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## Hydrothermal synthesis of defective TiO2 nanoparticles for long-wavelength visible light-photocatalytic killing of cancer cells

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Defective TiO<sub>2</sub> nanoparticles (*d*-TiO<sub>2</sub> NPs) were successfully synthesized using a simple hydrothermal technique through the formation of liposome-TiO<sub>2</sub> composites using P25 TiO<sub>2</sub> powder as the starting material. The morphology and vibrational structures of the *d*-TiO<sub>2</sub> NPs were characterized by various techniques, including X-ray diffraction, transmission electron microscopy, and Fourier transform-infrared and Raman spectroscopic analyses. The synthesized *d*-TiO<sub>2</sub> NPs were composed of both the rutile and anatase phases of TiO<sub>2</sub> with a diameter of approximately 25 nm, and they exhibited absorption of broad visible light including near-infrared wavelengths, from 400 nm to 1000 nm, which was confirmed by diffuse reflectance spectroscopy and X-ray photoelectron spectroscopy. The agglomeration size of the *d*-TiO<sub>2</sub> NPs in aqueous solution was lesser (224 nm) than that of P25 TiO<sub>2</sub> (1440 nm). Therefore, the synthesized *d*-TiO<sub>2</sub> NPs could more easily infiltrate the membrane and cytoplasm of cancer cells *via* endocytosis than could P25 TiO<sub>2</sub>. Consequently, upon irradiation with long-wavelength visible light (400-800 nm), the *d*-TiO<sub>2</sub> NPs could generate reactive oxygen species, including singlet oxygen, leading to the destruction of cancer cells. These results suggest that the newly fabricated *d*-TiO<sub>2</sub> NPs are promising nanomaterials for future *in vivo* photodynamic therapy of cancer.

their photoactivity under visible-light excitation. Nevertheless, the doping

materials may have adverse effects on biomolecules due to their toxicity, and the doped  $TiO_2$  NPs absorb only a short range of visible light (400-600

nm), which has limited penetration distance in the tissue. The NIR range

(700 ~1000 nm) light shows the maximum penetration of light into tissues.<sup>18</sup>

In addition to light responses, determining the appropriate size and

morphologies of TiO2 NPs is an important prerequisite for their efficient

infiltration into biological cells,<sup>19</sup> because conventional TiO<sub>2</sub> NPs are easily

aggregated in an aqueous environment. Thus, it is necessary to develop

longer-wavelength visible light (up to the NIR range)-sensitive TiO<sub>2</sub>NPs with

In recent years, many researchers have attempted a defect-engineering

method with an electron beam (EB) to avoid the use of hazardous

dopants.<sup>20-22</sup> EB irradiations have been applied to form TiO<sub>2</sub> NPs containing

defect sites such as oxygen vacancies, which can lead to the creation of

unpaired electrons or  $Ti^{3+}$  centers. These  $Ti^{3+}$  defects can form a shallow

donor level just below the conduction band, which could facilitate both

visible light and NIR responses.<sup>23-24</sup> Thus, owing to efficient interfacial

electron transfer, EB-generated defective TiO<sub>2</sub> NPs (d-TiO<sub>2</sub> NPs) have been

successfully applied to induce visible light-driven photocatalytic activity for water splitting and CO<sub>2</sub> reduction.<sup>21</sup> However, *d*-TiO<sub>2</sub> NPs have not yet been employed for biomedical applications such as PDT. Further, the EB

technique requires the use of a high-energy EB accelerator under high-

vacuum conditions, which makes it a costly procedure. Hereby, we

developed a simple method for the hydrothermal synthesis of d-TiO<sub>2</sub> NPs

through the formation of liposome-TiO2 nanocomposites, and investigated

their optical and structural properties for exploring their photocatalytic

effects on the viability of cancer cells<sup>25</sup> under visible light irradiation.

minimum aggregation, biocompatibility, and non-toxicity.

#### Introduction

In the past decade, the use of various semiconductor nanomaterials in biomedicine has increased owing to their controllable size, shape, and dimensionality, making them suitable for several application, including biosensing, cell imaging, drug/gene delivery and cancer therapy.<sup>1-4</sup> In particular, TiO<sub>2</sub> nanoparticles (NPs) show advantages of strong photocatalytic activity, non-toxicity, high photostability, and low cost, and have thus attracted much attention as alternative phototherapy (PDT) agents<sup>5-10</sup> to replace conventional dye sensitizers such as porphyrin derivatives.<sup>11</sup> The majority of pristine TiO<sub>2</sub> NPs are active under ultraviolet (UV) light excitation due to their large band gap energy (~3.2 eV). However, UV light induces damage to biological components, and its penetration into a biological tissue is very limited; thus, UV light is often unable to reach the cancer cells located far from the tissue surface. Consequently, UV-sensitive TiO<sub>2</sub> NPs are not useful for successful in vivo PDT, which requires longwavelength visible-light or near-infrared (NIR) irradiation. Thus, TiO<sub>2</sub> NPs have been modified by doping carbon, nitrogen,<sup>12-14</sup> and some transition metals,<sup>15-17</sup> which can narrow their band gap energies and in turn enhance

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Fig. 1 Schematic flow chart for the synthesis of d-TiO<sub>2</sub> NPs

#### **Results and discussion**

#### Structural and optical properties

Fig. 2 (A) shows the XRD pattern of the as-synthesized d-TiO<sub>2</sub> NPs, which exhibited the same pattern as that of P25  $TiO_2$  and EB-induced d-TiO<sub>2</sub> NPs. This indicates that these d-TiO<sub>2</sub> NPs possess a mixture of both the rutile and anatase phases.<sup>20, 30</sup> The XRD intensities of the *d*-TiO<sub>2</sub> NPs slightly decreased compared to those of P25 TiO<sub>2</sub>, indicating that the defects and surface modification of the *d*-TiO<sub>2</sub> NPs reduced the crystallinity.<sup>20</sup> The average crystallite size of the NPs was calculated using the Scherrer equation,  $D = k\lambda$  $/\beta cos\vartheta$ , where k is the shape factor of the particles and has a typical value of 0.89,  $\lambda$  is the wavelength of the X-ray (1.5406 Å),  $\beta$  is the peak broadening in radians (full width at half maximum; FWHM), and  $\vartheta$  is the Bragg angle.<sup>31</sup> The average particle sizes of the anatase phase obtained using this formula were approximately 18.2 nm (d-TiO<sub>2</sub> NPs) and 17.5 nm (P25 TiO<sub>2</sub>), and the average crystallite sizes of the rutile phase were approximately 27.0 nm (d-TiO<sub>2</sub> NPs) and 24.6 nm (P25 TiO<sub>2</sub>). The increase in the particle size under the hydrothermal reaction confirmed the transformation of the phase and defects at the surface of the  $TiO_2 NPs$ .<sup>20, 32</sup>

In order to confirm the surface defects, vibrational structures of the *d*-TiO<sub>2</sub> NPs were evaluated by Raman spectroscopy. In general, there are six (144 cm<sup>-1</sup> (E<sub>g</sub>), 197 cm<sup>-1</sup> (E<sub>g</sub>), 397 cm<sup>-1</sup> (B<sub>1g</sub>), 518 cm<sup>-1</sup> (A<sub>1g</sub>+ B<sub>1g</sub>), 640 cm<sup>-1</sup> (E<sub>g</sub>)) and four (144 cm<sup>-1</sup> (B<sub>1g</sub>), 488 cm<sup>-1</sup> (E<sub>g</sub>), 613 cm<sup>-1</sup> (A<sub>1g</sub>), 827 cm<sup>-1</sup> (B<sub>2g</sub>)) Raman active modes for anatase and rutile TiO<sub>2</sub>, respectively.<sup>33</sup> The *d*-TiO<sub>2</sub> NPs showed the typical Raman bands, although they were broader, as shown in Fig. 2 (B). The FWHM of the *d*-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub> were 9.57 and 8.34 cm<sup>-1</sup> at 144 cm<sup>-1</sup> (E<sub>g</sub>), respectively. The broadening of the Raman peaks of the *d*-TiO<sub>2</sub> NPs indicates that various defects had formed, such as oxygen vacancies and Ti<sup>3+</sup>.<sup>20</sup>



Fig. 2 X-ray diffraction pattern (A) and Raman spectra (B) of d-TiO<sub>2</sub> NPs (a) and P25 TiO<sub>2</sub> (b)

The FT-IR spectra of the *d*-TiO<sub>2</sub> NPs were recorded in KBr phase in the frequency region of 400–4000 cm<sup>-1</sup> (Fig. 3). The functional groups of the recorded FT-IR spectra were compared with the standard spectra of the TiO<sub>2</sub> NPs. The vibration bands observed at ~534 cm<sup>-1</sup> in all spectra were attributed to the vibration of the Ti-O bond in the TiO<sub>2</sub> lattice. The absorption bands at 1640 cm<sup>-1</sup> and 1500 cm<sup>-1</sup> were caused by the bending vibrations of Ti-O-H and Ti-O-Ti, respectively. Furthermore, a broad peak appearing at 3100-3600 cm<sup>-1</sup> caused by the fundamental stretching vibration of O-H hydroxyl groups reduced in the *d*-TiO<sub>2</sub> NPs. Moreover, additional vibrational bands were observed at 3746 and 3850 cm<sup>-1</sup>, indicating the existence of octahedral ( $_{6}Ti^{3+}$ -OH) and tetrahedral ( $_{4}Ti^{4+}$ -OH) coordinated vacancies.<sup>34, 35</sup> These results indicate that the *d*-TiO<sub>2</sub> NPs have more oxygen vacancies than P25 TiO<sub>2</sub>.

Further detailed structures, particle sizes, and structural characterization of the *d*-TiO<sub>2</sub> NPs were determined using field-emission TEM combined with the selected area electron diffraction (SAED) pattern. As shown in Fig. 4, the observed lattice spacing was approximately 0.35 nm, close to the anatase [101] planes.<sup>36</sup> The size distribution determined from the TEM image was similar to that calculated using the Scherrer equation, revealing that the particles size was in the range ~25 nm.



Fig. 3 FT-IR spectra of d-TiO<sub>2</sub> NPs (a) and P25 TiO<sub>2</sub> (b)



**Fig. 4** TEM images of d-TiO<sub>2</sub> NPs (a) and P25 TiO<sub>2</sub> (c); HRTEM images of d-TiO<sub>2</sub> NPs (b) and P25 TiO<sub>2</sub> (d). The insets in (a) and (c) show the electron diffraction pattern of the corresponding selected area.

To investigate the optical properties of the d-TiO<sub>2</sub> NPs, the diffuse reflectance UV-visible absorption spectrum was measured at room



**Fig. 5** UV-visible reflectance absorption spectra (A) and ensemble-averaged fluorescence spectra (B) of d-TiO<sub>2</sub> NPs (a) and P25 TiO<sub>2</sub> (b); the excitation wavelength was 325 nm.

temperature, as shown in Fig. 5(A). Compared with the absorption spectrum of P25 TiO<sub>2</sub> (curve b), the absorbance of the d-TiO<sub>2</sub>NPs (curve a) was higher in the broad-visible region from 400 nm up to the NIR range (700 - 1000 nm) due to the surface defects. The surface defects are formed through reaction of Ti<sup>3+</sup> with a certain amount of oxygen under hydrothermal equilibrium condition,26 and their optimum amount could be obtained at 180 °C as proved by the temperature dependence of the redshifted absorption (Fig. S1). The band gap energies of d-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub> were 2.77 eV and 3.11 eV, respectively. This decrease in the band gap energy may be due to the localized surface states induced by Ti<sup>3+</sup>, which can form donor levels in the electronic structure of  $TiO_2$ , as well as the oxygen vacancies, which can affect the electron-hole (e-h) recombination process.<sup>20,34,37-38</sup>The bulk defects only act as charge-carrier traps where e-h recombine, whereas the surface defects can also facilitate charge transfer to adsorbed species, which can prevent e-h recombination, as well as serve as charge-carrier traps.<sup>23,39</sup> This speculation is supported by the observation that the fluorescence emission intensity of the d-TiO<sub>2</sub> NPs was guenched compared to that of P25  $\text{TiO}_2$  under the same excitation condition at 325 nm (Fig. 5(B)). This is likely due to the fact that d-TiO<sub>2</sub>NPs have more surface defects than P25 TiO<sub>2</sub>. Thus, it is expected that the *d*-TiO<sub>2</sub> NPs would have higher photocatalytic activities under visible light radiation.

For further confirmation of the structural and defect properties, we analyzed the chemical bonding of TiO<sub>2</sub> NPs by XPS. Fig. 6 shows the XPS spectra of the Ti 2p<sub>3/2</sub> and O 1s core levels recorded from d-TiO<sub>2</sub> NPs (spectra a and c) and P25  $\text{TiO}_2$  (spectra b and d). The Ti  $2p_{3/2}\,\text{and}$  O 1s XPS peaks of the d-TiO<sub>2</sub> NPs were observed at 458.8 eV (Ti<sup>4+</sup>2p<sub>3/2</sub>), 458.0 eV (Ti $^{3+}2p_{3/2}$ ), 530.0 eV [TiO<sub>2</sub> (Ti $^{4+}$ )], 531.1 eV [Ti<sub>2</sub>O<sub>3</sub> (Ti $^{3+}$ )], and 532.1 eV [water adsorbed on the  $TiO_2$  surface].<sup>20,22,33</sup> The XPS peaks of P25 TiO\_2 were observed at 458.6 eV (Ti^{4+}2p\_{3/2}), 457.5 eV (Ti^{3+}2p\_{3/2}), 529.8 eV [TiO<sub>2</sub> (Ti<sup>4+</sup>)], 531.1 eV [Ti<sub>2</sub>O<sub>3</sub> (Ti<sup>3+</sup>)], and 532.2 eV [water adsorbed on the TiO<sub>2</sub> surface]. The amount of Ti<sup>3+</sup> on the TiO<sub>2</sub> surface plays an important role, as previously reported for TiO<sub>2</sub> doped with metal atoms. The photogenerated electrons can be trapped in Ti<sup>3+</sup>, thereby preventing the recombination of the majority and minority carriers. Moreover, Ti<sup>3+</sup> formation is related to the oxygen vacancies.<sup>23</sup> Therefore, the increased  $Ti^{3+}$  of the *d*-TiO<sub>2</sub> NPs indicates that the *d*-TiO<sub>2</sub> NPs have more defects.



**Fig. 6** X-ray photoelectron spectra of Ti 2p ((a) and (b)) and O 1s ((c) and (d)) for *d*-TiO<sub>2</sub> NPs ((a) and (c)) and P25 TiO<sub>2</sub> ((b) and (d))

To test whether the d-TiO<sub>2</sub> NPs can readily infiltrate the biological cells, we examined their distribution and sizes in aqueous solution according to the zeta potential and DLS measurements. The zeta potentials of the *d*-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub> were strongly negatively charged at pH 7.0 in deionized water, at around -22.5 mV and -21.8 mV, respectively (Fig. S2). Such large zeta potential values can affect the distribution of particles. In other words, if particles have a small zeta potential, there is no force to prevent the particles from coming together, leading to their aggregation. Further details of the aggregation of the *d*-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub> were monitored via particle size measurement using DLS to determine their stability in deionized water (Fig. S3). When dispersed in water at pH = 7.0 at a concentration of 1 mg/mL, the *d*-TiO<sub>2</sub>NPs showed reduced agglomeration (224 nm) compared to that of P25 TiO<sub>2</sub> (1440 nm). These results indicate that the synthesized d-TiO<sub>2</sub> NPs can disperse better than P25 TiO<sub>2</sub> in solution. Therefore, the cellular uptake of *d*-TiO<sub>2</sub>NPs is expected to be improved compared to that of P25 TiO2. 19,40-41

#### Photocatalytic action on cancer cells

To evaluate the photocatalytic action of the d-TiO<sub>2</sub> NPs on cancer cells, the viability of HeLa and Hep-G2 cells was measured using the WST method in the presence of various concentrations of TiO<sub>2</sub> NPs under visible light irradiation. The confocal DIC images of HeLa showed that the addition of the P25 TiO<sub>2</sub> and exposure to visible light caused negligible stress (Fig. 7). Similar cell morphology was observed when HeLa cells were treated with and without the P25 TiO<sub>2</sub> in dark and under visible light, indicating no major effect of either visible light or the P25 TiO<sub>2</sub> on cell viability. In contrast, upon visible light irradiation with d-TiO<sub>2</sub> NPs, the HeLa cells showed extensive cell death (about 70%, as estimated by cell counting).

These observations were confirmed more quantitatively by the cell viability measurements using the WST method (Fig. 8 (A)). In the dark, the viabilities of both HeLa and Hep-G2 cells remained strong, at over 90%, with the treatment of 25  $\mu$ g/mL *d*-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub>. Over 80% of the cells in both cell lines also survived under visible light irradiation

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without TiO<sub>2</sub> NPs. In the dark, the cell viabilities remained the same when treated with much higher concentrations of  $TiO_2 NPs$ , up to 100  $\mu g/mL$  as shown in Fig. 8 (B) and (C). However, under visible-light irradiation with TiO<sub>2</sub>NPs, the viabilities of both HeLa and Hep-G2 cells decreased to 30% and 40%, respectively; their viabilities further decreased with increasing  $TiO_2 NPs$  concentrations. This indicates that TiO<sub>2</sub> NPs photocatalyze the killing of cancer cells under visible light. In particular, the minimum concentrations of the d-TiO<sub>2</sub> NPs required to observe the photocatalytic killing of HeLa and Hep-G2 cells (70 % and 60 % death rates, respectively) were much lower (25  $\mu\text{g}/\text{mL}$  and 50  $\mu$ g/mL) than those of P25 TiO<sub>2</sub>, implying that the visible-light photocatalytic effect of the d-TiO<sub>2</sub> NPs is much stronger than that of P25  $TiO_2$ . The difference in the concentration dependence of the photocatalytic-killing effect between HeLa and Hep-G2 cells may also imply that HeLa cells can uptake d-TiO2 NPs more easily than Hep-G2 cells. Recently Li et al.<sup>12</sup> demonstrated the visible-light photocatalytic killing of HeLa cells up to 60% using N-doped  $TiO_2$  NPs at a concentration of 200  $\mu$ g/mL which is eight times higher than concentration used in our study under the same illumination condition. This comparison suggests that the visible-light photocatalytic activity of d-TiO<sub>2</sub>NPs in the killing of cancer cells is much more effective than that of N-doped TiO<sub>2</sub> NPs.



**Fig. 7** Confocal DIC images of HeLa cells without or with d-TiO<sub>2</sub> NPs (or P25 TiO<sub>2</sub>) under visible light irradiation or in the dark.

In general, it is known that the photocatalytic action of  $TiO_2$  NPs on cancer cells is mostly triggered by the photoinduced generation of various reactive oxygen species (ROS) such as hydroxyl radicals (·OH), hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>), and superoxide anion radicals (O<sub>2</sub>-<sup>-</sup>) on the surface of cell membranes and in the cytoplasm.<sup>9,25,40</sup> This occurs because TiO<sub>2</sub> NPs can attach to the cellular membranes and enter the cytoplasm via endocytosis.<sup>37,41</sup> From this perspective, the *d*-TiO<sub>2</sub> NPs are also expected to generate ROS in cancer cells upon visible light irradiation, as shown in the following equations:

- (1)  $TiO_2 + hv (\langle E_g \rangle \rightarrow h^+ + e^{it})$
- $(2) \quad H_2O + h^{\scriptscriptstyle +} \mathop{\rightarrow} ^{\scriptscriptstyle \bullet}OH + H^{\scriptscriptstyle +}$
- $(3) \quad O_2 + e^{-} \rightarrow O_2^{-}$
- $(4) \quad \mathsf{O}_2^{\cdot} + \mathsf{H}^* \to \mathsf{HO}_2^{\cdot}$
- (5)  $2HO_2 \rightarrow H_2O_2 + O_2$
- $(6) \quad H_2O_2 + O_2 \stackrel{\cdot}{\cdot} \rightarrow \stackrel{\cdot}{\cdot} OH + OH \stackrel{\cdot}{\cdot} + O_2$

The *d*-TiO<sub>2</sub>NPs absorb the visible light with energy larger than the band gap. Band gap excitation of the *d*-TiO<sub>2</sub>NPs results in e<sup>-</sup>(conduction band) – h<sup>+</sup> (valence band) separation. The high oxidative potential of the holes allows the formation of reactive intermediates such as H<sup>+</sup> and 'OH. The reactive 'OH can be formed by the decomposition of water. Meanwhile, the electrons are usually reactive with O<sub>2</sub> to yield O<sub>2</sub> <sup>-/-</sup>. Moreover, some HO<sub>2</sub><sup>-</sup> can form H<sub>2</sub>O<sub>2</sub>, and the reaction of H<sub>2</sub>O<sub>2</sub> can also result in the formation of 'OH.

In addition to conventional ROS, it was recently reported that  $TiO_2$  NPs can efficiently generate  ${}^1O_2$  by energy transfer *via* the defect sites if they absorb long-wavelength visible light and NIR wavelengths (700-1000 nm) in addition to an electron transfer mechanism (Nosaka's mechanism),<sup>24</sup> as shown in the next eqution.

(7) 
$$O_2^{\cdot} + h^+ \rightarrow {}^1O_2$$

Since the *d*-TiO<sub>2</sub>NPs were shown to absorb long-wavelength visible



**Fig. 8** (A) Histogram of the photokilling effects of TiO<sub>2</sub> ( $25\mu g/mL$ ) on the relative viabilities of HeLa and Hep-G2 cels under visible light irradiation or in the dark. Relative cell viabilities as a function of TiO<sub>2</sub> NP concentrations for HeLa (B) and Hep-G2 (C) cells in dark ((a) and (b)) and under visible light irradiation ((c) and (d)) with *d*-TiO<sub>2</sub> NPs ((a) and (c)) and P25 TiO<sub>2</sub> ((b) and (d)).

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light and NIR light, we attempted to confirm the photoinduced formation of  ${}^1\text{O}_2$  by using the SOSG reagent.  $^{^{28\text{-}29}}$  SOSG is highly selective in reactions with <sup>1</sup>O<sub>2</sub>, without any appreciable response to ·OH or ·O<sub>2</sub>. This indicator initially exhibits weak fluorescence; however, in the presence of  ${}^{1}O_{2}$ , it emits strong fluorescence. Fig. 9 (C) shows a plot of the SOSG fluorescence intensity measured at 529 nm with d-TiO<sub>2</sub> NPs (a) or P25 TiO<sub>2</sub> (b), depending on the visible-light (460-800 nm) irradiation time. The fluorescence intensity of SOSG with d-TiO<sub>2</sub> NPs was higher than that observed with P25 TiO<sub>2</sub>, indicating that the d-TiO<sub>2</sub>NPs generate  ${}^{1}O_{2}via$  their defect sites. Therefore, it can be concluded that the *d*-TiO<sub>2</sub> NPs may show enhanced photocatalytic killing of cancer cells owing to the generation of  ${}^{1}O_{2}$  in addition to the other ROS in the photooxidation of cell components. This potential mechanism is summarized in a schematic shown in Fig. 10.42 These results suggest that the *d*-TiO<sub>2</sub>NPs are promising materials for future in vivo PDT of cancer cells under long-wavelength visible and NIR light irradiation.



**Fig. 9** Fluorescence intensity of aqueous SOSG measured at the maximum of 529 nm upon excitation at 480 nm in the presence of or d-TiO<sub>2</sub> NPs (A) or P25 TiO<sub>2</sub> (B) after visible light irradiation by a xenon lamp. (C) Fluorescence intensity of the aqueous SOSG solution in the presence of d-TiO<sub>2</sub> NPs (a) or P25 TiO<sub>2</sub> (b) over time.



**Fig. 10** Schematic of the mechanism for the photokilling of cancer cells by the *d*-TiO<sub>2</sub> NPs.

**Experimental section** 

#### Chemicals

All of the chemicals used for the synthesis of d-TiO<sub>2</sub> NPs, including P-25 (Degussa-Huls), egg lecithin (L-R-phosphatidylcholine, >60%, Sigma-Aldrich), chloroform (>99.8%, Samchun, Korea), sodium hydroxide (>93%, Duksan, Korea), and ethanol (200-proof, >99.8%, Sigma-Aldrich), were of analytical grade and used without further purification.

#### Synthesis of d-TiO<sub>2</sub> NPs

The *d*-TiO<sub>2</sub> NPs were synthesized by the combination of sol-gel and hydrothermal reactions through the formation of liposome-TiO<sub>2</sub> composites.<sup>26</sup> A schematic representation of the synthesis procedure is shown in Fig. 1. Initially, the precursor solution was prepared by adding 50 mL of a 1.0 M NaOH aqueous solution into 0.1 g of P-25 powder in an ethanol (30 ml) mixture at room temperature under constant stirring. To prepare the liposome-TiO<sub>2</sub> composites, thin phospholipid films were prepared by evaporating a chloroform solution of egg lecithin lipid to dryness in a round-bottomed flask under reduced pressure. The lipid films were mixed with a precursory solution, followed by sonication for 30 min in an ultrasonic bath (Fisher Scientific model FS20, 70 W, 42 kHz) until the turbid liposome-TiO<sub>2</sub> composites were formed. The turbid solution of the liposome-TiO<sub>2</sub> composites was transferred into a Teflon-lined stainless steel autoclave and maintained at 180 °C for 24 h for the hydrothermal reaction. After the hydrothermal reaction, the autoclave was cooled naturally to room temperature. The solid matter was then collected and washed in distilled water and ethanol several times, followed by calcinations at 823 K for 24 h to form NPs.

#### Structural and optical characterization

The crystallinity and structure of the as-prepared NPs were characterized by X-ray diffraction (XRD) using a rotating anode X-ray diffractometer (D/MAX-2200 Ultima/PC) with CuKR radiation ( $\lambda$ ; 1.5405 Å). The crystalline phases and functional groups of the samples were also determined by Raman spectroscopy (Lab RAM HR-800, Horiba Jobin Yvon). The vibrational structures of the *d*-TiO<sub>2</sub> NPs were investigated using Fourier transforminfrared (FT-IR) spectroscopic analysis; the FT-IR spectra were recorded with the KBr pellet technique using a JASCO FT-IR 4100 spectrometer. The size and morphology of the synthesized *d*-TiO<sub>2</sub> NPs were examined by transmission electron microscopy (TEM; Tecnai G2 F30). The TEM sample was prepared by dip-coating Formvar/carbon film-Cu grids with a nanocolloidal solution obtained by sonication of the synthesized *d*-TiO<sub>2</sub> NPs in ethanol.

To evaluate the absorptions of the synthesized d-TiO<sub>2</sub> NPs in the visiblelight region, diffuse reflectance UV-VIS-NIR absorption spectra (DRS) were recorded on a Solid Spec-3700 double-beam spectrophotometer equipped with an integrating sphere. The steady-state fluorescence spectra were obtained at room temperature with a PL spectrometer at 325 nm (Lab RAM HR-800, Horiba Jobin Yvon). X-ray photoelectron spectra (XPS) were obtained using monochromatic Al K $\alpha$  X-ray radiation (1486.6 eV) with a power of 120 W (Kratos Analytical, AXIS Nova, UK).

The particle sizes and zeta potentials of the dispersed solution of the *d*-TiO<sub>2</sub> NPs and P25 TiO<sub>2</sub> were measured by dynamic light scattering (DLS) and laser Doppler velocimetry (LDV), respectively, using an electrophoretic light scattering spectrophotometer (Otsuka Electronics Co, Ltd; ELS-Z2). Confocal differential interference contrast (DIC) images of HeLa cells with or without the *d*-TiO<sub>2</sub> NPs and visible

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light irradiation were observed by laser-scanning confocal microscopy (LSCM; LSM5 live configuration Vario Two VRGB).

#### Cell culture

The human cervical carcinoma cells (HeLa) and human hepatocellular carcinoma cells (Hep-G2) were grown in 89% Dulbecco's modified Eagle's medium with 10% fetal bovine serum and 1% antibiotic-antimycotic solution. The cells were routinely maintained in the plastic tissue culture dishes at 37 °C under a humidified 5% CO<sub>2</sub>-containing atmosphere.

#### **Evaluation of cytotoxicity**

The cytotoxicity of the injected TiO<sub>2</sub> NPs was evaluated using the EZ-Cytox reagent (Daeil Lab Service, Seoul, South Korea) based on the water-soluble tetrazolium (WST) method.<sup>27</sup> HeLa or Hep-G2 cells were seeded at a density of  $1.5 \times 10^4$  cells per well in a 96-well microassay plate, and incubated for 24 h at 37 °C under a 5% CO<sub>2</sub> atmosphere. The *d*-TiO<sub>2</sub> NPs or P25 TiO<sub>2</sub> were added to the incubated cells at various concentrations, followed by further incubation for additional 24 h at 37 °C. Next, 10 µL of the EZ-Cytox reagent was added and the plates were incubated for 2 h at 37 °C. The absorbance of the EZ-Cytox reagent was measured at 450 nm using a microplate reader (VersaMax, Molecular Devices, USA). The cell viability (%) was calculated using the following equation: cell viability (%) = (OD <sub>450(cample)</sub> / OD <sub>450(control</sub>) × 100.

#### Photocatalytic killing of cancer cells

The cancer cells incubated with different concentration of the d-TiO<sub>2</sub> NPs or P25 TiO<sub>2</sub> were irradiated for 4 h with the visible light emitted from a 300-W xenon lamp (Asahi Spectra, Japan). The 400 nm longpass filter (GG 400) and heat-absorbing colored glass filter (KG 5) were used to cut off the UV and far-infrared wavelengths, respectively. All filters were purchased from Newport, USA. The visible-light power density at the liquid surface in the cell wells was 12 mW/cm<sup>2</sup>, as measured by a power meter (model 2936-C, Newport, USA). The irradiated cancer cells were incubated in the dark for another 24 h before analyzing the cell viability as described above.

#### Determination of singlet oxygen generation from TiO<sub>2</sub> NPs

Singlet oxygen  $({}^{1}O_{2})$  generation was monitored by using the singlet oxygen sensor green (SOSG) reagent.<sup>28-29</sup> Stock solutions of 5 mM SOSG were dissolved in methanol/water (3:97 v%) to obtain a final concentration of approximately 1µM. Subsequently, a 10 µM solution of *d*-TiO<sub>2</sub> NPs (or P25 TiO<sub>2</sub>) was added to the 1µM SOSG solution under stirring. This sample was irradiated under the visible-light region with a xenon lamp (Asahi Spectra; MAX-302) and a 460 nm long-pass filter every 5 min, followed by measuring the SOSG fluorescence changes (500-600 nm) upon excitation at 480 nm.

#### Conclusions

The defective TiO<sub>2</sub> nanoparticles (*d*-TiO<sub>2</sub> NPs) were synthesized using a simple hydrothermal gel/sol technique from P25 TiO<sub>2</sub> powder. Using various spectroscopic, diffraction, and microscopic techniques, we compared the structural properties and morphology of the *d*-TiO<sub>2</sub> NPs to P25 TiO<sub>2</sub>. Absorbance at long wavelengths, including NIR, was confirmed, and agglomeration was reduced (~224 nm) compared to that of P25 TiO<sub>2</sub> (~1440 nm) in aqueous solution, indicating that the *d*-TiO<sub>2</sub> NPs could infiltrate the cell membrane to access cancer cells. The improved cell-killing effect of the *d*-TiO<sub>2</sub> NPs in two cancer cell lines was confirmed, which was triggered by long-wavelength visible light to generate reactive oxygen species including

singlet oxygen  $({}^{1}O_{2})$ . Thus, *d*-TiO2 NPs show good potential for novel photodynamic therapy strategies in cancer treatment.

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## **Table of Contents**

## Hydrothermal synthesis of defective TiO<sub>2</sub> nanoparticles for longwavelength visible light-photocatalytic killing of cancer cells

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Newly fabricated *d*-TiO2 NPs were demonstrated to be efficient in long wavelength visible light-triggered killing of cancer cells.

