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1 **Formation potential of N-nitrosamines from soluble microbial**
2 **products (SMPs) exposed to chlorine, chloramine and ozone**

3

4 Beibei Zhang ^a, Qiming Xian ^{a*}, Gang Yang ^a, Tingting Gong ^a, Aimin Li ^a, Jianfang
5 Feng ^a

6

7 **Abstract**

8 Soluble microbial products (SMPs) are an important component of effluent organic
9 matter in wastewater treatment. This study investigated the N-nitrosamines formation
10 potential (NAsFP) from chlorination, chloramination and ozonation of SMPs. The
11 results show that more NAs were formed in chloramination than chlorination and
12 ozonation. Specially, the formation of NAs showed a good linear relationship with
13 disinfectant dosage and bromide level, and increased with reaction time, but
14 decreased at high temperature during chlorination, chloramination and ozonation. The
15 effects of pH on the NAsFP were different for different disinfection methods. The pH
16 values showed negative effects on the NAsFP in chlorination and positive effects on
17 the NAsFP in ozonation. Yet in chloramination study, the NAsFP showed a first
18 increasing and then a decreasing trend. Regression procedure revealed that bromide
19 level was the most important factor for NAs formation whether for chlorination,
20 chloramination or ozonation. The NAsFP of SMPs in the three disinfection methods

*Corresponding author. State Key Laboratory of Pollution Control and Resource Reuse, School of the Environment, Nanjing University, Nanjing 210023, P. R. China. E-mail address: xianqm@nju.edu.cn. Tel/fax: +86-25-89680259.

^a State Key Laboratory of Pollution Control and Resource Reuse, School of the Environment, Nanjing University, Nanjing 210023, P. R. China.

21 was compared between synthetic and real wastewater. Five NAs were detected, and
22 NDMA was the most NAs accounting for more than 50%. As for reducing NAs in the
23 effluent, ozonation was the best alternative to chlorination and chloramination, and
24 the bromide removal was the most important for chlorination, chloramination and
25 ozonation.

26

27 **Keywords**

28 Soluble microbial products (SMPs); N-nitrosamines (NAs); disinfection methods;
29 influence factors

30

31 **1. Introduction**

32 Nowadays, wastewater reuse has gained more and more attention to alleviate the
33 problem of freshwater resources shortage. As a cheap and effective disinfection
34 method, chlorine has been widely used to inactivate the pathogenic microorganisms in
35 water treatment. However, due to the strong oxidation of chlorine, it can also react
36 with some organic matters to form disinfection by-products (DBPs), such as
37 trihalomethanes (THMs), haloacetic acids (HAAs), halonitromethanes (HNMs), etc.^{1,2}
38 Organic matters from wastewater effluent are generated by microbial metabolism and
39 the main components are called soluble microbial products (SMPs),³ which consist of
40 macromolecules and cellular debris including proteins, polysaccharides, humic acids,
41 and DNA.⁴ According to previous studies, SMPs can undergo both carbonaceous
42 DBPs (C-DBPs) and nitrogenous DBPs (N-DBPs) formation when subjected to
43 chlorination, such as THMs, HAAs and dichloroacetonitrile (DCAN), and could
44 increase the DBPs formation in both wastewater effluent and surface water supply
45 after chlorination.⁵⁻⁷

46 With the great efforts of researchers,⁸ DBPs were associated with the teratogenic,
47 carcinogenic and mutagenic risks, and could also increase the ecotoxicity of the
48 receiving water body. In order to reduce the amount of regulated DBPs, chloramine
49 and ozone are chosen as the alternative disinfectant. However, some N-DBPs, which
50 were more toxic than C-DBPs may be formed by these alternative disinfectants. For
51 example, chloramine could decrease the amount of regulated DBPs, but more
52 N-DBPs, such as HNMs and N-nitrosamines (NAs), may be formed.^{9,10} Ozone can

53 significantly reduce or eliminate the formation of THMs and HAAs, however, it can
54 result in the formation of bromate and NAs.¹¹ In general, N-DBPs, especially NAs are
55 present in relatively low concentrations (ng/L) but actually more toxic than C-DBPs,¹²
56 therefore, more and more attentions are paying to the formation of N-DBPs.

57 The concentration and speciation of N-DBPs during disinfection were
58 significantly affected by water quality parameters and operating conditions. In the
59 chlorination process, increasing pH decreased the formation of DCAN but increased
60 the formation of HNMs, however, the opposite trend of DCAN and HNMs may occur
61 during the chloramination process.^{13,14} The higher disinfectant dose and bromide level
62 had a positive effect on haloacetonitriles (HANs) yields during chloramination but not
63 during chlorination.¹³ For both chlorination and chloramination, high disinfectant
64 dose, long reaction time and high bromide level increased the formation of HNMs.
65 ^{2,14,15} Chloramination generated more NAs than chlorination and ozonation, and the
66 formation of NAs increased with an increase of bromide level.^{11,16,17} However, in
67 the most of these previous studies, drinking water or surface water disinfection were
68 studied, and thus natural organic matter (NOM) was the main organic precursor which
69 generated the studied DBPs. Up to now, limited studies have been carried out to
70 investigate the formation of NAs formed by SMPs during disinfection,⁷ and the
71 influence factors on the formation of NAs in chlorination, chloramination and
72 ozonation of SMPs is even fewer. Since wastewater reuse has become a growing
73 portion of water supplies, chloramine and ozone have gained more and more
74 popularity in water disinfection, and NAs are of potentially greater health concern

75 than THMs, HAAs, HANs and HNMs, it is quite necessary to compare the formation
76 of NAs from SMPs under chlorination, chloramination and ozonation with various
77 conditions, and thus provide more information for the disinfection of reuse water.

78 The objectives of the study were, therefore, to investigate the effects of
79 disinfection methods (chlorination, chloramination and ozonation) and various factors
80 (temperature, pH, bromide level, disinfectant dosage and reaction time) on the
81 formation of NAs from SMPs, and to evaluate the main factors affecting the
82 formation of NAs from SMPs. The SMPs are chosen because almost all the soluble
83 organic matters of wastewater effluent are SMPs.

84

85 **2. Material and methods**

86 **2.1. Chemicals and reagents**

87 Sodium hypochlorite solution (NaClO, 5%), standard solutions of nine NAs (NDMA,
88 N-nitrosodimethylamine; NMEA, N-nitrosomethylethylamine; NDEA,
89 N-nitrosodiethylamine; NPyr, N-nitrosopyrrolidine; NMor, N-nitrosomorpholine;
90 NDPA, N-nitrosodipropylamine; NPip, N-nitrosopiperidine; NDBA,
91 N-nitrosodibutylamine; NDPhA, N-nitrosodiphenylamine) were supplied from
92 Sigma-Aldrich. Isotopically labelled standards [$6\text{-}^2\text{H}$] N-nitrosodimethylamine
93 (NDMA- d_6 , 98%) and [$14\text{-}^2\text{H}$] N-nitrosodipropylamine (NDPA- d_{14} , 98%) were
94 obtained from Cambridge Isotope Laboratories (Andover, MA) and used as surrogate
95 and internal standard for NAs, respectively. All other reagents were reagent grade.

96

97 2.2. SMPs collection

98 Activated sludge was collected from an aeration tank in a Municipal Wastewater
99 Treatment Plant, and used as inoculums for the laboratory-scale Sequencing Batch
100 Reactor (SBR). The seed activated sludge was added into the synthetic wastewater to
101 a final biomass concentration of about 2000 mg/L. Glucose (800 mg/L) was selected
102 as the only carbon and energy source, as it can be biodegraded completely leaving
103 only SMPs as the remaining organics in the solution.⁷ The other substances were as
104 following according to our previous study (in mg per L): (NH₄)₂SO₄ (189), KH₂PO₄
105 (35), CaCl₂ (0.37), MgSO₄ (5.07), MnCl₂ (0.27), ZnSO₄ (0.44), FeCl₃ (1.45), CuSO₄
106 (0.39), CoCl₂ (0.42), Na₂MoO₄ (1.26).¹⁸ The reactor was incubated for 6 h at 25°C
107 followed by a precipitation time of 30 min. Supernatant was then collected and
108 filtered through a 0.45 µm filter paper. The filtrate was defined as SMPs.^{18,19}

109 The characteristics of SMPs were determined. Dissolved organic carbon (DOC)
110 was measured with a TOC analyzer (TOC-VCH, Shimadzu, Japan). Glucose
111 (measured as chemical oxygen demand (COD)), Total nitrogen (TN), ammonia
112 nitrogen (NH₄⁺-N), nitrite nitrogen (NO₂⁻-N), and nitrate nitrogen (NO₃⁻-N) were
113 determined by HACH methods (www.hach.com/wah) using a DR2800 (HACH, USA).
114 Dissolved organic nitrogen (DON) was calculated by subtracting values of NO₂⁻-N,
115 NO₃⁻-N and NH₄⁺-N from the TN value. UV₂₅₄ absorption was analyzed with a visible
116 spectrophotometer (UV7595, Shanghai). Bromide was measured with an ion
117 chromatography (Dionex DX-600, German). The parameters of the obtained SMPs
118 were as follows: Glucose (measured as COD) = none, DOC = 20.4 - 25.2 mg/L,

119 $UV_{254} = 0.042 - 0.058 \text{ cm}^{-1}$, $TN = 12.4 - 15.0 \text{ mg/L}$, $NH_4^+-N = 2.9 - 3.3 \text{ mg/L}$,
120 $NO_2^- - N = 0.117 - 0.133 \text{ mg/L}$, $NO_3^- - N = 1.9 - 2.6 \text{ mg/L}$, $DON = 6.3 - 8.9 \text{ mg/L}$.
121 These parameters of SMPs were generally constant in different batches.

122 In addition, SMPs were also collected from the real wastewater effluent from two
123 domestic wastewater treatment plants of Nanjing. The parameters of SMPs were as
124 follows: $DOC = 21.5 - 23.2 \text{ mg/L}$, $UV_{254} = 0.137 - 0.179 \text{ cm}^{-1}$, $TN = 10.4 - 11.6$
125 mg/L , $NH_4^+ - N = 3.8 - 4.3 \text{ mg/L}$, $NO_2^- - N = 0.176 - 0.246 \text{ mg/L}$, $NO_3^- - N = 2.2 - 3.1$
126 mg/L , $Br^- = 108.4 - 112.6 \text{ } \mu\text{g/L}$, $DON = 3.3 - 4.9 \text{ mg/L}$, respectively.

127

128 **2.3. Disinfection of SMPs**

129 Chlorination, chloramination and ozonation were conducted as previously
130 describe.^{20,21} Briefly, SMPs from synthetic and real wastewater were chlorinated by
131 $NaClO$. Monochloramine was prepared daily by slowly adding sodium hypochlorite
132 into ammonium chloride solution at a Cl/N molar ratio of 0.7:1 with continuous
133 stirring. To minimize the disproportionation of monochloramine to dichloramine,
134 phosphate buffer (10 mmol/L) was used to maintain the pH above 8.5. After 30 min of
135 stirring, monochloramine solution was aged in the dark for at least 1 h. Both chlorine
136 and monochloramine solutions were standardized using the N,
137 N-diethylphenylene-1,4-diamine (DPD) colorimetric method before disinfection.²²
138 Ozone was produced from extra dry grade oxygen (with a minimum purity of 99.99%)
139 using a WH-H-Y10 ozone-generator (WAOHUANG, China). The ozone concentration
140 was determined using spectrophotometric methods.²¹

141 Chlorination, chloramination and ozonation were conducted in glass bottles with
142 Teflon inner plugs. Temperature was kept by a thermostatic reactor and pH was
143 adjusted with phosphate buffer. After disinfection, the residual chlorine, chloramine
144 and ozone were quenched using $\text{Na}_2\text{S}_2\text{O}_3$.

145 In order to compare and understand the formation of NAs from SMPs under
146 different conditions, except for the formation of NAs under different disinfectant
147 dosage and reaction time conditions, NAs formation potential (NAsFP), which was
148 conducted with relatively high disinfectant dosage for a long reaction time was used
149 in this study. The details of the prepared samples are shown in Table 1. All samples
150 were conducted in duplicate.

151

152 **2.4. Analysis of NAs**

153 NAs were measured using a gas chromatography-mass spectrometer (GC-MS)
154 (Thermo Polaris Q, USA), with a modified version of EPA method 521 reported by
155 Pozzi et al.²³ The recoveries of nine NAs ranged from 78% to 109%. The detection
156 limits of nine NAs ranged from 0.8 – 4.3 ng/L.

157

158 **2.5. Statistical analysis**

159 The linear relationship was carried out by Origin software (Version 8.0). A
160 multivariate regression procedure (stepwise) was used to investigate the key factors
161 influencing the formation of NAs in disinfection of SMPs, and this statistical analysis
162 was conducted with SPSS software (Version 17.0) reported by Hong et al.¹⁵ Briefly,

163 the individual NAs and total NAs were respectively designated as the dependent
164 variable, and the influence factors (temperature, pH, bromide level, disinfectant
165 dosage and reaction time) were defined as independent variables. The regression
166 placed independent variables into the equation in the order of their partial correlation
167 coefficients with the dependent variable. Thus, the key factors were identified using
168 this process.

169

170 **3. Results and discussions**

171 **3.1. Factors affecting NAsFP**

172 **3.1.1. Effect of temperature**

173 Five NAs were detected including NDMA, NDEA, NMEA, NMor and NPip when
174 SMPs reacted with the three disinfectants, sodium hypochlorite, chloramine and
175 ozone, respectively. Fig.1 shows NAsFP levels at three different temperatures (25, 30
176 and 40 °C). The formation potential of NDMA was the highest among these five
177 detected NAs. The total NAsFP in chlorination and chloramination was much more
178 than that in ozonation in these temperature conditions. The total NAsFP decreased
179 slightly with temperature increased from 25 to 40 °C in the three different disinfection
180 methods, but no significant difference except in chloramination. The previous study
181 indicated that temperature had a significant effect on the stability of sodium
182 hypochlorite, monochloramine and ozone, and the higher temperature, the more
183 unstable of them.¹⁴ Although increasing temperature accelerates the reaction rate, it
184 may also increase the decomposition rate of chlorine, chloramine and ozone, thus

185 reducing the amount of effective disinfectant and the formation of NAs.

186

187 **3.1.2. Effect of pH**

188 Fig. 2 illustrates the effect of pH values on the formation of NAs. During chlorination,
189 increasing pH significantly decreased the total NAsFP. While sodium hypochlorite
190 was used as a disinfectant, the concentration of effective chlorine decreased with the
191 increasing of pH.²⁴ Therefore, acidic condition could accelerate the formation of NAs
192 in chlorination.

193 For chloramination, the different trends of total NAsFP were observed in different
194 pH values (Fig. 2). The total NAsFP is the highest in pH = 8, and followed in pH = 6
195 and pH = 10. The previous study had indicated that pH affected the speciation of
196 chloramines and the hydrolysis of monochloramine to form free chlorine, which had
197 been suggested to play a significant role in DBPs formation.²⁵ Under alkaline
198 condition (pH = 10), monochloramine was the only species and its hydrolysis to free
199 chlorine was reduced, which resulted in the decreasing formation of total NAsFP in
200 spite of the promotion of some intermediate products, such as asymmetric secondary
201 hydrazine.²⁶ However in pH = 8, monochloramine can generate dichloramine by
202 disproportionation, so that monochloramine and dichloramine could coexist in the
203 reaction solution. Schreiber and Mitch had proved that the amount of NDMA
204 generated by dichloramine was 1 - 2 orders of magnitude more than that of
205 monochloramine.²⁷ Under pH = 6, the amount of dichloramine increased and became
206 the dominant species, and the concentration of NAs precursors decreased (mostly in

207 the protonated form) at this pH, which resulted in fewer NAs generated.²⁷

208 For ozonation, the total NAsFP increased with the pH increased from 5 - 9, and
209 showed significantly difference between pH = 7 and pH = 9 (Fig. 2). The results were
210 difficult to be explained with formaldehyde catalytic theory by Keefer et al,²⁸ but
211 could be explained with hydroxylamine way by Yang et al.²⁹ Under pH = 9, hydroxide
212 ions can accelerate the decomposition of ozone to generate hydroxyl radical and thus
213 generated more hydroxylamine.

214

215 **3.1.3. Effect of bromide level**

216 The effect of bromide level on NAsFP was studied by performing experiments at four
217 bromide levels (0, 0.2, 0.5, 1.0 mg/L, Fig. 3). Similar to previous reports that bromide
218 can catalyze the formation of NAs, the NAsFP improved as the bromide level
219 increased during chlorination, chloramination and ozonation.^{11,17,30} The linear
220 relationships were observed between the total NAsFP and bromide level ($R^2 > 0.98$).
221 The bromide significantly accelerate the formation of NAs in chloramination (slope =
222 3.4) more than that in chlorination (slope = 1.9) and ozonation (slope = 0.3) (Fig. 3).
223 For chlorination, when the water contains ammonia and bromide ions, HOCl will first
224 react with ammonia to form chloramine. Chloramine is unstable and degrades rapidly
225 when mixed with an excess of bromide, and could react with bromide ions to form
226 dibromamine and bromochloramine.¹⁷ Because of the higher nucleophilic and
227 reactivity of dibromamine and bromochloramine, more NAs could be generated. For
228 ozonation, bromide ions could be oxidized by ozone or hydroxyl radical to

229 hypobromous acid (HOBr), a more effective halogen-substituting agent, thus resulting
230 in increased levels of NAs.^{11,31}

231 The effect of bromide level on the species fraction of NAs was also investigated in
232 this study (Fig. 4). For chlorination, chloramination and ozonation, bromide hardly
233 affected on the species of NAs, and NDMA was always the most abundant NAs
234 species accounting for more than 40% of total NAsFP, even more than 50% in
235 ozonation. The proportions of both NMor and NPip were about 20-30%, but that of
236 NMEA and NDEA were less than 5%. The proportions of different NAs species were
237 influenced by bromide level (Fig. 4). NDMA and NMEA decreased but NDEA and
238 NPip increased as the increasing bromide level in chlorination. NDMA decreased with
239 the increasing bromide levels in chloramination but slightly increased in ozonation.

240

241 **3.1.4. Effect of disinfectant dosage**

242 Fig. 5 illustrates the NAs levels with different disinfectant dosage. The formation of
243 the total NAs increased as disinfectant dosage rising. Moreover, there was a good
244 linear relationship between the concentration of total NAs and disinfectant dosage
245 ($R^2 > 0.98$), implying that disinfectant dosage was an important factor for the
246 formation of NAs in the range of our study. The effects of the disinfectant dosage on
247 total NAs levels in chlorination (slope = 40.8) were more than that in chloramination
248 (slope = 28.2).

249

250 **3.1.5. Effect of reaction time**

251 Fig. 6 shows the effect of reaction time on the formation of NAs. The concentrations
252 of NAs increased with increasing reaction time for the three disinfection methods.
253 During chlorination and ozonation, the total NAs levels first significantly increased
254 (chlorination: 1-5 d; ozonation: 0.5-2 h) and then slightly increased as the reaction
255 time continuously prolonged (chlorination: 5-7 d; ozonation: 2-12 h). While during
256 chloramination, the total NAs levels showed an obvious ($p < 0.05$) enhancement as
257 the reaction time grew from 1-7 d. The results were consistent with the stability of
258 disinfectant that chloramine can be long-term presence in the water supply network
259 due to its strong stability.³² If the precursors of NAs were not removed during water
260 treatment, they would react with chloramine to form more NAs during water
261 distribution.

262

263 **3.2. Key factors affecting NAs formation**

264 Key factors affecting NAs formation from SMPs in chlorination, chloramination and
265 ozonation were searched using a multivariate regression procedure and the results
266 were shown in Table 2. The regression coefficients ranged from 0.701 to 0.979, and
267 all terms were significant ($p < 0.05$).

268 Generally, the higher the partial correlations coefficients, the more important the
269 factor is. Among these factors, bromide level was generally more significant than
270 other factors during chlorination, chloramination and ozonation. Disinfectant dosage
271 was also an important influence on the formation of the total NAs and individual NAs
272 during chlorination and ozonation, but only on the formation of the total NAs during

273 chloramination. Additionally, temperature and pH showed some effect on the total
274 NAs during chlorination, however, these factors showed little effect during
275 chloramination and ozonation. These results indicated that reducing bromide level and
276 lowering the disinfectant dosage will be effective strategies to control the total NAs
277 formation whether for chlorination, chloramination or ozonation.

278

279 **3.3. NAsFP of real wastewater**

280 SMPs from two real wastewater treatment plants (A and B) were treated by
281 chlorination, chloramination and ozonation, respectively. The NAs species and
282 formation potential were almost the same for the two wastewater treatment plants (Fig.
283 7A-B). During chlorination and chloramination, the NAs species included NDMA,
284 NMor, NPip, NMEA and NDEA. During ozonation, there were only NDMA, NMor,
285 NPip and NMEA generated. The total NAsFP formed by chloramination (14.6 and
286 16.7 ng/mg DOC) were twice more than that by chlorination (6.3 and 7.0 ng/mg DOC)
287 and one order of magnitude more than that by ozonation (0.7 and 0.8 ng/mg DOC).
288 The proportions of different NAsFP from wastewater A and B were also almost the
289 same with that from synthetic wastewater. These results might be owing to their
290 similar water quality parameters including DOC and TN between the real wastewater
291 and synthetic wastewater despite of higher UV_{254} and bromide but lower DON in real
292 wastewater.

293

294 **3.4. NAs speciation and formation from SMPs under different disinfection**

295 **methods**

296 Through the analysis of the concentrations of NAs in the
297 chlorinated/chloraminated/ozonated SMPs samples with different treatments (Figs.
298 1-7), the NAsFP during disinfection of SMPs was about twice more than that from
299 disinfection of NOM,^{16,33} probably due to the higher portion of low molecular weight
300 (MW) hydrophobic acids and higher DON content in SMPs, which were associated
301 with NAs formation.^{14,18,34,35}

302 Furthermore, chloramination was induced to produce more NAs than chlorination
303 and ozonation. The most important NAs formation pathways during chloramination
304 were unsymmetrical dimethylhydrazine (UDMH) and chlorinated UDMH
305 (Cl-UDMH).^{27,36} Chloramine oxidized the precursors of NAs to form UDMH or
306 Cl-UDMH, and then UDMH or Cl-UDMH oxidation again by chloramine formed
307 NAs. For chlorination, there are two possible pathways: first, HOCl oxidized the
308 nitrite and nitrate to form dinitrogen tetroxide (N_2O_4), a nitrosation reagent with a
309 high reaction activity, and then N_2O_4 reacted with the precursors to form NAs.²⁶
310 Second, HOCl could react with ammonia to form chloramine, and then NAs was
311 produced through the UDMH and Cl-UDMH pathways.³⁷ However, HOCl could react
312 faster with the precursors of NAs (such as dimethylamine (DMA), the precursor of
313 NDMA) to form organic chloramines (such as chlorinated- dimethylamine (CDMA)),
314 which were stable substances and could not be continuously oxidized. Therefore, the
315 NAs produced from chlorination were less than that from chloramination. As to
316 ozonation, besides formaldehyde catalytic nitration and N_2O_4 nitration,²⁸

317 hydroxylamine mechanism was also an important pathway to generate NAs.²⁹ Ozone
318 oxidized the precursor to hydroxylamine, and then hydroxylamine reacted with the
319 precursor again to form NAs. The precursor consumed in this process was two times
320 of others, thus resulting less NAs formation during ozonation. Combining these
321 results, it could be concluded that chloramination may be a good alternative to
322 chlorination in terms of reducing C-DBPs in wastewater disinfection,¹⁴ but not a
323 better choice to control the formation of NAs.

324 Additionally, NDMA was always the major NAs in the three disinfection methods.
325 This may be related to the species and formation yields of the precursors. For example,
326 the formation yield of the precursor was NDMA > NMEA > NDEA during
327 chloramination of DMA, methylethylamine (MEA) and diethylamine (DEA).²⁹

328

329 **4. Conclusions**

330 Overall, five NAs were investigated during chlorination, chloramination and
331 ozonation of SMPs from synthetic and real wastewater. NDMA was the major NAs,
332 and the formation potential of NDMA accounted for more than 50% of total NAsFP.
333 Compared with chlorine and ozone, chloramine as the disinfectant generally resulted
334 in higher NAsFP. Ozonation was the best choice to reduce the formation of NAs in
335 wastewater disinfection. Bromide level and disinfectant dosage were two key factors
336 to affect the total NAsFP of SMPs from wastewater. NDMA, as the most important
337 NAs in SMPs disinfection, should be paid more attention on the precursor and
338 formation mechanism.

339

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345 **References**

- 346 1 R. Sadiq and M. J. Rodriguez, *Sci. Total Environ.*, 2004, **321**, 21–46.
- 347 2 J. Hu, H. Song and T. Karanfil, *Environ. Sci. Technol.*, 2010b, **44**, 794–799.
- 348 3 C. Jarusutthirak and G. Amy, *Water Res.*, 2007, **41**, 2787–2793.
- 349 4 E. Namkung and B. E. Rittmann, *Water Res.*, 1986, **20**, 795–806.
- 350 5 Z. P. Wang and T. Zhang, *Water Res.*, 2010, **44**, 5499–5509.
- 351 6 H. Zhang, J. H. Qu, H. J. Liu and X. Zhao, *J. Hazard. Mater.*, 2009, 164,
352 1433–1438.
- 353 7 J. L. Liu, X. Y. Li, Y. F. Xie and H. Tang, *Sci. Total Environ.*, 2014, **472**,
354 818–824.
- 355 8 S. D. Richardson, M. J. Plewa, E. D. Wagner, R. Schoeny and D. M. DeMarini,
356 *Mutat. Res.*, 2007, **636**, 178–242.
- 357 9 X. Yang, Q. Shen, W. Guo, J. Peng and Y. Liang, *Chemosphere*, 2012, **88**, 25–32.
- 358 10 Q. Luo, D. H. Wang and Z. J. Wang, *Sci. Total Environ.*, 2012, **437**, 219–225.
- 359 11 U. von Gunten, E. Salhi, C. Schmidt and W. A. Arnold, *Environ. Sci. Technol.*,
360 2010, **44**, 5762–5768.

- 361 12 M. J. Plewa, M. Muellner, S. D. Richardson, F. S. Cafasanok, K. M. Bueeener, Y.
362 T. Woo, A. B. Mckague and E. D. Wagner, *Environ. Sci. Technol.*, 2008, **42**,
363 955–961.
- 364 13 X. Yang, C. Shang and P. Westerhoff, *Water Res.*, 2007, **41**, 1193-1200.
- 365 14 H. C. Hong, Y. J. Xiong, M. Y. Ruan, F. L. Liao H. J., Lin and Y. Liang, *Sci.*
366 *Total Environ.*, 2013, **444**, 196-224.
- 367 15 H. C. Hong, L. Y. Qian, M. Y. Ruan, Y. J. Xiong, Z. Q. Xiao, H. J. Lin and H. Y.
368 Yu, *Chemosphere*, 2015, **119**, 540-546.
- 369 16 I. Kristiana, J. Tan, C. A. Joll, A. Heitz, U. von Gunten and J. W. A. Charrois,
370 *Water Res.*, 2013, **47**, 535–546.
- 371 17 J. Le Roux, H. Gallard and J. P. Croue', *Environ. Sci. Technol.*, 2012, **46**,
372 1581-1589.
- 373 18 B. B. Zhang, Q. M. Xian, J. P. Zhu, A. M. Li and T. T. Gong, *Chem. Eng. J.*, 2015,
374 **279**, 258-263.
- 375 19 Y. Y. Wei, Y. Liu, Y. Zhang, R. H. Dai, X. Liu, J. J. Wu, Q. Zhang, *Environ. Sci.*
376 *Pollut. Res.*, 2011, **18**, 46-50.
- 377 20 I. Kristiana, H. Gallard, C. Joll and J. P. Croue, *Water Res.*, 2009, **43**, 4177–4186.
- 378 21 L. P. Padhye, J. H. Kim and C. H. Huang, *Water Res.*, 2013, **47**, 725-736.
- 379 22 W. Q. Qi and S. N. Zeng, *Water and Wastewater Monitoring Analysis Method*,
380 China Environmental Science Press, Beijing, 2002.
- 381 23 R. Pozzi, P. Bocchini, F. Pinelli and G. C. Galletti, *J. Chromatogr. A*, 2011, **1218**,
382 1808-1814.

- 383 24 S. Liu and X. N. Wan, *J. Soil Water Conserv.*, 2005, **12**, 268-270.
- 384 25 X. Zhang, S. Echigo, R. A. Minear and M. J. Plewa, in *Natural Organic Matter*
385 *and Disinfection By-Products: Characterization and Control in Drinking Water*,
386 ed. S. E. Barrett, S. W. Krasner and G. L. Amy, American Chemical Society,
387 Washington D. C., 2000, p. 299.
- 388 26 J. H. Choi and R. L. Valentine, *Environ. Sci. Technol.*, 2003, **37**, 4871-4876.
- 389 27 I. M. Schreiber and W. A. Mitch, *Environ. Sci. Technol.*, 2005, **39**, 3811-3818.
- 390 28 L. K. Keefer and P. P. Roller, *Science*, 1973, **181**, 1245-1247.
- 391 29 L. Yang, Z. L. Chen, J. M. Shen, Z. Z. Xu, H. Liang, J. Y. Tian, Y. Ben, X. Zhai,
392 W. X. Shi and G. B. Li, *Environ. Sci. Technol.*, 2012, **43**, 5481-5487.
- 393 30 J. Luh and B. J. Marinas, *Environ. Sci. Technol.*, 2012, **46**, 5085-5092.
- 394 31 S. D. Richardson, A. D. Thruston, C. Rav-Acha, L. Groisman, I. Popilevsky, O. V.
395 Juraev, Glezer, A. B. McKague, M. J. Plewa and E. D. Wagner, *Environ. Sci.*
396 *Technol.*, 2003, **37**, 3782-3793.
- 397 32 H. Song, J. W. Addison and J. Hu, *Chemosphere*, 2010, **79**, 174-179.
- 398 33 C. K. Wang, X. J. Zhang, J. Wang and C. Chen, *J. Environ. Sci.*, 2013, **25**, 85-95.
- 399 34 Z. H. Fan, S. Gong, X. Xu, X. H. Zhang, Y. Zhang and X. Yu, *Int. J. Hyg. Envir.*
400 *Heal.*, 2014, **217**, 300-306.
- 401 35 A. Dotson, P. Westerhoff and S. W. Krasner, *Water Sci. Technol.*, 2009, **60**,
402 135-143.
- 403 36 I. M. Schreiber and W. A. Mitch, *Environ. Sci. Technol.*, 2006, **40**, 6007-6014.
- 404 37 A. D. Shah, W. A. Mitch, *Environ. Sci. Technol.*, 2012, **46**, 119-131.

Table 1 Experimental design

Factors	Chlorination	Chloramination	Ozonation
Temperature (°C)	25, 30, 40	25, 30, 40	25, 30, 40
pH	5.0, 7.0, 9.0	6.0, 8.0, 10.0	5.0, 7.0, 9.0
Reaction time	1, 3, 5, 7 (d)	1, 3, 5, 7 (d)	0.5, 2, 12 (h)
Disinfectant dosage	0.2, 0.5, 1.0, 2.0*	0.2, 0.5, 1.0, 2.0*	1, 2, 3, 4 (mg/L)
Bromide (mg/L)	0, 0.2, 0.5, 1.0	0, 0.2, 0.5, 1.0	0, 0.2, 0.5, 1.0
Baseline conditions	25°C, pH=7, 7d	25°C, pH=8, 7d	25°C, pH=7, 12h
	2.0*, Bromide =none	2.0*, Bromide= none	4 mg/L, Bromide=none

*molar ratio of disinfectant (chlorine or chloramine)/dissolved organic carbon (DOC)

Table 2 Results of regression procedure for NAs

disinfectant	DBPs	partial correlations coefficients					regression coefficients	p values
		Temperature	pH	disinfectant dosage	reaction time	bromide level		
chlorine	NDMA		-0.853	0.938	0.798	0.951	0.949	<0.01
	NMEA	-0.701	-0.955	0.976	0.962	0.948	0.975	<0.05
	NDEA					0.938	0.870	<0.01
	NMor		-0.896	0.961	0.918	0.980	0.976	<0.01
	NPip			0.554		0.920	0.851	<0.05
	NAs	-0.643	-0.893	0.959	0.894	0.975	0.971	<0.05
chloramine	NDMA					0.883	0.761	<0.01
	NMEA					0.912	0.817	<0.01
	NDEA					0.974	0.945	<0.01
	NMor					0.851	0.701	<0.01
	NPip					0.863	0.724	<0.01
	NAs			0.822		0.885	0.855	<0.01
ozone	NDMA			0.911	0.737	0.985	0.975	<0.01
	NMEA		0.827	0.765		0.991	0.979	<0.01
	NMor		0.635	0.908	0.725	0.958	0.940	<0.05
	NPip			0.732		0.952	0.914	<0.01
	NAs			0.891	0.689	0.977	0.963	<0.05

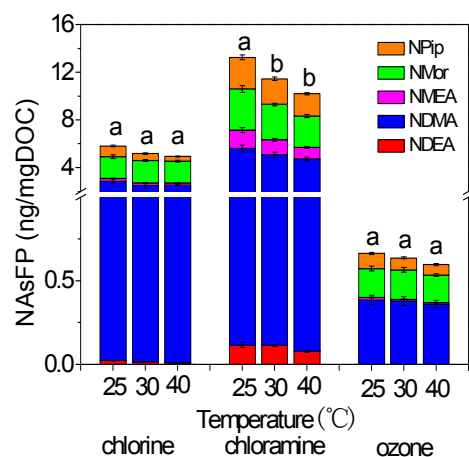


Fig. 1 NAsFP as a function of temperature after chlorination (pH=7, bromide=none, molar ratio of chlorine/DOC=2.0, reaction time=7d), chloramination (pH=8, bromide=none, molar ratio of chloramine/DOC=2.0, reaction time=7d) and ozonation (pH=7, bromide=none, ozone=4mg/L, reaction time=12h). Means with the same letter were not significantly different ($p > 0.05$) according to One Way Anova test (Duncan)

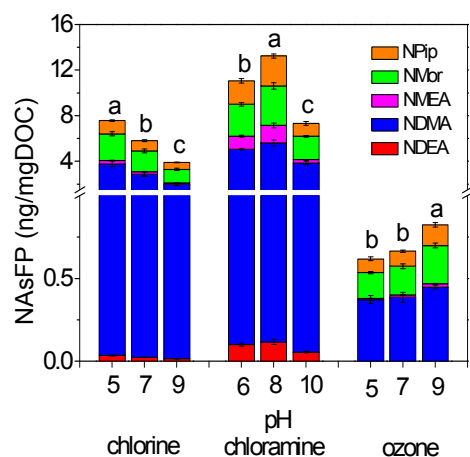


Fig. 2 NASFP as a function of pH after chlorination (temperature =25 °C, bromide=none, molar ratio of chlorine/DOC=2.0, reaction time=7d), chloramination (temperature =25 °C, bromide=none, molar ratio of chloramine/DOC=2.0, reaction time=7d) and ozonation (temperature =25 °C, bromide=none, ozone=4mg/L, reaction time=12h). Means with the same letter were not significantly different ($p>0.05$) according to One Way Anova test (Duncan)

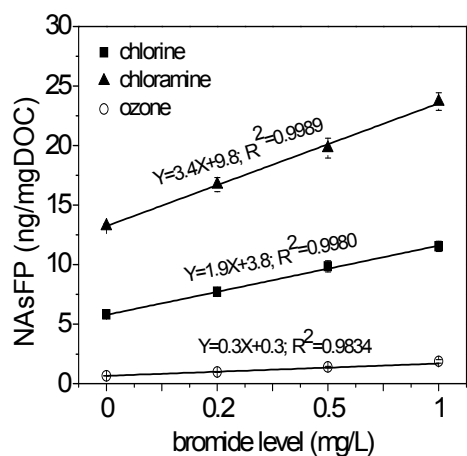


Fig. 3 NASFP as a function of bromide level after chlorination (temperature =25 °C, pH=7, molar ratio of chlorine/DOC=2.0, reaction time=7d), chloramination (temperature =25 °C, pH=8, molar ratio of chloramine/DOC=2.0, reaction time=7d) and ozonation (temperature =25 °C, pH=7, ozone=4mg/L, reaction time=12h). Means with the same letter were not significantly different ($p>0.05$) according to One Way Anova test (Duncan)

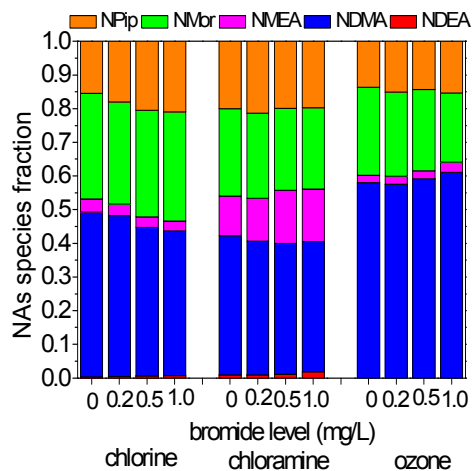


Fig. 4 NAs species fraction as a function of bromide level after chlorination (temperature =25 °C, pH=7, molar ratio of chlorine/DOC=2.0, reaction time=7d), chloramination (temperature =25 °C, pH=8, molar ratio of chloramine/DOC=2.0, reaction time=7d) and ozonation (temperature =25 °C, pH=7, ozone=4mg/L, reaction time=12h). Means with the same letter were not significantly different ($p>0.05$) according to One Way Anova test (Duncan)

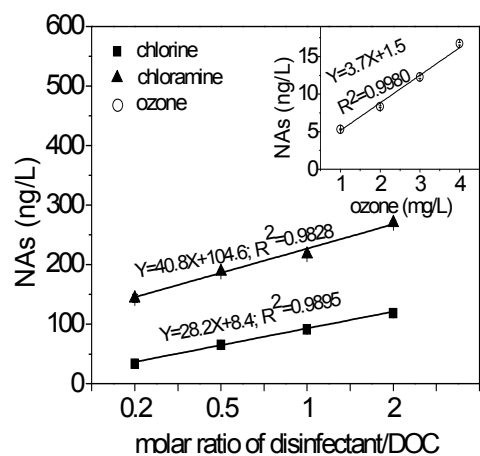


Fig. 5 NAs levels as a function of disinfectant dosage after chlorination (temperature =25 °C, pH=7, bromide level=none, reaction time=7d), chloramination (temperature =25 °C, pH=8, bromide level=none, reaction time=7d) and ozonation (temperature =25 °C, pH=7, bromide level=none, reaction time=12h). Means with the same letter were not significantly different ($p>0.05$) according to One Way Anova test (Duncan)

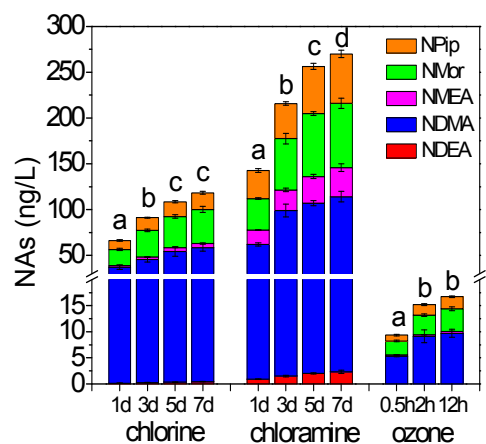


Fig. 6 NAs levels as a function of reaction time from chlorination (temperature =25 °C, pH=7, bromide level=none, molar ratio of chlorine/DOC=2.0), chloramination (temperature =25 °C, pH=8, bromide level=none, molar ratio of chloramine/DOC=2.0) and ozonation (temperature =25 °C, pH=7, bromide level=none, ozone=4mg/L). Means with the same letter were not significantly different ($p>0.05$) according to One Way Anova test (Duncan)

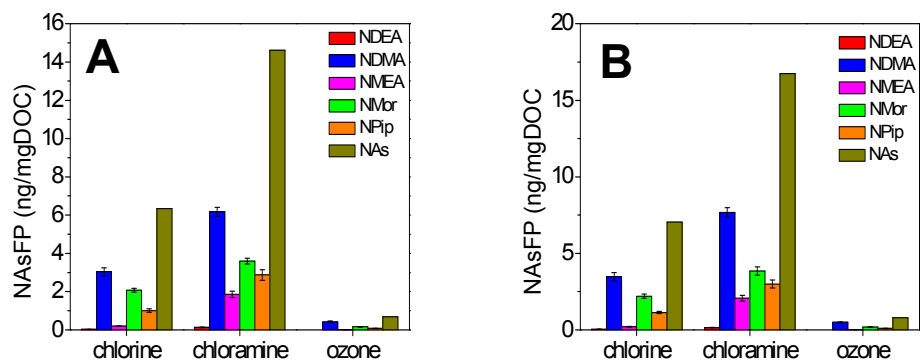
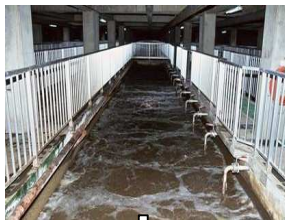
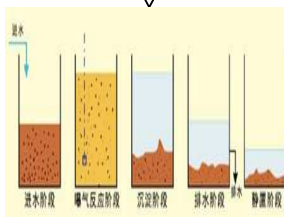


Fig. 7 NAsFP of two real domestic wastewater treatment plants (A and B) upon chlorination (temperature =25 °C, pH=7, bromide level=none, molar ratio of chlorine/DOC=2.0, reaction time=7d), chloramination (temperature =25 °C, pH=8, bromide level=none, molar ratio of chloramine/DOC=2.0, reaction time=7d) and ozonation (temperature =25 °C, pH=7, bromide level=none, ozone=4mg/L, reaction time=12h)



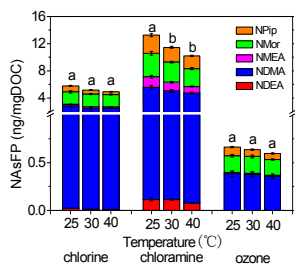
synthetic wastewater



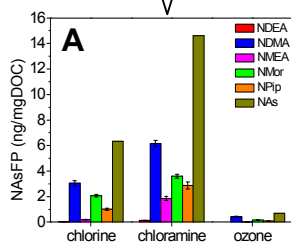
SBR treatment



SMPs $\xleftarrow{\text{disinfect}}$



NAsFP under different disinfection methods and factors



NAsFP of the real domestic wastewater treatment plants

partial correlations coefficients of NAs ^o					
disinfectant	Temperature ^o	pH ^o	disinfectant ^o dosage ^o	reaction time ^o	bromide ^o level ^o
chlorine ^o	ϕ	-0.643 ^o	0.959 ^o	0.894 ^o	0.975 ^o
chloramine ^o	ϕ	ϕ	0.822 ^o	ϕ	0.885 ^o
ozone ^o	ϕ	ϕ	0.891 ^o	0.689 ^o	0.977 ^o

Key factors affecting DBPs faormation