained with mobile phases containing different acids

416 Mobile phase, (a) 5 mmol L⁻¹ tartaric acid - 50% acetonitrile (v/v); (b) 3 mmol L⁻¹ methane
417 sulfonic acid - 30% acetonitrile (v/v). Peaks (50 mg L⁻¹) : 1, tetrabutyl ammonium ([TBA]⁺) ; 2,
418 tetrabutyl phosphonium ([TBP]⁺)
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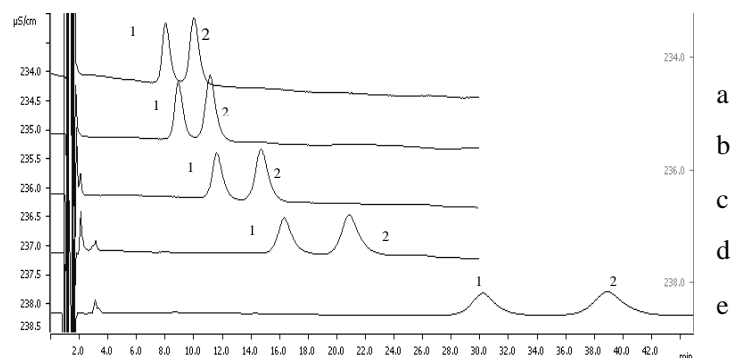
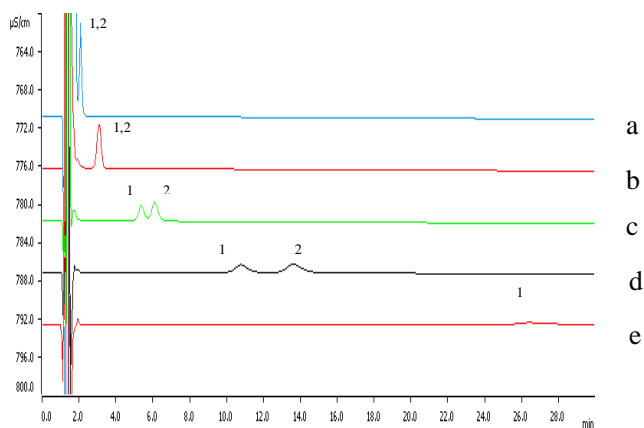


Fig. 3 Chromatograms obtained with mobile phases containing different concentrations of methanesulfonic acid

(a) 5 mmol L⁻¹; (b) 4 mmol L⁻¹; (c) 3 mmol L⁻¹; (d) 2 mmol L⁻¹; (e) 1 mmol L⁻¹. Mobile phase: different concentrations of methanesulfonic acid - 30% acetonitrile (v/v). Peaks (50 mg L⁻¹) : 1, [TBA]⁺; 2, [TBP]⁺

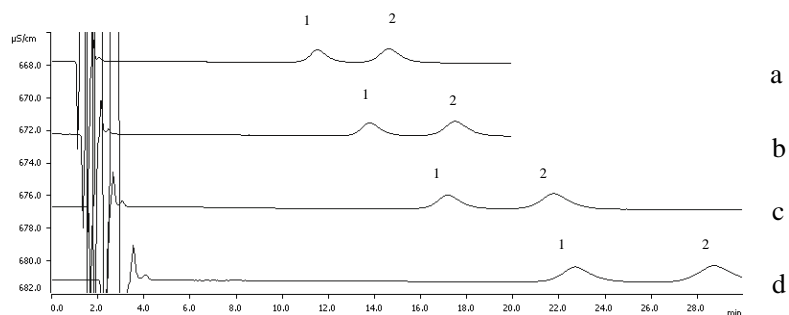
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Fig. 4 Chromatograms obtained with mobile phases containing different volume fractions of acetonitrile
(a) 50%; (b) 40%; (c) 30%; (d) 20%; (e) 10%. Mobile phase: 3 mmol L⁻¹ methanesulfonic acid - different volume fractions of acetonitrile (v/v). Peaks (50 mg L⁻¹) : 1, [TBA]⁺; 2, [TBP]⁺

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Fig. 5 Chromatograms of different flow rates

(a) 1.2 mL/min; (b) 1.0 mL/min; (c) 0.8 mL/min; (d) 0.6 mL/min. Peaks (50 mg L^{-1}): 1, $[\text{TBA}]^+$;
2, $[\text{TBP}]^+$

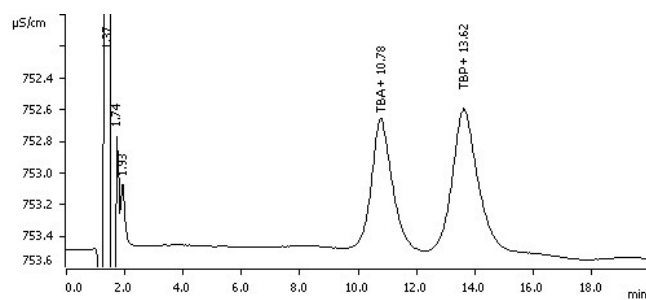


Fig. 6 Chromatogram of standard mixture solution

Sample pretreatment: solid-phase extraction column, UF-SCX (200mg/3mL) ; eluent, 0.02 mol L^{-1} hydrochloric acid - 98% methanol (v/v). Chromatographic conditions: column, Shodex IC YK-421; mobile phase, 3 mmol L^{-1} methanesulfonic acid - 20% acetonitrile; flow rate, 1.2 mL min^{-1} ; column temperature, room temperature; direct conductivity detection; inject volume, 20 μL . Peaks (50 mg L^{-1}) : 1, [TBA]⁺ ; 2, [TBP]⁺

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Table 1 Recoveries of solid-phase extraction

Eluent (mol L ⁻¹)	Recovery (%)	
	[TBA] ⁺	[TBP] ⁺
Hydrochloric acid-methanol		
0.02	90.5	97.1
0.05	74.7	5.0
0.07	58.6	8.0
0.09	69.4	17.8
0.12	52.8	19.9
0.24	46.0	20.1
0.36	30.5	21.0

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Table 2 Linear regression equations, limits of detection (LOD) and relative standard deviations of retention time (RSD_t) and peak area (RSD_s)

Ion	Linear regression equation $y = \text{area}$ $x = \text{concentration, mg L}^{-1}$	Correlation coefficient ($r, n = 5$)	LOD ($S/N=3,$ mg L^{-1})	Linear range (mg L^{-1})	RSD_t/RSD_s (%, $n = 5$)
[TBA] ⁺	$y = 0.0142x - 0.0132$	0.9996	0.502	1.67–100	1.22/0.24
[TBP] ⁺	$y = 0.0195x - 0.0265$	0.9997	0.448	1.49–100	1.23/0.25

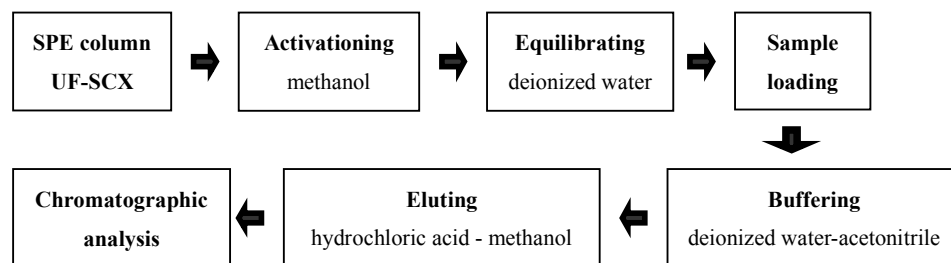
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Table 3 Contents and spiked recoveries of tetrabutyl ammonium cation and tetrabutyl phosphonium cation in environmental water sample

Sample	Ion	Original (mg/L)	Added (mg/L)	Total found (mg/L)	Recovery (%)
Environmental water	[TBA] ⁺	0.0	40.0	41.3	103.2
	[TBA] ⁺	0.0	50.0	48.5	97.0
	[TBA] ⁺	0.0	60.0	58.3	97.2
	[TBP] ⁺	0.0	40.0	40.4	100.9
	[TBP] ⁺	0.0	50.0	47.5	95.0
	[TBP] ⁺	0.0	60.0	56.9	94.8

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Table of contents entry



Solid phase extraction and ion chromatography for simultaneous determination of tetrabutyl ammonium and tetrabutyl phosphonium in environmental water sample