

Journal of Materials Chemistry B

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1 **Titania as driving agent for DHICA polymerization: a novel strategy**
2 **for the design of bioinspired antimicrobial nanomaterials**

3
4
5 Giuseppe Vitiello,^{a,b} Alessandro Pezzella,^c Anna Zanfardino,^d Mario Varcamonti,^d Brigida
6 Silvestri,^a Aniello Costantini,^a Francesco Branda^a and Giuseppina Luciani^{a,*}

7
8
9
10 ^a Dipartimento di Ingegneria Chimica, dei Materiali e della Produzione Industriale, Università di
11 Napoli “Federico II”, p.le V. Tecchio 80, 80125Napoli, Italy.

12 ^b CSGI, Consorzio Interuniversitario per lo Sviluppo dei Sistemi a Grande Interfase, Italy.

13 ^c Dipartimento di Scienze Chimiche, Università di Napoli “Federico II” via Cintia 4, 80126 Napoli,
14 Italy.

15 ^d Dipartimento di Biologia, Università di Napoli “Federico II” via Cintia 4, 80126Napoli, Italy.

16

17

18 * Corresponding author. Tel.: +390817682433; Fax: +390817682595.

19 E-mail address: giuseppina.luciani@unina.it

20

21

1 **Abstract**

2 Organic materials are widely employed to tune surface chemistry and/or as structuring agent of
3 inorganic materials. Here, we propose a novel synthetic approach whereby TiO₂ not only promotes
4 5,6-dihydroxyindole-2-carboxylic acid (DHICA) polymerization but also acts as templating agent
5 for the forming eumelanin itself. Hybrid TiO₂-DHICAmelanin nanostructures have been produced,
6 showing biocide activity even under visible light condition (activation). Hybrid nanostructures have
7 been analyzed and characterized by multiple techniques, proving that both organic and inorganic
8 phases strongly affect each other during *in-situ* formation, as far as it concerns both morphology
9 and microstructure, conferring unique biocide properties to the resulting nanomaterials. This
10 strategy ensures much more far-reaching implementation in the synthesis of hybrid nanosystems,
11 opening new perspectives in the design of multifunctional materials.

12

13

14 **Keywords:** nanomaterials, TiO₂, DHICA, eumelanin, radicals, antimicrobial

15

16

17

1 Introduction

2 An increasing interest has been recently growing on organic-inorganic hybrids as creative
3 alternative for obtaining new multifunctional nanomaterials. These hybrids made by coupling
4 organic molecules to inorganic nanostructures, not only combine the often dissimilar properties of
5 inorganic and organic components in one material, but also actually provide the opportunity to
6 invent a huge set of new multifunctional materials with a large spectrum of known and as yet
7 unknown properties.¹ A large number of recent literature data actually focus on
8 modification/modulation of surface chemistry of inorganic materials in order to improve their
9 functional properties.^{2,3} Alternatively, organic components play a key role in structuring the
10 inorganic phase.³

11 Herein, we provide a novel synthetic approach whereby inorganic materials are employed as active
12 and templating agents for formation of organic polymers. This bioinspired synthetic strategy has
13 been exploited to produce eco-friendly hybrid nanostructures with antimicrobial activity even under
14 visible light induction condition, following a bioinspired approach.

15 Eumelanins are negatively charged, hydrophobic pigments of high molecular weight, widely
16 dispersed in mammals, plants and microorganisms.^{4,5} Many studies have recently shown that
17 melanins interfere with numerous host defense mechanisms: they are highly immunogenic⁶ and
18 have anti-inflammatory properties⁷, protecting organisms against UV-radiation⁸⁻¹⁵ as well as
19 microbial lysis.^{16,17} Indeed, melanization of skin and other tissues is an important component of the
20 innate immune defense system: in this, the function of eumelanin in skin seems to inhibit the
21 proliferation of bacterial, fungal and other parasitic infections of the dermis and epidermis.^{18,19} In
22 particular, eumelanins²⁰ are produced by oxidation of tyrosine through tyrosinase to DOPA (o-
23 dihydroxyphenylalanine) and dopachrome, further the cyclization mediates to form 5,6-
24 dihydroxyindole (DHI) or 5,6-dihydroxyindole-2-carboxylic acid (DHICA).⁴ Since isolation of
25 natural eumelanins is a very difficult task, it is possible to mimic their formation by oxidizing in

1 vitro DHI²⁰ and DHICA²¹ obtaining synthetic eumelanin models quite similar to the natural
2 pigment.^{4,20}

3 Titanium dioxide (TiO₂, titania) is an inexpensive, environmentally friendly, nontoxic and
4 photostable material, as well as high performance catalyst in photo-oxidative processes.^{22,23}

5 In this paper, this TiO₂ property has been actually exploited to catalyze DHICA polymerization
6 through oxidative reactions. In details, TiO₂-DHICAmelanin hybrids were prepared through *in-situ*
7 methodology based on hydrothermal route. Morphology of nanostructures was investigated through
8 transmission electron microscopy (TEM). The specific surface area and pore size distribution were
9 also evaluated by BET porosimetry. Influence of DHICA-melanin on crystal structure of TiO₂ was
10 assessed by X-ray diffraction (XRD). Furthermore, electron paramagnetic resonance (EPR)
11 spectroscopy provided information about the presence of intrinsic radical species and their
12 distribution within the nanostructures. Optical properties of these materials were also evaluated
13 through Diffuse Reflectance UV-VIS (DRUV) spectroscopy. Finally, tests on bacterial pathogens in
14 the absence and presence of UV-irradiation proved antimicrobial activity of these nanomaterials
15 even under visible light. By in-depth overview, we linked antimicrobial activity to the physico-
16 chemical properties of these nanosystems.

17 This research is inspired by the growing interest on hybrid titania-based materials. Most of studies
18 reported in literature are focused on narrowing the band gap of TiO₂ in order to have photocatalytic
19 behavior even under visible light, as well as avoiding fast recombination of charge carriers (e⁻/h⁺)
20 in order to improve photo catalytic performance.²⁴ To the best of our knowledge, this study sets up
21 a novel role for TiO₂ which acts as both catalyst and structuring direct agent for the growing
22 polymeric/organic phase.

23 In this scenario, the proposed methodology defines a new potential synthetic strategy for the design
24 and production of hybrid nanomaterials. In addition, the proposed hybrid systems have intrinsic
25 potent antimicrobial activity even under environmental light. Biocide properties as well as non-
26 toxic and bio-friendly nature of both organic and inorganic components make these materials very

1 intriguing as a synthetic platform to produce antimicrobial formulations for biomedical and food
2 packaging applications.

3

4

1 **Experimental Section**

2 *Materials*

3 Titanium isopropoxide (TTiP), isopropanol, acetic acid, and triethylamine (TEA) were purchased
4 from Sigma Aldrich and used as received. DHICA monomer was prepared as described
5 elsewhere.²⁵

7 *Synthesis of TiO₂ nanoparticles*

8 TiO₂ nanoparticles were prepared by hydrothermal synthesis.^{26,27} A typical precursor solution was
9 obtained by adding dropwise 6 mL of Tiisopropoxide (TTiP)/isopropanol solution (Sol-1, 3.38 M in
10 TTiP) to 31.3 mL of water solution at pH 1.5 achieved by means of acetic acid (Sol-2). After Sol-1
11 addition a white precipitate was obtained that upon stirring at room temperature for two days led to
12 a yellowish colloidal solution, thus indicating resuspension of the precipitate and reduction of the
13 particles size below 20 nm.²⁸ Triethylamine (TEA) was then added dropwise to the TiO₂ colloidal
14 solution till pH=7. The obtained white precipitate suspension was then sealed within a Teflon
15 recipient (the liquid volume corresponding to 75% of the whole), placed into a circulating oven and
16 kept at 120 °C for 24 h. TiO₂ powders were obtained by centrifugation and repeated washing (3
17 times with distilled water). Then, obtained precipitates were re-suspended water.

19 *Synthesis of hybrid DHICA-TiO₂ nanoparticles*

20 Hybrid TiO₂-DHICA nanoparticles were prepared *in situ* following almost the same procedure as
21 bare nanoparticles. Briefly, Sol-1(6 mL) was added drop-wise to Sol-2 (31.3 mL) and the mixture
22 was kept stirred for two days leading to TiO₂ colloidal solution. Then, 10 mg of DHICA for 1.193
23 mL of TTiP were added just before neutralization with TEA till pH 7. Solution turned brown just
24 after DHICA addition. This sample will be named in the following as TiO₂-DHICA_{ti}. Finally, the
25 obtained suspension was sealed within a Teflon recipient (the liquid volume corresponding to 75%
26 of the whole), placed into a circulating oven and kept at 120 °C for 24h.²⁷

1 TiO₂-DHICA_{ti} nanoparticles recovered after hydrothermal treatment were submitted to oxidation
2 by exposure to an oxidizing atmosphere (e.g. oxygen atmosphere and ammonia vapors).²⁹ In the
3 general procedure, the appropriate suspension (hybrid nanoparticles or DHICA) was incubated in
4 the oxygen/ammonia atmosphere at controlled temperature (25-40 °C). The ammonia vapors were
5 produced by equilibration of the atmosphere with ammonia solution (28% to 7% NH₃ in H₂O) in a
6 sealed camera at 1atm pressure. Exposure times varied in the range 2 to 18 h. These samples will be
7 named as TiO₂-DHICA_{polym} in the following. Bare DHICA-melanin nanoparticles, as a
8 reference, were also prepared using the same procedure.

9

10 *Physico-chemical characterization*

11 In order to perform oxidative degradation³⁰ of hybrid DHICA-titania nanoparticles, a suspension of
12 the appropriate sample (10 mg) in 1M K₂CO₃ (3 mL of water solution) was treated with 30% v/v
13 H₂O₂ (20 μL of water solution) and vigorously stirred at 25 °C for 3 h. The reaction was stopped by
14 addition of 5% w/w NaHSO₃ (500 μL). The mixture was acidified to around pH=2 with 1M HCl
15 and extracted with ethyl acetate (4×1 mL). The combined organic layers were dried over sodium
16 sulfate and evaporated to dryness. The residue was taken up in water (1 mL) and analyzed by HPLC
17 using as the mobile phase a mixture of 0.05M citric acid and 0.3M formic acid containing 3% v/v
18 acetonitrile brought to pH=2.5 with conc. ammonium hydroxide.

19 Morphological analysis of the prepared nanoparticles was carried out by Thermal Electron
20 Microscopy (TEM). Samples for TEM were prepared by dispersing the obtained powders in
21 aqueous solution and then placing a drop of suspension on one side of the transparent polymer
22 coated 200 mesh copper grid. Samples were successively coated by carbon to increase the thermal
23 and the electric conductivities of powders. TEM images have been taken with a PHILIPS EM208S
24 microscope equipped with a Mega View camera for digital acquisition of images.

1 Crystalline phases of the titania-based nanosystems were identified by X-ray diffraction (XRD)
2 experiments performed by using a PANalytical diffractometer with a nickel filter and Cu
3 $K\alpha$ radiation.

4 The specific surface area (S_{BET}) and the pore volume (V_{P}) of hybrid nanosystems were evaluated by
5 generating seven-point isotherms at 77 K for N_2 adsorption (Autosorb-1, Quantachrome) using an
6 amount of char sample capable to provide a specific surface area equal to 5 m^2 in the sample cell.
7 The mesopore volume (V_{BJH}), the average pore radius (r_{P}) and the pore size distributions were
8 estimated by the Barreto-Joyner-Halenda (BJH) method applied to the desorption branch of the
9 isotherm.³¹

10 Electron Paramagnetic Resonance (EPR) spectroscopy experiments were carried out by means of
11 X-band (9 GHz) Bruker Elexys E-500 spectrometer (Bruker, Rheinstetten, Germany), equipped
12 with a super-high sensitivity probe head. Solid samples were transferred to flame-sealed glass
13 capillaries which, in turn, were coaxially inserted in a standard 4 mm quartz sample tube.
14 Measurements were performed at room temperature. The instrumental settings were as follows:
15 sweep width, 100 G; resolution, 1024 points; modulation frequency, 100 kHz; modulation
16 amplitude, 1.0 G. The amplitude of the field modulation was preventively checked to be low
17 enough to avoid detectable signal overmodulation. Preliminarily, EPR spectra were measured with a
18 microwave power of $\sim 0.6 \text{ mW}$ to avoid microwave saturation of resonance absorption curve.
19 Several scans, typically 32, were accumulated to improve the signal-to-noise ratio. Successively, for
20 power saturation experiments, the microwave power was gradually incremented from 0.02 to 160
21 mW. The g -factor value was evaluated by means an internal standard (Mg/MnO)³² which was
22 inserted in the quartz sample tube co-axially with the capillary containing the samples. Free radical
23 concentration in the sample was estimated by using a specific amount of MnO sample as reference.
24 The area under the EPR absorption curves was estimated by double integration of their first
25 derivatives.

1 Diffuse Reflectance UV–VIS (DRUV) measurements have been performed by using a Jasco
2 spectrophotometer and BaSO₄ as a reference in the 190–850 nm wavelengths range.

3

4 *Antimicrobial assay*

5 A single colony of *Escherichia coli* DH5 α strain was re-suspended in 5 mL of LB medium (Becton
6 Dickinson) and over-night incubated at 37 °C. When the culture reached an OD₆₀₀ of 1 unit, it was
7 diluted 1:100 in 20 mM, pH 7.0 NaP buffer. Samples were prepared by adding 40 μ L of bacterial
8 cells and the different compounds to test at a final concentration of 200 μ g/mL, 1 mL final volume
9 was reached with 20 mM pH 7.0 NaP buffer. Negative controls were represented by cells incubated
10 without any compound or with BSA at the same concentration of tested molecules; cells incubated
11 with ampicillin (0.05 mg/mL) were used as positive control. Samples were incubated at 37 °C for 4
12 hours and dilutions (1:100 and 1:1000) of all the samples were placed on solid LB medium and
13 incubated over-night at 37 °C. The following day the survived cells were estimated by colonies
14 counting on each plate and compared with the controls. All compounds were tested in triplicate
15 experiments, standard deviations were always <5% for each experiment. Finally, all samples, TiO₂,
16 DHICA-melanin, TiO₂-DHICA_{ti}, TiO₂-DHICA_{polym}, were also UV irradiated for 1 minute at
17 wavelength of 254 and 366 nm before their inoculation into bacterial cells.

18

19 **Results and Discussion**

20 *Melanin content determination*

21 In order to validate chemical composition of hybrid-titania nanoparticles, a classical analytical
22 procedure for eumelanin characterization was pursued.³⁰ The procedure involves the use of alkaline
23 hydrogen peroxide as the oxidizing agent to induce degradation of the eumelanin pigment into easy
24 to detect and quantify pyrrolecarboxylic acids to obtain indirect estimation of eumelanin content
25 (Table 1).

26

	hybrid TiO ₂ nanoparticles	DHICA-melanin
Yield (nmol/mg sample) ^a PTCA	10.0	28.5
Melanin content estimation	28 %	100 %

^aDetermined by HPLC, average of three experiments, S.D.b5%.

Table 1. Yields^a of pyrrole acids by oxidative degradation of hybrid nanoparticles and DHICA-melanin.

Data obtained do confirm that the eumelanin pigment is formed within the hybrids. In order to confirm the key role played by TiO₂ in polymerization of DHICA, hydrothermal synthesis was also carried out without the addition of TTiP to the mixture, as a control experiment. No particles were recovered from solution after the hydrothermal treatment, as demonstrated by the EPR spectrum (see Figure ESI3) which does not show any signal.

A possible mechanism of titania driven melanin formation can be hypothesized. It is well known that TEA leads to the precipitation of TiO₂ amorphous gel from Ti⁴⁺ aqueous solutions.^{27,33} TiO₂ nanoparticles can be sensitized to visible light through adsorption of organic molecules (dye-sensitizers) onto the semiconductor surface. In the case of high molecular weight molecules, upon photoexcitation, an electron is transferred from the HOMO of the adsorbed molecule to the conduction band of the semiconductor, either directly or indirectly, via the LUMO of the adsorbed molecule. Alternatively, for small organic molecules, such as catechol and dopamine, the modifier adsorption leads to the formation of an adsorbate-to-nanoparticle (ligand-to-metal) charge-transfer complex, which absorbs visible light, inducing a direct injection of an electron from the ligand to the oxide conduction band.³⁴⁻³⁶ Electrons injected into the conduction band by charge transfer complexes, have molecular oxygen reduced to superoxide ion O₂⁻, a very reactive species in oxidative processes, that may involve even oxidation of the organic ligand. Indeed, it is known that catechol adsorbed onto titania surfaces, polymerizes under visible light.³⁴ Similarly, DHICA can adsorb onto TiO₂ and act as a ligand for Ti⁴⁺ ions, through its catechol group. Consequently, the

1 formed charge transfer complex activates oxidative polymerization of DHICA even under visible
2 light. Moreover it is well established that metal ions³⁷ can promote 5,6-dihydroxyindole aerobic
3 oxidation and it is expected Ti does behave as its close metal elements.³⁸

4 Rough comparison with a DHICA melanin sample allows to estimate the pigment content in hybrid
5 particles in the range of 20% w/w for both TiO₂-DHICA_ti and TiO₂-DHICA_polym.

6

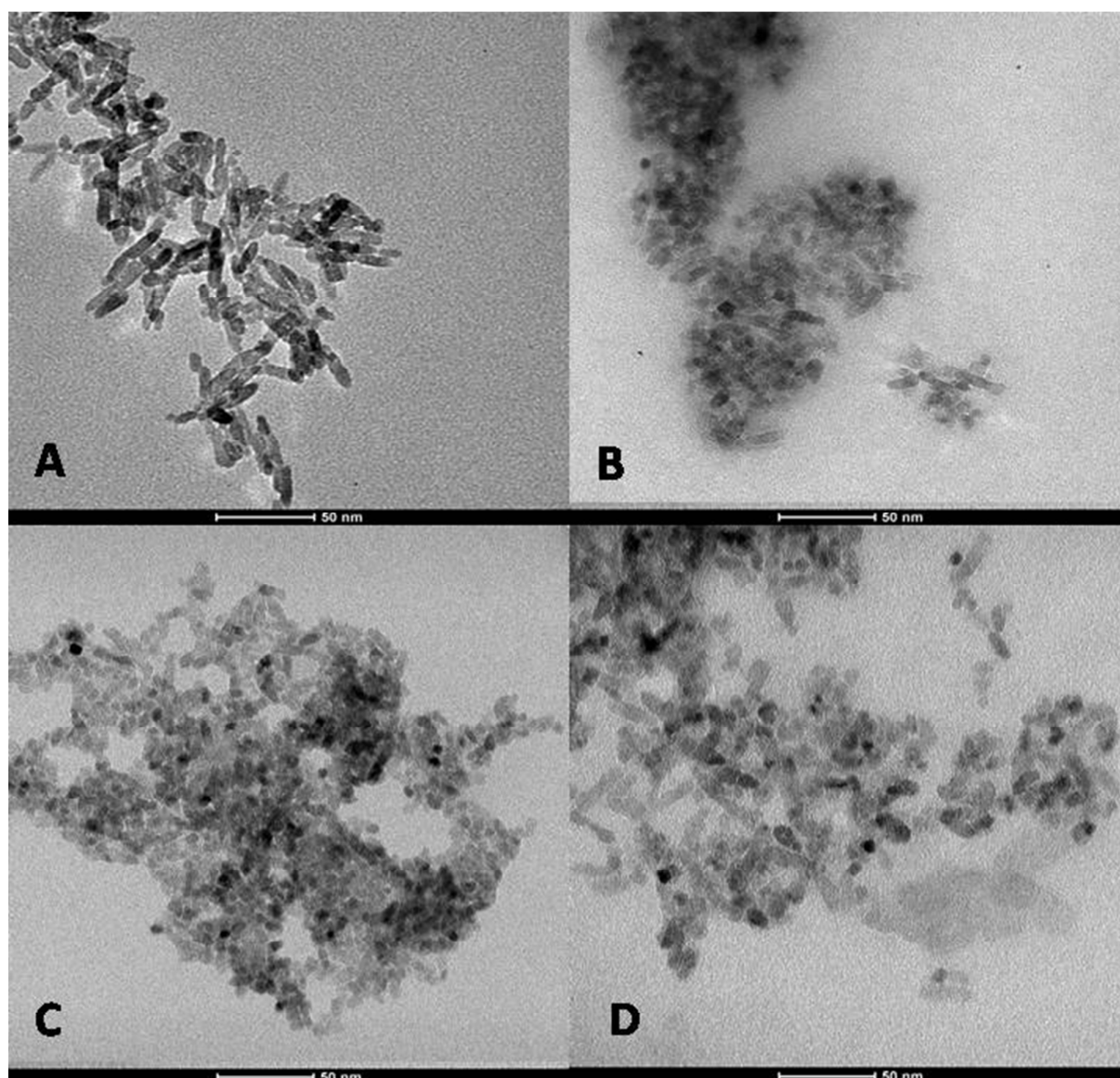
7 *TEM analysis*

8 Morphology of the bare TiO₂ and hybrid TiO₂-DHICAnanoparticleswas observed with TEM (Fig.
9 1). DHICA-melanin micrography is also reported for comparison (Fig.1B).

10 As shown in Fig. 1A, bare TiO₂ nanosystems show rod-like structures (3x12 nm in size), while
11 DHICA-melanin presents a cubic shape, about 30 nm in size. In contrast, different morphologies
12 can be observed for TiO₂-DHICA_ti and TiO₂-DHICA_polym samples with respect to TiO₂: *in situ*
13 procedure led to brownish rounded shaped particles, about 10 nm in size, due to a DHICA
14 involvement in the growth of hybrid nanoparticles.²⁶ DHICA can actually act as a complexing agent
15 for Ti⁴⁺ ions, through its catechol group. Furthermore, DHICA molecules may preferentially absorb
16 onto specific planes during the growth of the particles.³⁹

17 TEM micrographs (Figs. 4C and 4D) clearly indicate that no differences are detectable for the two
18 TiO₂-DHICA_ti and TiO₂-DHICA_polym systems, as far as it concerns shape and size, suggesting
19 that morphology of nanomaterials is defined by the hydrothermal synthesis.

20



1
2 **Fig. 1** TEM micrographs of bare TiO₂ (A), DHICA-melanin (B), TiO₂-DHICA_{ti} (C) and TiO₂-
3 DHICA_{polym} (D) nanostructures.

4
5 *XRD analysis*

6 XRD patterns of the hybrid samples, reported in Electronic Supporting Information (Figure ESI1),
7 are in agreement with the standard anatase profile (JCPDS 84–1286), both in terms of peak
8 positions (i.e., the diffraction angles 2θ) and relative peak intensities⁴⁰ exhibiting strong diffraction
9 peaks at 25° and 48°. Therefore, titania crystallized in the anatase structure in all samples.⁴¹ Worth
10 of note is that, the anatase pattern is featured also by TiO₂-DHICA_{ti} and TiO₂-DHICA_{polym}.

1 The latter shows sharper width at half height than both bare TiO₂ and TiO₂-DHICA_ti, suggesting
 2 larger dimensions of the crystals within the particles (about 4.0 nm determined by Sherrer formula).

3

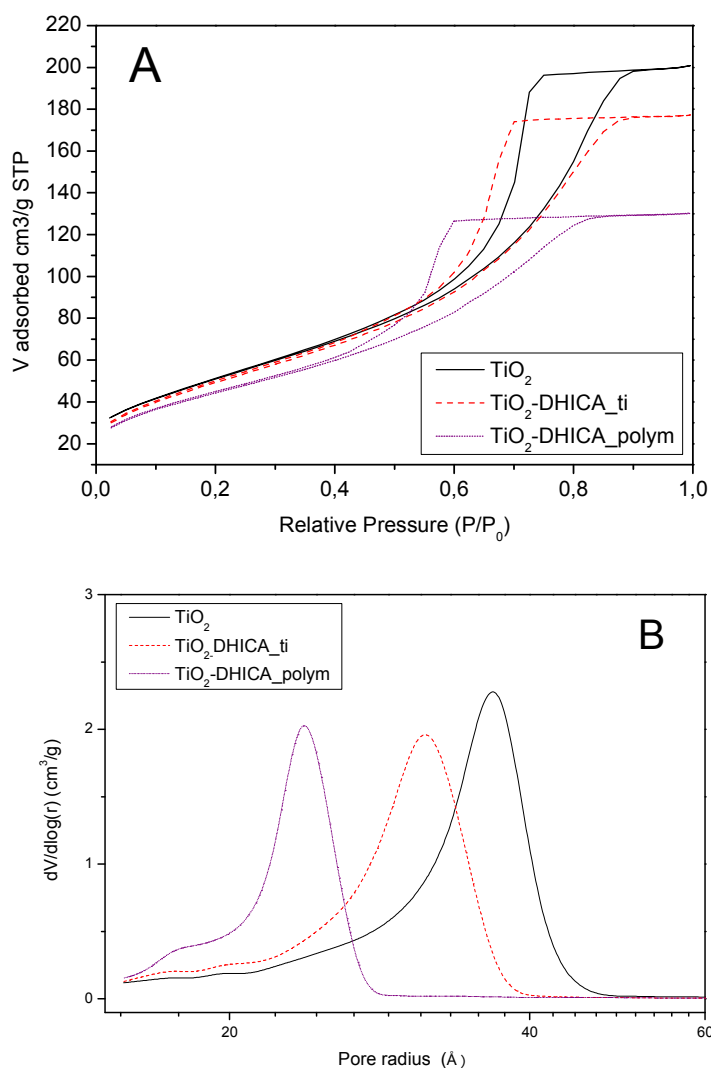
4 *Surface area and pore distribution*

5 Nitrogen adsorption-desorption isotherms of bare TiO₂ and hybrid TiO₂-DHICA_melanin
 6 nanostructures are shown in Fig. 2A. All samples show a type-IV isotherm, with a type-H2
 7 desorption hysteresis loop⁴², typical of mesoporous solids. This is confirmed by pore size
 8 distribution curves (Fig. 2B), indicating that all systems have mesopores with size in the range 2-4
 9 nm (Table 2). Furthermore, pore size distribution of TiO₂-DHICA_polym shows a pronounced
 10 shoulder at about 1.8 nm, suggesting the presence of micropores within the sample. Mesopores size
 11 is narrower in the TiO₂-melanin hybrids, suggesting that *in situ* formation of both melanin and
 12 titania during the hydrothermal process produces an intimate mixture of the organic and inorganic
 13 components. Moreover, the further polymerization step of DHICA after hydrothermal treatment
 14 (TiO₂-DHICA_polym) is supposed to produce high molecular weight oligomers within the pores,
 15 thus decreasing their size, and increasing the micropores, as shown in Fig. 2B. Finally, DHICA
 16 polymerization within the pores must partially seal porosity and lead to a decrease of specific
 17 surface area (see Table 2). As reference, the surface area of DHICA-melanin nanostructures is also
 18 determined, showing a very low value equal to 28.53 m²/g.

	TiO ₂	TiO ₂ -DHICA_ti	TiO ₂ -DHICA_polym
Surface Area (m ² /g)	190.3	191.4	171.7
Pore Volume (cc/g)	0.30	0.27	0.19
Pore Radius (Å)	37.1	31.4	23.7

19 **Table 2.** BET surface area and pore properties of the resulting hybrid nanoparticles.

20



1

2

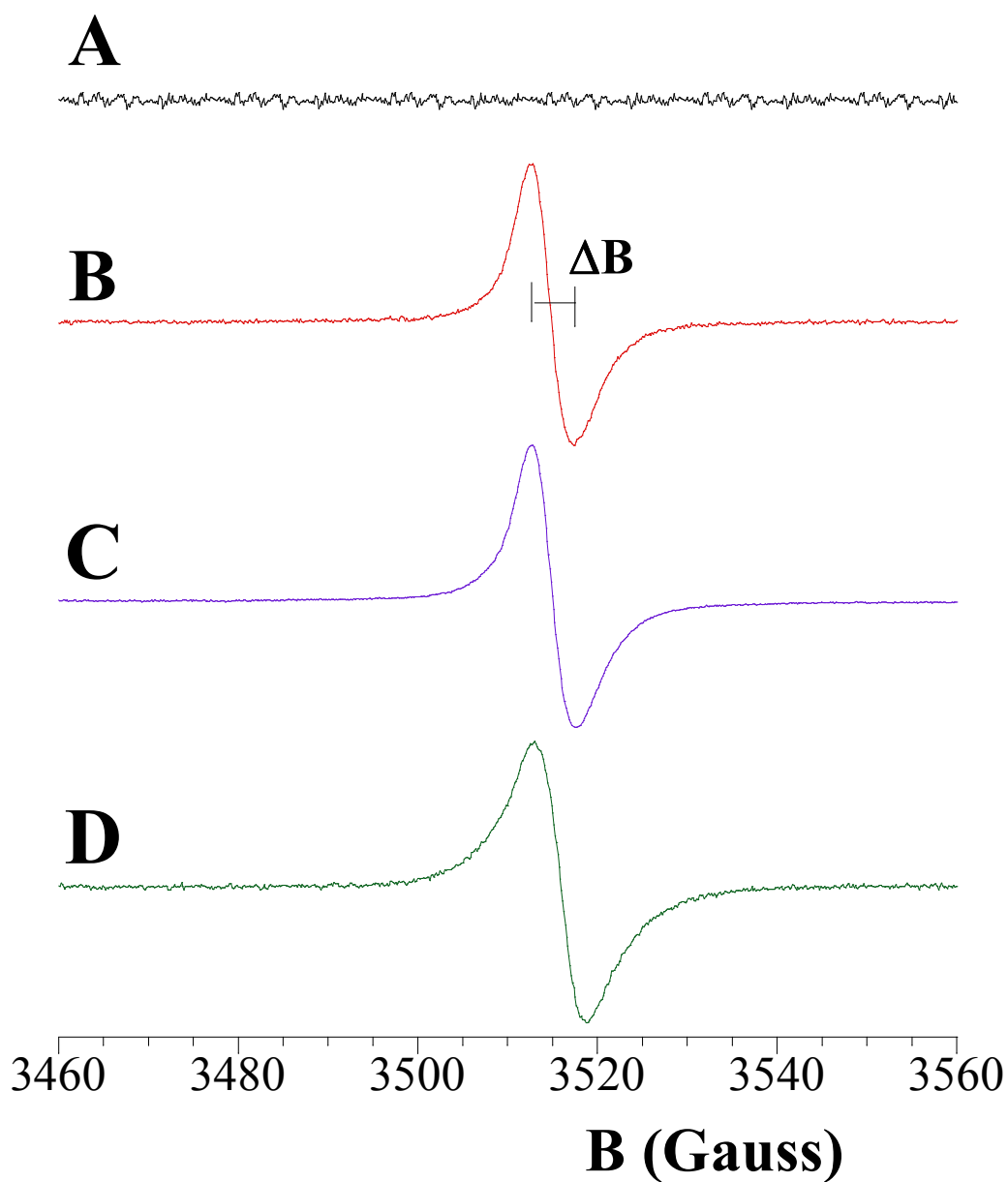
3 **Fig. 2** Adsorption-desorption isotherms of nitrogen (A). BJH distribution curves for bare TiO₂ and
4 hybrid melanin-based nanoparticles (B).

5

6 *EPR analysis*

7 EPR measurements on TiO₂ and DHICA-hybrid TiO₂nanosystems were performed by following an
8 experimental procedure recently used in the characterization of melanins.^{43,44} Analysis of this EPR
9 signal provides significant information on the nature of the paramagnetic centers and on the
10 supramolecular properties of solid materials. In this study, we undertook an EPR characterization of
11 powdered samples synthesized as previously described. All EPR spectra are reported in Fig. 3,

1 whereas the spectral parameters calculated from them are summarized in Table 3. By observing Fig.
2 3A, no signal was detected for neat TiO₂ (spectrum A), indicating no oxygen vacancy or Ti³⁺ in
3 these powder which is consistent with previous reports that indicates no EPR signal for pure
4 TiO₂.^{22,43} In Fig.3, the spectra B, C and D represent the spectra of TiO₂-DHICA_ti, TiO₂-
5 DHICA_polym and DHICA-melanin, respectively.



6
7 **Fig.3** EPR spectra of TiO₂ (A), TiO₂-DHICA_ti (B) TiO₂-DHICA_polym (C) and DHICA-melanin
8 (D) nanosystems.

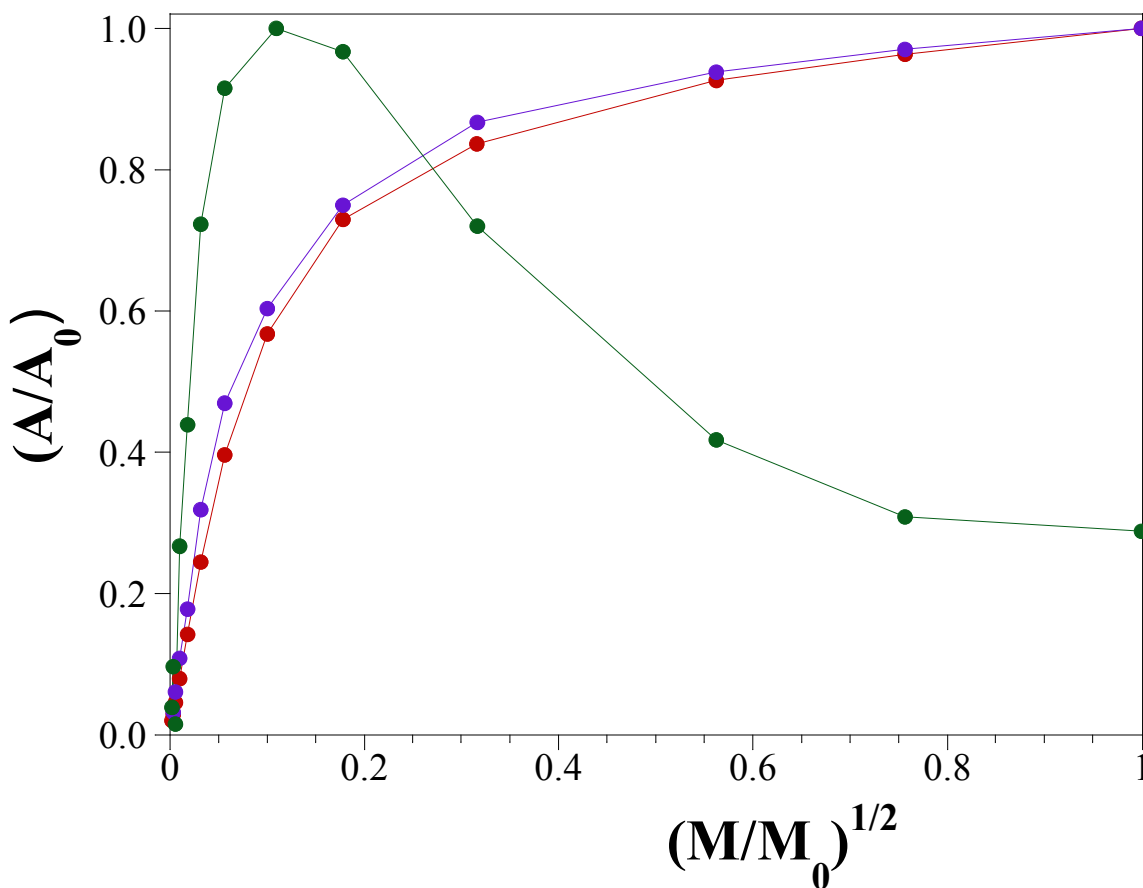
1 They present a similar lineshape: a single, roughly symmetric signal at a g value of ~ 2.0035 (see
2 Table 3), typical of carbon-centered radicals derived from the DHICA polymerization process, in
3 agreement with the literature.^{45,46} However, a deeper analysis of the spectra of composites shows
4 that the TiO₂-DHICA_ti and TiO₂-DHICA_polym spectra are evidently broader than the DHICA-
5 melanin one. The difference in lineshapes was corroborated by the quantitative determination of the
6 signal amplitude (ΔB). This parameter, detectable by the experimental spectra (see Fig. 3), is related
7 to the mean distance between the radical centers. The ΔB values determined from TiO₂-DHICA_ti
8 and TiO₂-DHICA_polym spectra are similar and significantly lower than the value obtained from
9 the spectrum of DHICA-melanin (Table 3). This evidence indicates that in the hybrid nanoparticles,
10 the radical centers are nearest in their spatial distribution within the nanostructures, probably due to
11 the presence of TiO₂ which modulates the macromolecular organization.

	TiO ₂ -DHICA_ti	TiO ₂ -DHICA_polym	DHICA-melanin
g -factor	2.0034 \pm 0.0004	2.0033 \pm 0.0004	2.0035 \pm 0.0004
ΔB (G)	4.9 \pm 0.2	5.0 \pm 0.2	5.6 \pm 0.2
Spin \times g ⁻¹	5.2 \times 10 ²¹	5.1 \times 10 ²²	4.3 \times 10 ²³

12 **Table 3** EPR spectral parameters for hybrid TiO₂-DHICA nanosystems.

13

14 To ascertain physical reasons for this experimental evidence, the EPR spectra of the same samples
15 were acquired setting a higher microwave power. An inspection of the normalized power saturation
16 profiles, reported in Fig. 4, shows that, as the microwave power is increased, the spin density
17 increases.



1

2 **Fig.4** Plot of normalised amplitude vs. power intensities of free radicals of TiO₂-DHICA_{ti} (red
3 circles), TiO₂-DHICA_{polym} (purple circles) and DHICA-melanin (greencircles) nanosystems.

4

5 Also, the power saturation curves of TiO₂-DHICA_{ti} and TiO₂-DHICA_{polym} go to a plateau, in
6 contrast to that observed in the case of DHICA-melanin. Despite very similar *g* values, the
7 quantitative determination of the signal amplitude ΔB and the power saturation profiles suggest a
8 different spatial distribution of the radical centers in the supramolecular organization of the hybrid
9 TiO₂-DHICA_{ti} and TiO₂-DHICA_{polym} systems with respect to the DHICA-melanin. Moreover,
10 the presence of TiO₂ influences the DHICA organization, causing an apparent loss of the typical
11 homogeneous distribution of free-radicals in DHICA melanin⁴⁶ associated with a broadening of the
12 signal amplitude in the TiO₂-DHICA_{ti} and TiO₂-DHICA_{polym} spectra. It is worth to note, here,
13 that the presence of TiO₂ in DHICA-based samples produces power saturation profiles very similar

1 to that obtained for the pure DHI melanin⁴⁶ suggesting an increase in the variety of the radical
2 species in the hybrid nanostructures with respect to the bare DHICA-melanin.

3 Finally, the quantitative analysis of the spectra intensities demonstrates that TiO₂-DHICA_{polym}
4 system presents a higher spin density than TiO₂-DHICA_{ti} (see Table 3).

5

6 *Diffuse Reflectance UV–VIS analysis*

7 For UV-Vis spectra, the measured intensity was expressed as the value of the Kubelka-Munk
8 function $F(R)$. The energy gap values were calculated by linearization of the plot of $(F(R)h\nu)^{1/2}$
9 against $h\nu$, as reported in Fig. ESI2. For the bare TiO₂, the evaluated optical band gap is 3.2 eV for
10 the transition from the valence band (VB) to conduction band (CB), as well reported in literature.⁴⁷

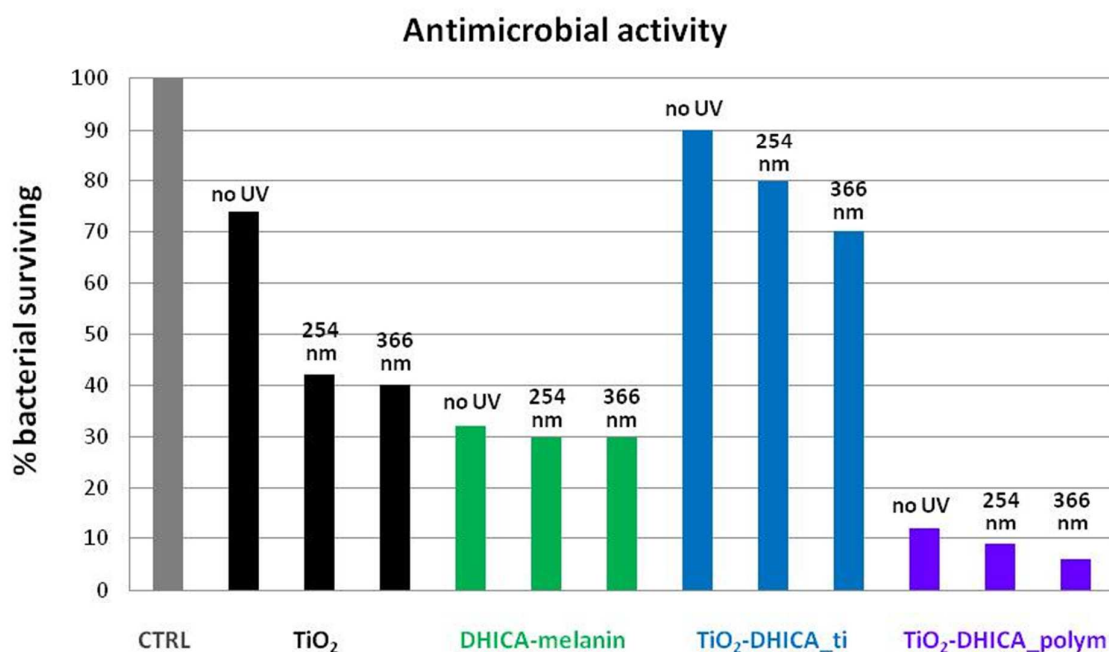
11 At the same time, the hybrid DHICA-based nanosystems show a decrease in the TiO₂ band-gap of
12 0.5 eV. Yet, a decrease of intensity of absorption band of TiO₂ within 200 and 400 nm can be
13 appreciated in hybrid systems. This must be due to a shielding effect of melanin towards TiO₂.

14

15 **Antimicrobial assay**

16 Antimicrobial activity of TiO₂, DHICA and their hybrid nanostructures was tested against
17 *Escherichia coli* DH5 α . A concentration of 200 μ g/mL of nanoparticles suspension was added to
18 the bacterial cell cultures. Bare TiO₂ was not very active against the indicator strain, as already
19 previously reported,⁴⁸ while DHICA-melanin showed a significant amount of activity, leaving the
20 30% of bacterial cells alive. This result is in agreement with the described function of melanin as
21 part of the innate immune defense system.¹⁸ The combination of TiO₂ and DHICA in the hybrid
22 nanosystems has also been investigated. In the case of TiO₂-DHICA_{ti} nanoparticles, no
23 antibacterial activity was detected, instead TiO₂-DHICA_{polym} nanoparticles have shown an
24 excellent antibacterial activity, leaving only the 10% of bacterial cells alive, as observable in Fig. 5.
25 This result can be ascribable to the higher radicals concentration in TiO₂-DHICA_{polym} than TiO₂-
26 DHICA_{ti}, as determined by EPR, according to the hypothesis of an antimicrobial mechanism

1 based on the free radical-induced membrane damage.⁴⁹⁻⁵¹ Indeed, TiO₂-DHICA_polym shows an
 2 higher biocide activity than bare DHICA-melanin, even with a lower radical concentration. This
 3 must be due to a broader free-radical species distribution within TiO₂-DHICA_polym, as evidenced
 4 by EPR results. This peculiar distribution might improve interaction of these radicals with cells
 5 wall, resulting in enhanced antimicrobial activity. Furthermore, higher surface area (Table 2) of
 6 TiO₂-DHICA_polym nanostructures may also account for its higher biocide efficacy.



7

8 **Fig. 5** Antibacterial activity of the different TiO₂-based nanoparticles against *Escherichia coli*
 9 DH5α strain. On y-axis is represented the % of bacterial surviving. CTRL bar corresponds to
 10 untreated bacterial cells. The other bars represent bacterial survival after treatment with nano-
 11 particles containing TiO₂, DHICA-melanin, TiO₂-DHICA_ti and TiO₂-DHICA_polym without
 12 irradiation (first bar of each set), with UV irradiation at 254 nm (second bar of each set) and UV
 13 irradiation at 366 nm (third bar of each set). The standard deviation was always lower than 5%.
 14 Values are reported as means±SEM of three different experiments. P< 0.05 (one-way ANOVA
 15 followed by Tukey post-test).

16

17 The same antibacterial tests were also performed by using all the considered nanoparticles after an
 18 UV-irradiation at both 254 and 366 nm. A significant increase of antibacterial activity was observed

1 overall for bare TiO₂ nanoparticles, as already reported in literature where TiO₂ biocide activity
2 under UV irradiation has been widely assessed.^{52,53} On the other hand, the variations in the
3 antimicrobial activity of the other nanosystems were mainly in the range of the standard deviations
4 (see Fig. 4). These results clearly evidence that, with this melanin content (20% w/w), TiO₂ is no
5 longer active as antimicrobial agent even under UV irradiation. This could be ascribed to a UV-
6 shielding effect of DHICAmelanin towards TiO₂,⁵⁴ as confirmed by DRUV results.

7

1 **Conclusion**

2 In the present work, we demonstrated that TiO₂ can catalyze the formation of eumelanin biopolymer
3 from DHICA, leading to the formation of hybrid nanostructures with an eumelanin content of about
4 20% w/w. These nanomaterials show higher antimicrobial activity towards *Escherichia Coli*
5 bacteria cultures than DHICA-melanin itself, even under visible light. This property must be strictly
6 related to the physico-chemical properties of the investigated systems. In particular, we observed
7 that the DHICA presence during the hydrothermal treatment strongly affects the nanoparticles
8 morphology. All hybrid nanostructures keep the typical anatase crystalline structure and show a
9 mesoporous character, with a higher surface area than DHICA-melanin. Indeed, the presence of
10 TiO₂ influences the DHICA supramolecular organization inducing the generation of a free-radicals
11 variety with respect to the bare DHICA-melanin. This might be associated with a more effective
12 interaction of the available different radical species with cell targets, resulting in an improved
13 antimicrobial activity.

14 The proposed methodology settles a new role for the inorganic phase, as an active agent in
15 promoting formation of hybrid nanosystems and driving structure of the organic component,
16 opening new perspectives in the design of multifunctional materials.

17

18

19

20

21

22

23

24

25

26

1 **References**

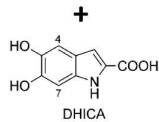
- 2 1 G. Kickelbick, in: G. Kickelbick (Ed.), *Hybrid Materials*, Wiley-VCH, Weinheim, 2007, **1**.
- 3 2 C. Sanchez, B. Julian, P. Belleville, M. Popall, *J. Mater. Chem.*, 2005, **15**, 3559-3592.
- 4 3 C. Sanchez, P. Belleville, M. Popall, L. Nicole, *Chem. Soc. Rev.*, 2011, **40**, 696-753.
- 5 4 G. Prota, *Melanins and Melanogenesis*; Academic Press: San Diego, CA, 1992.
- 6 5 J. J. Nordlund, R. E. Boissy, V. J. Hearing, R. A. King, W. S. Oetting, J. P. Ortonne, Eds.;
7 Blackwell Publishing: Malden, MA, 2.
- 8 6 J. D. Nosanchuk, A. L. Rosas, A. Casadevall, *J. Immunol.*, 1998, **160**, 6026-6031.
- 9 7 N. Avramidis, A. Kourounakis, L. Hadjipetrou, V. Senchuk, *Arzneim Forsch./Drug*
10 *Res.*, 1998, **48**, 764-771.
- 11 8 P. Meredith, J. Riesz, *Photochem. Photobiol.*, 2004, **79**, 211-216.
- 12 9 S. P. Nighswander-Rempel, J. Riesz, J. Gilmore, J. P. Bothma, P. Meredith, *J. Phys. Chem. B*,
13 2005, **109**, 20629-20635.
- 14 10 S. P. Nighswander-Rempel, *Biopolymers*, 2006, **82**, 631-637.
- 15 11 J. B. Nofsinger, T. Ye, J. D. Simon, *J. Phys. Chem. B*, 2001, **105**, 2864-2866.
- 16 12 J. Riesz, T. Sarna, P. Meredith, *J. Phys. Chem. B*, 2006, **110**, 13985-13990.
- 17 13 T. Ye, J. D. Simon, *J. Phys. Chem. B*, 2003, **107**, 11240-11244.
- 18 14 A. Corani, A. Huijser, T. Gustavsson, D. Markovitsi, P. A. Malmqvist, A. Pezzella, M.
19 D'Ischia, V. Sundström, *J. Am. Chem. Soc.*, 2014, **136**, 11626-11635.
- 20 15 A. Huijser, A. Pezzella, V. Sundström, *Phys. Chem. Chem. Phys.*, 2011, **13**, 9119-9127.
- 21 16 B. J. Bloomfield, M. Alexander, *J. Bacteriol.*, 1964, **93**, 1276-1280.
- 22 17 M. J. Kuo, M. Alexander, *J. Bacteriol.*, 1967, **94**, 624-629.
- 23 18 J. A. Mackintosh, *J. Theor. Biol.*, 2001, **211**, 101-113.
- 24 19 P. Zhao, J. Li, Y. Wang, H. Jiang, *Insect Biochem. Mol. Biol.* 2007, **37**, 952-959.
- 25 20 M. D'Ischia, K. Wakamatsu, A. Napolitano, S. Briganti, J. C. Garcia-Borrón, D. Kovacs, P.
26 Meredith, A. Pezzella, M. Picardo, T. Sarna, J. D. I. Simon, *Pigm. Cell Melanoma Res.* 2013,
27 **26**, 616-633.
- 28 21 A. Pezzella, D. Vogna, G. Prota, *Tetrahedron*, 2002, **58**, 3681-3687.
- 29 22 M. Alfè, D. Spasiano, V. Gargiulo, G. Vitiello, R. Di Capua, R. Marotta, *Appl. Catal. A*, 2014,
30 **487**, 91-99.
- 31 23 S. Satyro, R. Marotta, L. Clarizia, I. Di Somma, G. Vitiello, M. Dezotti, G. Pinto, R. F. Danats,
32 R. Andreozzi, *Chem. Engin. J.* 2014, **251**, 257-268.
- 33 24 G. Mallocci, M. Binda, A. Petrozza, A. Mattoni, *J. Phys. Chem. C*, 2013, **117**, 13894-13901.
- 34 25 R. Edge, *Pigm. Cell. Res.*, 2006, **19**, 443-450.

- 1 26 B. Jiang, H. Yin, T. Jiang, *Mater. Chem. Phys.*, 2006, **98**, 231-235.
- 2 27 A. Pezzella, L. Capelli, A. Costantini, G. Luciani, F. Tescione, B. Silvestri, G. Vitiello, F.
3 Branda, *Mater. Sci. Engin. C*, 2013, **33**, 347-355.
- 4 28 G. Oskam, A. Nellore, R. Lee Penn, P. C. Searson, *J. Phys. Chem. B*, 2003, **107**, 1734-1738.
- 5 29 A. Pezzella, M. Barra, M. Musto, A. Navarra, M. Alfè, P. Manini, S. Parisi, A. Cassinese, V.
6 Criscuolo, M. d'Ischia, *Mater. Horiz.*, 2015, DOI: 10.1039/C4MH00097H.
- 7 30 A. Napolitano, A. Pezzella, *Tetrahedron*, 1996, **52**, 8775-8780.
- 8 31 E. P. Barret, L. G. Joynt, P. P. aHalenda, *J. Am. Chem. Soc.*, 1951, **73**, 373-380.
- 9 32 N. D. Yordanov, V. Gancheva, V. A. Pelova, *J. Radioanalyt. Nucl. Chem.*, 1999, **240**, 619-622.
- 10 33 T. M. Stawski, S. A. Veldhuis, R. Besselink, H. L. Castricum, G. Portale, D. H. A. Blank, J. E.
11 Elshof, *J. Phys. Chem. C*, 2012, **116**, 425-434.
- 12 34 T. Lana-Villarreal, A. Rodes, J. M. Pérez, R. Gómez, *J. Am. Chem. Soc.*, 2005, **127**, 12601-
13 12611.
- 14 35 S. Varaganti, G. Ramakrishna, *J. Phys. Chem. C*, 2010, **114**, 13917-13925.
- 15 36 D. C. Grauer, A. P. Alivisatos, *Langmuir*, 2014, **30**, 2325-2328.
- 16 37 A. Napolitano, M. G. Corradini, G. Prota, *Tetrahedron Letters* 1985, **26**, 2805-2808.
- 17 38 G. M. Martin, E. P. Beneditt, N. Eriksen, *Nature*, 1960, **186**, 884-885.
- 18 39 H. Zhu, K. Yao, Y. Wo, N. Wang, L. Wang, *Semicond. Sci. Technol.*, 2004, **19**, 1020-1023.
- 19 40 JCPDS, International Centre for Diffraction Data 1998.
- 20 41 H. Zhang, M. Finnegan, J. F. Banfield, *Nano Lett.*, 2001, **1**, 81-85.
- 21 42 J. Sing, K. S. W. Everett, D. H. Haul, R. A. W. Moscou, L. Pierotti, R. A. Rouquerol, J. T.
22 Siemieniewska, *Pure and Appl. Chem.* 1985, **57**, 603-619.
- 23 43 E. Cesareo, L. Korkina, G. D'Errico, G. Vitiello, M. S. Aguzzi, F. Passarelli, J. Z. Pedersen, A.
24 Facchiano, *Plos One*, 2012, **7**, e48849.
- 25 44 N. F. Della Vecchia, A. Luchini, A. Napolitano, G. D'Errico, G. Vitiello, N. Szekely, M.
26 D'Ischia, L. Paduano, *L. Langmuir*, 2014, **30**, 9811-9818.
- 27 45 F. Zuo, L. Wang, T. Wu, Z. Zhang, D. Borchardt, P. Feng, *J. Am. Chem. Soc.*, 2010, **132**,
28 11856-11857.
- 29 46 L. Panzella, L.; Gentile, G.; D'Errico, G.; Della Vecchia, N. F.; Errico, M. E.; Napolitano, A.;
30 Carfagna, C.; d'Ischia, M. *Angew. Chem. Int. Ed.* 2013, **52**, 12684-12687.
- 31 47 K. M. Reddy, S. V. Manorama, A. R. Reddy, *Mater. Chem. Phys.*, 2002, **78**, 239-245.
- 32 48 R. Ahmad, M. Sardar, *Int. J. Inn. Res. in Sci., Eng. Tech.*, 2013, **2**, 3569-3574.
- 33 49 M. A. Kohanski, D. J. Dwyer, B. Hayete, C. A. Lawrence, J. J. Collins, *J. Cell.*, 2007, **130**, 797-
34 810.

- 1 50 J. S. Sung Kim, E. Kuk, K. N. Yu, J. H. Kim, S. J.Park,H. J. Lee, S. H. Kim,Y. K.Park, Y.
2 H.Park,C. Y. Hwang, Y. K.Kim, Y. S.Lee,D. H. Jeong, M. H.Cho, *Nanomed.: Nanotech., Biol.*
3 *Med.*, 2007, **3**, 95-101.
- 4 51 M. A. Kohanski, M. A.DePristo,J. J. Collins,*Molecular Cell.*, 2010, **37**, 311-320.
- 5 52 A.Kubacka, M. Ferrer, M. Fernández-García, C. Serrano, M. L. Cerrada, M. Fernández-García,
6 *Appl. Catal. B*, 2011, **104**, 346-352.
- 7 53 L. Korösi, D. Dömötör, S.Beke, M. Prato, A. Scarpellini, K. Mezker, G. Schneider, T. Kovács,
8 A. Kovács, S. Papp, *Sci. Adv. Mat.*, 2013, **5**, 1184-1192.
- 9 54 M. D'Ischia, O. Crescenzi, A. Pezzella, M. Arzillo, L. Panzella, A. Napolitano, V. Barone,
10 *Photochem. Photobiol.* 2008, **84**, 600-607.
- 11

1 Graphical Abstract

Acetic Acid/Titanium
isopropoxide/Isopropanol
aqueous mixture

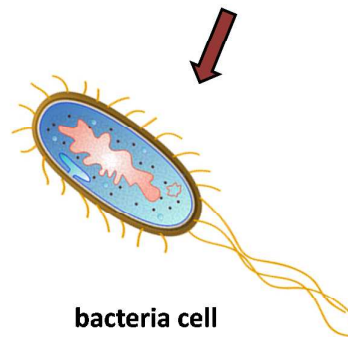
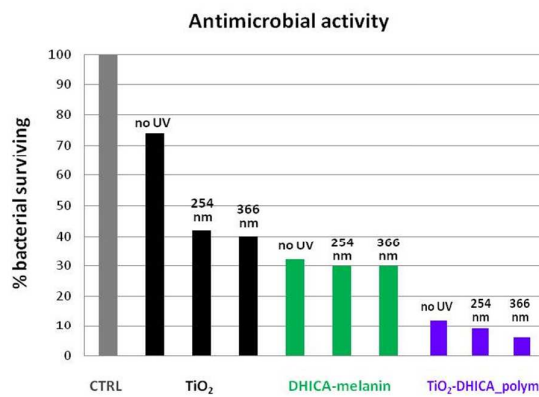
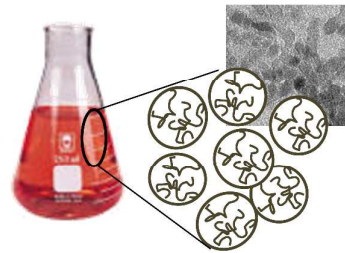


Triethylamine (TEA)

In-situ oxidative
polymerization catalyzed by Ti-sol

under hydrothermal
conditions (120 °C, 24hs)

TiO₂-DHICA_polym NPs



2

3