

Cholestatic liver injury model of bile duct ligation and the protection of Huang-Lian-Jie-Du decoction by NMR metabolomic profiling

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25 Abstract

Cholestatic liver injury has been increasingly recognized as a cause for high morbidity 26 and mortality of some diseases in human. This model could be established by bile duct 27 ligation (BDL), which led to the toxic accumulation of bile acids in animals, resulting 28 in cholestatic liver injury. In this study, rats were intragastrically administrated with an 29 extract of Huang-Lian-Jie-Du decoction once a day for seven consecutive weeks to 30 study its therapeutic effect. Serum and urine samples were collected and subjected to 31 ¹H NMR-based metabolomic analysis. Perturbations on energy metabolism, amino 32 acid metabolism, gut bacteria metabolism and oxidative stress were observed in BDL 33 rats. The metabolomic pattern showed a distinct biphasic feature of BDL model. Most 34 of these metabolic disturbances occured in acute phase (week 1) were greatly 35 attenuated in the long run. HLJDD ameliorated the disturbed metabolism throughout 36 this model, showing bilateral adjustment of some metabolites varied in opposite 37 direction in the two phases. This study demonstrated that ¹H NMR-based 38 metabolomics approach is a powerful and feasible tool to study the pathological 39 changes of a disease model dynamically and holistically and for the understanding of 40 the therapeutic effects of complex Chinese herbal medicine formula. 41

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43 **1. Introduction**

Chronic cholestatic liver disease is one of the major risks responsible for the
development of liver cirrhosis and end-stage liver disease culminating in liver failure.
Cholestatic liver disease occurs when there is a decrease in bile flow,^{1, 2} which is a

common pathological condition that can be reproduced in rodents by common bile 47 duct ligation (BDL) during surgical laparotomy.³ Bile acids (BAs) are the active 48 constituents of bile and essential for absorption and solubilization of dietary lipids in 49 the digestive tract.⁴ Of them, hydrophobic bile acids could induce damage of 50 mitochondrial membrane structure and increase of oxidative stress, leading to 51 apoptosis in liver cells.^{5, 6} The obstruction of bile flow results in an increased 52 accumulation of potentially harmful hydrophobic bile acids in the liver and blood, and 53 liver dysfunction.⁷ 54

Cholestasis is associated with many liver diseases, and a well-established experimental
animal model is BDL in rodents, in which hydrophobic bile acid mediated liver injury.
This model develops in a biphasic manner, including acute (phase 1) and chronic
(phase 2) cholestasis.⁸

Huang-Lian-Jie-Du decoction (HLJDD) is a herbal formula of Traditional Chinese 59 Medicine, consisting of Coptidis rhizoma, Radix Scutellariae, Cortex Phellodendri 60 and Frucuts Gardeniae. With marked anti-inflammatory activities and the ability to 61 reduce oxidative stress,⁹⁻¹¹ it has been used to treat hepatitis and liver dysfunction.¹² 62 The conventional clinical chemistry and histopathology methods are not 63 region-specific and the sensitivity is relatively low. However, NMR-based 64 metabolomics revealed a global profile of endogenous metabolites, thus performing an 65 overall assessment of the global metabolic state of the entire organism. In this study, 66 an animal model of cholestasis was constructed by BDL and the treatment effect of 67 HLJDD in the BDL model was investigated by a NMR-based metabolomics approach 68

69 complemented with histological inspection and biochemical evaluation.

70 2 Materials & Methods

71 2.1 Chemicals and kits

Component herbs of HLJDD (Rhizoma Coptidis, Radix Scutellariae, Cortex 72 *Phellodendri* and *Fructus Gardeniae*) were obtained from Jiangsu Medicine Company 73 (Nanjing, China) and authenticated by Professor Mian Zhang, Department of 74 Medicinal Plants, China Pharmaceutical University, Nanjing. The voucher specimens, 75 deposited at the herbarium of the Department of Natural Medicinal Chemistry, China 76 Pharmaceutical University, were 2012066-RC, 2012067-RS, 2012068-CP and 77 2012069-FG for Rhizoma Coptidis, Radix Scutellariae, Cortex Phellodendri and 78 Fructus Gardeniae, respectively. 3-Trimethylsilylpropionic acid (TSP) was obtained 79 from Sigma-Aldrich (St. Louis, MO) and deuterium oxide (D₂O, 99.9%) was 80 purchased from QingDao TengLong WeiBo Technology Co. Ltd (QingDao, China). 81 Ultra-pure distilled water was prepared from a Milli-Q purification system. The serum 82 clinical enzymatic chemistry kits of aspartate aminotransferase (AST), alanine 83 aminotransferase (ALT), alkaline phosphatase (ALP), creatinine (CR), total protein 84 (TP), albumin (ALB) and globulin (GLB) were commercially available from Beckman 85 Coulter Inc (Harbor Boulevard, Fullerton, California, 92834 USA), while the serum 86 radioimmunoassay kits of hyaluronic acid (HA), type IV collagen (CIV), type III 87 precollagen (PCIII) and laminin (LN) were bought from Beijing North biology 88 technique institute (Beijing, China). 89

90 2.2 Preparation of HLJDD

Rhizoma Coptidis, Radix Scutellariae, Cortex Phellodendri and Fructus Gardeniae 91 were mixed in a ratio of 3:2:2:3, reaching a total weight of 500 g. Then they were 92 extracted with 70% ethanol (1:10, 1:10 and 1:5, w/v) under reflux for three times, 1h 93 each, and the extracted solution was filtered through 5 layer gauzes. The decoction 94 was concentrated to dryness to afford 142.5 g HLJDD (yield: 28.5%) using a rotary 95 vacuum evaporator, then stored in refrigerator at 4 °C. The dried extracts were 96 suspended in 0.5% (w/v) sodium carboxy-methylcellulose (CMC-Na) before 97 intragastric administration and the doses were calculated as raw material weights for 98 the animal experiments. 99

100 **2.3 Bile duct ligation (BDL) operation**

BDL was performed using a standard technique.¹³ Briefly, animals were anesthetized by injected intraperitoneally (ip) with chloral hydrate (350 mg/kg) and kept under anesthesia with additional ip injection throughout the experiment. After a midline incision under sterile conditions, a single ligature with silk suture was done with 4-0 nylon sutures, followed by careful suturing of the peritoneum and muscle layers as well as the skin wound. The sham-operated rats underwent the same surgical operation except for ligation of bile duct.

108 **2.4 Animals handling procedure**

Forty-two adult male Sprague-Dawley rats (220-240 g), of Specified-Pathogens Free (SPF) grade, were obtained from the Experimental Animal Center of Yangzhou University (Yangzhou, China). Rats were group-housed in polysulfone cages (5 rats to one cage) with bedding material, and were housed in a room with controlled humidity

113 $(50 \pm 10\%)$ and temperature $(25 \pm 3 \text{ °C})$ under a 12/12-h light/dark cycle. The animals 114 were given free access to standard diet and water and were allowed to acclimate for 7 115 days before treatment. The studies were in accordance with the standard guidelines for 116 the Care and Use of Laboratory Animal from the National Institute of Health (NIH) 117 and were approved by the Animal Ethics Committee of the China Pharmaceutical 118 University.

Rats were randomly divided into three groups (n=14): sham-operated (NC), bile duct ligation (BDL) and BDL with HLJDD treatment (BHD). BHD rats were intragastrically administered with HLJDD at doses of 2.7 g/kg body weight, and NC and BDL rats were administered with the same volume of 0.5% CMC-Na for each administration once a day for seven consecutive weeks.

124 **2.5 Collection of serum and urine**

On weeks 1, 3, 5 and 7 after the treatment, blood samples were taken from the ocular vein of rats after 12 h fasting. The serum samples were obtained by centrifugation at 13282 g for 10 min, and stored at -80 °C before next experiments. Centrifugation was performed on Beckman Coulter Microfuge® 22R refrigerated microcentrifuge using F241.5 rotor, with a radius of 8.25 cm and the maximum RCF at 21591 g.

Rats were housed in metabolic cages for a 24 hour interval and the urine samples were collected at weeks 1, 3, 5 and 7. The samples were then centrifuged at 13282 g for 10 min to afford the supernatants and stored at -80 °C before NMR spectroscopic analysis.

134 **2.6 Histopathology and serum biochemical analysis**

At the end of week 7, rats were fasted overnight and sacrificed after deep anesthetization with chloral hydrate (350 mg/kg, i.p.). The livers and kidneys were removed at the time of death, flushed with cold phosphate buffer solution to remove residual blood. The liver and kidney tissues obtained were immediately immersed in 10% neutral-buffered formaldehyde and embedded in paraffin to be stained with hematoxylin eosin (HE).

Serum samples harvested at different time-points were used for clinical chemistry
evaluation. To assess liver and renal function, the concentrations of AST, ALT, ALP,
TP, GLB, ALB and Cr were measured using commercially available kits from Nanjing
Jiancheng Biotech Inc. On week 7, HA, PCIII, LN and CIV were detected by
radioimmunoassay (RIA).

146 2.7 ¹H NMR spectroscopic measurement of serum and urine

The serum and urine samples were thawed at room temperature and 300 μ L of each was added with 300 μ L D₂O (0.2 mol L⁻¹ Na₂HPO₄ and 0.2 mol L⁻¹ NaH₂PO₄, pH 7.4, containing 0.05 % TSP). TSP acted as a chemical shift reference (δ 0.0) and D₂O provided a lock signal. The samples were vortexed and centrifuged at 13282 g for 10 min at 4 °C to remove insoluble material. The supernatants were then pipetted out into 5 mm NMR tubes for NMR recording.

¹⁵³ ¹H NMR spectra of the samples were recorded on a Bruker AV 500 MHz spectrometer at 300 K. For each serum sample, the transverse relaxation-edited Carr–Purcell– Meiboom–Gill (CPMG) spin-echo pulse sequence (RD-90°-(τ -180°- τ) n-ACQ) with a total spin-echo delay (2nt) of 40 ms was used to suppress broad signals from

macromolecules, therefore the signals of micromolecules were clearly observed. ¹H 157 NMR spectra were measured with 128 scans into 32 K data points over a spectral 158 width of 10000 Hz. Prior to Fourier transformation, an exponential window function 159 with a line broadening of 0.5 Hz was used to the free induction decays (FIDs). For 160 urine, a nuclear overhauser effect spectroscopy (NOESY) pulse sequence (relaxation 161 delay-90°-µs-90°-tm-90°-acquire-FID) was used to attenuate the residual water signal. 162 FIDs were collected into 32 k data points over a spectral width of 10000 Hz with an 163 acquisition time of 2.04 s. The FIDs were weighted by an exponential function with a 164 0.3 Hz line-broadening factor prior to Fourier transformation. 165

166 **2.8 Spectral pre-processing and data analysis**

The spectra for all samples were manually phased and baseline corrected, and 167 referenced to TSP at 0.0 ppm, using Bruker Topspin 3.0 software (Bruker GmbH, 168 Karlsruhe, Germany). The ¹H NMR spectra were automatically exported to ASCII 169 files using MestReNova (Version 8.0.1, Mestrelab Research SL), which were then 170 imported into "R" (http://cran.r-project.org/), and aligned with an in-house developed 171 R-script to further reduce phase and baseline distortions. The one-dimensional (1D) 172 spectra were converted to an appropriate format for statistical analysis by 173 automatically segmenting each spectrum into 0.005 ppm integrated spectral regions 174 (buckets) between 0.2 and 10 ppm. The region of the residual water and affected 175 signals (4.2–5.7 ppm) was removed. To account for different dilutions of samples, all 176 binned spectra were probability quotient normalized and then mean-centered before 177 further multivariate analysis. 178

The mean-centered and Pareto-scaled NMR data were analyzed by principal 179 component analysis (PCA) and orthogonal partial least-squares discriminant analysis 180 (OPLS-DA). PCA is an exploratory unsupervised method to maximize the separation 181 by providing model-free approaches for determining the latent or intrinsic information 182 in the dataset.¹⁴ However, no clustering was observed when variables were not 183 selected. OPLS-DA determines PLS components that are orthogonal to the grouping 184 and was used to concentrate group discrimination into the first component with 185 remaining unrelated variations contained in subsequent components.¹⁵ All OPLS-DA 186 models were validated by a repeated two-fold cross-validation method and 187 permutation test. The parameters of R^2 and Q^2 reflected the goodness of fitness and the 188 predictive ability of the models, respectively. The p value of the permutation test 189 denoted the number of times that the permutated data yielded a better result than the 190 one using the original labels. The fold change values of metabolites among different 191 groups were calculated. The Benjamini & Hochberg method¹⁶ was used to adjust the 192 related p-values for controlling the false discovery rate in multiple comparisons 193 applying scripts written in R language, which is available freely, open-source software 194 package. 195

196 **3 Results**

197 **3.1 Animal monitoring**

There was no significant difference in the weight of rats in NC, BDL and BHD group on week 7. The serum and urine collected from BDL rats were visually yellow, and after dissection rats undergoing BDL had yellow obstructive jaundice skin, thicken liver especially in the ventral lobe and enlarged bile duct.

3.2 Liver and kidney histopathology

H&E staining of representative liver sections were observed and shown at 200 \times 203 magnifications. The most obvious character of liver impairment after BDL was the 204 severe proliferation of bile duct around the pre-existing interlobular ducts and 205 surrounded by a connective-tissue sheath. Additional diffuse collagen fiber was also 206 noted in interstitial and the portal zones with formation of pseudo lobe. Inflammatory 207 cell infiltration localized specifically to the periportal zones of the livers of BDL rats, 208 was observed in the vicinity of proliferating bile ducts (Figure 1A), as compared with 209 normal livers in the NC group (Figure 1B). Consecutive administration of HLJDD for 210 7 weeks resulted in remarkable amelioration in these pathological changes (Figure 1C). 211 Histopathological inspection revealed a slight increase of the volume of glomerular in 212 BDL group (Figure 1D) as compared with the NC group (Figure 1E); BHD group 213 showed almost no difference from the normal group (Figure 1F). 214



Figure 1. Histopathological study of liver and kidney of NC, BDL and BHD rats at 200 \times 216 magnifications. (A) Liver of BDL rats 49 days after operation: exhibiting severe hyperplasia of 217 bile-ducts, inflammatory cell infiltration, and additional diffuse collagen fiber. (B) Livers in sham 218 operation rats: showing no pathological changes. (C) Livers in BHD rats 49 days after intragastric 219 administration: showing no obvious change without any sign of cell degeneration or necrosis. (D) 220 Kidneys of BDL rats after 7 weeks, with slight increase in the volume of glomerular. (E) Kidneys in 221 sham operation group rats, with no pathological changes. (F) Kidneys in BHD group rats after 7 222 weeks, with no pathological changes. 223

3.3 Serum biochemical parameters

Serum levels AST, ALT, ALP, CR, TP, ALB and GLB were determined on week 1, 3, 5 225 and 7, and the results of the clinical chemistry are presented in Fig. 2. Levels of AST, 226 ALT and ALP in BDL group increased significantly on week 1 as compared with the 227 NC group, but the increase attenuated from week 1 onwards, reaching to a minimum 228 on week 7. The elevated AST, ALT and ALP levels indicated that cholestasis and liver 229 cell necrosis were already present one week after BDL¹⁷. The ALB concentration of 230 BDL rats significant decrease on week 1, 3 and 5, but without significant disturbance 231 on week 7 as compared with NC group. The TP and GLB in BDL group showed some 232 fluctuations throughout the experiments: markedly decreasing on week 1, but 233 markedly increasing on week 3 and 5 and finally kept at normal levels. ALB and GLB, 234 synthesized by liver cells, are indicators of hepatocellular function.^{18, 19} The increase 235 of AST, ALT, ALP and decrease of ALB in BDL rats on week 1 indicated a severe 236 cholestatic liver injury in the model.²⁰ However, variations of these parameters were 237

238 attenuated and reversed towards the normal status. On week 7, showing a partial recovery of liver function in BDL rats. The significantly decreased levels of TP, ALB 239 and GLB on week 1 indicated the impaired hepatocellular function in synthesis, and 240 also featuring an acute liver injury. In contrast, on week 3 and 5, both the decrease of 241 ALB and increase of GLB were observed, characterizing chronic liver damage. 242 Similarly, both of them reversed to normal on week 7, indicating the alleviation of 243 liver injury in the long run. The levels of those index indicated liver dysfunction, but 244 HLJDD could restore most of the fluctuations of AST, ALT, ALP, TP, ALB and GLB 245 in the serum. CR in BDL group did not show any significant difference at all time 246 periods. 247



Figure 2. Boxplots of serum levels of creatinine (Cr), total protein (TP), albumin (ALB), alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), globulin (GLB) and the levels of ratio of albumin and globulin (ALB/GLB). The boxes cover 25% quartile

and 75% quartile of the data. The line in the box represents the median value. The extended whiskers show the extent of the rest of the data. (A) AST. (B) ALT. (C) AST_ALT. (D) ALP. (E) CR. (F) TP. (G) ALB. (H) GLB. (I) ALB/GLB. Outliers are shown as open circle. Values were expressed as mean \pm SD (n = 10-14). * *p* < 0.05, ** *p* < 0.01 and *** *p* < 0.001 vs. NC rats; * *p* < 0.05, ** *p* < 0.01 and *** *p* < 0.001 vs. BDL rats.

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To asses liver fibrosis, we measured the levels of hyaluronic acid (HA), laminin (LN), type III precollagen (PCIII) and type IV collagen (CIV) in serum (Fig. 3). HA, LN, PCIII and CIV were markers for liver fibrosis.²¹ The results revealed that levels of HA and PCIII in BDL rats significantly increased on week 7 (p < 0.05), compared with NC rats. The levels of LN and CIV in the serum of BDL group augmented slightly. The results suggested the initial formation of liver fibrosis on week 7. HLJDD could restore increased levels of HA, LN and PCIII in serum.



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Figure 3. Boxplots of serum levels of hyaluronic acid (HA), type IV collagen (CIV), type III precollagen (PCIII) and laminin (LN). The boxes and whiskers represent the same as in Figure 2. Boxplots of HA (A), LN (B), PCIII (C), and CIV (D), respectively. Outliers are shown as open circle. Values were expressed as mean \pm SD. * p < 0.05, ** p < 0.01 and *** p < 0.001 vs. NC rats; [#]p <0.05, ^{##}p < 0.01 and ^{###}p < 0.001 vs. BDL rats.

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272 **3.4 Metabolites identification in serum and urine**

Representative 500 MHz ¹H NMR spectra of serum and urine samples from NC, BDL 273 and BHD rats were shown in Figures 4 A and B with the assignment of metabolites. 274 Aided by Chenomx NMR suit (Version 8.1, Chenomx, Inc.) and a statistical total 275 correlation spectroscopy (STOSCY) technique,²² their assignments were made by 276 referencing reported data^{16, 23} and querying publicly accessible metabolomic databases, 277 such as HMDB (http://www.hmdb.ca), KEGG (http://www.kegg.jp), METLIN 278 (http://metlin.scripps.edu). The detailed information of the metabolites was listed in 279 Tables S1 and S2. 280



Figure 4. (A) Typical 500 MHz CPMG ¹H NMR spectra of the serum samples from NC, BDL and BHD rats on week 1. 1. very low density lipoprotein (VLDL) / low density lipoprotein (LDL); 2. valine; 3. β-Hydroxybutyrate (HB); 4. alanine; 5. lysine; 6. acetate; 7. adipate; 8. N-Acetyl glycoproteins (NAGP); 9. O-Acetyl glycoproteins (OAGP); 10. glutamate; 11.citrate; 12. cysteine; 13. creatine; 14. tyrosine; 15. trimethylamine-N-oxide (TMAO); 16. glycerophosphorylcholine (GPC); 17. taurine; 18. β-glucose; 19. α-glucose; 20. glycine; 21. glutamine; 22. lactate; 23.

288	phenylalanine; 24. histidine; 25. formate. (B) Typical 500 MHz NOESY ¹ H NMR spectra of the
289	urine samples from NC rats, BDL rats and BHD rats on week 1: 1. very low density lipoprotein
290	(VLDL) / low density lipoprotein (LDL); 2. isoleucine/leucine; 3. valine; 4. β-Hydroxybutyrate; 5.
291	α-hydroxyisobutyrate; 6. lactate; 7. alanine; 8. ornithine; 9. acetate; 10. proline; 11. N-Acetyl
292	glycoproteins (NAGP); 12. O-Acetyl glycoproteins (OAGP); 13. methionine; 14. adipate; 15.
293	acetoacetate; 16. glutamate; 17. oxalacetate; 18. succinate; 19. citrate; 20. dimethyl amine (DMA);
294	21. methylguanidine; 22. trimethyl amine (TMA); 23. dimethyl glycine (DMG); 24. pyruvate; 25.
295	2-oxoglutarate; 26. creatinine; 27. choline; 28. phosphocholine; 29. trimethylamine-N-oxide
296	(TMAO); 30. taurine; 31. glycine; 32. creatine/phosphocreatine; 33. fumarate; 34.
297	3-hydroxymandelate; 35. 4-aminohippurate; 36. gallate; 37. phenylalanine; 38. tryptophan; 39.
298	benzoate; 40. hippurate; 41. formate; 42. nicotinate; 43. N-methylnicotinamide. 44. NAD ⁺ .

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300 3.5. OPLS-DA score trajectory plot at all time points

The metabolic profiles were first subjected to PCA analysis. The three groups showed 301 partial separation from each other in PCA score plots (data not shown) due to the 302 unsupervised nature of PCA. The variations of ¹H NMR signals among groups were 303 not only arisen from interested grouping but also from other factors that were group 304 unrelated. To concentrate group discrimination into the first component and filter those 305 variations unrelated to class discrimination, supervised OPLS-DA was further 306 performed.¹⁶ The dynamic metabolic events in the rats were visualized by the 307 OPLS-DA score trajectory plots (Fig.2), where each spot represented the mean 308 position of each group at one time. The direction denoted by arrows represented the 309

trend of the changing metabolite pattern. The shift of the metabolic patterns in both serum and urine were similar in that BDL group showed the furthest deviation to NC group on week 1, and then moved towards the normal (Fig. 5 A and B). The score trajectory showed a distinct biphasic course of the BDL model. Data on week 1 (acute stage) and week 3 to week 7 (recovery stage) were analyzed independently.



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Fig. 5 OPLS-DA score trajectory plots of rats serum (A) and urine (B) of different groups on week 1,
3, 5 and 7 after BDL operation. Symbols of ● (black filled circles), ■ (red filled squares) represented
NC and BDL group respectively.

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320 **3.6** ¹H NMR metabolomics profiles on week 1

321 3.6.1 Metabolic changes in BDL rats

On week 1, the BDL and NC group showed a complete separation in score plots of

serum and urine (Fig. 6 A and C), suggesting a severe metabolic disturbance in BDL

groups by a supervised OPLS-DA with a well goodness of fit ($R^2 = 0.93$, $Q^2 = 0.79$; R^2

= 0.98, $Q^2 = 0.93$ for the serum and urine, respectively) displayed in Fig. 6 G and H. 325 The corresponding S-plots and loading plots revealed the differential metabolites in 326 serum and urine. In the S-plots, where points in different color and shape represented 327 variables (metabolites), the more further away from the center of a variable, the more 328 contribution of the variable to the class separation of the groups. The loading plots 329 were color-coded with the absolute value of correlation coefficients where red (high 330 coefficients) indicates more marked contribution to the separation than blue (low 331 coefficients) one. The S-plots (Fig.6 B and D) and loading plots (Fig.6 E and F) 332 revealed elevated levels of acetate, creatine, trimethylamine-N-oxide (TMAO), 333 glycerophosphorylcholine (GPC), histidine, phenylalanine in serum; elevated levels of 334 3-methylglutarate, α -hydroxyisovalerate, isoleucine, leucine, valine, isobutyrate, 335 α -hydroxyisobutyrate, alanine, acetate, trimethylamine (TMA), taurine, 336 N-methylnicotinamide, 3-hydroxymandelate, NAD⁺, pyruvate in urine, and lower 337 levels of High density lipoprotein (HDL), alanine, β -glucose, α -glucose, glycine in 338 serum; lower levels of succinate, citrate. dimethylglycine (DMG), 339 creatine/phosphocreatine, creatinine, 2-oxoglutarate, 4-aminohippurate, nicotinate, 340 gallate, hippurate, benzoate, acetylsicylate in urine of BDL rats. These important 341 differential metabolites were further tested for their between-group difference using 342 univariate analysis, and found to be mostly significant as visualized in the heat map 343 (Fig. 8) and fold change plots (Fig. S1). 344

345 **3.6.2 Effect of HLJDD on the metabolic profiles of BDL rats**

To explore the influence of HLJDD on BDL rats, ¹H NMR data of serum and urine of

NC, BDL and BHD group were analyzed together. They were clearly separated in the 347 OPLS-DA score plots (Fig. 7 A and C), and BHD groups were in the middle and close 348 to the NC groups, declaring HLJDD can reduce the changes of metabolites caused by 349 the administration of BDL. To find out metabolites that were directly associated with 350 the treatment effect of HLJDD on BDL rats, the metabolic profiles of BDL and BHD 351 groups were analyzed by OPLS-DA (Fig. 7 B and D). The BHD rats were clearly 352 separated from BDL rats with a well goodness of fit (Fig. 7 G and H). 353 The loading plots (Fig. 7 E and F) revealed great increase of β -glucose, α -glucose, 354 glycine, caprate, glutamine in serum; higher levels of isobutyrate, N-Acetyl 355

glycoproteins (NAGP), O-Acetyl glycoproteins (OAGP), methylguanidine, 356 creatine/phosphocreatine, trimethylamine-N-oxide (TMAO). creatinine, 357 2-oxoglutarate, threonine, glycine, creatinine, lactate, acetoacetate, β -hydroxybutyrate 358 (3-HB), fumarate, phenylalanine, 4-aminohippurate, 2-oxoglutarate, gallate, NAD⁺, 359 benzoate, threonine in urine, and lower levels of LDL, VLDL, adipate, 360 trimethylamine-N-oxide (TMAO) in serum; lower levels of α -hydroxyisovalerate, 361 isoleucine, leucine, valine, alanine, acetate, proline, adipate, dimethyl glycine (DMG), 362 taurine. tryptophan, hippurate. nicotinate. formate. N-methylnicotinamide, 363 3-hydroxymandelate, NAD⁺ in urine of BHD groups. The important differential 364 metabolites selected based on loading plots of OPLS-DA, and found to be mostly 365 significant as visualized in the heat map (Fig. 8) and fold change plots (Fig. S1). 366



Fig. 6 On week 1, cross-validated OPLS-DA scores plots (A for serum and C for urine), the corresponding S-plots (B for serum, D for urine) and loadings plots (E for serum, F for urine) derived from ¹H NMR spectra for NC, BDL and BHD rats. OPLS-DA scatter plot from serum (G) and urine (H) of the statistical validations obtained by 200 times permutation tests, with R^2 and Q^2 values in the vertical axis, the correlation coefficients (between the permuted and true class) in the horizontal axis, and OLS line representing the regression of R^2 and Q^2 on the correlation coefficients.



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Fig. 7 On week 1, OPLS-DA analysis of ¹H NMR data in serum and urine for NC, BDL and BHD group. Scores plots on NC, BDL and BHD rats (A and C), and on BDL and BHD rats (B and D), loadings plots (E and F) for OPLS-DA. OPLS-DA scatter plot from serum (G) and urine (H) of the statistical validations obtained by 200 times permutation tests, with R^2 and Q^2 values in the vertical axis, the correlation coefficients (between the permuted and true class) in the horizontal axis, and OLS line representing the regression of R^2 and Q^2 on the correlation coefficients.



Fig. 8 Heatmap visualization of the z-scored levels of metabolites in serum (A) and urine (B) with stars denoting the differential significance. Row represent metabolites and column represent groups.

"BDL" and "BHD" mean "BDL group compare with NC group" and "BHD group compare with BDL group", the number "1", "3", "5" and "7" mean week one, three, five and seven. Color key indicates metabolite quantities value, white: no significant change, deep blue: highest, deep red: lowest, P< 0.05 represented statistically significant threshold. * P< 0.05, ** P< 0.01 and *** P< 0.001.

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391 **3.7** ¹H NMR metabolomics profiles on week 3, 5, 7

On week 3, 5 and 7, OPLS-DA method was carried out in the BDL, NC, BHD groups. 392 From the score plots of BDL and NC groups (Fig. S2 C and D), revealed a good 393 separation. Compared with NC group, these findings according to the corresponding 394 loading plots (Fig. S3 A, B and C) were observed in BDL group: elevated levels of 395 very low density lipoprotein (VLDL)/low density lipoprotein (LDL), caprate, 396 3-methyladipate/succinyl acetone in serum; elevated levels of citrate, 2-OG in urine, 397 and reduced levels of alanine, lysine, adipate, acetate, glycine in serum; reduced levels 398 of 3-methylglutarate, isoleucine, leucine, valine, α -hydroxyisobutyrate, glutamate in 399 urine of BDL groups. What's more, the OPLS-DA score plots of the serum and urine 400 (Fig. S2 A and B) showed a partial separation of BDL group from NC and BHD group, 401 suggesting damage caused by BDL is gradually restored on week 3, 5 and 7, and NC 402 and BHD group were much overlap in the score plots indicated HLJDD can reduce the 403 changes of metabolites caused by the administration of BDL. Most of the disturbances 404 of metabolites in BDL rats were reversed after treatment of HLJDD (Fig. S2 E,F and 405 Fig. S3 D, E, F). 406

407

408 **4 Discussion**

In this study, ¹H NMR-based metabolomics approach combined with clinical 409 chemistry and histopathology inspection was used to investigate cholestatic liver 410 damage caused by bile duct ligation (BDL) and the treatment effects of HLJDD on 411 BDL rats. Histopathology inspection indicated initial formation of fibrosis in the liver 412 which was testified by the increase of ALT, AST, ALP and decrease of ALB in serum 413 of BDL rats, and the increase of hyaluronic acid (HA), laminin (LN), type III 414 precollagen (PCIII) and type IV collagen (CIV). Slight edema of glomerular was 415 found in the kidney of BDL rats. OPLS-DA analyses of the serum and urinary NMR 416 data of the three groups on week 1, 3, 5 and 7 were performed. The metabolic profiles 417 in BDL rats were severely disturbed, the furthest away from the control on week 1 418 (acute phase) and then gradually recovered on week 3, 5 and 7 (chronic phase). 419

420 One week after BDL operation, rats showed a series of metabolic perturbations, 421 including energy and amino acid metabolism, oxidative stress and intestinal flora 422 disruption.

423 **4.1 Acute phase**

424 **4.1.1 Energy metabolism**

Significantly decreased levels of urinary citrate, 2-OG, succinate and serum β -glucose, a-glucose together with slightly increased pyruvate level in urine were observed in BDL group as compared with the NC group. Pyruvate is a key intermediate that takes part in both glycolysis and the tricarboxylic acid (TCA) cycle. Generated by

decomposition of glucose, pyruvate can be converted into acetyl-CoA by 429 decarboxylation and enter the TCA cycle under aerobic conditions.²⁴ The increased 430 level of pyruvate in urine suggested a hampered conversion of pyruvate to acetyl-CoA, 431 which together with notably decreased other intermediates of TCA cycle, citrate, 2-OG 432 and succinate, suggested an inhibition of the TCA cycle.^{25, 26} TCA cycle is the most 433 efficient and major source of energy supply. Its inhibition caused energy deficiency, so 434 other means, such as glycolysis, come to rescue. By glycolysis, glucose is converted to 435 lactate by lactate dehydrogenase (LDH) or to alanine by alanine aminotransferase 436 (ALT), resulting in increased lactate and alanine levels.²⁷ The marked decrease of 437 serum glucose, and increase of lactate and alanine in serum of BDL rats demonstrated 438 an enhanced glycolysis in BDL group. 439

However, glycolysis is inefficient in energy production. Therefore, ketone bodies 440 metabolism, another means of energy production, have to be enhanced to ameliorate 441 the shortage of energy. Ketone bodies, including acetoacetate and 3-HB, are 442 well-known metabolites of fatty acids in liver mitochondria. Ketones could be 443 transported from serum to muscles, tissues and organs, where ketone bodies could be 444 oxidized to produce energy. The obvious increase of ketone bodies in urine as well as 445 slight decrease of LDL or VLDL in serum of BDL group, suggested an enhanced fatty 446 acid oxidation to produce ketone bodies. As a result, the concentration of acetate, the 447 end product of fatty acid oxidation, was significant increase in the BDL rats. Serum 448 creatine and urinary creatinine were increased in BDL rats. Creatine-phosphate can 449 transfer high-energy phosphate to ADP and produce ATP for energy demand.²⁸ 450

Therefore, the significant increase of creatine and its degradation product creatinine suggested a facilitated utilization of creatine-phosphate to replenish energy demand. The increase of creatine and creatinine was also deemed as a sign of hepatic injury,²⁴ in consistent with pathological result in this study.

BDL rats after HLJDD treatment exhibited markedly enhanced levels of 2-OG, and slightly decreased amount of pyruvate, alanine, 3-HB and acetate, which showcased the ability of HLJDD to ameliorate the disturbed energy supply induced by BDL, possibly by restoration of TCA cycle.

459 **4.1.2 Oxidative stress**

Bile acids (BAs) were secreted by hepatocytes, transported into the extracellular 460 matrix in liver and then excreted to duodenum for the absorption and solubilization of 461 dietary lipids. BDL prevented bile flow from liver to duodenum, leading to an 462 accumulation of BAs in liver that generate reactive oxygen species (ROS).²⁹ The 463 imbalance between the generation of ROS and antioxidant defenses induce oxidative 464 stress.³⁰ We observed significantly decreased levels of glutamine and glycine and 465 slightly decreased levels of glutamate and cysteine in serum, the precursors of GSH, in 466 BDL rats. GSH as a major natural antioxidant, can react with free radicals directly,³¹ 467 thus resisting the damage caused by ROS. The lowered levels of these GSH precursors 468 demonstrated an accelerated GSH synthesis as a consequence of excessive depletion 469 of GSH to counteract ROS.³² 470

471 Urinary choline and phosphocholine were decreased obviously in BDL group472 compared with the NC group. Phospholipid, consisting of choline and phosphocholine,

is the major component of cell membrane and essential for the maintenance of its 473 integrity.³³ ROS could attack membrane phospholipids, leading to the damage on the 474 construction and function of membranes, and ultimate the rupture of cell and 475 organelles, such as mitochondria.³⁴ As an evidence, serum levels of NAGP and OAGP 476 were decreased slightly in BDL group since that they were synthesized in membranes 477 of endoplasmic reticulum and golgi apparatus. The lowered levels of choline and 478 phosphocholine indicated an accelerated use of them to renovate the membranes 479 damaged by ROS, thus representing a self-repair mechanism. 480

HLJDD significantly decreased the elevated urinary level of taurine, and markedly
increased the lower serum levels of glutamine, glycine and taurine and urinary choline
and phosphocholine in BDL rats, showcasing its protection on BDL induced oxidative
injury.

485 4.1.3 Amino acid metabolism

Taurine and glycine were decreased in serum but increased in urine of BDL rats. BAs 486 could be conjugated to either taurine or glycine in order to reduce the toxicity caused 487 by accumulated BAs. The conjugated BAs were hydrolyzed into free BAs, liberating 488 taurine or glycine under the activation of intestinal bacteria. Normally, most BAs were 489 reabsorbed and, taurine and glycine were excreted into urine.³⁵ Therefore, the opposite 490 change of taurine and glycine in serum and urine of BDL rats demonstrated 491 accelerated their conjugation with BAs to attenuate the toxicity of accumulated BAs. 492 Leucine, isoleucine and valine (branched-chain amino acids, BCAAs) were significant 493

⁴⁹⁴ increased in urine of BDL rats. ROS induced the decomposition of proteins, resulting

⁴⁹⁵ in the damage of cell membrane.³⁶ BCAAs are important precursors for protein ⁴⁹⁶ synthesis,³⁷ and thus essential for the repairment of the damaged cell membranes. The ⁴⁹⁷ increased excretion of BCAAs in urine of BDL rats showed either an inhibited protein ⁴⁹⁸ synthesis or an enhanced protein degradation.³⁸ In addition, these BCAAs can be ⁴⁹⁹ reabsorbed by glomerular in normal status and their increase may also suggested the ⁵⁰⁰ dysfunction of glomeruli reabsorption,^{39, 40} which also supported by histopathological ⁵⁰¹ examination.

Phenylalanine was markedly increased and tyrosine was slightly decreased in serum of BDL rats. phenylalanine is an essential amino acid that has to be obtained from food directly. Tyrosine is a semi-essential amino acid as it can only be synthesized by the hydroxylation of phenylalanine under the catalysis of phenylalanine hydroxylase (PAH). The increased conversion of phenylalanine to tyrosine ratio in BDL group has been observed in patients incurred with hepatitis C virus suffering hepatic damage,⁴¹ and thus may also indicated liver injury induced by BDL operation.

The increased urinary BCAAs, and enhanced conversion of Phe to tyrosine ratio due to BDL operation exhibited a downward trend towards a normal status by the intervention of HLJDD, showing its ability to ameliorate amino acid metabolism and protect rats from BDL induced liver injury.²⁶

513 **4.1.4 Intestinal flora metabolism**

The levels of hippurate and benzoate were significantly decreased in urine of BDL rats. Benzoate was synthesized from plant phenolics and aromatic amino acids, by intestinal microflora.⁴² Benzoate could be absorbed by intestinal tract, eventually

entering into the liver through the portal vein by systemic circulation. BDL prevented 517 the entrance of BAs to intestinal tract, leading to deficiency of intestinal bile salts, and 518 inevitably alteration of gut bacteria,⁴³ as supported by the obvious decrease of 519 benzoate in urine of BDL rats. As a sequence, the synthesis of hippurate was decreased 520 in BDL group since that hippurate was synthesized by conjugation of glycine with 521 benzoate in the mitochondrial matrix of liver^{44, 45} A series of studies have also 522 concluded that the decrease of urinary levels of benzoate and hippurate could be 523 ascribed to the disruption of intestinal flora.⁴⁶⁻⁴⁸ 524

The disturbance of gut microbes could also be evidenced by the observed significant increase of trimethylamine (TMA) and partial decrease of trimethylamine (TMAO), since that TMAO is the oxidation product of TMA through the action of gut microbes.¹⁷

HLJDD significantly increased the levels of urinary TMAO and hippurate,
demonstrating a great amelioration of gut microbiota metabolism by HLJDD.

531 **4.2 Chronic cholestasis phase**

The severe disturbance in energy metabolism, amino acid metabolism, oxidative stress and gut bacteria in acute phase was greatly attenuated in the long run, characteristic of BDL model. The BDL rats employed a self-repairing process with compensation to address the hampered bile flow by construction of the bypass of bile ducts, which was fully established in BDL rats on week 7.⁴⁹

Interestedly, with no good reasons, levels of urinary citrate, 2-OG and adipate in BDL group were significantly higher than those in NC group, suggesting the complex of the

body in response to pathological changes. Surprisingly, these metabolites could also be
reversed towards normal levels by HLJDD, in opposite direction to its performance in
acute phase, exhibiting a bilateral adjustment of HLJDD.

542 **5 Conclusion**

¹H NMR-based metabolomics approach was applied to explore global metabolic 543 features in serum and urine of BDL-induced cholestasis in rats and the treatment 544 effects of HLJDD. The metabolomic pattern showed a distinct biphasic feature of BDL 545 model: acute phase (week 1) and chronic phase (week 3-7). BDL brought severe 546 disturbance in energy and amino acid metabolism, alteration of intestinal flora and 547 oxidative stress in acute phase. The metabolomic results combined with clinical 548 chemistry indicated conspicuous liver dysfunction and the damage of renal glomerular 549 function at acute phase, which were greatly attenuated at chronic phase due to the 550 self-protection mechanism of the body. HLJDD showed bilateral adjustment of some 551 metabolism disturbance. These results demonstrated sensitivity and superiority of 552 metabonomics in disease subtypes and diagnosis, and in the understanding of complex 553 mechanism of a Chinese herbal medicine formula. This integrated metabolomics 554 approach might help to develop a systematic view of BDL-induced injury process and 555 assess its therapy. 556

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564	References
564	References

565	1.	M. J. Pollheimer, P. Fickert and B. Stieger, Molecular aspects of medicine, 2014, 37, 35-56.
566	2.	M. Galicia - Moreno, L. Favari and P. Muriel, Fundamental & clinical pharmacology, 2013, 27, 308-318.
567	3.	S. Ahmadi, Z. Karami, A. Mohammadian, F. Khosrobakhsh and J. Rostamzadeh, Neuroscience, 2015, 284, 78-86.
568	4.	V. Shivanna, Y. Kim and K. O. Chang, <i>Virology</i> , 2014, 456-457 , 268-278.
569	5.	D. Sokolovic, J. Nikolic, G. Kocic, T. Jevtovic-Stoimenov, A. Veljkovic, M. Stojanovic, Z. Stanojkovic, D. M.
570		Sokolovic and M. Jelic, Drug and chemical toxicology, 2013, 36, 141-148.
571	6.	T. P. M. d. Sousa, R. E. Castro, S. N. Pinto, A. Coutinho, S. D. Lucas, R. Moreira, C. M. P. Rodrigues, M. Prieto and
572		F. Fernandes, Biophysical Journal, 2015, 108, 242a.
573	7.	B. L. Copple, H. Jaeschke and C. D. Klaassen, Seminars in liver disease, 2010.
574	8.	S. Heinrich, P. Georgiev, A. Weber, A. Vergopoulos, R. Graf and PA. Clavien, Surgery, 2011, 149, 445-451.
575	9.	C. B. Li, X. X. Li, Y. G. Chen, H. Q. Gao, P. L. Bu, Y. Zhang and X. P. Ji, <i>PloS one</i> , 2013, 8 , e67530.
576	10.	J. Lu, JS. Wang and LY. Kong, Journal of ethnopharmacology, 2011, 134 , 911-918.
577	11.	Xb. Wang, Y. Feng, N. Wang, F. Cheung and Cw. Wong, Recent patents on food, nutrition & agriculture, 2012,
578		4 , 91-106.
579	12.	X. Ye, Y. Feng, Y. Tong, KM. Ng, S. Tsao, G. K. Lau, C. Sze, Y. Zhang, J. Tang and J. Shen, Journal of
580		ethnopharmacology, 2009, 124 , 130-136.
581	13.	H. Uchinami, E. Seki, D. A. Brenner and J. D'Armiento, Hepatology, 2006, 44, 420-429.
582	14.	S. M. Holand, <u>http://strata</u> . uga. edu/software/pdf/pcaTutorial. pdf. Last accessed, 2008, 12 , 2011.
583	15.	J. Boccard and D. N. Rutledge, Analytica chimica acta, 2013, 769, 30-39.
584	16.	J. C. Lindon, R. D. Farrant, P. N. Sanderson, P. M. Doyle, S. L. Gough, M. Spraul, M. Hofmann and J. K. Nicholson,
585		Magnetic Resonance in Chemistry, 1995, 33 , 857-863.
586	17.	M. I. Shariff, A. I. Gomaa, I. J. Cox, M. Patel, H. R. Williams, M. M. Crossey, A. V. Thillainayagam, H. C. Thomas, I.
587		Waked and S. A. Khan, Journal of proteome research, 2011, 10, 1828-1836.
588	18.	M. Bernardi, C. S. Ricci and G. Zaccherini, Journal of clinical and experimental hepatology, 2014, 4, 302-311.
589	19.	R. Garcia-Martinez, F. Andreola, G. Mehta, K. Poulton, M. Oria, M. Jover, J. Soeda, J. Macnaughtan, F. De Chiara,
590		A. Habtesion, R. P. Mookerjee, N. Davies and R. Jalan, Journal of hepatology, 2015, 62, 799-806.
591	20.	S. Sathesh Kumar, B. Ravi Kumar and G. Krishna Mohan, Journal of ethnopharmacology, 2009, 123, 347-350.
592	21.	FR. Yang, World Journal of Gastroenterology, 2010, 16, 1458.
593	22.	DD. Wei, JS. Wang, PR. Wang, MH. Li, MH. Yang and LY. Kong, Journal of pharmaceutical and biomedical
594		analysis, 2014, 98 , 334-338.
595	23.	A. W. Nicholls, J. C. Lindon, S. Caddick, R. D. Farrant, I. D. Wilson and J. K. Nicholson, Xenobiotica, 1997, 27,
596		1175-1186.
597	24.	Cy. Jiang, Km. Yang, L. Yang, Zx. Miao, Yh. Wang and Hb. Zhu, <i>PloS one</i> , 2013, 8 , e66786.
598	25.	S. Satapati, N. E. Sunny, B. Kucejova, X. Fu, T. T. He, A. Mendez-Lucas, J. M. Shelton, J. C. Perales, J. D. Browning
599		and S. C. Burgess, Journal of Lipid Research, 2012, 53, 1080-1092.
600	26.	N. Wang, Y. Feng, H. Y. Tan, F. Cheung, M. Hong, L. Lao and T. Nagamatsu, Journal of ethnopharmacology, 2015,

601		164 , 309-318.
602	27.	H. Li, L. Wang, X. Yan, Q. Liu, C. Yu, H. Wei, Y. Li, X. Zhang, F. He and Y. Jiang, Journal of proteome research, 2011,
603		10 , 2797-2806.
604	28.	M. Wyss and R. Kaddurah-Daouk, <i>Physiological reviews</i> , 2000, 80 , 1107-1213.
605	29.	P. Ljubuncic, Z. Tanne and A. Bomzon, <i>Gut</i> , 2000, 47 , 710-716.
606	30.	X. Cheng, DX. Gao, JJ. Song, FZ. Ren and XY. Mao, <i>Rsc Advances</i> , 2015, 5 , 4511-4523.
607	31.	I. S. Ovey and M. Naziroglu, Neuroscience, 2015, 284, 225-233.
608	32.	YT. Huang, YC. Hsu, CJ. Chen, CT. Liu and YH. Wei, Journal of Biomedical Science, 2003, 10, 170-178.
609	33.	K. Chingin, J. Liang and H. Chen, <i>Rsc Advances</i> , 2014, 4 , 5768-5781.
610	34.	N. Nakamura, <i>Cells</i> , 2013, 2 , 732-750.
611	35.	L. Yang, A. Xiong, Y. He, Z. Wang, C. Wang, Z. Wang, W. Li, L. Yang and Z. Hu, Chemical research in toxicology,
612		2008, 21 , 2280-2288.
613	36.	H. Yang, T. W. Li, Y. Zhou, H. Peng, T. Liu, E. Zandi, M. L. Martinez-Chantar, J. M. Mato and S. C. Lu, Antioxidants
614		& redox signaling, 2015, 22 , 259-274.
615	37.	A. D. Lake, P. Novak, P. Shipkova, N. Aranibar, D. G. Robertson, M. D. Reily, L. D. Lehman-McKeeman, R. R.
616		Vaillancourt and N. J. Cherrington, Amino Acids, 2015, 47, 603-615.
617	38.	M. Forestier, M. Solioz, F. Isbeki, C. Talos, J. Reichen and S. Krahenbuhl, Hepatology, 1997, 26, 386-391.
618	39.	R. Williams and E. Lock, <i>Toxicology</i> , 2005, 207 , 35-48.
619	40.	B. U. Bradford, T. M. O'Connell, J. Han, O. Kosyk, S. Shymonyak, P. K. Ross, J. Winnike, H. Kono and I. Rusyn,
620		Toxicology and applied pharmacology, 2008, 232 , 236-243.
621	41.	H. Zoller, A. Schloegl, S. Schroecksnadel, W. Vogel and D. Fuchs, Journal of Interferon and Cytokine Research,
622		2012, 32 , 216-220.
623	42.	H. J. Lees, J. R. Swann, I. D. Wilson, J. K. Nicholson and E. Holmes, Journal of proteome research, 2013, 12,
624		1527-1546.
625	43.	R. S. Lord and J. A. Bralley, Alternative Medicine Review, 2008, 13, 292-306.
626	44.	C. Lu, Y. Wang, Z. Sheng, G. Liu, Z. Fu, J. Zhao, J. Zhao, X. Yan, B. Zhu and S. Peng, Toxicology and applied
627		pharmacology, 2010, 248 , 178-184.
628	45.	JY. Cho, T. Matsubara, D. W. Kang, SH. Ahn, K. W. Krausz, J. R. Idle, H. Luecke and F. J. Gonzalez, Journal of
629		Lipid Research, 2010, 51 , 1063-1074.
630	46.	J. R. Swann, K. M. Tuohy, P. Lindfors, D. T. Brown, G. R. Gibson, I. D. Wilson, J. Sidaway, J. K. Nicholson and E.
631		Holmes, Journal of Proteome Research, 2011, 10 , 3590-3603.
632	47.	Y. L. Wang, H. R. Tang, J. K. Nicholson, P. J. Hylands, J. Sampson and E. Holmes, Journal of Agricultural and Food
633		Chemistry, 2005, 53 , 191-196.
634	48.	J. Y. Cho, T. Matsubara, D. W. Kang, S. H. Ahn, K. W. Krausz, J. R. Idle, H. Luecke and F. J. Gonzalez, Journal of lipid
635		research, 2010, 51 , 1063-1074.
636	49.	E. Biecker, A. De Gottardi, M. Neef, M. Unternährer, V. Schneider, M. Ledermann, H. Sägesser, S. Shaw and J.
637		Reichen, Journal of Pharmacology and Experimental Therapeutics, 2005, 313, 952-961.
638		
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