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1 **Growth Responses and Accumulation of Cadmium in Switchgrass (*Panicumvirgatum* L.)**
2 **and Prairie Cordgrass (*Spartinapectinata* Link)**

3

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22 Abstract

23 Phytoextraction could be an efficient technique to remediate heavy metals from contaminated
24 soils. Identifying bioenergy crops that can be produced successfully on marginal lands, such as
25 those polluted by heavy metals, also reduces the pressure to produce energy crops on land that
26 would otherwise be used to produce food crops. The objective of this study was to determine the
27 phytoremediation capability of two warm season perennials, prairie cordgrass (*Spartinapectinata*
28 Link, ‘Savoy’) and switchgrass (*Panicumvirgatum* L., ‘Cave-In-Rock’(CIR)) through their
29 growth response to cadmium (Cd). Growth rate, Cd tolerance, accumulation and translocation
30 were measured at concentrations of 0 (control), 5, 10, 30, and 50 mg L⁻¹ of Cd mixed with
31 Hoagland’s solution in an aerated hydroponic system. Although a reduction of plant growth was
32 observed when Cd concentration was higher than 10 mg L⁻¹ for both species, there were
33 significant differences in Cd tolerance, translocation and accumulation between species. The
34 tolerance index (T_i) was between 72.5 and 107.1 in Savoy and 48.7 and 75.7 in CIR under Cd
35 concentration of 50 mg L⁻¹ and 5 mg L⁻¹, respectively. The translocation factor (T_f) for both
36 species was increased with increasing Cd concentration in solution, but the T_f of Savoy was
37 higher than CIR. The highest bio-concentration factor (BCF) of the roots reached 325.7 for
38 Savoy and 144.5 for CIR when the Cd concentration was 5 mg L⁻¹ and the BCF of the shoots in
39 both species was consistently low (13.7 to 16.8 and 4.1 to 6.0 for Savoy and CIR, respectively)
40 indicating higher Cd retention in the roots than shoots. It was concluded that both species could
41 be utilized in phytoremediation when the Cd concentration is less than 10 mg L⁻¹, however
42 Savoy has the higher tolerance, translocation and accumulation capabilities which makes it a

43 better candidate for phytoremediation and biomass production on Cd polluted soils.

44 **Keywords:** Energy crops, Cadmium, Tolerance, Phytoremediation, Bio-concentration factor,

45 Translocation index

46

47

48 Introduction

49 Cadmium is a heavy metal that is toxic to humans, animals and plants and is a widespread,
50 highly toxic environmental pollutant with a long biological half-life (10-30 years in animals).¹
51 Taken up in excess by plants, Cd directly or indirectly inhibits physiological processes, such as
52 respiration, photosynthesis, cell elongation, plant and water relationships, nitrogen metabolism
53 and mineral nutrition, resulting in poor growth and low biomass.² Response of plants to Cd stress
54 is only partially understood. Cadmium may affect the uptake, transport and use of different
55 macronutrients and micronutrients, especially Fe and Zn.^{3,4} Cadmium is not essential to plant
56 growth, and Cd accumulation in plant tissue can cause various phytotoxic symptoms including
57 leaf chlorosis, root putrescence, and growth inhibition. The tolerable range of Cd concentration in
58 leaf tissue (dry weight) of various agronomic crops is 0.05-0.2 mg kg⁻¹, and it is considered to
59 reach excessive or toxic level within a range of 5-30 mg kg⁻¹.⁵ The accumulation of Cd within
60 the aquatic environment, sediments, and soils is a concern because the Cd taken up by plants can
61 be passed on to humans through the food chain. Therefore, it is important to develop methods to
62 remediate Cd-polluted soils.

63 Phytoremediation is a process where plants are used to degrade, extract, contain or
64 immobilize contaminants from soil and water.^{6,7} It is a cost-effective, environmentally friendly,
65 and technically applicable technology used to restore polluted sites in situ.⁸⁻¹⁰ A plant species
66 with an exceptional capacity to accumulate metals is the key for phytoremediation.⁹⁻¹¹ Potential
67 candidates must be high biomass producers and have strong developed root systems, excellent
68 transpiration, and the ability to remove heavy metals from contaminated soils.^{12,13} Although over

69 450 plant species have been identified as hyperaccumulators of heavy metals, only a few are
70 known to hyperaccumulate Cd.^{6,13-18} Grasses such as *Festuca ovina* L., *F. rubra* L.,
71 *Agrostis capillaris* L., *A. delicatula* Pourr.ex Lapeyr and *A. stolonifera* L. have high degrees of
72 metals tolerance but they do not hyperaccumulate metals.¹⁹

73 Both prairie cordgrass and switchgrass are C₄ perennial rhizomatous grasses and have been
74 recognized as excellent energy crops for marginal lands.²⁰⁻²² Several studies have demonstrated
75 the tolerance of prairie cordgrass and certain switchgrass cultivars for high soil salinity.²³⁻²⁵

76 Prairie cordgrass is well adapted to soils that are wet throughout the growing season. Even
77 though there is limited information available on stress tolerances of prairie cordgrass including
78 abiotic stress and heavy metal toxicity, *Spartina* spp is known as highly stress tolerant. Two other
79 species in the genus, austral cordgrass (*S. densiflora* Brongon) has proven to be a vigorous plant,
80 which can grow well in salt marshes²⁶ and is tolerant of very high levels or continues exposure to
81 Zn.²⁷ Prairie cordgrass has been reported to have a high tolerance for conditions of water logging,
82 high salinity, and low pH.^{28,29} Certain switchgrass cultivars have been reported to be tolerant of
83 Cd when grown in sand.³⁰ Switchgrass can be grown on Cd-contaminated sites to produce
84 acceptable levels of biomass; however, the plant material produced should not be used as animal
85 feed without testing and verification of the Cd concentration.³⁰

86 However, no direct comparison has been made between switchgrass and prairie cordgrass
87 for Cd tolerance or performance on marginal land. Since prairie cordgrass and switchgrass play
88 an important role for feedstock production on marginal land, direct comparison between two
89 species will provide useful information for developing the future energy crops for sustainable

90 bioenergy feedstock on marginal land. The objective of this study was to determine the effects of
91 Cd on plant growth, biomass distribution, and Cd accumulation in ‘Cave-in-Rock’ switchgrass
92 (*Panicum virgatum* L.) and ‘Savoy’ prairie cordgrass (*Spartina pectinata* Link), there by
93 assessing Cd toxicity tolerance and ability to accumulate Cd in biomass.

94

95 **Materials and Methods**

96 The seeds of Savoy prairie cordgrass (Savoy) originated from central Illinois, USA and were
97 obtained from our breeding program, and Cave-in-Rock switchgrass (CIR) seeds were purchased
98 from a commercial seed source (Millborn Seeds Inc., Brookings, SD, USA).

99 A hydroponic experiment was conducted in a controlled greenhouse setting at the University
100 of Illinois, Urbana, IL, USA. A 20-day-old seedling was transplanted into a 3 L pot containing
101 20% Hoagland’s nutrient solution³¹ without Cd. After 5 days of seedling culture, the solution
102 was replaced by a 100% Hoagland’s nutrient solution without Cd for another 5 days of seedling
103 culture, and then exposed to a 100% Hoagland’s nutrient solution with 0, 5, 10, 30, 50 or 100
104 Cd²⁺ mg L⁻¹. The solutions with different Cd²⁺ concentrations were refreshed every three days.
105 The six treatments for each species were performed in a randomized complete block design and
106 each treatment was replicated four times and the experiment was repeated twice. The pH of
107 solution was maintained at 6.0±0.5 modification with 1 mol L⁻¹ HCl or 1 mol L⁻¹ NaOH every
108 day. Air was pumped with EcoPlus Air Pumps for 30min every 3 hr each day in order to keep the
109 dissolved oxygen in solution. The plants were harvested 20 d after exposure to the Hoagland’s
110 nutrient solutions containing Cd. Biomass was measured as the fresh weight (g F.W) of plant

111 from both before and after treatment.

112 The longest roots and shoots were recorded at the initial treatment with Cd and at harvest.
113 The elongation of the longest roots and height of the shoots were calculated using the final length
114 at harvest minus the initial length. The number of tillers was observed before treated with Cd and
115 at harvest. The increase in the number of tillers was calculated as the tillers at treatment with Cd
116 subtracted from the total tillers at harvest. The fresh biomass of energy crops was recorded at
117 harvest and the plant samples were washed with tap water and rinsed with distilled water three
118 times. The washed samples were separated into roots and shoots, then the roots were immersed
119 into a 500 ml solution of Na₂-EDTA (10 mmol L⁻¹) for 10 min to remove the Cd attached to the
120 root surface. The samples were washed again with distilled water before further processing. The
121 roots and shoots were dried at 105°C for 30 min and then at 65°C for 2 d, and the dried root
122 weight and shoot weight were measured. Then, the dry plant tissues were ground to pass a 1mm
123 screen to determine the concentration of Cd.

124 The plant tissue samples were digested with microwave assisted acid digestion.³¹ 0.2500 g
125 representative plant samples, 9 mL of concentrated nitric acid and 3 mL hydrofluoric acid were
126 placed in a 50mL inert polymeric microwave vessels, and then the vessels were sealed and
127 heated in the microwave system for 15 minutes at 180±5 °C temperature. After cooling, the
128 vessel contents were filtered and then decanted, diluted to 100mL, and analyzed by inductively
129 coupled plasma-atomic emission spectrometry (Optima8000, PE Company, USA).³²

130 The tolerance index (T_i) was calculated to measure the ability of the plant to grow in the
131 presence of a given concentration of Cd.^{33,34}

$$T_i(\%) = \frac{W_{Cd}}{W_{CR}} \times 100$$

132

133 where W_{Cd} is the total dry biomass of plants grown in Hoagland's nutrient solution with
134 different Cd^{2+} concentrations (mg) and W_{CR} is the total dry biomass of plants grown in the
135 control solution without Cd (mg).

136 The translocation factor (T_f) was calculated to evaluate the capability of the plant to
137 accumulate Cd in the roots and shoots:³⁴

$$T_f(\%) = \frac{C_{Cd,shoots}}{C_{Cd,roots}} \times 100$$

138

139 where $C_{Cd,shoots}$ is the Cd concentration in the shoots ($mg\ kg^{-1}$) and $C_{Cd,roots}$ is the Cd
140 concentration in the roots ($mg\ kg^{-1}$).

141 The Cd bio-concentration factor (BCF) was calculated as follows:³⁵

$$BCF = \frac{C_{Cd,plant}}{C_{Cd,solution}}$$

142

143 where $C_{Cd,plant}$ is the Cd concentration in the plant ($mg\ kg^{-1}$) and $C_{Cd,solution}$ is the Cd
144 concentration in the solution ($mg\ L^{-1}$).

145 Analysis of variance (ANOVA) for Cd concentration, species, and their interaction was
146 performed with a significance level of $P < 0.05$ using the GLM procedure of SAS software (SAS
147 Institute, Cary, NC). The effect of experiments was considered random and was not significant.
148 The data presented are the least squared mean values and is compared as the mean separations
149 using Tukey's Studentized Range (HSD) test ($P\text{-value} < 0.05$).

150

151 Results

152 *Cadmium tolerance and growth response*

153 The tolerance of plants to toxic metals is frequently measured by comparing the rates of
154 root and shoot growth in culture solutions with and without the addition of a target heavy metal.
155 ³³ Plant growth responses including root and shoot growth, number of tillers, and biomass under
156 Cd stress for 20 days are shown in Figures 1, 2, 3, and 4, respectively, which indicate that growth
157 was diminished by Cd stress. Negative effects of Cd on plant growth responses of both grasses
158 were increased by increasing Cd concentration in the solution. Root growth of Savoy was not
159 significantly different from the control when the concentration of Cd in Hoagland's solution was
160 less than 10 mg L⁻¹ (Fig. 1). However, root growth of Savoy was significantly inhibited when the
161 concentration of Cd was more than 30 mg L⁻¹. Roots of Savoy stopped growing and began to
162 decay when the concentration of Cd was more than 50mg L⁻¹, but Savoy continued to survive
163 at 100 mg L⁻¹. Cave-in-Rock was more sensitive to Cd and its growth was inhibited when the
164 concentration of Cd was 5 mg L⁻¹ or higher (Fig. 1; Fig. 5). Root elongation of CIR stopped
165 growing and began to decay when the concentration of Cd was more than 10 mg L⁻¹.
166 Cave-in-Rock plants died when the concentration of Cd was higher than 50 mg L⁻¹ and therefore no
167 data was shown at this concentration.

168 The plant height was also affected by Cd concentration in solution. Shoot growth of Savoy
169 and CIR was significantly decreased under all Cd concentrations (Fig. 2; Fig. 5). Shoot growth in
170 both grasses maintained 50% level of the control when the concentration of Cd was 5mg L⁻¹, and
171 growth was negligible when the concentration of Cd reached higher than 10mg L⁻¹, as compared

172 with the control (Fig. 2).The number of tillers of Savoy and CIR responded differently to Cd
173 concentrations in solution (Fig. 3). The number of tillers of CIR continuously decreased as Cd
174 concentration increased. However, the number of tillers of Savoy under any given Cd
175 concentrations was not different, though they were lower than the control. The biomass yield of
176 both grasses decreased under Cd stress, but biomass accumulation patterns between grasses were
177 different (Fig.4).For Savoy, biomass with the control and 5 mg L⁻¹ was not different, decreased
178 from 5 mg L⁻¹ to 10 mg L⁻¹, and were not different under Cd concentrations between 10 and 50
179 mg L⁻¹. However, CIR biomass accumulation continuously decreased as Cd concentration
180 increased and no biomass accumulated above 30 mg L⁻¹.

181 From the growth response of both grass species to Cd stress, including the elongation of
182 longest roots, height of shoot, the number of tillers and total biomass, it was generally observed
183 that the tolerance to Cd of Savoy was higher than that of CIR. The difference between the two
184 species with regards to Cd tolerance was further confirmed by the analysis of the tolerance index,
185 T_i (Fig.6). On the basis of the total dry biomass of each species, the T_i revealed that on average
186 Savoy had a greater tolerance to Cd than CIR. The $T_i\%$ of Savoy and CIR was 107.13% and
187 75.74%, respectively, when the Cd concentration was 5 mg L⁻¹ which indicated that the growth
188 was not affected. But the growth of both Savoy and CIR was affected when Cd concentration
189 was 10mg L⁻¹ or above.

190 ***Accumulation of Cd***

191 The concentration of Cd detected in the roots and shoots of Savoy and CIR after exposure
192 for 20 days to various concentrations of Cd is shown in Fig. 7. In the control, the Cd

193 concentration in the shoots of both grasses was below the threshold of detection. High
194 concentrations of Cd were detected in the roots and shoots of both grasses and Cd concentrations
195 increased as the Cd concentration increased in the solution. The roots and shoots of Savoy
196 accumulated Cd ranging from 1628.7 to 2912.0 mg kg⁻¹ and 74.8 to 416.2 mg kg⁻¹, respectively
197 for the various Cd treatments. The roots and shoots of CIR accumulated Cd ranging from 722.7
198 to 1695.7 mg kg⁻¹ and 21.1 to 145.6 mg kg⁻¹, respectively. Root accumulation of Cd represented
199 approximately 87% to 96% and 92% to 97% of total accumulation in biomass of Savoy and CIR,
200 respectively. The Cd concentration in Savoy was 2.2 to 4.0 times higher in the shoots and 1.7 to
201 2.8 times higher in the roots than in CIR at the same Cd treatment.

202 *The Cd bio-concentration factor (BCF) and translocation factor (T_f)*

203 To evaluate the ability of Savoy and CIR to extract and accumulate Cd in the plant, the
204 bio-concentration factor (BCF) was calculated. The root BCF was higher than the shoot BCF in
205 both Savoy and CIR, being 7 to 22 times and 12 to 34 times higher, respectively and Savoy BCF
206 was higher than CIR (Fig. 8). In the roots of both species, the BCF decreased as the Cd
207 concentration increased. However, in the shoots of both species, the BCFs were not different
208 among Cd treatments.

209 The capability of Savoy and CIR to accumulate Cd in the above ground tissues was further
210 confirmed by calculating the translocation factor (T_f), which indicated the percentage of the
211 amount of absorbed metal that reached the shoots with respect to the amount present in the roots
212 (Fig. 9). The T_f in both energy crops showed different translocation capabilities. The higher T_f
213 was observed in Savoy, and the T_f of both Savoy and CIR increased as the concentration of Cd

214 increased in the solution.

215

216 **Discussions**

217 Metal tolerance, and consequently the protection of the integrity and functionality of the
218 primary physiological and metabolic processes, is an essential pre-requisite for a plant to be
219 utilized in phytoremediation.³⁶ In this study, the responses of Savoy and CIR to Cd stress, which
220 were analyzed by growth parameters (elongation of longest roots, height of the shoot, the number
221 of tillers, total biomass) and Cd uptake, indicated there were significant differences in tolerance,
222 accumulation and translocation of Cd. These fundamental aspects should form the criteria used
223 to screen plants for selection in phytoremediation. However, phytoremediation needs to be a
224 combination of heavy metal accumulation and reduction of damaging effects on biomass
225 production, and not merely due to metal extraction.³⁴ Metal tolerance could be estimated based
226 on root elongation and growth since tolerance at the root level represents the first step in metal
227 absorption and loading into the xylem vessels.^{37,38} Previous research has shown that 40 μM (4.49
228 mg L^{-1}) and 1 μM (0.11 mg L^{-1}) the solution with Cd^{2+} can reduce growth of barley and soybean,
229 respectively, by 50%.^{39,40} Indian mustard (*Brassica juncea* L. Czern.) showed a decline in growth
230 vigor under 25 μM (2.81 mg L^{-1}) the solution with Cd^{2+} concentration even though it is
231 considered a Cd hyperaccumulator.^{41,42} In a similar study, *P. australis* showed a 50% depression
232 in root elongation when grown in Cd concentrations ranging from 2.85 to 3.20 μM (0.32 mg L^{-1}
233 to 0.36 mg L^{-1}), and the T_i was reduced to less than 80% when the Cd concentration was above
234 8.8 μM (0.99 mg L^{-1}).³⁵ Reed³⁰ indicated that different switchgrass cultivars showed different

235 levels of tolerance to Cd in solution, and approximately one third of the control biomass yield
236 was lost when grown at 16 mg L⁻¹ Cd concentration. In this study, Savoy and CIR were tolerant
237 to the Cd concentration of 5 mg L⁻¹ Cd, with a T_i of 92.2% and 87.9%, while Cd concentrations
238 above 10 mg L⁻¹ reduced the T_i to 60.8% and 57.7%, respectively. These results suggested that
239 both energy crops were tolerant to Cd stress.

240 Based on the total dry biomass of each energy crop, the T_i revealed that on average Savoy
241 absorbed much more Cd than CIR. Most vascular herbaceous species accumulate greater
242 concentrations of heavy metals in the roots than that in shoots.^{43,44} Under our experimental
243 conditions, the roots and shoots of Savoy accumulated Cd to the highest concentrations of 2912.0
244 mg kg⁻¹ and 416.2 mg kg⁻¹, respectively, with a BCF ranging 325.7 and 16.8 respectively, for the
245 different Cd treatments. The roots and shoots of CIR accumulated Cd to the highest
246 concentrations of 1695.7mg kg⁻¹ and 145.6mg kg⁻¹ with a BCF ranging from 56.5 to 144.5 and
247 4.2 to 6.0, respectively for the different Cd treatments. Our results demonstrated that, even
248 though the majority of Cd was found in the roots, substantial amounts were also in the shoots.
249 The Cd concentration in both grasses was higher than 100 mg kg⁻¹ dry mass. This level reached
250 the threshold concentration defined by Van der Ent for a Cd hyperaccumulator.¹⁸ In this study,
251 the bioconcentration factor (BCF) values under lower concentration of Cd treatment indicated
252 that Savoy had double or triple the capacity to remove metal from the solution compared to CIR.
253 In a study comparing two wooden species (poplar and willow) seedlings performance under a
254 similar condition, Zacchini reported a range of BCF values from 52 to 290 in roots and 2.5 to
255 14.5 in shoots, respectively.⁴⁵ Other studies on herbaceous plants showing higher BCF values

256 compared to the current experiment were largely conducted under lower cadmium concentrations
257 or trace element supply.^{35,36} The ability to accumulate heavy metal in the shoots and roots can be
258 better illustrated by calculating the translocation factor or (T_f). In this study, the range of the T_f of
259 Savoy and CIR was 4.6 to 14.3 and 2.9 to 8.6, respectively. Due to a different experimental
260 setting, the T_f measured in this study is lower than that indicated in other plants cultivated in
261 soil.^{46,47} In general, Savoy showed a higher Cd accumulation capability in its harvestable parts
262 and accumulated more Cd per plant base since it produced more biomass than CIR under each
263 soluble Cd concentration. Wetland plants are generally not hyperaccumulators according to the
264 definition by Baker & Brooks⁴⁸ and Wei & Zhou⁴⁹. However, perennial wetland plants can store
265 metals in below ground parts other than roots. In a study comparing four emergent rhizome type
266 wetland plants, Zhang found that a significant amount of Cd was accumulated by rhizome under
267 soluble Cd concentration 20 mg L⁻¹.⁵⁰ In engineering a remediation system on marginal land,
268 especially wetlands, the perennial nature of these two species can provide both roots and
269 rhizomes as preferable alternatives to the destructive mechanical harvesting of aboveground
270 tissue. Moreover, both species in the current study were able to survive under Cd concentration
271 in solution at 10 mg L⁻¹ which already causes net weight loss for other hyperaccumulators such
272 as water hyacinth (*Eichhorniacrassipes*) and duckweed (*Lemnaminor L.*).^{51, 52}

273 In conclusion, the evaluation of the T_i , T_f and BCF for Savoy prairie cordgrass and CIR
274 switchgrass in this study confirms that both grass species have a considerable potential to remove
275 Cd from a contaminated environment with Savoy having a greater potential to remediate polluted
276 soils because it has a higher tolerance, greater ability to translocate heavy metals to its shoots,

277 and is a good Cd accumulator. The high accumulation in belowground tissues of both species
278 could be used either as rhizofiltrations to remediate wastewater or phytostabilization to limit Cd
279 flowing into water.

280

281 Acknowledgements

282 Research was supported by the National Natural Science Foundation of China
283 (No.41461091) and the National Natural Science Foundation of Guangxi, China
284 (2014GXNSFAA18039, 2015GXNSFEA139001) and was funded by the Energy Biosciences
285 Institute.

286 References

- 287 1. M. Arabi and A. A. Mohammadpour. *Veterinary Research Communications*, 2006, 30(8),
288 943-951.
- 289 2. L. S. Di Toppi and R. Gabbrielli. *Environmental and Experimental Botany*, 1999, 41(2),
290 105-130.
- 291 3. P. Das, S. Samantaray and G. R. Rout. *Environmental Pollution*, 1997, 98(1), 29-36.
- 292 4. P. Vollenweider, C. Cosio, M. S. Günthardt-Goerg and C. Keller, *Environmental and*
293 *Experimental Botany*, 2006, 58(1), 25-40.
- 294 5. A. Kabata-Pendias, H. Pendias, *Trace elements in soils and plants*. 3rd ed. Boca Raton, Florida:
295 CRC Press, 2001.
- 296 6. I. Raskin, R. D. Smith and D. E. Salt, *Current Opinion in Biotechnology*, 1997,8(2), 221-226.
- 297 7. M. Mench, N. Lepp, V. Bert, J. P. Schwitzguébel, S.W. Gawronski, P. Schröder, J.
298 Vangronsveld, *Journal Soils Sediments*, 2010,16: 876-900.
- 299 8. W. H. O. Ernst, CRC Press, Boca Raton, Florida ,1990, pp. 211-237
- 300 9. D. E. Salt, M. Blaylock, N. P. Kumar, V. Dushenkov, B. D. Ensley, I. Chet and I. Raskin,
301 *Bio/Technology*, 1995, (13), 468-474.
- 302 10. A. Kabata-Pendias, *Geoderma*, 2004, 122(2), 143-149.
- 303 11. M. Ghosh, & S. P. Singh, *Environmental Pollution*, 2005, 133(2), 365-371.
- 304 12. E. Lombi, F. J. Zhao, S. J. Dunham and S. P. McGrath, *Journal of Environmental Quality*,
305 2001,30(6), 1919-1926.
- 306 13. I. D. Pulford and C. Watson, *Environment International*, 2003, 29(4), 529-540.
- 307 14. A. J. M. Baker, S.P. McGrath, C. M. D. Sidoli, R.D. Reeves, *Resources Conservation and*
308 *Recycling*, 1994, 11, 41-49
- 309 15. H. Küpper, E. Lombi, F. J. Zhao and S. P. McGrath, 2000, *Planta*, 212(1), 75-84.
- 310 16. J. T. Li, B. Liao, C. Y. Lan, Z. H. Ye, A. J. M. Baker and W. S. Shu, *Journal of Environmental*
311 *Quality*, 2010,39(4), 1262-1268.
- 312 17. S. Tian, L. Lu, J. Labavitch, X. Yang, Z. He, H. Hu and P. Brown, *Plant physiology*, 2011,
313 157(4), 1914-1925.
- 314 18. A. Van der Ent, A. J. M. Baker, R. D. Reeves, A. J. Pollard, H. Schat , *Plant and Soil*, 2013,
315 362,319-334.
- 316 19. W. H. O. Ernst, *Applied geochemistry*,1996, 11(1), 163-167.
- 317 20. A. Boe, V. Owens, J. Gonzalez-Hernandez, J. Stein, D. K. Lee and B. C. Koo, *GCB*

- 318 Bioenergy, 2009,1(3), 240-250.
- 319 21. D. H. Vaughan, J. S. Cundiff and D. J. Parrish, Biomass, 1989, 20(3), 199-208.
- 320 22. D. J. Parrish and J. H. Fike, Critical Reviews in Plant Sciences, 2005, 24(5-6), 423-459.
- 321 23. S. Kim, A. L. Rayburn, T. Voigt, A. Parrish and D. K. Lee, BioEnergy Research, 2012, 5(1),
322 225-235.
- 323 24. E. K. Anderson, T. B. Voigt, S. Kim and D. K. Lee, Industrial Crops and Products, 2015, 64,
324 79-87.
- 325 25. Y. Liu, X. Zhang, J. Miao, L. Huang, T. Frazier and B. Zhao, BioEnergy Research, 2014, 7(4),
326 1329-1342.
- 327 26. P. M. Kittelson and M. J. Boyd, Estuaries, 1997,20(4), 770-778.
- 328 27. E. Mateos-Naranjo, S. Redondo - Gómez, J. Cambrollé, T. Luque and M. E. Figueroa, Plant
329 Biology, 2008, 10(6), 754-762.
- 330 28. M. B. Montemayor, J. S. Price, L. Rochefort and S. Boudreau, Environmental and
331 Experimental Botany, 2008, 62(3), 333-342.
- 332 29. M. B. Montemayor, J. S. Price, L. Rochefort and S. Boudreau, Environmental and
333 Experimental Botany, 2010, 69(2), 87-94.
- 334 30. R. L. Reed, M. A. Sanderson, V. G. Allen and R. E. Zartman, Communications in Soil
335 Science and Plant Analysis, 2002,33(7-8), 1187-1203.
- 336 31. EPA, Method 3052, Revision 0, Dec. 1996.
- 337 32. EPA, Method 6010B, Revision 2, Dec. 1996.
- 338 33. D. A. Wilkins, New Phytologist, 1978, 80(3), 623-633.
- 339 34. M. M. Lasat, Journal of Environmental Quality, 2002, 31(1), 109-120.
- 340 35. A. Zayed, S. Gowthaman and N. Terry, Journal of Environmental Quality, 1998,27(3),
341 715-721.
- 342 36. N. A. Ali, M. P. Bernal and M. Ater, Aquatic Botany, 2004, 80(3), 163-176.
- 343 37. M. H. Wong and A. D. Bradshaw, New Phytologist, 1982, 91(2), 255-261.
- 344 38. D. L. Godbold and C. Kettner, Journal of Plant Physiology, 1991,138(2), 231-235.
- 345 39. A. L. Page, F. T. Bingham and C. Nelson, Journal of Environmental Quality, 1972,1(3),
346 288-291.
- 347 40. I. V. Seregin and V. B. Ivanov, Russian Journal of Plant Physiology, 2001, 48(4), 523-544.
- 348 41. G. S. Banuelos and D. W. Meek, Journal of Environmental Quality, 1990, 19(4), 772-777.
- 349 42. A. Haag-Kerwer, H. J. Schäfer, S. Heiss, C. Walter and T. Rausch, Journal of Experimental
350 Botany, 1999,50(341), 1827-1835.

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- 351 43. W. F. Punz and H. Sieghardt, *Environmental and Experimental Botany*, 1993, 33(1), 85-98.
- 352 44. Y. Q. Zu, Y. Li, J. J. Chen, H. Y. Chen, L. Qin and C. Schwartz, *Environment International*,
- 353 2005,31(5), 755-762
- 354 45. M. Zacchini, F. Pietrini, G. S. Mugnozza, V. Iori, L. Pietrosanti and A. Massacci, *Water, Air,*
- 355 *and Soil Pollution*, 2009,197(1-4), 23-34.
- 356 46. M. I. Mattina,W. Lannucci-Berger, C. Musante, and J. C.White, *Environmental Pollution*,
- 357 2003,124(3), 375-378.
- 358 47. H. Deng, Z. H. Ye and M. H. Wong, *Environmental Pollution*,(2004,132(1), 29-40.
- 359 48. A. J. M. Baker and R. Brooks, *Biorecovery*, 1989, 1(2), 81-126.
- 360 49. S. Wei and Q. Zhou, *Progress in Natural Science*, 2004, 14(6), 495-503.
- 361 50. Z. Zhang, Z. Rengel and K. Meney, *Water, Air, & Soil Pollution*, 2010, 212(1-4), 239-249.
- 362 51. Y. L. Zhu, A. M. Zayed, J. H. Qian, M. De Souza and N. Terry, *Journal of Environmental*
- 363 *Quality*, 1999,28(1), 339-344.
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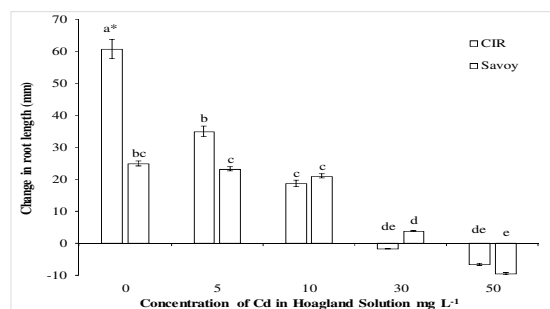
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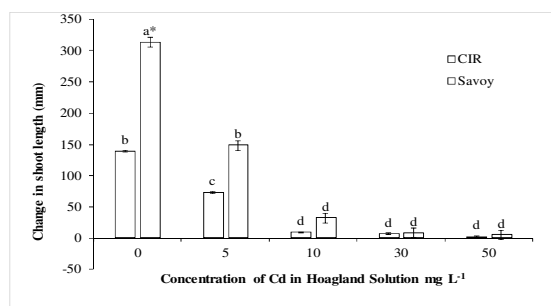
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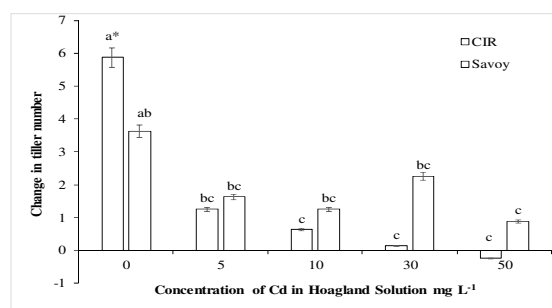
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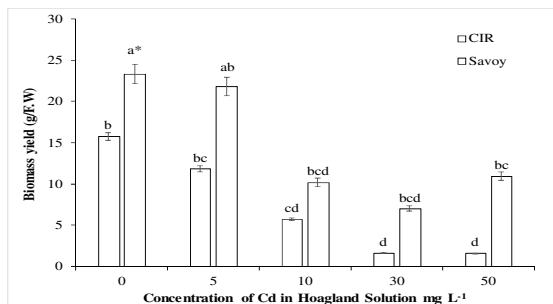


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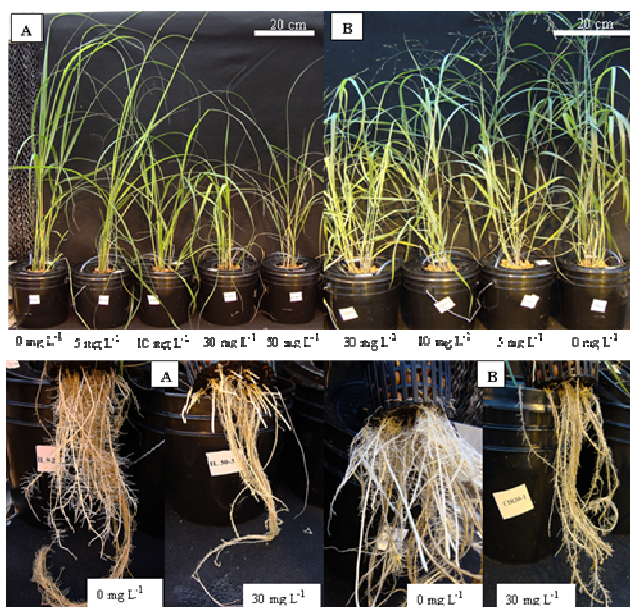
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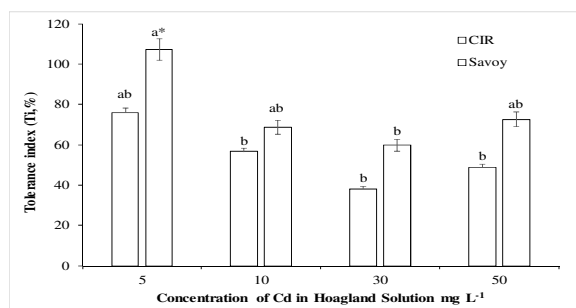
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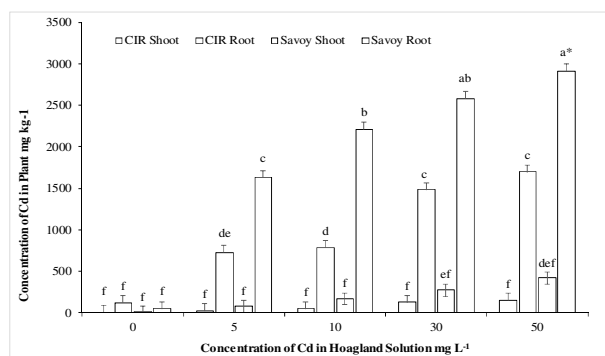
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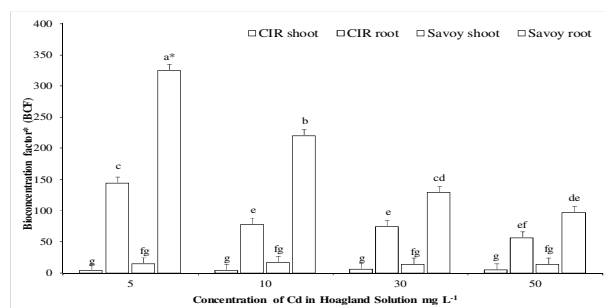
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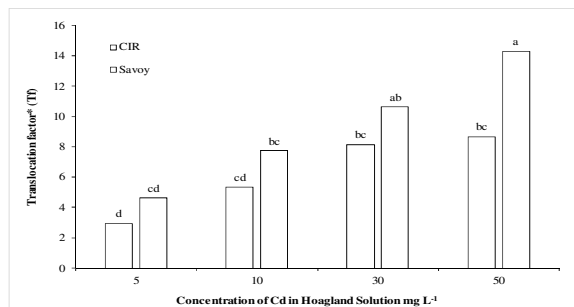
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