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Heterogeneous Fenton-like reaction using Fe@ **Bacillus Subtilis**

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A novel heterogeneous catalyst Fe@ Bacillus subtilis has been synthesized though the impregnation method with Iron (III) chloride hexahydrate. Field emission scanning electron microscopy (FE-SEM), energy dispersive spectrometry (EDS), energy dispersive X-ray (EDX) mapping, X-ray photoelectron spectroscopy (XPS) and Fourier transform infrared spectroscopy (FT-IR) were used to characterize the materials. The as-prepared materials were employed as a heterogeneous Fenton's reagent with the addition of H_2O_2 for degradation of Tetracycline Hydrochloride (TC). This new heterogeneous Fenton-like system resulted in a nearly total elimination of TC and a negligible release of iron leaching from the catalyst was achieved. The catalytic performance could be maintained in three consecutive runs without a significant drop. This behavior was attributed to the synergistic structural and functional effect of the combined B. subtilis and iron ions. The FTIR and XPS characterizations of the catalyst before and after Fenton-like reaction showed that the no structural deformation of the particles was occurred.

1 Introduction

2 comprising a significant amount of Antibiotics, 3 pharmaceutical compounds, are widely used in 4 concentrated animal feeding operations around the world to 5 treat the diseases and improve the growth rate of animals¹, ². However, the majority of antibiotics are excreted in forms 6 7 of feces and urine without digestion and metabolism by 8 animals³. The large number of antibiotic residues leads to 9 the emergence of environmental issues, such as the threat to 10 aquatic life, indigenous microbial populations and the risk 11 posed by antibiotic-resistant pathogens etc.⁴. Thus, it is of great significance to develop efficient and cost-effective 12 13 technologies to remove antibiotics from aquatic 14 environment. In practice, the antibiotics residues are hard to 15 be treated by the traditional methods such as physical 16 adsorption and biological degradation due to its 17 antibacterial nature and the difficulty faced on the post-18 treatment of sorbents ⁵.

19 The Advanced Oxidation Processes (AOPs), as a 20 promising method, have presented the predominance to remove the recalcitrant and non-biodegradable compounds 21 22 through the generation of highly reactive radicals (e.g. 23 hydroxyl free radicals) ⁶. In particular, homogeneous 24 Fenton (H_2O_2/Fe^{2+}) and Fenton-like (H_2O_2/Fe^{3+}) systems-25 which are based on ferrous ion and hydrogen peroxide-are 26 proven to be effective technologies for destruction of a 27 large number of hazardous and organic pollutants ⁷⁻¹⁰. It 28 may also be operated at or near room temperature and 29 atmospheric pressure ⁸. Moreover, the oxidant used 30 (hydrogen peroxide) breaks down into environmental friendly species like water and oxygen ⁹. However, some 31 32 significant disadvantages, such as narrow pH range 33 required, the production of iron-containing waste sludge, 34 and high concentration of iron needed for successful 35 mineralization which introduces secondary pollution, 36 impede the large scale application of such technologies at the industrial level ^{10, 11}. In order to overcome these 37 38 disadvantages, much effort has been made to develop 39 heterogeneous Fenton-like catalysts such as iron-containing 40 mesoporous materials with similar catalytic activities as homogeneous Fenton system ¹²⁻¹⁶. Fe-ions have been 41 reported to be immobilized by support materials like 42 membrane¹², C-fabrics¹³, zeolite¹⁴, clays¹⁵ 43 and montmorillonite¹⁶. 44

45 Recently, microbial cells as support materials have 46 attracted burgeoning interests due to the major advantages, 47 including abundant resources, easy accessibility, renewable 48 and environmentally friendly and easy to remove ¹⁷. 49 Bacillus subtilis, known as a member of the genus Bacillus, 50 is an extremely common bacterium found in soil, water, air,

and decomposing plant matter ¹⁸. Due to its abundant 1 2 resources and special physicochemical/biological properties, 3 B. subtilis exhibits an ideal candidate as a microbial cell 4 template for the formation of micro-nano photocatalytic composite materials ¹⁹. Specifically, *B. subtilis* cells has a 5 6 rigid and peritrichous structure cell wall which comprised 7 virtually of 46 % peptidoglycan and 54 % teichoic acids 8 and is abundant in functional groups like hydroxyl, acylamino, carboxyl and amino groups ²⁰. Therefore, such 9 10 coexistence of peritrichous structure and the hydrophilic 11 functional groups in the framework of B. subtilis cell wall 12 provides plentiful absorption sites and excellent 13 absorbability, which makes a good reason to utilize B. subtilis as a support to synthesize micro-nano 14 photocatalytic composite materials ^{17, 21}. However, no one 15 16 has reported the synthesis of Fe-catalyst materials with B. 17 subtilis cells as support material. Therefore, based on what 18 has been illustrated above, we firstly present a Fe(a) B. 19 subtilis composite material loaded with Fe-ions as an 20 immobilized catalyst being applied to decompose 21 antibiotics by Fenton-like reactions. The as-prepared Fe@ 22 B. subtilis samples were characterized using FE-SEM, EDS, 23 EDX, FT-IR and XPS, respectively. Tetracycline 24 hydrochloride (TC) was selected as an object pollutant to 25 examine the catalytic performances of the designed Fe@ B. 26 subtilis catalyst.

27 Experimental

28 Materials preparation

29 In our works, Fe@B. subtilis composite was firstly 30 synthesized through the impregnation method using Iron (III) 31 chloride hexahydrate (FeCl₃ 6H₂O) as a single iron source and 32 B. subtilis as support of the active phase. Briefly, 1.000 g of B. 33 subtilis powder was washed with distilled water and ethanol for 34 three times. Then, the washed B. subtilis was wet impregnated at 35 room temperature using 50 mL saturated iron chloride solution 36 as precursor with the help of magnetic stirring for 1h. After that, 37 the above mixture was left for 24.0 h at ambient temperature and 38 pressure to form the Fe@ B. subtilis composite. The resulting 39 particles were collected by centrifugation from the mixture, 40 followed by two cycles of distilled water rinsing. The obtained 41 Fe@ B. subtilis particles were dried in air at 50 °C for 6 h and 42 stored in a sealed bottle for further use.

43 Materials characterization

44 Several techniques were used to characterize the composite 45 microspheres. After sputtering Pt onto the samples, the 46 morphology of the sample was examined by fieldemission 47 scanning electromicroscopy (FE-SEM, JEOL-6300F). A 48 detailed composition characterization was carried out by energy 49 dispersive spectroscopy (EDS) analysis and energy dispersive 50 X-ray (EDX) mapping. Fourier-transform infrared (FT-IR) 51 spectra of samples were recorded on a Bio-Rad FTS135 52 spectrometer in the range 400-4000 cm⁻¹ using a KBr wafer technique. X-ray diffraction (XRD) patterns were conducted on
X. Pert Pro diffractometer using Cu Kα radiation (λ = 0.15418
nm) at a scanning rate of 10°per min to investigate the crystal
structures and the phase compositions of the samples. The X-ray
photoelectron spectroscopy (XPS) spectra were obtained with an
ESCALab220i-XL electron spectrometer (VG Scientific) using
300 W Al-Ka radiation.

60 Catalytic performance

61 Catalytic experiments were performed in a batch glass reactor 62 with semi-batch operation mode, at atmospheric pressure and 63 ambient temperature, under continuous stirring. In a typical run, 64 the samples were introduced into the reactor, loading with 100 mL 25 mg L⁻¹ of TC solution, with magnetic stirring to maintain 65 66 a uniform suspension. After allowing 160 min for the 67 adsorption/desorption of TC to reach equilibrium, 2 mL H₂O₂ 68 (30%) was added into the reactor and time logged. Samplings 69 were taken at regular intervals during the reaction and analyzed 70 immediately after centrifugation to remove suspended particles. 71 A UV-vis Spectrophotometer ((UV-752, Shanghai)) was used to 72 follow the TC concentration histories, i.e., concentration 73 evolution along reaction time, at $\lambda_{max} = 356$ nm.

74 In this work, the effect of hydrogen peroxide dose to TC 75 degradation has also been investigated and experiments were 76 conducted, applying an aqueous TC solution (100 mL in volume 77 containing 30 mg L^{-1} of TC). The catalyst and H_2O_2 were added 78 to the reaction system at the start of each run.

79 Stability and reusability

80 To test the stability and reusability of the Fe@ *B. subtilis*81 catalyst, three runs were conducted with the same reaction
82 conditions. The used Fe@ *B. subtilis* catalyst was removed, and
83 then washed before each runs. Meanwhile, the conversions of
84 TC as well as iron leaching concentration in the solution were
85 monitored.

86 Settling performance

87 At room temperature 50.0 mg Fe@ *B. subtilis* samples, *B. subtilis* and Iron chloride hexahydrate were dispersed into 89 50 mL of distilled water without any additional addictive in 90 a vertical cylindrical burette, respectively. The falling 91 height was determined at regular time intervals. The 92 sedimentation ratio (SR) was measured by: 93 $SR = \frac{a}{100\%} \times 100\%$ (1)

$$SR = \frac{a}{a+b} \times 100\%$$
(1)
s the length of the clear fluid and b the length of the

a is the length of the clear fluid and b the length of theturbid fluid, respectively.

96 Results and discussion

97 Characterization

98 Figure 1shows the SEM images of the naked *B. subtilis* and
99 Fe@ *B. subtilis* composite particles. In Figure 1a and b, the
100 primitive *B. subtilis* cells were ordered rod-shaped with the

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1 length of approximately $1.4 \pm 0.2 \ \mu m$ and width of $0.6 \pm 0.1 \ \mu m$ 2 (Figure S1). From Figure 1c and d, the Fe@ B. subtilis 3 composite particles maintained the shape of the primitive B. 4 subtilis cores and possessed relatively better monodispersity. 5 The insert images in Figure 1b and 1d presented the EDS 6 analysis of B. subtilis and Fe@ B. subtilis composite particles. 7 The detection of Fe element (the insert image in Figure 1d) 8 obviously provided an evidence of the existence of iron ions on 9 the surface of the B. subtilis cells. By comparing the weight 10 concentration of C, N, O, S, Cl and Fe in Table 1(Table S1), the 11 C, N, O and S weight concentration in the Fe@ B. subtilis 12 decreased, which might be attributed to the attachment of ferric

13 trichloride onto the surfaces of *B. subtilis*.

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Fig. 1 SEM images and EDS spectrum of naked *B. subtilis* (a, b)
17 and Fe@*B. subtilis* (c, d).

18 The homogeneity of chemical composition was 19 investigated by two-dimensional X-ray mapping of selected 20 Fe@ B. subtilis zones (Figure 2). The clear C, O and Fe 21 elemental mapping images of Fe@ B. subtilis in Figure 2(b-22 d) indicated the homogeneous dispersions of C, O, and Fe 23 elements on the surface of B. subtilis support. Moreover, 24 the elaborative observation of Figure 2d confirmed that iron 25 ions were almost uniformly deposited on the B. subtilis 26 surface.

27 Figure 3 exhibited the FT-IR spectra of original B. subtilis, 28 and Fe@ B. subtilis composite particles. For B. subtilis, the 29 broad and intense peak located at about 3410 cm⁻¹ could be 30 assignable to the -OH stretching vibration. The peaks of amide 31 groups in *B. subtilis* protein can be observed at 1650 cm⁻¹and 1542 cm^{-1 22}. The adsorption bands around 2924 and 2852 cm⁻¹ 32 33 can be attributed to the -CH₂- asymmetric and symmetric 34 stretching. In addition, the characteristic bands at 1070 cm⁻¹, 35 1230 and 1382 were assigned to the stretching vibration of C-O group, C-O-C vibration in cyclic ether moieties and O-H 36 vibration in carboxyl acid, respectively ²³⁻²⁵. As for Fe@ B. 37 38 subtilis in Figure 3a, the characteristic absorption peaks at 3400, 39 2924, 2852, 1650, 1542, 1382, 1230 cm⁻¹ decreased and shifted 40 dramaticly, compared with the spectrum of original B. subtilis, 41 implying that the functional groups on the *B. subtilis* have some 42 kind of interaction with the iron ions.



Fig. 2 Selected zones of Fe@*B. subtilis* samples (a) and corresponding X-ray mapping, (b) for C, (c) O, and (d) Fe elements.

49 XRD spectra of the bare B. subtilis and Fe@Bacillus subtilis 50 composite particles were exhibited in Fig. 3b. As depicted Fig. 51 3b, the broad peak around $2\theta=20^{\circ}$ was attributed to the 52 amorphous phase of B. subtilis. After anchoring the iron ions on 53 the Bacillus subtilis, the broad peak almost disappear. The 54 surface element compositions and the valence states of the Fe(a)55 B. subtilis catalyst were revealed by XPS. Figure 3c shows the 56 fully scanned spectra in the range of 200-800 eV. As seen in 57 Figure 3c, the wide scan spectra of the Fe@B. subtilis exhibited 58 photoelectron lines at binding energies of ~200, 285, 399, 530 59 and 711 eV which are ascribed to Cl 2p, C 1s, N 1s, O 1s and 60 Fe2p, respectively. The binding energy peaks at 710.9 and 724.5 61 eV in the high resolution Fe 2p scan (Figure 3d) correspond to Fe 2p3/2 and Fe 2p1/2, respectively ²⁶. The peak (719.1 eV) 62 63 located approximately 8 eV higher than the main Fe 2p3/2 peak was the satellite peak of Fe 2p3/2, indicating the ionic state of 64 $Fe^{3+27,28}$. Moreover, the elemental concentration in atomic % 65 (at. %) of C, N, O, Cl and Fe determined by relative sensitivity 66 67 factors (RSFs) and the spectra intensities were 76.4, 1.88, 16.7, 68 2.86 and 2.24 %, respectively (Table S2).

69 The results obtained by N2 adsorption/desorption 70 measurement (77 K) for Bacillus subtilis and Fe@B. 71 subtilis are depicted in Fig. 4. All the isotherms exhibited a 72 type IV pattern according to the International Union of Pure 73 and Applied Chemistry (IUPAC) classification, which 74 corresponds to mesoporous materials, and this can also be 75 affirmed by the pore size distribution (Fig. 4 inset). Their 76 textural properties such as surface area, pore size are 77 summarized in Table S3. The surface areas for Bacillus 78 subtilis and Fe@ B. subtilis were 4.94 and 6.95 m² g⁻¹, 79 respectively. The Barrett-Joyner-Halenda (BJH) pore size 80 distribution (Fig.4 inset, using the desorption branches) 81 analyses revealed that the average pore sizes of Bacillus 82 subtilis and Fe@B. subtilis were 5.9 and 6.1 nm, 83 respectively, and were in the mesoporous range of 2-50 nm according to the IUPAC classification ^{29, 30}. 84

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1 Settling test

2 The sedimentation performance of Fe@ B. subtilis in 3 aqueous solutions was also evaluated to demonstrate the unique 4 features of Fe@ B. subtilis in comparison with B. subtilis and 5 iron ions. It can be clearly observed in Fig. 5 that the suspension 6 ability of Fe@ B. subtilis (56 %) is better than B. subtilis (13%) 7 and iron ions (nearly zero). It was also observed in the 8 experimental process that slight agitation or vibration could 9 make the settled Fe@ B. subtilis re-suspend in aqueous solution. 10 Therefore, in practical applications, lower stirring speeds may 11 ensure the full contact between the Fe@ B. subtilis and the 12 contaminants. Moreover, one of the main merits of applying 13 heterogeneous catalyst is to promote the separation operation. 14 Thus, the relatively large particles in suspensions can allow easy 15 separation and reuse of the catalyst.





18 Binding Energy (eV)
19 Fig.3 (a) FT-IR spectra and (b) XRD patterns of *B. subtilis* and
20 Fe@*B. subtilis;* (c) wide scan XPS spectra and (d) high
21 resolution Fe 2p spectra of Fe@*B. subtilis.*

22 Catalytic performance

23 Fenton-like reaction based on ferric ions and H₂O₂ has 24 been reported as an effective method to degrade organic 25 pollutants by oxidation in aqueous solutions. Iron-26 containing materials are commonly used as catalysts for 27 Fenton-like reactions and have shown relatively high activities ^{13, 15, 31}. In this study, Fenton-like reactions were 28 29 carried out for tetracycline hydrochloride degradation, the 30 antibiotic second highest in production and use, to test the 31 activities of the prepared Fe@B. subtilis. No acid or base 32 solutions were used to adjust the pH value of the reaction 33 system and all experiments were carried out in the dark to 34 avoid the effect of light.

As shown in Fig. 6, before the addition of H_2O_2 , the self-degradation of the tetracycline hydrochloride is negligible and so did iron ions. *B. subtilis* and Fe@B. subtilis showed certain ability to remove nearly 36.0 and 46.8 % of tetracycline hydrochloride concentration due to the synthetic effect of textural properties, surface functional groups and electrostatic attraction. Firstly, based on the FT- 42 IR spectrum, the proved surface functional groups on the 43 Bacillus subtilis and Fe@ B. subtilis may bond with the 44 tetracycline hydrochloride molecules. Secondly, the 45 experimental pH (~6.0) was between the pKa1 (3.3) and 46 pKa2 (7.7) of tetracycline hydrochloride. At this pH value, 47 TC would be partially in its cationic form, which is 48 important for the electrostatic attraction since both Bacillus 49 subtilis and Fe@B. subtilis showed negatively charge at 50 this pH value (as illustrated in Fig. S2). In the presence of 2 51 mL of H₂O₂, the oxidation of tetracycline hydrochloride in 52 parallel system was still negligible, whereas the iron ions 53 system exhibited certain ability to decrease the tetracycline 54 hydrochloride concentration during the first 30 min, as 55 nearly 44% concentration decrease was observed. However, 56 the concentration of tetracycline hydrochloride remained 57 almost unchanged from then on, indicating that the 58 concentration decrease was due to the Fenton-like oxidation 59 of tetracycline hydrochloride molecules initiate by ferric 60 ions. This phenomenon also demonstrated that the 61 oxidation ability in the iron ions system can only maintain a 62 short time and thus showed a restricted application.

63 Moreover, nearly 93.5% of tetracycline hydrochloride 64 was removed by Fe@ B. subtilis. The great progress 65 happened to Fe@ B. subtilis compared to bare B. subtilis 66 and iron ions can be attributed to the integration of 67 adsorption by the B. subtilis bodies with Fenton-like 68 oxidation by the ferric ions attached on the B. subtilis 69 surface. Specifically, B. subtilis can adsorb the TC 70 molecules from the bulk solution and enrich them on the 71 surface of the catalyst, resulting in a higher reactant 72 concentration, which was similar to the role of mesoporous SiO₂ shell reported by Cui et al. ³². Moreover, the removal 73 74 of TC via adsorption of the B. subtilis surface is preserved 75 by keeping the adsorption sites unsaturated through the 76 decomposition of the molecules by Fe³⁺/H₂O₂ Fenton-like 77 system, which increases the active sites for generation of 78 hydroxyl (HO·) and perhydroxy (HOO·). Li et al. have 79 investigated the radical intermediates involved in the 80 Fenton reaction system by using radical scavengers ³³. 81 Their experiment indicated that hydroxyl (HO) and are active radical 82 perhydroxy (HOO·) radicals 83 intermediates involved in the reaction, but they are not the 84 only species. The singlet oxygen ¹O₂ produced from 85 HOO and HO is directly participated in the degradation of organics. Thus, the relevant radical reactions can be 86 proposed as shown in Eqs. $(2)-(9)^{33,34}$. 87

96 Simultaneously, the recovered adsorption sites provide97 a durative supply of TC molecule for Fenton-like system.98 All in all, the combination of both adsorption and Fenton-

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like reaction could be regarded as cleaner, greener, favored, and promising technology for removing organics from

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water.



6 Relative pressure P/P_0 7 Fig.4 N₂ adsorption/desorption Isotherms and BJH pore 8 size distributions (inset) of (a) Bacillus subtilis and (b) 9 Fe²⁺@B. subtilis. 10

11 Effect of H₂O₂ dosage

12 Hydrogen peroxide concentration is an important 13 parameter for the degradation of the organic pollutants in 14 the heterogeneous Fenton-like reaction system. Fig. 7a 15 shows the removal efficiency of tetracycline hydrochloride 16 as a function of the H₂O₂ (30%) dosage in the solution and 17 the results showed that 2mL was the best dosage of H₂O₂ 18 assayed when the initial concentration of TC was 30mgL⁻¹ and the amount of catalyst 0.5 g L^{-1} . The reaction went 19 20 more slowly when the concentration was lower (1 mL) or 21 higher (5 mL). At low concentration, H₂O₂ cannot generate 22 enough HO· radicals and the oxidation rate is logically slow ^{14, 33}. The increase of the oxidant dosage from 1 to 2 23 24 mL leads to an increase in the reaction rate, as expected, 25 because more radicals formed. Nevertheless, for a relatively 26 higher H₂O₂ dosage (5 mL), the performance decreases. 27 This is because OH radicals efficiently react with excess 28 hydrogen peroxide in reaction system (Eq. (4)), which contributes to the ·OH-scavenging ³⁵. It is noteworthy that 29 30 Eq.4 shows the generation of another radical HO_2 , but its 31 oxidation ability is significantly lower than that of the OH 32 radicals, leading to negligible contribution to the organic degradation ³⁶. As a consequence, an optimum value of 33 34 about 2mL for the hydrogen peroxide dosage was obtained 35 when the initial concentration of the 36 tetracycline hydrochloride was 30 mg L^{-1} .

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41 **Fig.6** TC removal by self-photolysis, bare *B. subtilis*, ferric 42 **Fig.6** TC removal by self-photolysis, bare *B. subtilis*, ferric 43 ion (0.003mM) and Fe@*B. subtilis*(0.5g/L) (inset: chemical 44 structure of TC). Reaction conditions: initial concentration 45 of TC, 25mg L⁻¹; H_2O_2 2mL, room temperature, agitation 46 speed 130 rpm.

47 Iron leaching and stability

48 At the termination of each experimental run under optimum 49 H₂O₂ concentration, the supernatant of the reaction solution was 50 analyzed by atomic absorption spectroscopy and the equilibrium 51 solubility of iron in the solution is about 2 mg L^{-1} which 52 represents 2% of the total amount of iron loaded on the Fe@ B. 53 subtilis catalyst (Fig.7b). Such a low extent of iron release may be regarded as the iron resistance to the leaching process ³⁵. 54 55 Apart from the catalytic activity of the catalyst in the 56 heterogeneous Fenton-like process, another important property 57 is its long-term stability. To test the long-term stability of the 58 catalyst, Fe@ B. subtilis was re-collected by filtration from the 59 reaction solution, washed with distilled water and then tested 60 again under the same reaction conditions. The results showed 61 that the catalytic behaviour of Fe@ B. subtilis could be 62 maintained in three consecutive runs without a significant drop 63 (96.0~85%) in the degradation efficiency (Fig.7b). Besides, as 64 the pH variation depicted in Fig. 7b, the solution showed a 65 decrease of the initial TC aqueous solution pH (~6.0), which 66 was normally attributed to the formation of low molecular 67 weight organic acid ³⁷.

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2 Fig.7 (a) Effect of H_2O_2 concentrations on the removal of TC. 3 Reaction conditions: initial concentration of TC, $30 \text{mg } \text{L}^{-1}$; catalyst dosage 0.5 g L^{-1} , room temperature, agitation speed 130 4 5 rpm. (b) Iron leaching, stability and pH variation test. Reaction conditions: initial concentration of TC, 30mg L⁻¹; catalyst 6 dosage 0.5 g L⁻¹, H₂O₂ addition, 2 mL; room temperature, 7 8 agitation speed 130 rpm. (c) FT-IR spectra of Fe@B. subtilis 9 composites before and after Fenton-like reaction; (d) XPS 10 spectrum of iron after Fenton-like reaction.

11 In addition, FT-IR spectra of the catalyst before and 12 after the three consecutive runs were recorded (Fig. 7c). It 13 could be seen that there was no obvious change in the 14 chemical structure of the Fe@B. subtilis before and after 15 the treatment process. To investigate the structure of the 16 catalyst after the Fenton reaction, XPS was obtained after 3 17 repeated reuses (Fig. 7d). This result indicates the presence 18 of two oxidation states for the surface iron species after 19 being used. The Fe $2p_{3/2}$ (Fe $2p_{1/2}$) spectrum was resolved into two peaks at 710.9 and 712.9 eV (724.5 and 726.9 eV), 20 21 which compared well with the Fe²⁺ octahedral species and Fe³⁺ tetrahedral species ^{27, 33, 38}. This indicated that part of 22 Fe³⁺ in the outermost layer of the catalyst was deoxidized 23 into Fe²⁺ during the Fenton reaction, which was in 24 25 accordance with reaction mechanism illustrated in Eq.(2).

26 27

28 Conclusions

29 For the first time, the heterogeneous catalytic oxidation 30 of Tetracycline Hydrochloride, using an iron-impregnated B. 31 subtilis as the catalyst, was investigated. EDS, EDX and 32 XPS was evidenced that the iron in the samples was ferric 33 ions. The as-prepared Fe@ B. subtilis showed excellent catalytic performance in TC degradation, and the optimal 34 35 dosage of H₂O₂ was 2 mL under the reaction conditions. 36 The Fe@ B. subtilis exhibited good reusability and stability 37 because its catalytic performance could be maintained in 38 three consecutive runs with a slight drop from 96% to 88%. 39 The FTIR and XPS characterizations of the catalyst before 40 and after Fenton-like reaction showed that the no structural 41 deformation of the particles was occurred. The catalyst used

42 has evidenced no short term decrease of activity, and was
43 one of the first being reported for application in
44 heterogeneous Fenton-like degradation of TC. Moreover,
45 the low iron leaching will not lead to the iron-containing
46 waste sludge, which might be a great progress in the
47 industrial application.

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60 Notes and references

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Graphical Abstract



In this work, a novel heterogeneous catalyst Fe@*Bacillus subtilis* was firstly fabricated via a simple impregnation method and its catalytic efficiency was examined for removal of Tetracycline Hydrochloride.