# Chemical Science

# Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/chemicalscience

### ARTICLE

Received 00th January 20xx,

Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/



cyclisations.<sup>[7]</sup>

# Stereoselective Synthesis of Protected L- and D-Dideoxysugars and Analogues *via* Prins Cyclisations

Ryan J. Beattie, Thomas W. Hornsby, Gemma Craig, M. Carmen Galan\* and Christine L. Willis\*

substrates

A *de novo* approach for the rapid construction of orthogonally protected L- and D-deoxysugars and analogues is described. A novel and robust silicon-acetal undergoes Prins cyclisations with a series of homoallylic alcohols in high yield and excellent stereocontrol. Modified Tamao-Fleming oxidation of the resulting silyltetrahydropyrans gives direct access to deoxyglycoside analogues and the approach was showcased in the synthesis of protected L-oliose, a component of the anticancer agent aclacinomycin A.

Reddy, Yadav and co-workers have used sugar derivatives as

Success of our proposed approach to deoxyglycosides relied

upon use of a suitable electrophile bearing a hydroxyl

surrogate (X in Scheme 1) which would need to be both stable

to the acidic conditions required for the cyclisation and would

readily be converted to a suitable functional group (e.g.

acetate 3) for use in glycosylation reactions. An orthoformate

was considered as the electrophile to directly introduce a 1-O-

alkyl side-chain, but these have rarely been used in Prins

cyclisations and are limited to substrates in which the reaction

A trialkylsilane was considered a suitable hydroxyl surrogate

as, following cyclisation, a Tamao-Fleming oxidation would

lead to the required acetal. Whilst dimethylphenylsilanes have

been widely used,<sup>[10]</sup> Hosomi and co-workers reported the benzyldimethylsilyl group (BDMS) as an attractive alternative that is readily oxidised to alcohols.<sup>[11]</sup> An important criterion

for our synthetic strategy was that the electrophile should be

stable and readily prepared on a synthetically valuable scale.

Thus, novel silyl acetal 5 was prepared in two steps and 76%

overall yield via treatment of 2-lithio-1,3-dithiane<sup>[12]</sup> with

benzyldimethylsilyl chloride (BDMSCI) to give dithiane 4

Prins

in

proceeds via a tertiary carbocation.<sup>[8],[9]</sup>

Scheme 1. Proposed synthetic approach to deoxysugars.

**Results and discussion** 

#### Introduction

Deoxyglycosides are important components of a wide variety of natural products isolated from plants, fungi and bacteria including compounds exhibiting anticancer and antibiotic activities. Some have proved effective for use in the clinic (e.g. the antibiotic vancomycin and the anthracycline antibiotic altromycin B) or as lead compounds to pharmaceuticals.<sup>[1]</sup> In addition, deoxyglycans are also prevalent in bacterial membrane glycoproteins, thus being a viable target for drug discovery and vaccine development.<sup>[2]</sup>

The ability to fully understand and exploit the glycobiology of rare deoxysugars and analogues is hindered by the challenges of isolating pure materials in reasonable quantities from natural sources.<sup>[11]</sup> In addition, synthetic approaches from naturally-abundant carbohydrates often require lengthy synthetic routes which make rare sugars very expensive.<sup>[3]</sup> An alternative and potentially more versatile approach is the *de novo* asymmetric synthesis of deoxy sugars.<sup>[4],[5]</sup> An ideal synthetic strategy would be efficient, robust and readily adapted for the construction of a series of deoxysugars and derivatives. To this end, we have developed a new approach for the enantioselective synthesis of differentially-protected L-and D-deoxyglycosides and analogues *via* Prins cyclisations and its utility exemplified by the synthesis of 2,4- and 2,6-dideoxyglycosides including protected L-oliose.

Prins cyclisations involve acid-mediated reactions of homoallylic alcohols **1** (or derivatives thereof) to form an oxycarbenium ion I which cyclises *via* carbocation II and is subsequently trapped by a nucleophile, giving

School of Chemistry, University of Bristol, Cantock's Close, Bristol BS8 1TS, U.K. E-mail: m.c.aalan@bristol.ac.uk, chris.willis@bristol.ac.uk

This journal is © The Royal Society of Chemistry 20xx

Electronic Supplementary Information (ESI) available: [details of any supplementary information available should be included here]. See DOI: 10.1039/x0xx00000x

tetrahydropyrans 2 with excellent stereocontrol (Scheme 1).<sup>[6]</sup>

followed by mercuric-mediated deprotection in ethanol (Scheme 2).<sup>[13]</sup> The reaction was conducted on a multigram scale and the acetal is stable with no apparent decomposition after 6 months on the bench.



Scheme 2. Synthesis of acetal 5

Initially, the key Prins cyclisation was optimised using the known (*R*)-homoallylic alcohol **6** prepared from dihydrocinnamaldehyde *via* a Nokami crotyl transfer reaction.<sup>[14]</sup> Several methods have been reported for the introduction of oxygen nucleophiles,<sup>[15]</sup> and in this case treatment of alcohol **6** and acetal **5** with trifluoroacetic acid (TFA) at 0  $^{\circ}$ C,<sup>[16]</sup> then hydrolysis of the resultant ester gave alcohol **7** in 97% yield (Scheme 3). A single diastereomer was isolated in which all four substituents were equatorial.

Our ultimate targets, 2,6- and 2,4-dideoxysugar analogues, lack a substituent at C-2 and their synthesis requires a substrate with a terminal alkene. Hence (*R*)-homoallylic alcohol **8** (prepared *via* a Brown allylation<sup>[17]</sup>) was treated with acetal **5** under the optimised reaction conditions to give alcohol **9** in 93% yield. It is known that the mechanism of Prins cyclisations is not simple and, depending on the nature of the substrate and reaction conditions, competing processes may occur.<sup>[18]</sup> To ensure that there was no loss of stereochemical integrity during the cyclisation, the enantiopurity (97.5:2.5 e.r) of **9** was confirmed by chiral SPC.



Scheme 3. Cyclisation of homoallylic alcohols 6 and 8.

An alternative synthetic approach to the silvl tetrahydropyrans was to incorporate the silvl moiety into the alkene coupling partner which on reaction with an aldehyde would enable the facile introduction of a range of side-chains at C-5 of the target deoxysugars (Scheme 4). Several methods were investigated for the synthesis of  $\alpha$ -silyl-homallylic alcohol **10** via acidmediated allylation of acetal 5 (e.g. in the presence of InCl<sub>3</sub>, AgNO<sub>3</sub>, SnCl<sub>2</sub>) but none of the required product was isolated. In contrast, when silyl acetal 5 was treated with allyltributylstannane and LiBF<sub>4</sub> in wet acetonitrile, alcohol 10 was isolated in 61% yield.<sup>[19]</sup> The TFA mediated reaction of 10 with either acetaldehyde or 3-benzyloxypropanal followed by hydrolysis of the resultant ester gave silyltetrahydropyrans 11 and 12 in 84% and 77% overall yields respectively from acetal 5. By varying the reaction conditions the analogous acetates 13 and 14 were readily prepared. Further studies are ongoing to investigate the enantioselective allylation of acetal 5.



Scheme 4. Alternative cyclisation strategy to silyltetrahydropyrans.

The second stage of our synthetic strategy required oxidation of the benzyldimethylsilyl group; Trost and Donohoe have reported the use of tetrabutylammonium fluoride (TBAF) followed by hydrogen peroxide for similar transformations.<sup>[20]</sup> Following detailed investigations we established suitable conditions for the successful oxidation of silane **7** (Scheme 5). It was evident that two steps are involved. First addition of TBAF converted silane **7** to silanol **15** which could be isolated and characterised.<sup>[21]</sup>



Scheme 5. Oxidation of silane 7.

However, it was not necessary to isolate **15** as it was converted *in situ* to hemiacetal **16** *via* a urea hydrogen peroxide oxidation and then directly acetylated to give **17** in 73% yield from silane **7**. It proved vital to keep the temperature of the fluoride activation step in the range 0-5 °C, as at higher temperatures disiloxanes were formed from the condensation of two silanols, which were only slowly oxidised under the reaction conditions.<sup>[22]</sup>

To confirm that the oxidation/acetylation protocol was compatible with different protecting groups commonly used in carbohydrate chemistry, the secondary alcohols in 7 and 9 were converted to acetates (18 and 19) and benzyl ethers (20 and 21) in high yields using standard reaction conditions (Scheme 6). Oxidation of each silane gave the corresponding anomeric acetates (17, 22-24) in 64-80% isolated yield.



Scheme 6. Oxidation of acetoxy and benzyloxy derivatives.

Page 3 of 5

Next the optimised cyclisation/oxidation/acetylation strategy was applied to the preparation of protected 2,4dideoxyglycosides (Scheme 7). Homoallylic alcohol 26 was prepared in 88% overall yield from (S)-glycidol via protection of the alcohol as silvl ether 25 and ring opening of the oxirane with vinylmagnesium and CuCN. Treatment of 26 with silyl acetal **5** and TESOTf in acetic acid<sup>[23]</sup> gave silyltetrahydropyran 27 in 65% yield which was readily converted to diacetate 28 as a 1:2 mixture of anomers using the oxidation/acetylation protocol. Interestingly, treatment of the mixture of homoallylic alcohol 26 and acetal 5 with TFA, our standard cyclisation conditions, gave none of the expected product, instead the analogous ethyl ether 29 was isolated. [24] Some deoxysugars indeed have ethers at C-3, for example, D-oleandrose is a component of the highly potent and selective anticancer agent apoptolidin<sup>[25]</sup> and L-cymarose, found in the DNA-helicase inhibitor, heliquinomycin<sup>[26]</sup> and so this unexpected result has potential significant synthetic value.



Scheme 7. Preparation of 2,4-dideoxysugar 28.

To access orthogonally protected 2,4-dideoxyglycosides, benzyl-protected tetrahydropyran **30** was prepared *via* a similar protection/vinyl addition/cyclisation strategy from (*S*)-glycidol benzyl ether in 71% overall yield (Scheme 8). Oxidation of **30** gave diacetate **31** which subsequently was used to glycosylate cyclohexanol in the presence of BF<sub>3</sub>.OEt<sub>2</sub> giving **32** as exclusively the  $\alpha$ -anomer in 72% yield. The synthetic approach was extended to 2,4-dideoxyglycosides with an axial C-3 oxygenated substituent *via* hydrolysis of acetate **30** and Mitsunobu inversion to give **33** which was oxidised to protected glycoside **34** in 75% yield.



Scheme 8. Synthesis of protected 2,4-dideoxyglycosides 31 and 34.

Next we turned out attention to the synthesis of protected 2,6-dideoxysugars as all diastereoisomers of D- and L-2,6-

dideoxyhexoses have been found in biologically active natural products e.g. D-olivose is a component of angucylcline antibiotic landomycin  $A^{[27]}$  whilst D-digitoxose is present in the steroidal glycoside digitoxin.



Figure 1. Examples of 2,6-dideoxyhexoses.

Initial studies revealed that whilst homoallylic alcohol **35** was readily prepared *via* Brown allylation of allyl ethyl ether,<sup>[28]</sup> reaction of **35** with silyl acetal **5** under our standard TFA conditions gave the 5-membered ring aldehyde **36** in 46% yield (Scheme 9). Aldehyde **36** is likely to be formed *via* a Prinspinacol reaction<sup>[29]</sup> involving oxonia-Cope rearrangement of oxycarbenium ion **III** to enol ether **IV** followed by cyclisation to tetrahydrofuran **V** and finally *O*-alkyl cleavage to generate the carbonyl group.<sup>[30]</sup> Hence to favour formation of a tetrahydropyran over a tetrahydrofuran we reasoned that an electron withdrawing group rather than an ether was required and a carbamate protecting group was selected.<sup>[31]</sup>



Scheme 9. Cyclisation to tetrahydrofuran 36 and nOe correlations.

Thus an asymmetric synthesis of homoallylic alcohol **40** was required which could be readily adapted for both the L- or D-protected 2,6-dideoxysugars since, for example, L-oliose is a component of aclarubicin, clinically used for the treatment of acute leukaemias,<sup>[32]</sup> whilst D-oliose is present in the antitumour drugs mithramycin and chromocyclomycin,<sup>[33]</sup> as well as the HIV-inhibitor durhamycin A.<sup>[34]</sup>



Scheme 10. Enantioselective synthesis of protected L-oliose 42.

Singh and Guiry reported that Sharpless asymmetric epoxidation (SAE) of divinylcarbinol, followed by Mitsunobu inversion of the resulting alcohol gives epoxide **37** (Scheme

This journal is © The Royal Society of Chemistry 20xx

#### ARTICLE

10).<sup>[35]</sup> Importantly, choice of (-)-DIPT or (+)-DIPT in the SAE step allows access to the D- and L- series, respectively. Protection of alcohol **37** with *N*,*N*-diisopropylcarbamoyl chloride gave a mixture of chlorohydrin 38 and the required epoxide 39. Chlorohydrin 38 was readily converted to 39 by treatment with NaOH in THF at room temperature within a matter of minutes. Reductive ring opening of the oxirane with DIBALH gave mono-protected syn allylic diol 40 which cyclised with acetal 5 to give the required tetrahydropyran 41 with an equatorial C-3 alcohol and the axial C-4 protected hydroxyl. Finally oxidation of silane 41 gave protected L-oliose 42 in 77% yield as a 1:1 mixture of anomers. The methodology could be extended to the synthesis of L-olivose via protected alcohol 44 which was prepared from epoxide 43 using the same conditions as for assembly of the diastereomer 40 (Scheme 11). Interestingly, reaction of 44 with acetal 5 under the standard conditions gave a mixture of products due to migration of the carbamoyl group but on reduction of the mixture with LiAlH<sub>4</sub>, diol 45 was isolated in 57% yield. It is possible that neighbouring group participation by the carbamoyl group traps the intermediate carbocation I in the Prins cyclisation giving II and resulting in migration of the carbomyl group but this has not been proven.



Scheme 11. Enantioselective synthesis of L-olivose precursor 45.

#### Conclusions

In conclusion, a *de novo* approach for the rapid construction of a series of orthogonally protected L- and D-dideoxyglycosides and analogues is described from simple starting materials. A stable acetal 5 was prepared in two high yielding steps and used in a series of acid-mediated Prins cyclisations with different homoallylic alcohols to give the corresponding tetrahydropyrans good yield and in excellent diastereoselectivity. These reactions are readily performed on gram scales. A modified Tamao-Fleming oxidation/acetylation protocol gave the target 2,4-dideoxysugars with an acetyl group at the anomeric position. Extending the utility of the new methodology to the synthesis of 2,6-dideoxysugars revealed the importance of the choice of protecting group to avoid formation of tetrahydrofuranals. The enantioselective synthesis of protected L-oliose is described using N,Ndiisopropylcarbamoyl as a protecting group. Silane 41 has potential value for the synthesis of other 2,6-dideoxyhexoses for example methylation of the free hydroxyl group will lead to protected L-diginose while Mitsunobu inversion will give Lboivinose derivatives and subsequent methylation to Lsarmentose and these investigations are ongoing in our laboratories.

#### Acknowledgements

We thank the Bristol Chemical Synthesis Centre for Doctoral Training for the provision of PhD studentships EP/G0367641/1 (RJB, TWH) and EPSRC EP/L001926/1 and EP/J002542/1(MCG).

#### Notes and references

- For reviews see: a) A. C. Weymouth-Wilson, *Nat. Prod. Rep.* 1997, **14**, 99-110; b) Křen, V.; Řezanka, T. *FEMS Microbiol. Rev.* 2008, **32**, 858; c) A. Kirschning, A. F. W. Bechthold, J. Rohr, *Vol.* 188, Springer Berlin / Heidelberg, 1997, pp. 1-84.
- For recent reviews on the importance of oligosaccharides see: a) P. Stallforth, B. Lepenies, A. Adibekian, P. H. Seeberger, J. Med. Chem. 2009, 52, 5561-5577; b) M. C. Galan, D. Benito-Alifonso, G. M. Watt, Org. Biomol. Chem. 2011, 9, 3598-3610; c) M. C. Galan, P. Dumy, O. Renaudet, Chem. Soc. Rev. 2013, 42, 4599-4612.
- 3 Recently some elegant strategies have been reported for the synthesis of D-deoxyglycosides starting from readily available carbohydrates, e.g. M. Emmadi, S. S. Kulkarni, *Org. Biomol. Chem.* 2013, **11**, 3098-3102.
- 4 For selected reviews see: a) A. Kirschning, M. Jesberger, K. U. Schoning, Synthesis-Stuttgart 2001, 507-540; b) R. M. De Lederkremer, C. Marino, in Advances in Carbohydrate Chemistry and Biochemistry, Vol 61, Vol. 61 (Ed.: D. Horton), 2008, pp. 143-216; c) A. Z. Aljahdali, P. Shi, Y. Zhong, G. A. O'Doherty, in Advances in Carbohydrate Chemistry and Biochemistry, Vol 69, (Eds.: D. Horton), 2013, pp. 55-123.
- 5 For selected examples see: a) H. Guo, G. A. O'Doherty, Angew. Chem. Int. Ed. 2007, 46, 5206-5208; b) R. S. Babu, Q. Chen, S.-W. Kang, M. Zhou, G. A. O'Doherty, J. Am. Chem. Soc. 2012, 134, 11952-11955; c) D. Leonori, P. H. Seeberger, Beilstein J. Org. Chem. 2013, 9, 332-341; d) R. Pragani, P. Stallforth, P. H. Seeberger, Org. Lett. 2010, 12, 1624-1627.
- 6 For reviews see: a) I. M. Pastor, M. Yus, *Curr. Org. Chem.* 2007, **11**, 925-957; b) C. Olier, M. Kaafarani, S. Gastaldi, M. P. Bertrand, *Tetrahedron* 2010, **66**, 413-445.
- 7 a) B. V. S. Reddy, A. V. Ganesh, A.s. Krisna, G. G. K. S. N. Kumar, J. S. Yadav, *Tetrahedron Lett.*, 2011, *52*, 3342-3344;
  b) J. S. Yadav, B. V. S. Reddy, A. P. Singh, D. N. Chaya, D. Chatterjee, *Tetrahedron Lett.*, 2010, *51*, 1475-1478.
- 8 a) F. Perron-Sierra, M. A. Promo, V. A. Martin, K. F. Albizati, J. Org. Chem. 1991, 56, 6188-6199; b) G. E. Keck, J. A. Covel, T. Schiff, T. Yu, Org. Lett. 2002, 4, 1189-1192.
- 9 Prins cyclisations have been used to assmble tetrahyropyrans with a hydroxymethyl side-chain which then can be oxidatively cleaved to give lactones, see for example, J. S. Yadav, M. S. Reddy, A. R. Prasad, *Tetrahedron Lett.* 2005, 46, 2133-2136.
- 10 K. Tamao, Proc. Jpn. Acad. Ser. B. 2008, 84, 123-133.
- 11 K. Miura, T. Hondo, T. Takahashi, A. Hosomi, *Tetrahedron Lett.* 2000, **41**, 2129-2132.
- 12 a) E. J. Corey, D. Seebach, R. Freedman, J. Am. Chem. Soc. 1967, 89, 434-436; b) A. G. Brook, J. M. Duff, P. F. Jones, N. R. Davis, J. Am. Chem. Soc. 1967, 89, 431-434
- 13 A. Degl'Innocenti, P. Scafato, A. Capperucci, L. Bartoletti, C. Spezzacatena, R. Ruzziconi, Synlett 1997, 361, 362.
- 14 a) J. Nokami, M. Ohga, H. Nakamoto, T. Matsubara, I. Hussain, K. Kataoka, J. Am. Chem. Soc. 2001, **123**, 9168-9169;
  b) C. S. Barry, N. Bushby, J. R. Harding, C. L. Willis, Org. Lett. 2005, **7**, 2683-2868.
- 15 a) J. J. Jaber, K. Mitsui, S. D. Rychnovsky, J. Org. Chem. 2001,
  66, 4679-4686; b) E. H. Al-Mutairi, S. R. Crosby, J. Darzi, J. R. Harding, R. A. Hughes, C. D. King, T. J. Simpson, R. W. Smith, C. L. Willis, Chem. Commun. 2001, 835-836; c) K. Tadpetch, S. D. Rychnovsky, Org. Lett. 2008, 10, 4839-4842; d) J. S. Yadav,

4 | J. Name., 2012, 00, 1-3

This journal is © The Royal Society of Chemistry 20xx

B. V. Subba Reddy, G. G. K. S. Narayana Kumar, S. Aravind, *Synthesis* 2008, **3**, 395-400; e) J. S. Yadav, B. V. S. Reddy, A. V. Ganesh, G. G. K. S. Narayana Kumar, *Tetrahedron Lett.* 2010, **51**, 2963-2966.

- 16 a) C. S. J. Barry, S. R. Crosby, J. R. Harding, R. A. Hughes, C. D. King, G. D. Parker, C. L. Willis, *Org. Lett.* 2003, **5**, 2429-2432;
  b) C. S. Barry, J. D. Elsworth, P. T. Seden, N. Bushby, J. R. Harding, R. W. Alder, C. L. Willis, *Org. Lett.* 2006, **8**, 3319-3322; c) P.T. Seden, J. P. H. Charmant, C. L. Willis, *Org. Lett.* 2008, **10**, 1637-1640.
- 17 H. C. Brown, P. K. Jadhav, J. Am. Chem. Soc. 1983, 105, 2092-2093.
- S. Marumoto, J. J. Jaber, J. P. Vitale, S. D. Rychnovsky, Org. Lett., 2002, 4, 3919-3922; R. Jasti, S. D. Rychnovsky, J. Am. Chem. Soc., 2006, 128, 13640-13648; C. S. Barry, N. Bushby, J. R. Harding, R. A. Hughes, G. D. Gregory, R. Roe, C. L. Willis, Chem. Comm., 2005, 3727-2729.
- 19 B. H. Lipshutz, D. F. Harvey, F. Synth. Commun. 1982, 12, 267.
- 20 a) B. M. Trost, Z. T. Ball, *J. Am. Chem. Soc.* 2005, **127**, 17644-17655; b) B. M. Trost, M. R. Machacek, Z. T. Ball, *Org. Lett.* 2003, **5**, 1895-1898; c) T. J. Donohoe, P. C. M. Winship, B. S. Pilgrim, D. S. Walter, C. K. A. Callens, *Chem. Commun.* 2010, **46**, 7310-7312.
- 21 a) S. E. Denmark, R. F. Sweis, D. Wehrli, J. Am. Chem. Soc. 2004, **126**, 4865-4875; b) S. E. Denmark, D. Wehrli, J. Y. Choi, Org. Lett. 2000, **2**, 2491-2494.
- 22 E. J. Rayment, N. Summerhill, E. A. Anderson, J. Org. Chem. 2012, **77**, 7052-7060.
- 23 S. K. Woo, M. S. Kwon, E. Lee, Angew. Chem. Int. Ed. 2008, 47, 3242-3244.
- 24 For selected examples where the source of nucleophilic trapping originates from the electrophile, see: a) K. Zheng, X. Liu, S. Qin, M. Xie, L. Lin, C. Hu, X. Feng, J. Am. Chem. Soc. 2012, **134**, 17564-17573; b) J. S. Yadav, B. V. Subba Reddy, G. Mahesh Kumar, C. V. S. R. Murthy, *Tetrahedron Lett.* 2001, **42**, 89-91.
- 25 Y. Hayakawa, J. W. Kim, H. Adachi, K. Shin-ya, K.-i. Fujita, H. Seto, *J. Am. Chem. Soc.* 1998, **120**, 3524-3525.
- 26 M.Brasholz, S. Sörgel, C. Azap, H.-U. Reißig, *Eur. J. Org. Chem.* 2007, **23**, 3801-3814.
- R. T. Crow, B. Rosenbaum, R. Smith Iii, Y. Guo, K. S. Ramos, G. A. Sulikowski, *Bioorg. Med. Chem. Lett.* 1999, **9**, 1663-1666.
- 28 H. C. Brown, P. K. Jadhav, K. S. Bhat, J. Am. Chem. Soc. 1988, 110, 1535-1538.
- 29 Prins-pinacol reactions have been used in the synthesis of tetrahydrofurans, a) N. Hanaki, J. T. Link, D. W. C. MacMillan, L. E. Overman, W. G. Trankle, J. A. Wurster, *Org. Lett.*, 2000, 2, 223-226; b) L. E. Overman, L. D. Pennington *J. Org. Chem.*, 2003, *68*, 7143-7157.
- 30 a) L. D. M. Lolkema, H. Hiemstra, C. Semeyn, W. N. Speckamp, *Tetrahedron* 1994, **50**, 7115-7128; b) L. D. M. Lolkema, C. Semeyn, L. Ashek, H. Hiemstra, W. N. Speckamp, *Tetrahedron* 1994, **50**, 7129-7140.
- 31 Carbamates have been used to good effect in intramolecular Samurai cyclisations, B. Leroy, I. E. Markó, *Tetrahedron Lett.* 2001, **42**, 8685-8688.
- 32 W. Kersten, H. Kersten, W. Szybalski, M. Fiandt, *Biochemistry* 1966, **5**, 236-244.
- 33 H. Jayasuriya, R. B. Lingham, P. Graham, D. Quamina, L. Herranz, O. Genilloud, M. Gagliardi, R. Danzeisen, J. E. Tomassini, D. L. Zink, Z. Q. Guan, S. B. Singh, J. Nat. Prod. 2002, 65, 1091-1095.
- 34 W. Lu, C. Leimkuhler, M. Oberthür, D. Kahne, C. T. Walsh, *Biochemistry* 2004, 43, 4548-4558.
- 35 S. Singh, P. J. Guiry, J. Org. Chem. 2009, 74, 5758-5761.