

# RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

*Accepted Manuscripts* are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



23 **Abstract:**

24 It was found that the ultra-weak chemiluminescence(CL) emission from sodium  
25 bisulfite( $\text{NaHSO}_3$ )- $\text{H}_2\text{O}_2$  system could be enhanced by gold nanoclusters (Au NCs). The  
26 as-prepared Au NCs was applied to the  $\text{NaHSO}_3$ - $\text{H}_2\text{O}_2$  system for the first time. And a  
27 decreased CL was observed in the presence of trypsin. This novel CL system based on Au  
28 NCs- $\text{NaHSO}_3$ - $\text{H}_2\text{O}_2$  was developed for the trypsin determination. Herein, UV-visible  
29 spectroscopy, fluorescence spectra coupled with radical scavengers were used to explore  
30 the possible mechanism. The enhanced CL could be attributed to the catalysis of Au NCs  
31 and the decreased CL should be ascribed to the decomposition of Au NCs. Finally, the  
32 proposed method was successfully utilized to detect trypsin in human urine samples with  
33 good accuracy and precision.

34

35 **Keywords:**

36 Gold nanoclusters; ultra-weak; chemiluminescence; sodium bisulfite; trypsin.

37

38

39

40

41

42

43

44

## 45 1. Introduction

46 Protease is an enzyme that can catalyze the breakdown of proteins, which widely exists  
47 in the animal internal organs, plant leaves, fruits and microorganisms, and plays an  
48 essential and irreplaceable role in biological and physiological processes.<sup>1</sup> As a kind of  
49 protease, trypsin is the most important digestive enzyme in the pancreas zymogen, which  
50 can be used as a dependable and specific diagnostic biomarker for pancreatitis, cystic  
51 fibrosis and cancer.<sup>2-3</sup> Therefore, it is of great importance meaning to detect trypsin in  
52 human metabolic processes.

53 A number of analytical methods have been employed to determine trypsin, such as  
54 fluorescence methods,<sup>4-5</sup> raman,<sup>6</sup> colorimetric,<sup>7,8</sup> electrophoresis,<sup>9</sup> electrochemical.<sup>10-12</sup>  
55 liquid chromatography.<sup>13</sup> However, the fluorometric detection of trypsin usually needs to  
56 interact with a proper fluorescent probe or sensor.<sup>4,5</sup> The electrochemical methods for  
57 trypsin detection require a complicated electrodes preparation procedure.<sup>10-12</sup> Liquid  
58 chromatography methods for trypsin detection possess a high selectivity, while this method  
59 suffers from a tedious sample preparation process. Hence, these conventional methods  
60 maybe require sophisticated instrumentation or complicated operation process.

61 Chemiluminescence (CL) is known as a desirable analytical technique due to its high  
62 sensitivity, wide linear range, low detection and simple instrument.<sup>14-17</sup> CL detection  
63 technology has been applied to the analysis of many substances.<sup>18-23</sup> The traditional  
64 luminescence reagents such as Luminol, TCPO, Ru(bipy)<sub>3</sub><sup>2+</sup>, Acridinium ester were widely  
65 used in many CL systems and applied in analytical chemistry.<sup>24-27</sup> Unfortunately, these

66 reagents are expensive or poisonous to the environment. Thus, it is an attractive research  
67 area to develop a new CL system with relatively green and cheap reagents.

68 In recent years, the potential applications of the ultra-weak CL systems have gradually  
69 received research interest.<sup>28-31</sup> Nevertheless, the development of weak chemiluminescence  
70 was limited because its intensity was not strong enough for detecting demand. Therefore, it  
71 is necessary to find out the ways to increase its sensitivity.

72 Being an intriguing research field, noble metal nanoclusters (NCs), especially Au NCs  
73 have gained great attention because of their remarkable optical properties.<sup>32</sup> Until now, the  
74 applications of Au NCs in analytical fields mainly focused on their fluorescence  
75 properties.<sup>32-33</sup> Hence, it is highly desirable to find that Au NCs have effect on the  
76 ultra-weak CL reaction of  $\text{NaHSO}_3$  and  $\text{H}_2\text{O}_2$ .

77 In this paper, we report that the weak CL emission from  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  was  
78 significantly enhanced by Au NCs. To the best of our knowledge, there is no report about  
79 Au NCs-enhanced CL from  $\text{NaHSO}_3\text{-H}_2\text{O}_2$ . The possible mechanism of enhanced CL  
80 signal was also discussed. In the presence of trypsin, the CL signal greatly decreased due to  
81 the decomposition of Au NCs. Under optimum conditions, the CL intensity was linear with  
82 trypsin concentration, which led to a novel sensing platform based on the system of Au  
83 NCs-enhanced  $\text{NaHSO}_3\text{-H}_2\text{O}_2$ . And then the proposed method has been applied to detect  
84 trypsin in human urine samples with desirable accuracy and precision.

85

## 86 **2. Experimental**

### 87 *2.1 Reagents and materials.*

88 All chemicals and reagents were of analytical grade and used as received without  
89 further purification, and ultrapure water was used throughout. Bovine serum albumin (BSA)  
90 and trypsin were purchased from Sangon Biotech Co., Ltd. (Shanghai, China). 30% (v/v)  
91 H<sub>2</sub>O<sub>2</sub> and nitro blue tetrazolium (NBT) were purchased from Kelong Reagent Co., Chengdu,  
92 China. NaHSO<sub>3</sub> was purchased from Beijing J & K Chemical Co., Ltd. Thiourea and  
93 ascorbic acid (AA) were commercially obtained from Chongqing Chemical Regent  
94 Company (Chongqing, China).

95 Stock solutions of trypsin ( $1.0 \times 10^{-3}$  g mL<sup>-1</sup>) were prepared by dissolving 0.1000g  
96 trypsin in 100mL Tris-HCl buffer (0.05mol L<sup>-1</sup>, PH=8). Trypsin working solution  
97 (containing  $5 \times 10^{-3}$  M CaCl<sub>2</sub>) was prepared by diluting trypsin stock solution with the  
98 previous buffer. Stock solutions of NaHSO<sub>3</sub> (1 mol L<sup>-1</sup>) were prepared by dissolving  
99 5.2030g NaHSO<sub>3</sub> in 50mL ultra-pure water. Working solutions of H<sub>2</sub>O<sub>2</sub> were prepared fresh  
100 daily by dilution of 30% H<sub>2</sub>O<sub>2</sub> with water.

101

## 102 2.2 Apparatus

103 The CL experiments were carried out with a Ultra-Weak Luminescence analyzer (Xi'an  
104 Remax company, Xi'an, China). CL spectra was obtained by a photomultiplier tube  
105 (opened at -900Kv). UV-vis absorption spectra were taken on a Model UV-2550s  
106 Spectrophotometer (Shimadzu, Japan). Fluorescence spectra were obtained by means of  
107 F-4500 spectrofluorophotometer (Hitachi, Japan). Determination of trypsin was performed  
108 based on the net CL intensity of  $\Delta I = I_0 - I_s$ , where  $I_0$  and  $I_s$  denote the CL intensity in the  
109 absence and presence of trypsin, respectively.

### 110 *2.3 Preparation of BSA-Au NCs*

111 BSA modified Au NCs were prepared according to the previous literature.<sup>34</sup> HAuCl<sub>4</sub>  
112 solution (15mL, 10mmol L<sup>-1</sup>, 37°C) introduced into BSA solution (15mL, 50mg mL<sup>-1</sup>,  
113 37°C), stirring for 2min. Then NaOH solution (1.5mL, 1 mol L<sup>-1</sup>) was added to the mixture.  
114 The mixture reaction allowed incubating at 37°C for 24h. The color of the solution  
115 changed from light yellow to light brown, and then to deep brown. The as-prepared Au  
116 NCs were then dialyzed in ultra-pure water for 48h. The final solution was stored at 4°C  
117 for further work.

118

### 119 *2.4 Sample preparation*

120 Human urine sample was analyzed without any pretreatment. The obtained sample  
121 was diluted 10-folds with Tris-HCl buffer (pH=8.0). The above solution was then used for  
122 trypsin detection.

123

### 124 *2.5 CL system based on Au NCs for detection of trypsin*

125 In a typical experiment, 2mL Au NCs was incubated with 500 u L trypsin at 37°C for  
126 40min to destroy BSA-Au NCs and the as-prepared solution was placed to wait for the next  
127 measurement. In a quartz glass cuvette, 100 u L Au NCs solution was first mixed with  
128 200uL NaHSO<sub>3</sub> solution, and then 200uL H<sub>2</sub>O<sub>2</sub> was injected by a syringe. The CL profile  
129 and intensity was captured for a 0.2 s interval with the voltage of -900Kv.

130

## 131 **3. Results and discussion**

### 132 3.1 Enhancement of $\text{NaHSO}_3\text{-H}_2\text{O}_2$ CL

133 In order to evaluate the feasibility of the method, contrast experiment was conducted.  
134 As shown in Fig. 1 (curve a), the weak CL from  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  was recorded. The  
135 enhancement effect of Au NCs on the  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  was studied. As show in curve b, the  
136 CL intensity of  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  could be enhanced remarkably about 11 folds after adding  
137 Au NCs. When trypsin exists in Au NCs-enhanced system, the CL intensity decreased  
138 significantly (curve c).

139

### 140 3.2 Optimization of the reaction conditions

141 The reaction conditions were optimized for the  $\text{NaHSO}_3\text{-H}_2\text{O}_2\text{-Au NCs}$  CL system as  
142 shown in Fig. 2. The effect of the  $\text{NaHSO}_3$  concentration on the CL was examined in the  
143 range from 0.01 to 0.25 mol  $\text{L}^{-1}$ . The result was shown in Fig. 2a. The maximal signal was  
144 obtained at 0.15 mol  $\text{L}^{-1}$ . Therefore, the concentration of 0.15 mol  $\text{L}^{-1}$  was selected for  
145 subsequent investigating.

146 As a basic reaction solution,  $\text{H}_2\text{O}_2$  reacts with  $\text{NaHSO}_3$  to produce CL emission. The  
147 effect of the  $\text{H}_2\text{O}_2$  concentration was investigating over the range 0.3 to 0.6 mol  $\text{L}^{-1}$ . As  
148 shown in Fig. 2b. The CL intensity increased with increasing  $\text{H}_2\text{O}_2$  concentration in the  
149 range of 0.3 to 0.55 mol  $\text{L}^{-1}$ . At the concentration above 0.55 mol  $\text{L}^{-1}$ , there is no obvious  
150 change for CL intensity. Consequently, 0.55 mol  $\text{L}^{-1}$   $\text{H}_2\text{O}_2$  was chosen as the optimal for  
151 further experiments.

152 As a CL enhancer, the effect of Au NCs concentration on the CL was studied in the  
153 range from  $1 \times 10^{-4}$  to  $7.5 \times 10^{-4}$  mol  $\text{L}^{-1}$ . As can be seen from Fig. 2c, the CL signal increased

154 on increasing the concentration of Au NCs from  $1 \times 10^{-4}$  to  $6 \times 10^{-4} \text{ mol L}^{-1}$ . Higher  
155 concentration result in an obvious decrease in CL response. Hence,  $6 \times 10^{-4} \text{ mol L}^{-1}$  Au NCs  
156 was recommended.

157 In order to test the effect of the incubating time on the CL intensity, various incubating  
158 time ranging from 20 to 100min was compared. As shown in Fig. 2d, with the increase of  
159 incubating time, the CL intensity increased sharply and then reached a plateau at 40min.  
160 Thus, the incubating time of 40min was selected in this system.

161

### 162 *3.3 Possible mechanism of CL system*

163 The CL-generation mechanism for  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  system had summarized as the  
164 following steps by Li et al.<sup>35</sup>

165 (1)  $\text{HSO}_3^-$  was oxidized by  $\text{H}_2\text{O}_2$  to produce sulfite radical ( $\bullet\text{SO}_3^-$ ), which then  
166 dimerized to give  $\text{S}_2\text{O}_6^{2-}$  ion.

167 (2) The emitting species( $\text{SO}_2^*$ ), which was generated by the decomposition of  $\text{S}_2\text{O}_6^{2-}$   
168 ion. The emission wavelength of  $\text{SO}_2^*$  is 260–480 nm. Due to the low emission quantum  
169 yield of  $\text{SO}_2^*$ , the CL intensity is very low. Thus, it is an extreme difficulty to capture the  
170 emission wavelength of  $\text{SO}_2^*$ .<sup>36</sup>

171 The UV-visible absorption spectra was conducted in order to confirm the possible  
172 catalysis of Au NCs. As shown in Fig. 3, the maximum absorption peaks of Au NCs is  
173 observed at around 280nm and  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  system at around 204nm. Therefore, the  
174 light absorption of the mixed system was approximately equal to the sum of the light

175 absorption of the two single systems, which suggests that no change was happened between  
176 the species after the reaction.

177 A F-4500 mode fluorescence spectrophotometer has been used to discuss the  
178 mechanism of chemiluminescence based on  $\text{NaHSO}_3\text{-H}_2\text{O}_2\text{-AuNCs}$  system in the presence  
179 of trypsin. As shown in Fig 4a, the fluorescence intensity of Au NCs decreased clearly after  
180 the CL reaction. It was further demonstrated that BSA-Au NCs was not the luminophor of  
181 the CL system. The fluorescence spectra of Au NCs and the mixture of Au NCs-trypsin  
182 were shown in Fig 4b. It can be seen that the fluorescence intensity of Au NCs decreased in  
183 the presence of trypsin, which ascribed to the cleft of BSA molecule from Au NCs.<sup>33</sup> We  
184 presumed that the hydrolysis reaction of trypsin with BSA make Au NCs lost the protective  
185 effect of BSA. Therefore, trypsin can be detected, making use of our desirable method.

186 The mechanism was further discussed by the quenching effect of different reactive  
187 oxygen species (ROS) on the CL system. Ascorbic acid (AA) is well known as an efficient  
188 ROS scavenger, which can terminate active oxygen radicals through electron transfer.<sup>37</sup> AA  
189 at a concentration of  $0.1 \text{ mmol L}^{-1}$  had adverse impact on the CL intensity, the intensity  
190 decreased by a factor of  $\sim 82.5$  ( Table 1). Thus, the CL reaction must happen in a radical  
191 way. NBT was often used for the detection of  $\text{O}_2^{\cdot-}$  radicals, which could be reduced to its  
192 deep blue diformazan form by  $\text{O}_2^{\cdot-}$ .<sup>38</sup> When  $1 \text{ mmol L}^{-1}$  NBT was added to the CL system,  
193 and then the CL intensity decreased by a factor of  $\sim 42.6$ . The result confirmed that  
194  $\text{O}_2^{\cdot-}$  was also intermediate in the CL process. Thiourea is an effective radical scavenger for  
195  $\text{OH}^{\cdot}$ .<sup>39</sup> When  $1 \text{ mmol L}^{-1}$  thiourea was added to CL system, a distinct inhibition is observed  
196 by a factor of  $\sim 57.6$ . It indicated that  $\text{OH}^{\cdot}$  and  $\text{O}_2^{\cdot-}$  were involved in the CL reaction

197 process. It was reported that H<sub>2</sub>O<sub>2</sub> decomposition on supported metal catalysts such as Au  
198 NPs, Ag NPs and CuO NPs involved the formation of hydroxyl radicals OH<sup>•</sup>.<sup>40-42</sup> In the  
199 same manner, we suggested that the O–O bond of H<sub>2</sub>O<sub>2</sub> might be broken up into double  
200 OH<sup>•</sup> radicals by virtue of the catalysis of Au nanoclusters. The OH<sup>•</sup> reacted with <sup>•</sup>SO<sub>3</sub><sup>-</sup> to  
201 form HSO<sub>4</sub><sup>-</sup>, and then HSO<sub>4</sub><sup>-</sup> was transformed to SO<sub>2</sub><sup>\*</sup>. Finally, SO<sub>2</sub><sup>\*</sup> returned back to  
202 ground state with the emission of light. Based on the above results, the whole enhanced  
203 mechanism is summarized in Scheme 1.

204

### 205 *3.4 Analytical performance*

206 Under the optimum conditions described above, CL intensity versus trypsin  
207 concentration shows good linearity ranging from 2.4-48 u g mL<sup>-1</sup> (Fig.5). The regression  
208 equation is  $\Delta I = 3024.9 + 53.94[\text{trypsin}]$  (u g mL<sup>-1</sup>). The linear correlation coefficient is  
209 0.9973, the limit of detection (LOD) for trypsin is 0.19 u g mL<sup>-1</sup>. The relative standard  
210 deviation (RSD) was less than 3% for 100 u g mL<sup>-1</sup> trypsin (n=10).

211

### 212 *3.5 Selectivity of the method to trypsin over other possible background species*

213 The selectivity of the method for the detection trypsin was examined by compare the  
214 CL intensity of trypsin and the mixture of trypsin and background species. As can be seen  
215 in Fig 6, the effect of 100 u g mL<sup>-1</sup> urea, uric acid, glucose, ascorbic acid, dopamine, BSA,  
216 Fe<sup>3+</sup>, NH<sub>4</sub><sup>+</sup>, Mg<sup>2+</sup>, Ca<sup>2+</sup> and I<sup>-</sup> on the detection of 10 u g mL<sup>-1</sup> trypsin. These background  
217 species concentration adopted for interference studies are higher than that in normal urine

218 samples. Most of the interferences have no influence on the determination of trypsin,  
219 indicating that the proposed CL system has good selectivity.

220

### 221 *3.6 Detection of spiked trypsin in human urine*

222 In order to test the applicability and reproducibility of the proposed method, recovery  
223 experiments were performed in human urine samples. These results were shown in Table 2.  
224 Desirable recoveries within the range of 87.5%-104.2% was obtained, showing that this  
225 method has good practicability for trypsin detection.

226

## 227 **4. Conclusion**

228 In this work, a novel method based on Au NCs-enhanced NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub> CL system  
229 was established for trypsin detection. The added Au NCs could increase the sensitivity of  
230 the weak CL system of NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>. In the presence of trypsin, Au NCs was destroyed  
231 because the protein template was enzymatically hydrolyzed, leading to the decreased CL  
232 response. The Au NCs-based method was successfully applied to the detection of trypsin in  
233 human urine samples with satisfactory accuracy and precision. With trypsin being an index  
234 for diseases diagnosis, our method possesses its potential application. Compared with  
235 conventional CL systems, the NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub> CL system is a simple, inexpensive, and  
236 relatively nontoxic. What's more, the study on the ultra-weak CL system of  
237 NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>-AuNCs is a new direction to explore a new CL system with nanocluster as  
238 catalysts.

239

240 **Acknowledgements**

241 This work was supported by science and technology commission foundation of Chongqing  
242 (CSTC, 2010BB8328)

243 We thank Prof. H. Z. Zheng and Prof. Y. M. Huang for measurements.

244

245

246 **Figure captions**

247 **Scheme.1.** Possible mechanism for the Au NCs-NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub> system.

248

249 **Fig.1.** Kinetic curves of CL systems: (curve a) NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>; (curve b) NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>-Au  
250 NCs; (curve c) NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>-Au NCs-trypsin. NaHSO<sub>3</sub>: 0.15 mol L<sup>-1</sup>, H<sub>2</sub>O<sub>2</sub>: 0.55 mol L<sup>-1</sup>,  
251 Au NCs: 6×10<sup>-4</sup> mol L<sup>-1</sup>, trypsin: 50 ug mL<sup>-1</sup>.

252

253 **Fig.2.** Effects of the reaction conditions on the NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>-AuNCs system. (a) Effect of  
254 NaHSO<sub>3</sub> concentration: H<sub>2</sub>O<sub>2</sub>: 0.5 mol L<sup>-1</sup>; Au NCs: 7.5×10<sup>-4</sup> mol L<sup>-1</sup>; Incubating time:  
255 60min. (b) Effect of H<sub>2</sub>O<sub>2</sub> concentration: NaHSO<sub>3</sub>: 0.15 mol L<sup>-1</sup>; AuNCs: 7.5×10<sup>-4</sup> mol L<sup>-1</sup>;  
256 Incubating time: 60min. (c) Effect of Au NCs concentration: NaHSO<sub>3</sub>: 0.15 mol L<sup>-1</sup>; H<sub>2</sub>O<sub>2</sub>:  
257 0.55 mol L<sup>-1</sup>; Incubating time: 60min. (d) Effect of incubating time: NaHSO<sub>3</sub>: 0.15 mol L<sup>-1</sup>;  
258 H<sub>2</sub>O<sub>2</sub>: 0.55 mol L<sup>-1</sup>; Au NCs: 6×10<sup>-4</sup> mol L<sup>-1</sup>.

259

260 **Fig.3.** UV-vis absorption spectra of (a) Au NCs; (b) NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>; (c) NaHSO<sub>3</sub>-H<sub>2</sub>O<sub>2</sub>-Au  
261 NCs.

262

263 **Fig.4.** (a) Fluorescence spectra of BSA-Au NCs before and after the CL reaction. (b)

264 Fluorescence spectra of BSA-Au NCs with and without trypsin.

265

266 **Fig.5.** The calibration curve for trypsin. Error bars represent the standard deviation of three  
267 paralleled measurements.

268

269 **Fig.6.** Relative CL intensity of (a) 10  $\mu\text{g mL}^{-1}$  trypsin (b-l): the mixture of trypsin and  
270 potential background species (100  $\mu\text{g mL}^{-1}$ ). From left to right: urea, uric acid, glucose,  
271 ascorbic acid, dopamine, BSA,  $\text{Fe}^{3+}$ ,  $\text{NH}_4^+$ ,  $\text{Mg}^{2+}$ ,  $\text{Ca}^{2+}$  and  $\Gamma$ . Error bars represent the  
272 standard deviation of three paralleled measurements.

273

274 **Tables:**275 **Table.1** Effect of different radical scavengers on the CL of  $\text{NaHSO}_3\text{-H}_2\text{O}_2$  in the presence  
276 of Au nanoclusters <sup>a</sup>

Scavengers	Intermediates	Concentration	Percent inhibition <sup>b</sup> (%)
Ascorbic acid	$\text{OH}^\cdot$ $\text{O}_2^{\cdot-}$	$0.1\text{mmol L}^{-1}$	82.5
Thiourea	$\text{OH}^\cdot$	$1\text{mmol L}^{-1}$	57.6
NBT	$\text{O}_2^{\cdot-}$	$1\text{mmol L}^{-1}$	42.6
$\text{NaN}_3$	$^1\text{O}_2$	$5\text{mmol L}^{-1}$	8

277

278 <sup>a</sup> Solution condition:  $0.55\text{mol L}^{-1}$   $\text{H}_2\text{O}_2$ ,  $6.0 \times 10^{-4}\text{mol L}^{-1}$  Au NCs,  $0.15\text{mol L}^{-1}$   $\text{NaHSO}_3$ .

279 <sup>b</sup> Average value of three determination.

280

281

282

283

284

285 **Table 2** Detection of spiked trypsin in human urine

286

Samples	Added (u g mL <sup>-1</sup> ) trypsin	Found (u g mL <sup>-1</sup> ) trypsin	Recovery (%) (n=3)
1	2.4	2.1	87.5%
2	7.2	6.9	95.8%
3	14.4	15	104.2%

287

288

289

290

291

292

293

294

295

296

297

298

299

300

301 **References**

- 302 [1] H. Neurath, *Proc. Natl. Acad. Sci.*, 1999, **96**, 10962-10963.
- 303 [2] X. Gao, G. C. Tang, Y. Li and X. G. Su, *Anal. Chim. Acta*, 2012, **743**, 131-136.
- 304 [3] P. Miao, T. Liu, X. X. Li, L. M. Ning, J. Yin and K. Han, *Biosens. Bioelectron.*, 2013, **49**,
- 305 20-24.
- 306 [4] F. F. Zheng, J. F. Wu and G. C. Zhao, *Anal. Methods*, 2012, **4**, 3932.
- 307 [5] K. H. Xu, F. Liu, J. Ma and B. Tang, *Analyst*, 2011, **136**, 1199.
- 308 [6] L. X. Chen, X. L. Fu and J. H. Li, *Nanoscale*, 2013, **5**, 5905.
- 309 [7] W. X. Xue, G. X. Zhang and D. Q. Zhang, *Analyst*, 2011, **136**, 3136.
- 310 [8] J. Kallos, D. Kahn and D. Rizok, *Can. J. Biochem.*, 1964, **42**, 235-241.
- 311 [9] J. Uriel and J. Berges, *Nature*, 1968, **218**, 578-580.
- 312 [10] R. P. Liang, X. C. Tian, P. Qiu and J. D. Qiu, *Anal. Chem.*, 2014, **86**, 9256-9263.
- 313 [11] B. A. Zaccaro and R. M. Crooks, *Anal. Chem.*, 2011, **83**, 1185-1188.
- 314 [12] Y. Chen, J. W. Ding and W. Qin, *Bioelectrochem.*, 2012, **88**, 144-147.
- 315 [13] M. M. Vestling, C. M. Murphy and C. Fenselau, *Anal. Chem.*, 1990, **62**, 2391-2394.
- 316 [14] R. K. Zhang, G. K. Li and Y. F. Hu, *Anal. Chem.*, 2015, **87**, 5649-5655.
- 317 [15] F. Z. Fu, L. G. Ke and Y. F. Hu, *Chin J Anal Chem*, 2015, **43**, 1322-1328.
- 318 [16] E. Wolyniec, J. Karpińska, S. Losiewska, M. Turkowicz, J. Klimczuk and A. Kojlo,
- 319 *Talanta*, 2012, **96**, 223-229.
- 320 [17] Y. Guo and B. X. Li, *Carbon*, 2015, **82**, 459-469.
- 321 [18] W. Y. Li, Q. F. Zhang, H. P. Zhou, J. Chen, Y. X. Li, C. Y. Zhang, and C. Yu, *Anal.*
- 322 *Chem.*, 2015, **87**, 8336-8341.

- 323 [19] H. Chen, R. B. Li, L. Lin, G. S. Guo and J. M. Lin, *Talanta*, 2010, **81**, 1688-1696.
- 324 [20] S. Ahmed, S. Fujii, N. Kishikawa, Y. Ohba, K. Nakashima and N. Kuroda, *J*  
325 *Chromatogr A*, 2006, **1133**, 76-82.
- 326 [21] Y. F. Zhang, J. F. Liu, T. Liu, H. B. Li, Q. W. Xue, R. Li, L. Wang, Q. L. Yue and S. H.  
327 Wang, *Biosens. Bioelectron*, 2016, **77**, 111-115.
- 328 [22] Y. Zhou, H. Chen, N. Ogawa and J. M. Lin, *Journal of Luminescence*, 2011, **131**,  
329 1991-1997.
- 330 [23] Z. Lin, H. Chen and J. M. Lin, *Analyst*, 2013, **138**, 5182.
- 331 [24] Q. L. Zhang, L. Wu, C. Lv and X. Y. Zhang, *J Chromatogr A*, 2012, **1242**, 84-91.
- 332 [25] S. Meseguer-Lioret, C. Molins-Lagua and P. Campins-Falcó, *Anal Chim Acta*, 2005,  
333 **536**, 121-127.
- 334 [26] N. A. Al-Arfaj, *Talanta*, 2004, **62**, 255-263.
- 335 [27] D. Dreveny, C. Klammer, J. Michalowsky and G. Gübitz, *Anal. Chim. Acta*, 1999, **398**,  
336 183-190.
- 337 [28] J. M. Lin and M. Yamada, *Anal. Chem*, 2000, **72**, 1148-1155.
- 338 [29] Y. Zhou, G. W. Xing, H. Chen, N. Ogawa and J. M. Lin, *Talanta*, 2012, **99**, 471-477.
- 339 [30] M. Wang, L. X. Zhao and J. M. Lin, *Luminescence*, 2007, **22**, 182-188.
- 340 [31] Y. D. Liang, J. F. Song and X. F. Yang, *Anal Chim Acta*, 2004, **510**, 21-28.
- 341 [32] Y. Yue, T. Y. Liu, H. W. Li, Z. Y. Liu and Y. Q. Wu, *Nanoscale*, 2012, **4**, 2251.
- 342 [33] L. Z. Hu, S. Han, S. M. Parveen, Y. L. Yuan, L. Zhang and G. B. Xu, *Biosens.*  
343 *Bioelectron*, 2012, **32**, 297-299.
- 344 [34] J. P. Xie, Y. G. Zheng and J. Y. Ying, *J. Am. Chem. Soc*, 2009, **131**, 888-889.

- 345 [35] R. B. Li, T. Kameda, Y. Li, A. Toriba, N. Tang, K. Hayakawa and J. M. Lin, *Talanta*,  
346 2011, **85**, 2711-2714.
- 347 [36] R. B. Li, H. Chen, Y. Li, C. Lu and J. M. Lin, *J. Phys. Chem. A*, 2012, **116**, 2192-  
348 2197.
- 349 [37] H. Dai, X. P. Wu, Y. M. Wang, W. C. Zhou and G. N. Chen, *Electrochim. Acta*, 2008,  
350 **53**, 5113–5117.
- 351 [38] B. H. J. Bielski, G. G. Shiue and S. Bajuk, *J. Phys. Chem*, 1980, **84**, 830–833.
- 352 [39] W. F. Wang, M. N. Schuchmann, H. P. Schuchmann, W. Knolle, J. V. Sonntag and C. V.  
353 Sonntag, *J. Am. Chem. Soc*, 1999, **121**, 238–245.
- 354 [40] Y. X. Li, P. Yang, P. Wang and L. Wang, *Anal Bioanal Chem*, 2007, **387**, 585–592.
- 355 [41] H. Chen, F. Gao, R. He and D. X. Cui, *J. Colloid Interface Sci*, 2007, **315**, 158-163.
- 356 [42] M. J. Chaichi and M. Ehsani, *J Fluoresc*, 2015, **25**, 861–870.

**Figures content:**

1. Fig.1
2. Fig.2 (a)
3. Fig.2 (b)
4. Fig.2 (c)
5. Fig.2 (d)
6. Fig.3
7. Fig.4 (a)
8. Fig.4 (b)
9. Scheme.1
10. Fig.5
11. Fig.6

Fig.1

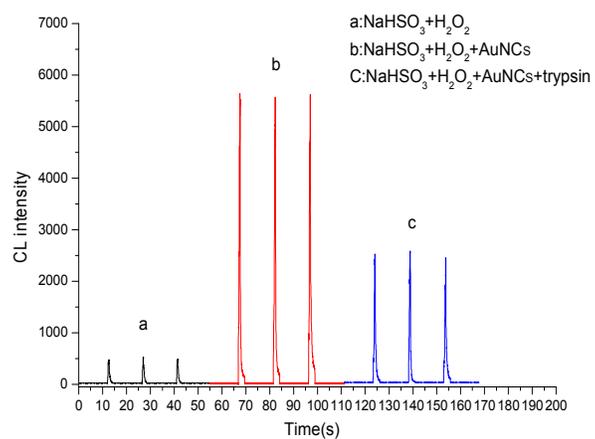


Fig.2(a)

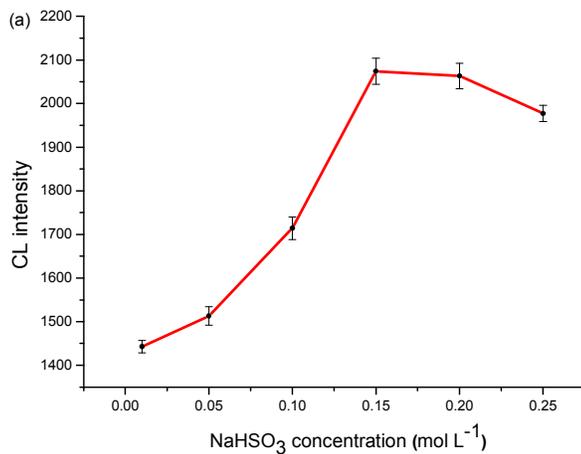


Fig.2(b)

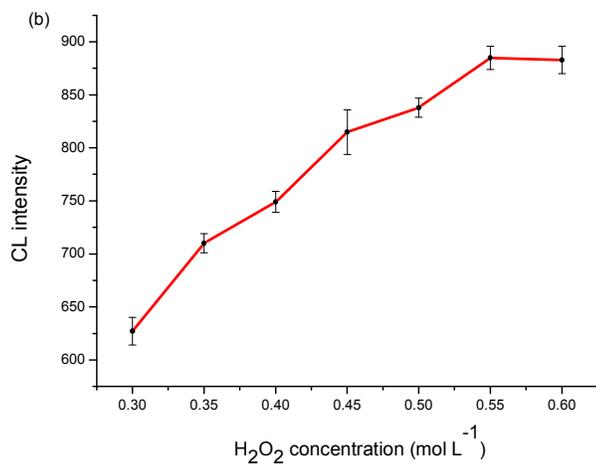


Fig.2(c)

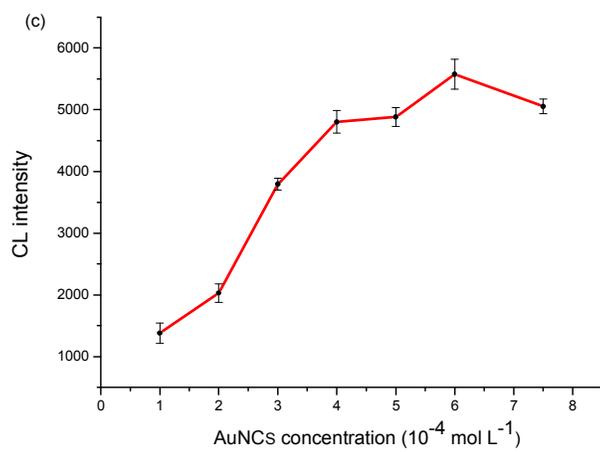


Fig.2(d)

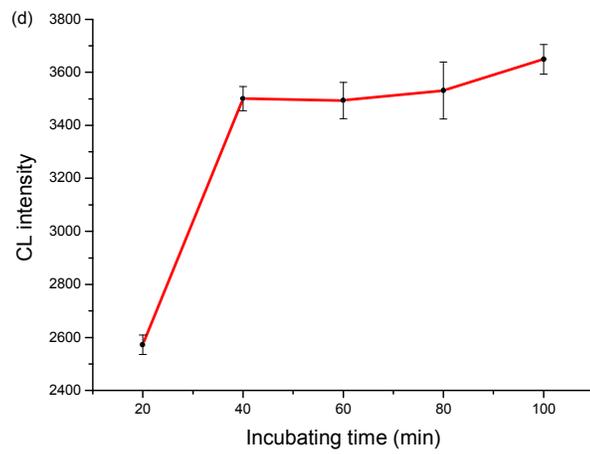


Fig.3

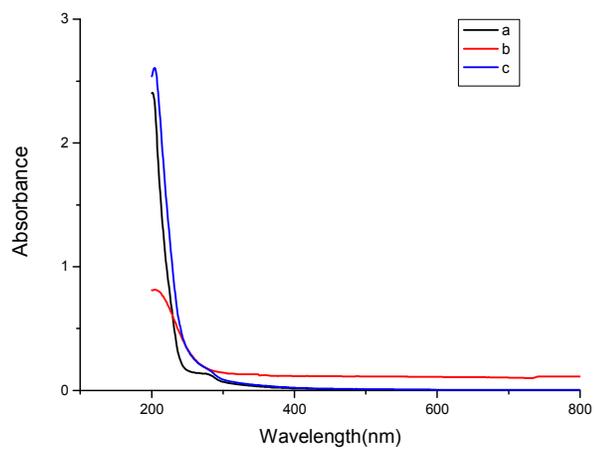


Fig.4(a)

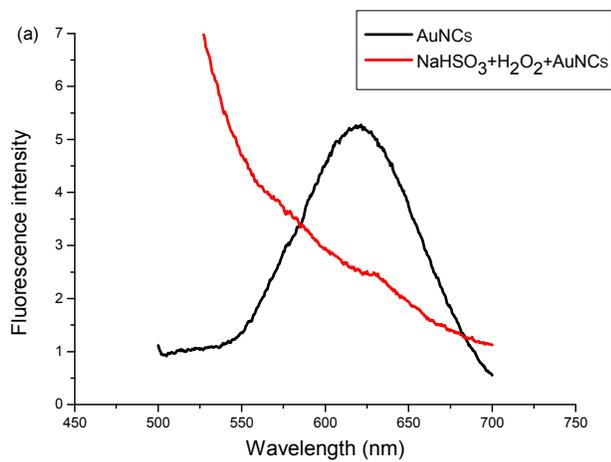
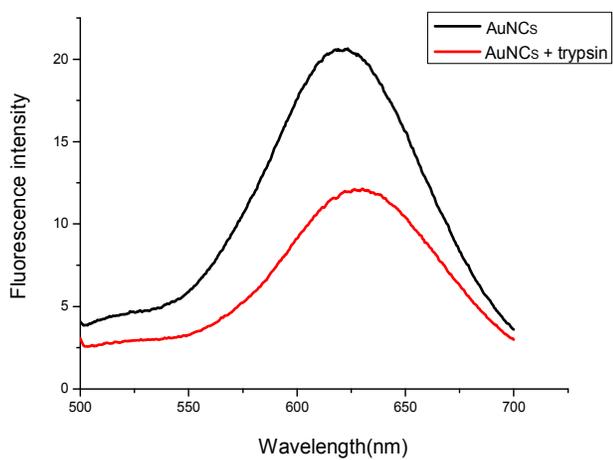


Fig.4(b)



Scheme.1

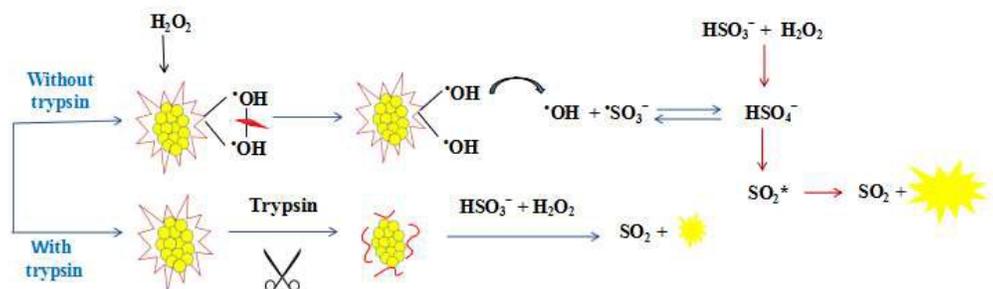


Fig.5

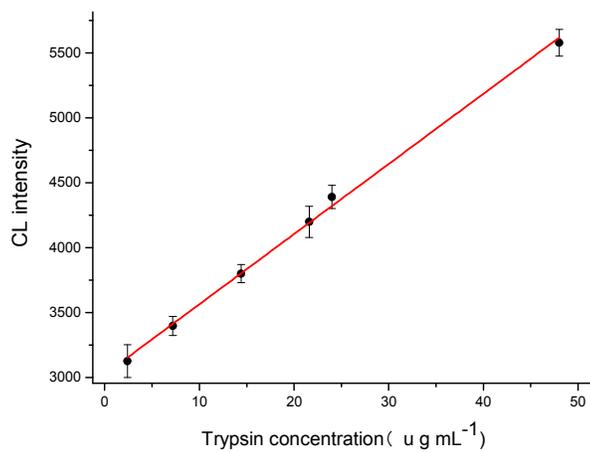


Fig.6

