

RSC Advances



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. This *Accepted Manuscript* will be replaced by the edited, formatted and paginated article as soon as this is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

1 **A urinary metabolomics study of the metabolic dysfunction and the**
2 **regulation effect of citalopram in the rats exposed to chronic**
3 **unpredictable mild stress**

4
5 Xinyu Yu^{a#}, Jia Luo^{b#}, Lijun Chen^a, Chengxiang Zhang^b, Rutan Zhang^a, Qi Hu^b,
6 Shanlei Qiao^{b✉}, Lei Li^{a✉}

7
8 *^aDepartment of Hygiene Analysis and Detection, School of Public Health, Nanjing Medical*
9 *University, Nanjing, Jiangsu 211166, P. R. China*

10 *^bThe Key Laboratory of Modern Toxicology, Ministry of Education, School of Public Health,*
11 *Nanjing Medical University, Nanjing 211166, Jiangsu, P.R. China.*

12
13
14 ✉ *Corresponding authors:*

15 *Lei Li, Department of hygiene analysis and detection, School of Public Health, Nanjing*
16 *Medical University, 101 Longmian Avenue, Nanjing 211166, P. R. China*

17 *Tel: +86-25-8686-8404; Fax: +86-25-8686-8499; E-mail: drleili@hotmail.com*

18 *Shanlei Qiao, The Key Laboratory of Modern Toxicology, Ministry of Education, School of*
19 *Public Health, Nanjing Medical University, Nanjing 211166, Jiangsu, P.R. China.*

20 *Tel: +86-25-8686-8402; Fax: +86-25-8686-8499; E-mail: alexqiao@139.com*

21

22

23

24

25

26

27

28

29 Abstract

30 Depression is a widespread mental disorder, however the molecular mechanism underlying depression and
31 antidepressive effects of diverse drugs remain largely unknown. An untargeted metabolic profiling based on Ultra
32 High Performance Liquid Chromatography coupled with Orbitrap mass spectrometry (UPLC-Orbitrap-MS) had
33 been performed to investigate the metabolic changes between chronic unpredictable mild stress (CUMS) rats and
34 healthy control rats, and screen for drug efficacy on these changed metabolites after administration of citalopram,
35 then provide a new perspective into the understanding for antidepressant effect of citalopram. Behavioral tests
36 were applied as a measurement of status of depression and the antidepressive effect. Multivariate statistics
37 including principal component analysis (PCA), partial least squares–discriminate analysis (PLS-DA) had been
38 applied to reveal the metabolic changes between control, model and antidepressant-treated group. Clear separation
39 among three groups was achieved and a total of 26 metabolites had been considered as the potential biomarkers
40 involved in the development of depression. Among them, 24 metabolites display the trend to return to normal level
41 which may correlate to the regulation of drug treatment on depression, and specifically, 10 of them could be the
42 key metabolites in the pathogenesis of depression and contribute to evaluate the effect of citalopram. These results
43 provided new insight into the pathophysiological mechanism of depression and indicated that citalopram mediated
44 synergistically abnormalities in metabolic network including tryptophan metabolism, energy metabolism and
45 synthesis of neurotransmitters, which may be helpful to evaluate its effectiveness by a metabolomics approach.

46 **Key words:** Depression; Metabolomics; Chronic unpredictable mild stress; UPLC-Orbitrap-MS

47

48 1. Introduction

49 Nowadays, depression has been known as one of the most common psychiatric illnesses with prevalence of
50 15%-20% around the world¹. A new WHO report predicts that depression will be the second most crucial cause of
51 suicide and disability by 2020². Different kinds of chronic stressful events have been proved to be the most
52 significant environmental factor in the etiology of the disease³. Currently, the common treatments for clinical
53 depression can be divided into two categories: psychotherapy and antidepressant medications. There are several
54 kinds of antidepressant medications such as the monoamine oxidase inhibitors (MOIs), the selective serotonin
55 reuptake inhibitors (SSRIs), and tricyclic antidepressants (TCAs)⁴. Among them the most widely and popular used
56 is the SSRIs which consists of citalopram, sertraline, fluoxetine *etc.*

57 Chronic unpredictable mild stress (CUMS) model has been proved a valid and reliable model of depression
58 for the reason that its symptoms are similar to human who suffer from depression⁵. Previous studies showed that

59 when after a period of CUMS exposure, animals would undergo a series of behavioral changes such as anxious and
60 fretful actions along with some metabolic changes, while these changes could be reversed by antidepressive drug
61 treatment^{5,6}. Therefore, the CUMS model has been widely used for the study of mechanism and the biochemical
62 changes in depression.

63 Previously, much of attention has been paid closely to the genomics (the gene expression) and the proteomics
64 (the protein function) due to the severity of depression^{7,8}. The emerging field of metabolomics, which devotes to
65 capturing the metabolic information at the global level, comes into our sight and offers us a promising chance to
66 come up with new hypotheses when study the molecular mechanisms of diverse diseases. Metabolomics places
67 emphasis on the induced biochemical perturbations according to changing metabolome in a specific period, which
68 can indicate our pathophysiological status⁹. It has been widely used to investigate the potential biomarker of
69 several diseases and evaluate the drug effects as a versatile tool^{10,11}.

70 It is common to combine multiple analytical techniques in order to get the biochemical information as much
71 as possible, and Nuclear Magnetic Resonance (NMR), gas chromatography/mass spectrometry (GC/MS) and
72 liquid
73 chromatography/mass spectrometry (LC/MS) are three common analytical instruments in metabolomics
74 research¹²⁻¹⁴. LC/MS has advantages in that LC provides better separation capabilities than GC with complex
75 biological samples and MS has greater sensitivity for qualitative and quantitative analysis than NMR. For complex
76 and mass data set acquired in metabolomics research, multivariate statistics have been applied to extract the
77 important information and visualize the differences between groups by the score map^{15,16}.

78 The primary goal of this work is to reveal the tiny changes of metabolites in the CUMS model, then screen for
79 the related pathways and investigate the therapeutic effects of citalopram according to the changes of metabolites
80 involved in the perturbed pathways by a metabolomics approach. Moreover, we also establish a method to rank the
81 level of depression by combining the results of behavioral tests and the changed metabolome, and this method is
82 validated by the antidepressant-treated group. It is very necessary and useful to discover the biomarkers which can
83 evaluate the efficiency of the therapeutic effect of antidepressive drugs at the early stage and assess the depressive
84 status according to the metabolome changing, so a high-resolution mass spectrometry (Orbitrap-MS) has been
85 applied to profile the urinary samples of control, model and antidepressant-treated group. Metabolites counted in
86 the distinction of three groups are identified as the potential biomarkers, and the regulated metabolites after drug
87 administration are taken for the biomarkers of the treatment effect of citalopram. This study is the first attempt to
88 combine the exploration of biomarkers of depression and evaluation of therapeutic effect of citalopram in CUMS

89 model rats by a metabolomics method, it can not only provide convincing biomarkers to evaluate drug effect of
90 citalopram, but also deepen the understanding of CUMS-induced metabolic perturbation and it help to access the
91 depressive state by combing the behavioral tests and the biomarkers.

92 **2. Experimental**

93 **2.1. Chemical and materials**

94 Methanol, acetonitrile, formic acid (HPLC grade) were purchased from Merck (Merck, Darmstadt, Germany).
95 Deionized water was acquired from Milli-Q50SP Reagent system (Millipore Corporation, MA, USA). Citalopram
96 hydrobromide was obtained from Sigma-Aldrich (MO, USA).

97 **2.2. Animals**

98 18 eight-week-old male Sprague-Dawley rats were commercially purchased from Shanghai Laboratory
99 Animal Co. Ltd. (SLAC, Shanghai, China). Rats were kept at a laboratory animal barrier system with required
100 environment (temperature of $24\pm 1^{\circ}\text{C}$, relative humidity of $45\pm 15\%$, and a 12h light/dark cycle) with access to food
101 and water ad libitum. The study was approved by national legislations of China and local guidelines and the
102 experiments were performed according to a protocol approved by the Nanjing Medical University Institutional
103 Animal Care and Use Committee.

104 **2.3. Chronic unpredictable mild stress (CUMS) procedure**

105 After 2 weeks of acclimatization, the rats were divided into three groups, control group (unstressed), model
106 group (stressed) and model group treated with citalopram according to the results of behavioral test and each group
107 contained 6 rats. The rats in three groups were housed individually. The control group had free access to the food
108 and tap water, excepted for a 20 hour food and water deprivation before each sucrose preference test. The model
109 group and the antidepressant-treated group were randomly exposed to a series of chronic unpredictable mild
110 stressors in the first three weeks, then the antidepressant-treated group received antidepressive drug administration
111 while both of two groups were still receiving the CUMS procedure. Citalopram was dissolved in physiological
112 saline and intraperitoneally injected at 9:00 daily in the morning at the dosage of 10 mg/kg. Meanwhile the rats in
113 another two groups were administered with the equivalent volume of physiological saline. The experimental
114 design is shown in Fig.1. The CUMS procedure was conducted according to protocol¹⁷, and stressors contained
115 food or/and water deprivation, 45°cage tilting along the vertical axis, paired housing, soiled cage (300 ml water
116 spilled into the padding), stroboscopic illumination (200 flashes/min), continuous overnight illumination, and
117 white noise (85db). The detailed schedule is displayed in Table.1.

118 **2.4. Behavioral test**

119 2.4.1. Sucrose preference test

120 The sucrose preference test (SPT) was conducted as a measurement of anhedonic effect in rats which exposed
121 to CUMS. In this test, after a 20-hour period of water and food deprivation, each rat was put into an individual
122 cage where placed two bottles of different solution, tap water and 1% sucrose solution. During the 2-hour test,
123 every rat stayed in a non-stressed environment and had open access to two kinds of solution. Sucrose preference
124 (SP) means the ratio of the amount of sucrose solution to that of total solution ingested in two hour [SP = sucrose
125 consumption/(sucrose consumption + water consumption) *100%] and it is defined as a measurement of
126 anhedonia¹⁷.

127 2.4.2. Open-field test

128 The open-field test (OPT) was done in a quiet environment (<65db) between 13:00 and 15:00 p.m. The open
129 field ground consisted of a black background floor which had been divided into 25 equal-size squares and a
130 40-cm-high side wall. Each rat was gently put into the central square under dim light and adapted for 30 seconds,
131 then took the 5-minutes-long test. The open-field was cleaned by 70% ethanol after each test. According to the
132 locomotor activity such as the time spent in the center square and the frequency of rearing, they were recorded as a
133 measurement to evaluate the status of rats¹⁸.

134 2.4.3. Forced swimming test

135 The forced swimming test (FST) is a valid test when evaluating the status of depression in the animals which
136 undergo the CUMS procedure and the antidepressant effects of drugs. The FST was conducted as described in the
137 previous study¹⁹. Briefly, the animals were individually placed in a plexiglas cylindrical (50cm in height and
138 18cm in diameter) filled with water (24±1°C) up to a height of 25 cm. Water was changed after each test. The total
139 immobility period during the 5-minutes-test was recorded with the help of stopwatch. The immobility status means
140 that when a rat just floats passively and just moves to keep its nose above water for survival.

141 2.5. Behavioral data statistical analysis

142 All the data acquired from behavioral test were organized into the form of mean±standard deviation. Data
143 were analyzed by using SPSS 17.0 software for Windows (Chicago, IL, USA). The behavioral data of SPT and
144 FST were analyzed by one-way analysis of variance (ANOVA) followed by post-hoc LSD test. The data of OPT
145 was a non-normal distribution, so the non-parametric tests (the Kruskal-Wallis test) was applied to analyze it. The
146 significance level was set at p<0.05.

147 2.6. Sample collection and preparation

148 After the final behavioral test at week 6, when the FST, OPT and SPT has been done, the collection of urine
149 sample then begin at 9:00 next morning. All rats were transferred into the metabolic cage in a non-stressed
150 environment to collect the urine samples. During the collection period, collecting tubes were put on ice to avoid
151 the enzymatic degradation. All samples were stored at -80°C immediately.

152 Prior to analysis, urine samples were thawed at room temperature and diluted at a ratio of 1:3 with methanol
153 (v/v) to remove the large-molecular-weight proteins, then were centrifuged at 11000g for 10 minutes. The
154 supernatants were diluted at a ratio of 1:3 with water (v/v) and then transferred to a vial for UPLC-Orbitrap-MS
155 analysis.

156 **2.7. UPLC-Orbitrap-MS analysis**

157 Metabolic profiling was performed on a UPLC Ultimate 3000 system (Dionex, Germering, Germany),
158 coupled to an Orbitrap mass spectrometer (Thermo Fisher Scientific, Bremen, Germany) in both positive and
159 negative mode simultaneously. Both the UPLC and the Orbitrap mass spectrometer system were controlled by the
160 Xcalibur 2.2 software (Thermo Fisher Scientific).

161 The chromatographic separation was performed on a 1.9 μ m Hypersile Gold C18 column (100mm \times 2.1mm)
162 (Thermo Fisher Scientific), and the column was maintained at 40°C. A multistep gradient consisted of 0.1% formic
163 acid in water (A) and 0.1% formic acid in acetonitrile (B) had been applied and the gradient operated at a flow rate
164 of 0.4 mL/min by linearly increasing solvent B from 5% to 95% over 15 min, then the column was washed with 95%
165 solvent B for 2 min and re-equilibrated in 5% solvent B. The UPLC autosampler temperature [Ultimate WPS-3000
166 UPLC system (Dionex, Germering, Germany)] was set at 4 °C and the injection volume for each sample was 5 μ L.

167 MS data were collected by the Orbitrap mass spectrometer equipped with a heated electrospray source (HESI)
168 at the resolution of 700,000. For both positive and negative mode, the operating parameters were as follows: a
169 spray voltage of 3 kV, the capillary temperature of 300°C, sheath gas flow of 40 arbitrary units, auxiliary gas flow
170 of 10 arbitrary units, sweep gas of 2 arbitrary units and S-Lens RF level of 50. In the full scan analysis (70 to 1050
171 amu), the resolution was set at 700,000 with an automatic gain control (AGC) target of 1×10^6 charges and a
172 maximum injection time (IT) of 120 ms. The energy of higher energy collision dissociation (HCD) is set at 70.0eV
173 in MS/MS experiment.

174 The quality control (QC) samples had been prepared by pooling same volume of urine from all samples and
175 analyzed every 10 samples to ensure the stability and repeatability. The mass spectrometry was calibrated every 24
176 hour while the profiling to ensure the mass accuracy.

177 **2.8. Data analysis**

178 All the profiling raw data files were produced by the SIEVE software (Thermo Fisher Scientific) where data
179 pretreatment procedures such as baseline correction, peak deconvolution and peak realignment had been done.
180 This progress produced a table organized in a three-dimensional matrix, including annotated peak indices (RT-m/z
181 pairs), sample names (observations), and intensity of each sample (i.e. peak area). The multivariate statistics was
182 performed by the SIMCA-P 13.0 software (Umetrics, Umea, Sweden)²⁰. All data were mean-scaled and imported
183 into the software for unsupervised PCA to identify the tiny differences among the samples. The PCA helped to
184 reduce the high dimensional spectral variation into a two or three principal components (PCs) without losing the
185 vast majority of information. The PCA score map was used to visualize the possible distribution the clustering or
186 the grouping in the observations. In order to improve the classification, offer pairwise comparison between three
187 groups, and search for the changed metabolites induced by CUMS procedure, the partial least squares discriminant
188 analysis (PLS-DA) had been performed in this study. The dataset acquired from model and control group was
189 chosen to conduct the PLS-DA model, then the dataset of antidepressant-treated group was used to validate and
190 test the predictive ability of the model. Then the variable importance in the projection (VIP) analysis was applied
191 to obtain the metabolites which counted most in the distinction of three groups and the variables with VIP values
192 greater than 1.0 were considered statistically significant in this model.

193 Furthermore, unpaired Student's t-test carried out by R language (<http://www.r-project.org/>) was used to
194 identify differential metabolites between groups. The q-value was employed to address the multiple testing²¹. The
195 q-value was calculated by the package q-value of R language and indicated that an estimated possibility of the
196 metabolites to that point were different among the three groups simply by chance. The compounds with q-value
197 less than 0.05 and VIP values greater than 1.0 were defined to be statistically significant.

198 **3. Results**

199 **3.1. Behavioral testing**

200 The SPT was done according to the experiment design mentioned above, and the SP of the three groups in
201 different periods was displayed in Fig.2. After first three weeks of CUMS exposure, significantly decreased SP,
202 depressive-like action such as anxious and aggressive behaviors were observed in model group and
203 antidepressant-treated group before drug administration. In the next three weeks, after drug administration and
204 continuously exposed to CUMS procedure, the SP of model group went down significantly ($p < 0.01$), while
205 antidepressant-treated group started to recover compared with model group ($p < 0.05$), which indicated the model
206 had been successfully conducted and citalopram did work.

207 For the OPT, the locomotor activity was measured twice during the CUMS period, time spent in center square
208 and the frequency of rearing were recorded as an evaluation of stress status¹⁷. The results of OPT at week 3 and
209 week 6 were shown in Fig.3. At week 3, rats in both model group and antidepressant-treated group before drug
210 administration spent much less time in the center square ($p < 0.05$ and $p < 0.05$, respectively). After received the
211 antidepressive drug treatment, the antidepressant-treated group showed a longer time staying in the center in
212 contrast with model group ($p < 0.05$). In the previous study, the time spent in the center square could be a
213 measurement of the degree of anxiety, which indicated animals displayed high activity level in the center could be
214 regarded as less anxious. For the frequency of rearing, it was greatly decreased in the model group compared with
215 the control group and the antidepressant-treated group ($p < 0.01$ and $p < 0.05$, respectively) at week 6. The reduction
216 of the rearing frequency and the shorter time in central area indicated a decrease in exploration ability in model
217 group¹⁸. These results demonstrated that depressive-like behaviors were observed in model group and these actions
218 improved due to the regulation of citalopram.

219 In the FST, the result was displayed in Fig.4. when after exposed to CUMS for three weeks, the immobility
220 time (floating state) was significantly increased in two groups compared with control group ($p < 0.05$, $p < 0.05$,
221 respectively), and then after antidepressive treatment while exposed to CUMS at the same time, the immobility
222 time of antidepressant-treated group began to decrease. At week 6, the immobility time in the model group showed
223 a significant increase compared with the control and antidepressant-treated group ($p < 0.01$, $p < 0.05$, respectively).
224 Additionally, the immobility time between control and antidepressant-treated group did not achieve statistical
225 significance ($p > 0.05$), which indicated the increase in the immobility time was closely related to the depressive
226 state.

227 3.2. Metabolomics profiling

228 The urinary metabolic profiling of control, model and antidepressant-treated group were obtained by
229 UPLC-Orbitrap-MS with the pre-described condition. Full scan mode was engaged to investigate the urinary
230 metabolites in both positive and negative modes simultaneously. A total of 3525 features, 2507 in positive mode
231 and 1018 in negative mode were obtained. A three-component PCA model was initially obtained with dataset
232 which was extracted and pretreated by SIEVE software. 66.7% of the total variations were accumulated within the
233 first three components. The score plot (Fig.5a.) showed a relative good separation between three groups which
234 preliminarily indicated the different metabolic pattern between groups.

235 **3.3. Prediction of depression ranking**The PLS-DA score plots showed that the metabolic profiles of model
236 rats deviated from those of control and antidepressant-treated group ($R^2Y = 0.969$, $Q^2 = 0.861$; Fig.5b.) and

237 suggested that significant biochemical changes were induced by CUMS. More specifically, we tried to combine
238 results of all the different behavioral test so as to get a comprehensive indicator to reflect the depression status, but
239 unfortunately, it is difficult to find a gold standard to give the weight of each result of the behavioral test. In order
240 to state the depression level as accurate as possible according to practical situation, we chose the SP as the
241 measurement of depression for its wide use in the CUMS model. So in the PLS-DA model, we defined the SP as a
242 Y variable, and the metabolic dataset of three groups was set as the X variable, then autofit the model in order to
243 study the relationship between the changed-metabolome and the depression status.

244 **3.4.** The score plot showed that the control and model group were clearly distinguished from each other and the
245 antidepressant-treated group also had distinctive pattern compared with the model group, while after drug
246 administration, the clusters of antidepressant-treated group was moving towards the control group especially
247 in the first component. The key parameters for assessing modeling quality ($R^2Y=0.910$ and $Q^2=0.733$) both
248 were larger than 0.5, which suggested that this model was predictive and robust. Furthermore, we applied a
249 permutation test to validate the PLS-DA model (Fig.5.c), and the validation plot showed that Q^2 and R^2 were
250 higher than those corresponding permuted Q^2 and R^2 values at left side. These results not only mean that the
251 PLS-DA model was robust and had a relative good predictive power when facing new observations, but also
252 demonstrate that it is practicable to rank the level of depression and evaluate the drug effect according to the
253 metabolome changes along with the behavioral test results by a metabolomics method.**Searching for identity**
254 **of biochemical changes**

255 The variable importance for projection (VIP) statistics was obtained from the PLS-DA and had been applied
256 for the selection of discriminational variables among the three groups. By considering the q-value, along with the
257 VIP-value, we revealed that there were 26 metabolites were significantly changed ($q\text{-value}<0.05$, $VIP\text{-value}>1.0$)
258 when exposed to the CUMS and then after drug administration, 24 metabolites changed before turned out a trend
259 to return to normal. Specifically, 10 of them were significantly changed ($q\text{-value}<0.05$) in the urine of
260 antidepressant-treated group compared with model group and had no statistically significant difference with
261 control group. These results indicated that the 26 metabolites could be the potential biomarkers of depression and
262 the 10 significant changed metabolites were the key metabolites and could be the biomarkers to evaluate the drug
263 effect of citalopram from the perspective of metabolomics.

264 **3.5. Metabolite identification**

265 The significantly changed metabolites indicated by q-value and VIP-value were presented in the form of
266 retention time and m/z pairs. A library consisting 493 authentic chemicals with high accurate m/z and retention

267 time in pre-described condition were established in our laboratory for identification of metabolomics analysis. The
268 library was conducted with the same column for metabolomics analysis and a model transfer protocol was created
269 to ensure the retention time shift when changing column. Briefly, high retention time reproducing chromatography
270 columns were carefully selected with no supplier change; the standards were carefully grouped and retention time
271 were re-acquired when columns changed. By comparing the retention time and high accurate m/z with specific
272 authentic standards, the identification was then delivered. For the high resolution of Orbitrap mass spectrometry, it
273 is exact enough for the identification by using the retention time and high accurate m/z and MS/MS spectra are
274 further compared with those in our library.

275 21 of the compounds were confirmed by comparing retention time and high accurate m/z with the local
276 library and 16 of them were further validated by MS/MS spectra. The rest 5 metabolites without authentic
277 chemicals was identified by searching the HMDB (<http://www.hmdb.ca/>) database for candidates with similar
278 molecular weight in the tolerance of 5ppm. According to the fold change and q-value, there were 10 metabolites
279 significantly changed after antidepressant treatment including glycerol, creatine, quinolinic acid, L-phenylalanine,
280 kynurenic acid, Gamma-aminobutyric acid, L-kynurenine, L-isoleucine, N-acetyl-L-aspartic acid, indoleacetic acid.
281 The list of 26 significantly changed metabolites after exposed to CUMS and the 10 key biomarkers that could be
282 useful to offer a better understanding of antidepressant effect of citalopram was shown in Table.2.

283 **4. Discussion**

284 **4.1. CUMS-induced metabolomic changes**

285 After the CUMS exposure, metabolic changes were observed and this was consisted with the behavioral tests.
286 In our research, compared with the control group, 26 metabolites were significantly altered in the model group and
287 the related metabolic pathways were shown in Fig.6.

288 **4.2. The therapeutic effect of citalopram in the rats exposed to CUMS**

289 When rats in antidepressant-treated group received the administration of citalopram, 24 of the 26
290 significantly-changed metabolites mentioned above displayed the trend to return to normal level, and specifically,
291 the level of glycerol, creatine, quinolinic acid, phenylalanine, kynurenic acid, gamma-aminobutyric acid,
292 kynurenine, isoleucine, N-acetyl-L-aspartic acid, indoleacetic acid was significantly changed compared with
293 model group due to the regulation of citalopram, which meant they were useful in evaluation of citalopram's
294 therapeutic effect.

295 Furthermore, the PLS-DA showed that the cluster of antidepressant-treated group was moving towards to the
296 control group especially in the first component, which was consist with the results of behavioral test, and this

297 indicated that the PLS-DA model has a good predictive ability and meant is was promising to combine the changed
298 metabolome and the behavioral test to rank the level of depression in the following research.

299 **4.3 Biochemical interpretation**

300 **4.3.1. Amino acids metabolism**

301 Tryptophan (Trp) and the related metabolites kynurenine (Kyn), kynurenic acid (KA), 5-hydroxyindoleacetic
302 acid (5-HIAA), 4,6-dihydroxyquinoline, quinolinic acid (QUIN), indoxyl sulfate, indole-3-carboxylic acid and
303 xanthurenic acid (XA) were significantly changed in the urine of rats in model group. Trp is an essential amino
304 acid and the metabolism of it is quite complicated in our body. Except for synthesizing proteins, there are three
305 main metabolic pathways: indoleacetic acid (IAA) pathway, Kyn pathway and serotonin (5-HT) pathway²². Trp
306 can be converted into formylkynurenine by L-tryptophan-2, 3-dioxygenase (TDO) or indoleamine-2,
307 3-dioxygenase (IDO), and then kynurenine formamidase converts it into Kyn. Kyn is the first key branch point in
308 Trp's catabolic pathway which undergoes a series of catabolic reactions producing KA, XA or the QUIN. Previous
309 study has reported that when suffering depression, the increasing proinflammatory cytokines can active IDO,
310 which results in the decrease of Trp²³. It is consistent with the results in our study, which shows a large decline of
311 Trp in urine of model group compared with the control group. 5-HIAA is a main breakdown product of serotonin
312 that is excreted in the urine, and the significant decline of 5-HIAA indicates the low level of its precursor 5-HT. As
313 a monoamine neurotransmitter, the lower level of 5-HT has been reported in the previous study which has the
314 relationship with the development with depression according to the monoamine neurotransmitter hypothesis^{24,25}.
315 The decrease of 5-HIAA and Trp, combined with the increasing concentration of Kyn, we can infer that the Kyn
316 pathway has been promoted which leads to reduce the availability of Trp for conversion into 5-HT. In the
317 xanthurenic pathway, the increased concentration of XA may indicate that the level of its precursor
318 L-3-hydroxykynurenine (L-3-HK) had also risen. An elevation of L-3-HK levels had been shown to constitute a
319 significant hazard in situations of excitotoxic injury and cause the death of neuronal cells which may be the
320 inducement of depression and some other CNS diseases, and the increasing level of XA founded in our research
321 was also consists with the results in the depression patients.

322 When Trp is catabolized into Kyn, the metabolism of Trp mainly comes into the kynurenine pathway in which
323 the product of it can affect the neuroprotective–neurodegenerative balance in the brain. Kyn can be further
324 metabolized into two main pathways, the toxic quinolinic pathway and the kynurenic pathway. Firstly, Kyn can be
325 converted into 3-hydroxy kynurenine which is the bioprecursor of QUIN by kynurenine-3-monooxygenase.
326 3-hydroxy kynurenine is a free radical generator, and it can cause neurons apoptosis and neurodegenerative

327 changes in brain. 3-hydroxy kynurenine can be further catabolized into QUIN, which is a kind of endogenous
328 excitotoxic material and can lead to many neurodegenerative disorders. However, Kyn can also be metabolized to
329 KA which proved to be a NMDA receptor antagonist and have protective effects against QUIN by kynurenine
330 aminotransferase²⁶. The metabolic abnormalities of Kyn can disequilibrate the balance of the neuroprotective–
331 neurodegenerative metabolites. In our research, the concentration of KA in model group was significantly lower
332 than control group, which suggested that the metabolism of Kyn was mainly going into the quinolinic pathway.
333 The lower level of Kyn and the up-regulation level of QUIN was consist with it. The increasing level of QUIN and
334 the decreasing level of KA led to the imbalance in the neuroprotective and neurodegenerative metabolites which
335 could be a hypothesis to further study the mechanism of depression and on this basis to evaluate the effect of
336 citalopram.

337 KA is an endogenous antagonist of the excitatory amino acid receptors which plays an important role in the
338 protection of nervous system and anticonvulsive activities. In some animal models of neurodegenerative diseases,
339 KA has been proved that it has the anticonvulsive and neuroprotective functions. As is mentioned above, the higher
340 level of QUIN could cause the apoptosis of astrocytes, and this would lead to the lower neuroprotective activity
341 against QUIN, and worse still, this may further caused the apoptosis of astrocytes, then result in consecutive
342 down-regulation of KA, which makes the balance of neuroprotective–neurodegenerative metabolites worse and
343 worse in the brain²⁷. As a neuroprotective metabolite, KA has been reported that it is closely connected to the
344 pathogenesis of some neurological diseases with age, such as the Parkinson's disease and Alzheimer's disease and
345 patients suffered severe metabolic disorder of KA in the ageing progress^{28, 29}. This remarkable profile of KA
346 metabolism alterations in the mammalian brain has been suggested to result from the development of the
347 organization of neuronal connections and synaptic plasticity, development of receptor recognition sites, maturation
348 and ageing. When after the drug administration, it was observed that the therapy could prevent the down-regulation
349 of KA which indicated that citalopram could have effect on the regulation of the dysfunction in the tryptophan
350 metabolism. When the Kyn metabolizes into the toxic quinolinic pathway, in which the excitotoxic QUIN is
351 produced and this may leads to an imbalance of neuroprotective and the excitotoxic metabolites. When released by
352 the activated macrophages, QUIN can act as a kind of endogenous excitotoxic material which is related to many
353 psychiatric disorders. Previous study proved that in the urine of depressed patients, there was a significant higher
354 level of 3-hydroxykynurenine (3-HK) that could result in the producing of QUIN than normal people³⁰. And in our
355 study, although we did not observe the up-regulated level of 3-HK, we found the level of QUIN in the CUMS
356 group was significant higher than the control group, and after treatment it had been down-regulated in

357 antidepressant-treated group compared with the model group, which indicated QUIN could be a biomarker of the
358 disorder of tryptophan metabolism and then served as a tool for investigation of the pathogenesis of depression and
359 pharmaceutical effect of citalopram. A lot of researches to study the mechanism of depression mechanism have
360 concentrated on the changed level of 5-HT, and have proved that depression has a close relation to the decline of
361 5-HT^{31,32}. Our research showed that the metabolic disturbance of tryptophan not only led to the decline of 5-HT,
362 but also a significant increase of QUIN. When the increased level QUIN reaches the pathological concentration, it
363 can over-activated the NMDA receptor and the metabotropic glutamate receptors which can cause the nerve
364 damage. Moreover, the up-regulation of QUIN can also inhibit the reabsorption of glutamic acid in the gliocyte,
365 which will intensify the excitotoxic effect in the CNS.

366 Isoleucine (Ile), as a kind of branched chain essential amino acid, plays a critical role in the human life and
367 has been proved particularly involved in stress, energy and muscle metabolism. In our body, Ile can provide the
368 amino when in the synthesis of glutamate, which has been regarded as an important neurotransmitter in the central
369 nervous system, by passing the blood-brain-barrier. It has been reported that the damaged homeostasis of
370 glutamate and glutamatergic neurotransmission may be closely linked to the development of depression³³.
371 Moreover, Ile can combine with glutamine to form the glutamyl-isoleucine, which is a kind of dipeptide and play
372 an important role in the locomotor behavior. Previous researches have shown that when the rats received the
373 injection of glutamyl-isoleucine into ventral tegmental area, which can transmit afferent glutamatergic
374 projections to the prefrontal cortex, and this can greatly improve the locomotor behavior. In our research, after
375 drug administration, the significantly down-regulated level of Ile had return to normal, and before treatment the
376 low Ile level was accompanied by the low degree of locomotor behavior in CUMS model group. On account of the
377 finding, it is necessary to further look into the relationship between Ile and the glutamatergic activity during the
378 development of depression. Besides, Ile can be converted into both carbohydrates and fats as it is an essential and
379 ketogenic amino acid, which means it can also affect the energy metabolism. In the citric acid circle, it can be
380 converted into succinyl CoA in the presence of alpha-ketoglutarate, which suggests that CUMS may also has
381 impact on the energy metabolism.

382 L-Serine can be biosynthesized from glycine which is a well-acknowledged inhibitory neurotransmitter.
383 Furthermore, a D/L-Serine racemase can link the formation of D-Serine and glycine to the L-Serine metabolism³⁴.
384 ³⁵. In our body, the function of L-Serine is not only offering the nucleotide precursors during the cell proliferation,
385 but also has the trophic effects³⁶. Glycine is one of the well-known inhibitory neurotransmitter and the role of
386 glycine is to regulate the locomotor behavior. It has been reported that in the plasma and urine of the patients who

387 suffered the schizophrenia, the concentration of serine and glycine had evaluated³⁷. However, the changes of the
388 serine and glycine in patients suffering psychiatric disorders have not been confirmed because there are some
389 different results in other study³⁸⁻⁴⁰, which means it is necessary to focus on the regulation of the pathways involved
390 and the specific activity of particular enzymes to get a better understanding and biochemical interpretation of
391 L-Serine, D-Serine and glycine.

392 **4.3.2. Synthesis of neurotransmitter**

393 In the metabolic pathway of phenylalanine (Phe) and tyrosine (Tyr), the concentration of Phe and its
394 metabolite phenylpyruvic acid was significantly increased, and the level of dopamine and its precursor L-dopa was
395 decreased in the model group compared with the control group. Phe is an essential amino acid which can be
396 incorporated into cellular proteins, or converted to phenylpyruvic acid, and it is the precursor for the amino acid
397 tyrosine (Tyr). Phe can be converted into Tyr in the liver and then, as a precursor for L-dopa, Tyr can be further
398 metabolized into the neurotransmitters dopamine, norepinephrine and epinephrine. Half of the Phe would go into
399 the biosynthesis of Tyr through phenylalanine hydroxylase. Research showed that depressed patients had a higher
400 ratio of Phe-Tyr, which meant there could be a dysfunction of phenylalanine hydroxylase^{41,42}. According to the
401 results, it could be found that more Phe had been metabolized into phenylpyruvic acid, along with the low level of
402 L-dopa and its metabolite dopamine, we could deduce that CUMS caused the metabolic disturbance of Phe and led
403 to the low level of Tyr. The reason why we didn't detect Tyr may be that Tyr had been rapidly metabolized, so it
404 was not found in large concentrations throughout the body. When the Tyr has been down-regulated, the
405 biosynthesis of its metabolites such as L-dopa, dopamine, norepinephrine and epinephrine could be disturbed, and
406 lower level of L-dopa is consist with this. When under stress, it is suggested that people need more Tyr so that the
407 supplement of it could prevent the stress-induced depletion of norepinephrine. The metabolic disturbance of Phe
408 and Tyr has closely relationship with depression, and could be the biomarkers to study the mechanism of
409 depression along with to evaluate the effect of the treatment for depression by citalopram.

410 Gamma-aminobutyric acid (GABA) is a chief inhibitory neurotransmitter in the nervous system, which can
411 act at inhibitory synapses in the brain. GABA works by binding to specific receptors in the membrane of both pre-
412 and postsynaptic neurons. GABA can be synthesized from glutamate with the action of L-glutamic acid
413 decarboxylase and pyridoxal phosphate. The drugs which can increase the available level of GABA or work as the
414 agonists of GABA receptors have been reported have anti-anxiety and anti-convulsive effects. It has been reported
415 that the dysfunction of GABA neurotransmitter system has great relationship with depression and has been
416 observed that there is an increased level of GABA in occipital cortex of depressed patients when receive the

417 treatment of SSRI^{43, 44}. Moreover, previous study also showed that GABA was involved in the pathogenesis of
418 some anxiety and mood disorders, while the mechanisms of the decreasing level of GABA was not quite sure⁴⁴.
419 The significantly reduced concentration of GABA observed in our research could be due to the dysfunction of
420 GABA synthesis which resulted from the reduction of glutamatergic stimulation along with the decreasing level of
421 necessary substrate involved in the synthesis of GABA. Moreover, the reduced level of GABA also can partly
422 reflect the abnormal activity of glutamic acid decarboxylase which has been observed that in depressed patients
423 and some environmental factors also can influence the regulation of the glutamic acid decarboxylase activity⁴⁵. In
424 this research, the result is consists with the previous study, which shows a significant down-regulation level of
425 GABA, and moreover, the GABA level then up-regulated after treatment, which meant the urinary level of GABA
426 could be very useful in the diagnosis of depression and be the trait biomarker for evaluating the effect of
427 citalopram by a metabolomics method.

428 N-Acetylaspartic acid (NAA) is a derivative of aspartic acid which is a major excitatory neurotransmitter and
429 can provide resistance to fatigue. NAA can be biosynthesized by aspartic acid and acetyl-CoA in neuronal
430 mitochondria. In the CNS, NAA is the second most concentrated molecule in the brain just after glutamate which
431 is the most abundant fast excitatory neurotransmitter. As a neuronal osmolyte, NAA can maintain the fluid balance
432 in the brain, and it is a source of acetate for lipid and myelin synthesis in oligodendrocytes, the glial cells that
433 myelinate neuronal axons. Moreover, it is also the precursor in the synthesis of N-acetylaspartylglutamate which is
434 an important neuronal dipeptide, and NAA play role in the energy and lipid metabolism. It has been observed that
435 the level of NAA is significantly down-regulated in depressed patients and in patients with brain atrophy⁴⁶.
436 However, the dysregulation of NAA in CUMS rats is not consistent with each other for the differences in
437 measurement equipment and samples⁴⁷⁻⁴⁹. In our study, we observed a significant down-regulation level of NAA in
438 CUMS group and then it had been corrected after drug administration, the difference among the CUMS group,
439 control group and drug administration group had statistical significance.

440 **4.3.3. Energy metabolism**

441 The pyruvic acid, citric acid, creatine, hypoxanthine and succinic acid are another five key metabolites which
442 play an important role in the discrimination of the three groups. These metabolites are crucial in the citric acid
443 cycle which is associated in the energy metabolism. The pyruvic acid is one of the most important metabolic
444 intermediates in the basic metabolisms in our body and it can link the transformation of carbohydrates, lipids and
445 amino acids by converting into acetyl-CoA and participating in the citric acid cycle. In the aerobic condition,
446 pyruvic acid can be converted into acetyl-CoA by pyruvate dehydrogenase system in mitochondria. The high level

447 of pyruvic acid along with the low level of citric acid and succinic acid indicated that the citric acid cycle had been
448 affected by the CUMS procedure and caused the energy metabolism perturbation. More specifically, the deficiency
449 of citric acid and succinic acid resulted in the energy deficiency which manifest as exhaustion and fatigue. One of
450 the most frequent symptoms of depression is fatigue and this is consistent with our results which indicate there is
451 an energy metabolism perturbation in the pathogenesis of depression.

452 Creatine is an amino acid that occurs in vertebrate tissues and in urine. In our body, creatine is synthesized
453 mainly in the liver and most of it is stored in skeletal muscle and the rest of it is stored in brain and heart. The main
454 function of creatine is participating in the transport of the cell's energy and the high energy phosphate group of
455 ATP is transferred to creatine to form phosphocreatine which is reversibly catalyzed by creatine kinase. In this
456 study, significantly down-regulated level of creatine was observed in the model group compared with the control
457 group, and when received the drug administration, the concentration of creatine in antidepressant-treated group
458 was associated with a significant rise at the side of the model group and had no statistical significance with the
459 control group. Although the mechanism of creatine is not clearly enough in depression, it has been proved that
460 creatine is closely associated with the energy deficiencies. While in depression, there are many common symptoms
461 such as extreme fatigue, psychomotor retardation and lethargy which is consistent with energy deficiencies. This
462 may be the joint result of the changed level of some energy-metabolism-related materials. The foregoing result
463 implies that creatine can be a potential biomarker for depression and also can evaluate the effect of citalopram by
464 the metabolomics method.

465 Glycerol is an important component of the phospholipids and triglycerides, which is a three-carbon substance
466 that forms the backbone of fatty acids in fats. When the body uses stored fat as a source of energy, glycerol and
467 fatty acids are released into the bloodstream. The glycerol component can be converted to glucose by the liver and
468 provides energy for cellular metabolism. Previous studies showed that there did occur the lipid metabolic disorder
469 in patients suffering from depression, along with the perturbations of some protein related to the lipid metabolism
470 in CUMS rats^{50,51}. So in this study the significantly down-regulated level of glycerol discovered in the model
471 group may due to the perturbation of lipid metabolism, and after treatment, the glycerol level returned to normal
472 which indicated it could be a biomarker to evaluate the drug effect.

473 **4.3.4. Intestinal flora variation**

474 In the model group, the concentration of indoleacetic acid (IAA) and indoxyl sulfate was significantly
475 decreased and up-regulated respectively. These changed metabolites was closely linked to the metabolism of
476 amino acids through the intestinal flora^{52,53}, and the perturbation of them may indicated the imbalance of intestinal

477 flora. The imbalance of intestinal flora could have an impact on the appetite, and previous research has reported
478 the symptom of the loss of appetite in depressed patients⁵⁴. IAA is a breakdown product of tryptophan metabolism
479 and is often produced by the action of bacteria in the mammalian. It may be produced by the decarboxylation of
480 tryptamine or the oxidative deamination of tryptophan. The reduced concentration of IAA had been observed in the
481 model group, and this may be due to the loss of appetite which is a clinical symptom in depressed patients. The
482 induced changes of IAA along with indoxyl sulfate could be partly explained by the disturbance of enteric flora
483 when facing CUMS procedure, which has been reported that there is a connection between irritable bowel
484 syndrome and depression. Previous research has reported that some depressed patients had gastrointestinal
485 symptoms due to the intestinal flora disturbance⁵⁵. Moreover, some research also reported a two-way link between
486 gut and brain which was the synergistic effect of immune system, endocrine system and gut^{56,57}. The two-way
487 link can associate depression with the alteration of intestinal flora, and the metabolism imbalance of IAA and
488 indoxyl sulfate in model group could provide a new insight to study the aetiopathogenesis of depression and learn
489 the effect of citalopram from a metabolomics method. Gut flora also plays an important in many essential
490 physiological functions even in the biosynthesis of 5-HT and the metabolism of tryptophan⁵⁸. Gut flora can
491 regulate the tryptophan metabolism by affecting the production of proinflammatory cytokine or cortisol. Recently,
492 an article in cell shows that most of 5-HT is produced in intestinal and stored in enterochromaffin cell⁵⁹. By
493 comparing the mice which don't have gut flora in intestinal with the healthy control, the level of 5-HT is about 60%
494 lower than normal mice, and when injected with the probably-related gut flora, 5-HT is significantly up-regulated.
495 All these results demonstrates that tryptophan metabolism can also be affected by gut flora, and this link the
496 intestinal flora variation with tryptophan metabolism which helps to give an overall understanding of depression.

497 **5. Conclusion**

498 In conclusion, our study focused on the metabolomic changes induced by CUMS procedure and firstly
499 studied whether the metabolome changes could correctly reflect the status of depression and obtained promising
500 results. Furthermore we searched for the potential biomarkers of depression and evaluate the therapeutic effect of
501 citalopram according to the regulated metabolites by a metabolomics method. A total of 26 metabolites accounted
502 for the difference between the control, model and antidepressant-treated group had been found and identified, and
503 then 24 of them appeared a trend to return to normal level. We also gave the appropriate interpretations for these
504 potential biomarkers. According to our study, the metabolic changes in the model group showed that the
505 depression induced by CUMS were mainly associated with the perturbation of amino acids metabolism, energy
506 metabolism, synthesis of neurotransmitter and intestinal flora disturbance. After drug administration, the

507 therapeutic effects of citalopram on CUMS rats was observed and resulted in the regulation of 10 key metabolites
508 involved in the perturbative pathways. Meanwhile, the PLS-DA model showed a good predictive ability which
509 meant it is feasible to evaluate the status of depression and the therapeutic effect of citalopram according to the
510 changed-metabolites at a specific period by a metabolomics method. This is the first attempt to combine the
511 exploration of biomarkers of depression and evaluating the effect of citalopram by a metabolomics method, and
512 then use the method to access the depression status according to the changed metabolome. It is promising to use
513 the metabolomics tools to evaluate the relationship between depression and relevant somatic symptoms by testing
514 the metabolic changes of the urine, and it has bright future in making a definite evaluation the therapeutic effect of
515 antidepressant drugs. Moreover, this untargeted metabolomics study can lay the foundation for our next targeted
516 metabolomics study of the patients suffering depression or other neurological disorders.

517

518 **Acknowledgements**

519 This work was supported by the Natural Science Foundations of China (81473020) and a project funded by
520 the Priority Academic Program Development of Jiangsu Higher Education Institutions (2010).

521

522 **References:**

- 523 1. P. F. Sullivan, M. C. Neale and K. S. Kendler, *American Journal of Psychiatry*, 2000,
524 **157**, 1552-1562.
- 525 2. K. Chaudhari, S. Khanzode, G. Dakhale, A. Saoji and S. Sarode, *Indian Journal of*
526 *Clinical Biochemistry*, 2010, **25**, 77-81.
- 527 3. D. J. Stein, K. C. Koenen, M. J. Friedman, E. Hill, K. A. McLaughlin, M. Petukhova, A.
528 M. Ruscio, V. Shahly, D. Spiegel and G. Borges, *Biological psychiatry*, 2013, **73**, 302-312.
- 529 4. R. Rizzoli, C. Cooper, J.-Y. Reginster, B. Abrahamsen, J. Adachi, M. Brandi, O.
530 Bruyère, J. Compston, P. Ducy and S. Ferrari, *Bone*, 2012, **51**, 606-613.
- 531 5. P. Willner, A. Towell, D. Sampson, S. Sophokleous and R. Muscat,
532 *Psychopharmacology*, 1987, **93**, 358-364.
- 533 6. P. Willner, R. Muscat and M. Papp, *Neuroscience & Biobehavioral Reviews*, 1992,
534 **16**, 525-534.
- 535 7. G. Naert, G. Ixart, T. Maurice, L. Tapia-Arancibia and L. Givalois, *Molecular and*
536 *Cellular Neuroscience*, 2011, **46**, 55-66.
- 537 8. V. Michopoulos, K. M. Reding, M. E. Wilson and D. Toufexis, *Hormones and*
538 *behavior*, 2012, **62**, 389-399.
- 539 9. R. Kaddurah-Daouk, S. Rozen, W. Matson, X. Han, C. M. Hulette, J. R. Burke, P. M.
540 Doraiswamy and K. A. Welsh-Bohmer, *Alzheimer's & dementia : the journal of the Alzheimer's*
541 *Association*, 2011, **7**, 309-317.

- 542 10. M. M. Koek, R. H. Jellema, J. van der Greef, A. C. Tas and T. Hankemeier,
543 *Metabolomics*, 2011, **7**, 307-328.
- 544 11. W. B. Dunn, A. Erban, R. J. Weber, D. J. Creek, M. Brown, R. Breitling, T.
545 Hankemeier, R. Goodacre, S. Neumann and J. Kopka, *Metabolomics*, 2013, **9**, 44-66.
- 546 12. W. B. Dunn, D. I. Broadhurst, H. J. Atherton, R. Goodacre and J. L. Griffin, *Chemical*
547 *Society Reviews*, 2011, **40**, 387-426.
- 548 13. M. Ernst, D. B. Silva, R. R. Silva, R. Z. Vêncio and N. P. Lopes, *Natural product*
549 *reports*, 2014, **31**, 784-806.
- 550 14. Y. Li, Z. Zhang, Z. Hou, L. Wang, X. Wu, L. Ju, X. Zhang and Y. Zhang, *RSC Advances*,
551 2015, **5**, 202-209.
- 552 15. J. Trygg, E. Holmes and T. Lundstedt, *Journal of proteome research*, 2007, **6**,
553 469-479.
- 554 16. H. Wu, A. D. Southam, A. Hines and M. R. Viant, *Analytical biochemistry*, 2008,
555 **372**, 204-212.
- 556 17. P. Willner, *Psychopharmacology*, 1997, **134**, 319-329.
- 557 18. R. N. Walsh and R. A. Cummins, *Psychological bulletin*, 1976, **83**, 482.
- 558 19. R. D. Porsolt, G. Anton, N. Blavet and M. Jalfre, *European journal of pharmacology*,
559 1978, **47**, 379-391.
- 560 20. O. Galtier, O. Abbas, Y. Le Dréau, C. Rebufa, J. Kister, J. Artaud and N. Dupuy,
561 *Vibrational Spectroscopy*, 2011, **55**, 132-140.
- 562 21. R. Kaddurah-Daouk, B. S. Kristal and R. M. Weinshilboum, *Annu. Rev. Pharmacol.*
563 *Toxicol.*, 2008, **48**, 653-683.
- 564 22. A. M. Myint, B. Bondy, T. C. Baghai, D. Eser, C. Nothdurfter, C. Schüle, P. Zill, N.
565 Müller, R. Rupprecht and M. J. Schwarz, *Brain, behavior, and immunity*, 2013, **31**, 128-133.
- 566 23. G. Anderson, M. Kubera, W. Duda, W. Lasoń, M. Berk and M. Maes,
567 *Pharmacological Reports*, 2013, **65**, 1647-1654.
- 568 24. M. Maes, K. Ringel, M. Kubera, M. Berk and J. Rybakowski, *Journal of affective*
569 *disorders*, 2012, **136**, 386-392.
- 570 25. M. Maes, B. Leonard, A. Myint, M. Kubera and R. Verkerk, *Progress in*
571 *neuro-psychopharmacology and biological psychiatry*, 2011, **35**, 702-721.
- 572 26. C. L. Raison, R. Dantzer, K. W. Kelley, M. A. Lawson, B. J. Woolwine, G. Vogt, J. R.
573 Spivey, K. Saito and A. H. Miller, *Molecular psychiatry*, 2010, **15**, 393-403.
- 574 27. A.-M. Myint, Y. K. Kim, R. Verkerk, S. Scharpé, H. Steinbusch and B. Leonard,
575 *Journal of affective disorders*, 2007, **98**, 143-151.
- 576 28. T. W. Stone, *Trends in pharmacological sciences*, 2000, **21**, 149-154.
- 577 29. Z. Hartai, A. Juhász, Á. Rimanóczy, T. Janáky, T. Donkó, L. Dux, B. Penke, G. K. Tóth,
578 Z. Janka and J. Kálmán, *Neurochemistry international*, 2007, **50**, 308-313.
- 579 30. J. O'connor, M. Lawson, C. Andre, M. Moreau, J. Lestage, N. Castanon, K. Kelley
580 and R. Dantzer, *Molecular psychiatry*, 2009, **14**, 511-522.
- 581 31. F. G. Graeff, F. S. Guimarães, T. G. De Andrade and J. F. Deakin, *Pharmacology*
582 *Biochemistry and Behavior*, 1996, **54**, 129-141.
- 583 32. F. Artigas, L. Romero, C. de Montigny and P. Blier, *Trends in neurosciences*, 1996,
584 **19**, 378-383.
- 585 33. G. Sanacora, G. Treccani and M. Popoli, *Neuropharmacology*, 2012, **62**, 63-77.

- 586 34. K. Rodgers, R. Dunlop, P.A. Cox, United States patent application publication,
587 20130156846 A1, 2013,1-12.
- 588 35. D. T. Balu and J. T. Coyle, *Current opinion in pharmacology*, 2015, **20**, 109-115.
- 589 36. S. Esposito, A. Pristera, G. Maresca, S. Cavallaro, A. Felsani, F. Florenzano, L.
590 Manni, M. T. Ciotti, L. Pollegioni and T. Borsello, *Aging cell*, 2012, **11**, 588-598.
- 591 37. D.-M. Otte, M. L. B. de Arellano, A. Bilkei-Gorzo, Ö. Albayram, S. Imbeault, H.
592 Jeung, J. Alferink and A. Zimmer, *PloS one*, 2013, **8**, e67131.
- 593 38. H. Mitani, Y. Shirayama, T. Yamada, K. Maeda, C. R. Ashby and R. Kawahara,
594 *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, 2006, **30**, 1155-1158.
- 595 39. G. L. Collingridge, S. Peineau, J. G. Howland and Y. T. Wang, *Nature Reviews*
596 *Neuroscience*, 2010, **11**, 459-473.
- 597 40. C. Altamura, M. Maes, J. Dai and H. Meltzer, *European*
598 *Neuropsychopharmacology*, 1995, **5**, 71-75.
- 599 41. M. Leyton, S. N. Young, R. Pihl, S. Etezadi, C. Lauze, P. Blier, G. Baker and C.
600 Benkelfat, *Neuropsychopharmacology*, 2000, **22**, 52-63.
- 601 42. J. P. Roiser, A. McLean, A. D. Ogilvie, A. D. Blackwell, D. J. Bamber, I. Goodyer, P. B.
602 Jones and B. J. Sahakian, *Neuropsychopharmacology*, 2005, **30**, 775-785.
- 603 43. H. Möhler, *Neuropharmacology*, 2012, **62**, 42-53.
- 604 44. A. Alcaro, J. Panksepp, J. Witzak, D. J. Hayes and G. Northoff, *Neuroscience &*
605 *Biobehavioral Reviews*, 2010, **34**, 592-605.
- 606 45. J.-P. Guilloux, G. Douillard-Guilloux, R. Kota, X. Wang, A. Gardier, K. Martinowich,
607 G. C. Tseng, D. A. Lewis and E. Sibille, *Molecular psychiatry*, 2012, **17**, 1130-1142.
- 608 46. E. Kozora, M. Brown, C. Filley, L. Zhang, D. Miller, S. West, J. Pelzman and D.
609 Arciniegas, *Lupus*, 2011, **20**, 598-606.
- 610 47. J. Li, G. Tang, K. Cheng, D. Yang, G. Chen, Z. Liu, R. Zhang, J. Zhou, L. Fang and Z.
611 Fang, *Molecular BioSystems*, 2014, **10**, 2994-3001.
- 612 48. Y. Ni, M. Su, J. Lin, X. Wang, Y. Qiu, A. Zhao, T. Chen and W. Jia, *FEBS letters*, 2008,
613 **582**, 2627-2636.
- 614 49. G. Chen, D. Yang, Y. Yang, J. Li, K. Cheng, G. Tang, R. Zhang, J. Zhou, W. Li and Z. Liu,
615 *Behavioural brain research*, 2015, **278**, 286-292.
- 616 50. F. Lamers, N. Vogelzangs, K. Merikangas, P. De Jonge, A. Beekman and B. Penninx,
617 *Molecular psychiatry*, 2013, **18**, 692-699.
- 618 51. B. Shi, J. Tian, H. Xiang, X. Guo, L. Zhang, G. Du and X. Qin, *Behavioural brain*
619 *research*, 2013, **241**, 86-91.
- 620 52. E. Banoglu, G. G. Jha and R. S. King, *European journal of drug metabolism and*
621 *pharmacokinetics*, 2001, **26**, 235-240.
- 622 53. A. Phipps, J. Stewart, B. Wright and I. WILSON*, *Xenobiotica*, 1998, **28**, 527-537.
- 623 54. P. Riederer, K. Toifl and P. Kruzik, *Clinica Chimica Acta*, 1982, **123**, 27-32.
- 624 55. S. M. O'Mahony, J. R. Marchesi, P. Scully, C. Codling, A.-M. Ceolho, E. M. Quigley, J.
625 F. Cryan and T. G. Dinan, *Biological psychiatry*, 2009, **65**, 263-267.
- 626 56. J. F. Cryan and S. O'Mahony, *Neurogastroenterology & Motility*, 2011, **23**,
627 187-192.
- 628 57. S. O. Fetissov and P. Déchelotte, *Current Opinion in Clinical Nutrition & Metabolic*
629 *Care*, 2011, **14**, 477-482.

630 58. L. Desbonnet, L. Garrett, G. Clarke, J. Bienenstock and T. G. Dinan, *Journal of*
631 *psychiatric research*, 2008, **43**, 164-174.

632 59. J. M. Yano, K. Yu, G. P. Donaldson, G. G. Shastri, P. Ann, L. Ma, C. R. Nagler, R. F.
633 Ismagilov, S. K. Mazmanian and E. Y. Hsiao, *Cell*, 2015, **161**, 264-276.

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

651

652 **Figure Legends**

653 **Fig.1** Experimental design for the present study.

654

655 **Fig.2** The sucrose preference at week3 (a), week6 (b) of the control, model, and treated group. Data are

656 represented as mean±SD.

657 * means a statistically significant difference at $p<0.05$, ** means a statistically significant difference $p<0.01$

658

659 **Fig.3** Measurement of locomotor activity in the open-field test. The time spent in the in the central area at week3
 660 (a), week6 (b), the frequency of rearing at week3 (c), week6 (d).

661 * means a statistically significant difference at $p < 0.05$, ** means a statistically significant difference $p < 0.01$

662

663 **Fig.4** The immobility time in the forced swimming test at week3 (a), week6 (b).

664 * means a statistically significant difference at $p < 0.05$, ** means a statistically significant difference $p < 0.01$

665

666 **Fig.5** (a) The score plot from a PCA model distinguishing the control, model and antidepressant-treated group
 667 (green circle, blue square, brown triangle) at week6. (b) PLS-DA score plots for pair-wise comparisons among the
 668 control, model and antidepressant-treated group (green circle, blue square, brown triangle) at week6.

669 (c)Correlation coefficient between original and permuted data.

670

671 **Fig.6** The perturbed metabolic pathways in response to CUMS and treatment of citalopram. The levels of potential
 672 biomarkers in the model group compared to the control group were labeled with (↑) down-regulated and (↓)
 673 up-regulated.

674

675

676

677

678

679

680

Table 1. Stressor weekly schedule

Time	Stressor	Lasting period
Monday	Cage cleaning, changing the soiled padding and weighing the rats	8:00
	Food and water deprivation	13:00
Tuesday	Sucrose preference test	9:00-11:00
	Paired housing	15:00-8:00
	Stroboscopic illumination	20:00-23:00
Wednesday	Remain single housing	8:00
	Soiled cage	15:00-8:00
	White noise	20:00-23:00

	Cage cleaning and changing	8:00
Thursday	the solid padding	
	Water deprivation	8:00-20:00
	Cage tilting	20:00-8:00
	Food deprivation	8:00-20:00
Friday	Stroboscopic illumination	20:00-23:00
	Cage tilting	23:00-11:00
Saturday	Paired housing	8:00-20:00
	Overnight illumination	20:00-8:00
Sunday	Soiled cage	15:00-8:00
	White noise	20:00-23:00

681

682

683

684

685

686

687

688

689

690

691

692

693

Table.2 List of candidate biomarkers for depression and evaluating the effect of citalopram

No	Metabolite	m/z (amu)	t _R (min)	VIP score	Model vs. Control		Model vs. Treat		Corresponding metabolic pathway
					fold change	q-value	fold change	q-value	
1	Glycerol ^{a,b}	92.0470594	1.34	2.84	0.51	0.008	0.66	0.021	Glycerolipid metabolism
2	4,6-Dihydroxyquinoline ^c	161.0472132	6.10	2.58	0.49	0.002	0.85	N.S.	Tryptophan metabolism
3	Succinic acid ^{a,b}	118.0262727	0.66	2.47	0.54	0.001	0.80	N.S.	Energy metabolism
4	Creatine ^{a,b}	131.0696566	5.49	2.30	0.24	0.002	0.44	0.016	Energy metabolism
5	Quinolinic acid ^{a,b}	167.0217195	0.99	2.20	2.34	<0.001	1.64	0.019	Tryptophan metabolism
6	L-Phenylalanine ^{a,b}	165.0797272	6.49	2.17	3.12	<0.001	1.34	0.031	Synthesis of neurotransmitter
7	Indole-3-carboxylic acid ^c	161.0470437	5.47	2.08	0.24	0.003	1.14	N.S.	Energy metabolism
8	Kynurenic acid ^a	189.0404926	4.87	2.03	0.26	0.001	0.43	0.006	Tryptophan metabolism
9	Indoxyl sulfate ^c	213.0090460	2.72	1.93	0.48	0.003	0.87	N.S.	Tryptophan metabolism
10	Gamma-Aminobutyric acid ^{a,b}	103.0635047	7.30	1.91	0.19	0.009	0.49	0.024	Synthesis of neurotransmitter
11	L-Kynurenine ^a	208.0842484	5.72	1.89	3.09	<0.001	0.43	0.015	Tryptophan metabolism
12	Dopamine ^{a,b}	153.0785807	5.25	1.85	0.26	0.004	0.72	N.S.	Synthesis of neurotransmitter
13	L-Tryptophan ^{a,b}	204.0902216	4.89	1.84	0.58	0.023	0.83	N.S.	Tryptophan metabolism
14	L-Isoleucine ^{a,b}	131.0952197	1.24	1.81	0.59	0.009	0.52	0.022	Isoleucine degradation
15	L-Serine ^{a,b}	105.0428964	1.11	1.80	0.49	0.002	0.86	N.S.	Glycine and serine metabolism
16	Hypoxanthine ^{a,b}	136.0380725	1.34	1.79	0.50	0.004	0.88	N.S.	Energy metabolism
17	5-hydroxyindoleacetic acid ^a	191.0586587	3.72	1.79	0.42	0.009	0.69	N.S.	Tryptophan metabolism
18	N-Acetyl-L-aspartic acid ^a	175.0476473	4.36	1.77	0.59	0.011	0.75	0.018	Synthesis of neurotransmitter
19	Pyruvic acid ^{a,b}	88.0158195	0.70	1.75	1.94	0.028	1.08	N.S.	Energy metabolism
20	Phenylpyruvic acid ^c	164.0469798	0.74	1.72	1.24	0.006	1.16	N.S.	Synthesis of neurotransmitter
21	Xanthurenic acid ^c	205.0369086	5.62	1.67	1.33	0.012	0.96	N.S.	Tryptophan metabolism
22	Phenylacetylglutamine ^a	193.0735057	5.53	1.66	1.39	0.008	1.23	N.S.	Glycine metabolism

23	Indoleacetic acid ^{a,b}	175.0629743	6.74	1.64	0.46	0.006	0.65	0.013	Tryptophan metabolism
24	Citric acid ^{a,b}	192.0264797	6.55	1.59	0.72	0.027	0.89	N.S.	Energy metabolism
25	L-Dopa ^{a,b}	197.0683015	5.35	1.54	0.60	0.007	0.90	N.S.	Synthesis of neurotransmitter
26	Glycine ^{a,b}	75.0323353	3.22	1.53	0.43	0.017	0.73	N.S.	Glycine and serine metabolism

694 a Metabolites identified by comparing with authentic standards.

695 b Metabolites further validated by MS/MS spectra with authentic standards.

696 c Metabolites identified by comparing with the HMDB database.

697

698

699

700

701

702

703

704

705

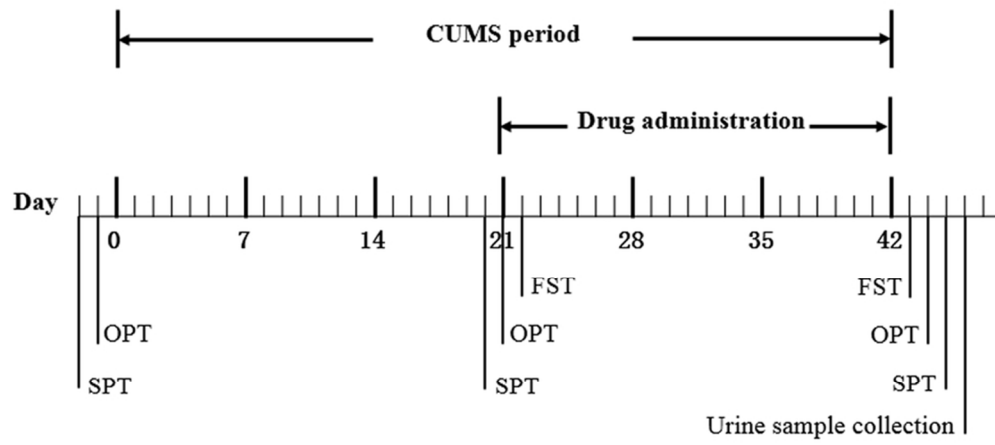


Fig.1 Experimental design for the present study.
82x42mm (300 x 300 DPI)

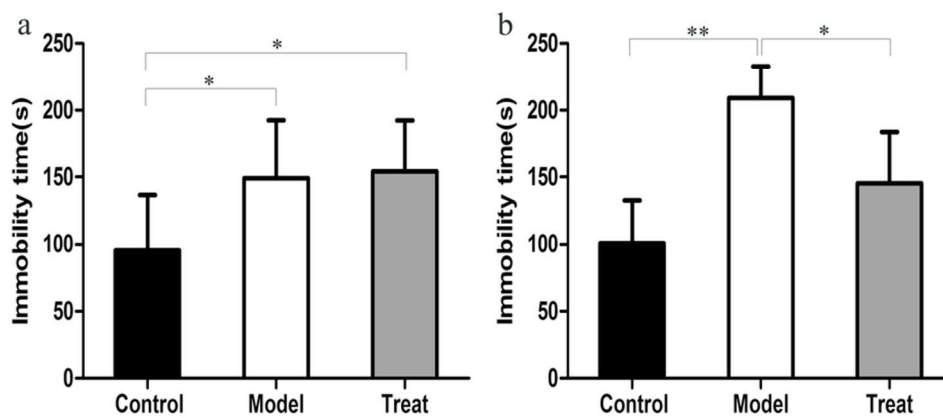


Fig.4 The immobility time in the forced swimming test at week3 (a), week6 (b).
* means a statistically significant difference at $p < 0.05$, ** means a statistically significant difference $p < 0.01$

82x36mm (300 x 300 DPI)

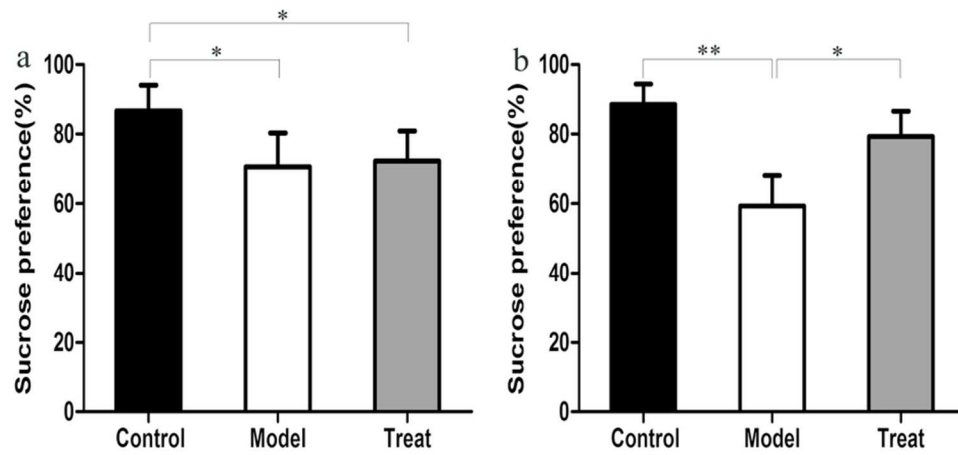


Fig.2 The sucrose preference at week3 (a), week6 (b) of the control, model, and treated group. Data are represented as mean \pm SD.

* means a statistically significant difference at $p < 0.05$, ** means a statistically significant difference $p < 0.01$

82x38mm (300 x 300 DPI)

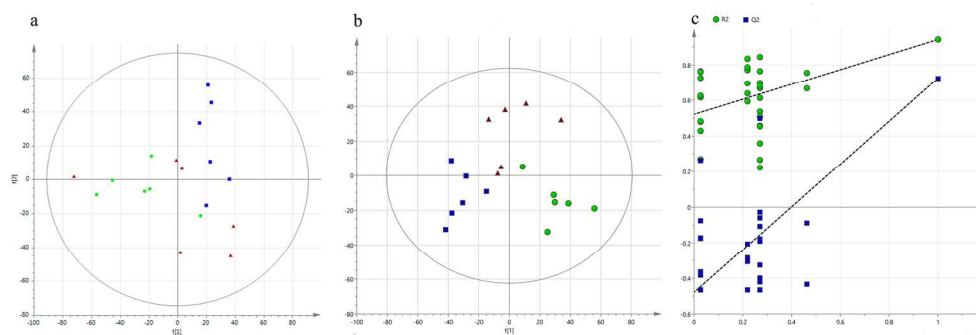


Fig.5 (a) The score plot from a PCA model distinguishing the control, model and antidepressant-treated group (green circle, blue square, brown triangle) at week6. (b) PLS-DA score plots for pair-wise comparisons among the control, model and antidepressant-treated group (green circle, blue square, brown triangle) at week6. (c)Correlation coefficient between original and permuted data.
82x28mm (600 x 600 DPI)

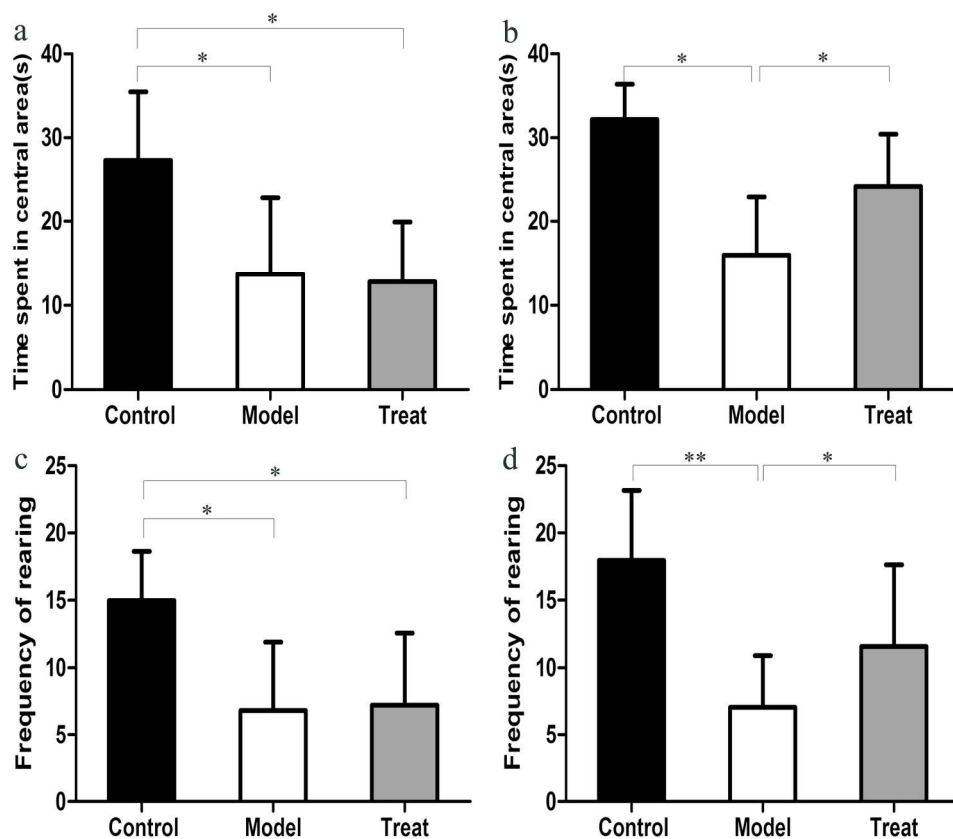


Fig.3 Measurement of locomotor activity in the open-field test. The time spent in the in the central area at week3 (a), week6 (b), the frequency of rearing at week3 (c), week6 (d). * means a statistically significant difference at $p < 0.05$, ** means a statistically significant difference $p < 0.01$

170x157mm (300 x 300 DPI)

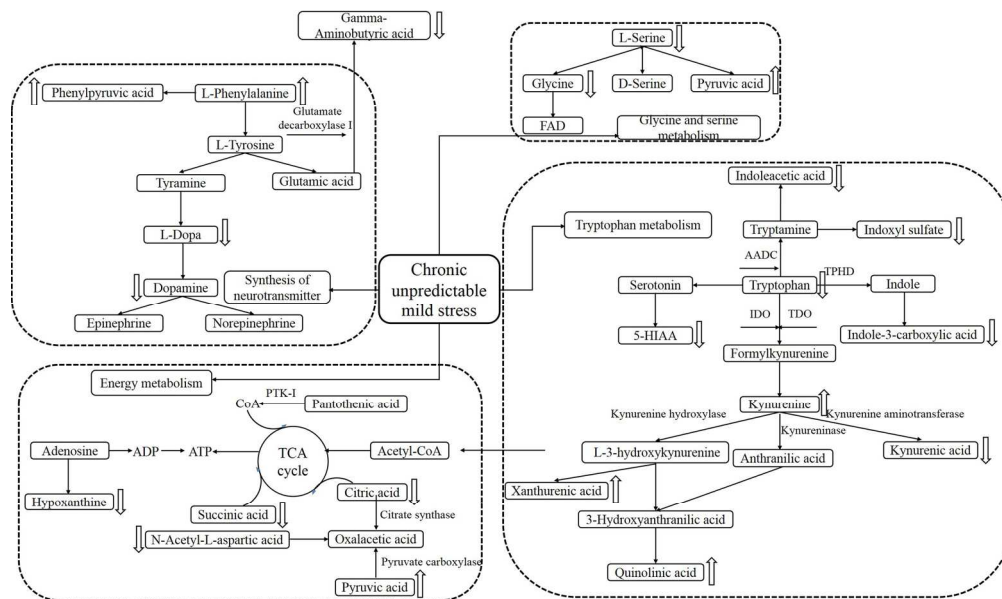


Fig.6 The perturbed metabolic pathways in response to CUMS and treatment of citalopram. The levels of potential biomarkers in the model group compared to the control group were labeled with (↑) down-regulated and (↓) up-regulated.
172x103mm (300 x 300 DPI)