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Journal:	RSC Advances
Manuscript ID:	RA-REV-04-2015-007927.R1
Article Type:	Review Article
Date Submitted by the Author:	02-Jun-2015
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Review article

Reactive oxygen species: friend or foe?

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Abstract

Reactive oxygen species (ROS) are an inevitable by-product of cellular metabolism. ROS generation associated with interaction of ionizing radiation with biological molecule, and devoted enzymes in phagocytic cells (NADPH oxidase and myeloperoxidase) or may be the result of imbalanced radical generating and scavenging systems. Albeit the myth about ROS as Pandora's Box, it has several innovative physiological roles in body. ROS serve as signalling messengers for activation of transcription factors from cytokine-receptor interactions. It facilitates the evolution and membrane fusion of spermatozoa and oocyte during fertilization. NADPH oxidase enzyme and nitric oxide (NO) are function as potent vasodilators and immunity booster. ROS has been suggested as a prevalent regulator of several nuclear factors including erythroid 2-related factor 2 (Nrf2), nuclear factor kappa-B cells (NFkB), mitogen-activated protein kinase (MAPK) and p53 which further associated with plenty of signalling cascades. Under physiological conditions the amount of ROS generated in body can be counterbalance by natural anti-oxidant of body. However, the aberrant augmented level of ROS predominantly leads to various defined disorders comprising myocardial infarction, autoimmune diseases, atherosclerosis, alzheimer, parkinson and emphysema diseases. Ordinarily, it has been observed that physiological roles of ROS are knocked down in front of their pathological action. But here a need of glimpse about the explicit line of margin between patho-physiological function of ROS. Worthy of this note is to reveal the beneficial responsibility of ROS in different cellular pathways and metabolic functions, over its injurious consequence.

Keywords: Mitochondria, ROS, RNS, Physiology, Pathology, Signalling mechanism.

Abbreviations: ROS, Reactive oxygen species; NO, Nitric oxide; Nrf2, Nuclear factor erythroid 2-related factor 2; NF κ B, Nuclear factor kappa-B cells; MAPK, Mitogen-activated protein kinase; TNF α , Tumor necrosis factor- α ; PTP, Permeability transition pore; NADPH-oxidase, Nicotinamide adenine dinucleotide phosphate-oxidase; eNOS, Endothelial nitric oxide synthase; BH4, Tetrahydrobiopterin; RNS, Reactive nitrogen species; JNK, c-Jun N-terminal kinases; PKB/Akt, Protein kinase B; mTOR, Mammalian target of rapamycin; GPCR, G protein-coupled receptors; TLR, Toll-like receptors; ICAM-1, Intercellular adhesion molecule-1; cAMP, Cyclic adenosine monophosphate; Keap1, Kelch-like ECH-associated protein 1.

Introduction

Mitochondria are the primary site of ROS generation. During aerobic metabolism electron transport chain reaction leads to instantaneous production of ROS as an unavoidable by-product [1]. Mitochondrial ROS generation is composed of four multi protein complexes (complex I-IV). Complex I and III are widely identified as the core site of ROS generation [2]. Endogenous production of ROS may initiate numerous signalling cascades for maintaining homeostasis in physicochemical or pathophysiological states [3]. Recent extensive evidences suggested that conventionally mitochondrial ROS plays a pivotal role in managing several transcription factors and receptor interactions [4-5]. Despite of physiological role several evidences show debilitating proposition about ROS at higher level. Over production of ROS due to damaged mitochondria is considered as major predictor for the development of acute and chronic disorders. Increased level of ROS involved in the activation of wide range of gene expression and associated pathogenesis [6]. As the brain consumed 20% oxygen of the total body's oxygen consumption, the vulnerability of oxidative damage is most prominent in this area. The accumulation of cellular ROS destroys the normal redox state in neural cells and leads to neurodegenerative diseases [7]. Ma et al. (2012) concluded the overproduction of these ambiguity molecules by mitochondria of β -cell activate various insidious metabolic disorder (diabetes mellitus followed by β -cell failure) [8]. Moreover, evidences suggested that high glucose concentration increased oxidative stress and further contribute to diabetes and associated complications [9-10]. In addition, ROS interfere with nuclear factor kappa-B (NF-κB) and tumor necrosis factor- α (TNF α) pathway and may leads to rheumatoid arthritis [11]. Indeed, free radicals react with all biological macromolecules (lipids, proteins, nucleic acids and carbohydrates) which contribute to oxidative stress and activate

various apoptotic pathways. Further, intracellular ROS also affect the mitochondria by activation of caspases with the rapid release of cytochrome c which ultimately leads to apoptosis [12]. TNF α can induce ROS by interacting with TNF receptor (TNF-RI) and leads to apoptosis/ necrosis. ROS induce apoptosis may affect 'Guardian of the genome' p53 which cause immense alteration in different signalling [13-14]. However, mitochondrial low oxidative stress or mitochondrial hormesis/mitohormesis prompts an adaptive reaction results including lower glucose metabolism, improve stress resistance, and influence mitochondrial signalling and metabolism [5]. Inaddition, Perez-Matute et al. (2009) also supports that ROS act as a loval second messenger at low level and regulates vital cellular functions [4]. NO is an imperative mediator of endothelial function and cardiovascular physiology because of its vasodilatory, anti-platelet, anti-proliferative, and anti-inflammatory potential. Increased bioavailability of NO in the blood vessels may improve the inflammation, atherosclerosis, hypertension, vascular endothelial dysfunction and associated cardiovascular diseases [15]. Moreover, recent evidence has shown that oxidative stress generated by reactive oxygen and nitrogen species monitor the array of physiological signalling in cardiovascular system [16]. The contentious facts on patho-physiological action of ROS can differentiate only upon its level in cells. Despite of the consciousness about these two parts of the coin, an overt line of boundary between physiological and pathological level of ROS is still unclear. Thus, there is a need of revisiting to understand the relation of oxidative stress level to precise health effect. The present review delineates positive liability of ROS in several cellular and metabolic pathways, more than its deleterious effects and put a requisite of measurable level of ROS, which may explain that how the physiological state get convert into pathological state.

Generation of reactive oxygen species

To begin with Richard Altmann, an elementary organism has been illustrated within the cells which conclusively recognized as 'bioblast' or powerhouse of the cell or mitochondria [17]. Primarily mitochondria was regarded as only an energy generator, but in last few decades' number of evidences revealed its extensive role in generating ROS and -associated fascinating mechanisms [18]. Mitochondria are the foremost site for *in vivo* production of free radical in continuous chain reaction mode, which extensively involved in various patho-physiological pathways. Superoxide is generated on the innermost membrane of mitochondria (matrix) [19]. This superoxide consecutively monitors mitochondrial functions including mitochondrial biogenesis, mitochondrial membrane permeability transition. lipid peroxidation. and mitochondrial DNA damage [20-21]. Mitochondrial ROS generation is organized by four multi-protein complexes (complex I-IV) [2]. Mitochondria comprise almost eight sites from where superoxide may generate [22]. Interestingly, out of them seven sites liberates generates superoxide within the matrix, but eighth (site IIIQo) on complex III and glycerol 3-phosphate dehydrogenase have an ability to discharge produced superoxide in inter membrane space. ROS moving in inter membrane space can easily cross the mitochondrial membrane in contrast to ROS present in the matrix [23]. Therefore, the ROS generated by site IIIQo on complex III and glycerol 3-phosphate dehydrogenase has more impact to participate in cellular signalling. In-addition, complex I (NADH CoQ reductase) has ability to stimulate the transfer of electron from NADH to co-enzyme Q which is responsible for production of significant amount of ROS. The translocation of proton from matrix to the inter membrane space revealed the potential of complex I to generate and release ROS as complex III. Moreover, complex II enzyme (succinate dehydrogenase) may reduce CoQ and

showing its involvement in production of superoxide [24-25]. Overall studies indicate that complex I, II and III have huge potential to generate ROS, but major contributors are complex I and complex III [2]. These contributions of producing ROS by each enzyme complex are variations in different tissue and pathologic conditions [26] [Figure 1].

Nicotinamide adenine dinucleotide phosphate-oxidase (NADPH-oxidase) is a multisubunit (membrane associated components, i.e., p22 phagocytes oxides, gp91phox and cytosolic component enzyme, i.e., p47phox, p40phox, p67phox, small GTPase Rac) enzyme which plays an integral role to catalyze the reduction of molecular oxygen to form superoxide. NADPH oxidase activation through translocation and accumulation of cytosolic proteins with the membrane turn out electron transference, which further combines with molecular oxygen and generate superoxide anion [27-28].

Endoplasmic reticulum (ER) is another major site of ROS production. It contains several proteins and enzymes including endoplasmic reticulum oxidoreductase, reduced glutathione and protein disulfide isomerase which participate in ER stressinduced ROS production. Protein folding is précised system which initiates the translaction of accurate folded protein to intended site. Any adversity in this system process leads to misfolding or immature protein aggregate and ER stress -associated ROS. Protein folding is carried out by the formation of disulfide bond which produce a specific oxidising environment. Alteration in the disulfide bond formation results to protein misfolding which leads to generate significant amount of ROS due to depletion of glutathione (GSH). Repetitive cycle of breakage and formation of disulfide bond degrade the level of GSH. Each cycle produce more and more ROS as

by-product. Moreover, accumulated unfolded proteins cause Ca⁺² leakages in cytosol which further responsible for mitochondrial ROS production [29-30].

Redox signalling plays a vital role in physiological and patho-physiological regulation. Cellular redox reactions maintain the equilibrium between oxidized and reduced state of cysteines and methionines. Extracellular fluid infused protein may oxidized by reacting with thiol group present in cystein residue. This thiol reaction with protein can modify critical cellular signalling mechanism. Disufide bonding between thiol group of cystein and extracellular protein is the major source of ROS and radition induced cellular death. Regulation of protein redox state through thiol/disulfide bonding may maintain physiologic functions [29-31].

Endothelial nitric oxide synthase (eNOS), is an exceptional enzyme that has the ability to generate both free radicals (Peroxynitrite) and free radical scavenging (NO) agent. Coupling of eNOS with cofactor tetrahydrobiopterin (BH4) produces NO during the conversion of L-arginine to L-citrulline in the presence of NADPH-dependent enzyme. NO, is a key regulator of vascular and cardiac functions having anti-inflammatory, anti-platelet, anti-proliferative and anti-migratory properties [32]. However, uncoupled eNOS produces reactive nitrogen species (RNS). Superoxide reacts with NO and leads to versatile nitrogen species; Peroxynitrite may further convert in peroxynitrous acid by protonation and may yield nitrogen dioxide and a hydroxyl radical [33-34]. Peroxynitrite and peroxynitrous acid can cause lipid peroxidation and membrane damage in vessel wall [35].

Pathological role of ROS

Healthy mitochondria persistently generate low levels of superoxide during conventional respiration. On the other hand Injured and unsynchronized mitochondria

generate large amount of superoxide which further cause detriment to mitochondrial components including lipids, proteins, DNA and eventually cell death [1, 36]. Numerous studies have demonstrated the pathological role of ROS in different metabolic syndrome. Indeed, ROS regulate autophagy process by directly modifying major proteins including Atg4, Atg5 and Beclin. Higher level of ROS indirectly alters the vital signalling pathway including c-Jun N-terminal kinases (JNK), p38 which leads to autophagy. Intriguingly, ROS mediated autophagy process allied with numerous diseases, comprising neurodegenerative diseases, diabetes mellitus, myodegenerative diseases, crohn's disease, and different type of cancer. Elevated ROS also obstruct the key signalling of protein kinase B (PKB/Akt) and down regulate mammalian target of rapamycin (mTOR) [37-39]. Clinical evidences support the subclinical based concept of DNA damage associated activation of mutagenic and apparently carcinogenic factor [40]. Additionally, high oxidative stress has acknowledged as key mediator in the occurrence and development of diabetes mellitus and -associated complications. Furthermore, diabetes related oxidative alteration would affect cellular metabolism, functions, and gene expression [41-42]. Oxidative stress is the major pitfall of vascular endothelial dysfunction and associated cardiovascular pathology. eNOS-generated NO is an imperative endothelial factor which can obstruct pathological event for cardiovascular disorders, including platelet aggregation, inflammation, vascular proliferation, hypertension, and leukocyte adhesion [43-44]. Excessive extent of NADPH-oxidase derived superoxide reacts with NO and converts this into versatile nitrogen species "Peroxynitrite". Peroxynitrite further reduced the bioavailability of NO by initiating eNOS uncoupling by oxidizing BH4. Thus, a deficiency in production or bioavailability of NO leads to endothelial atherosclerotic events. vascular dysfunction and -associated

cardiovascular diseases [45-46]. Moreover, in vitro evidence revealed that ample amount of ROS triggered apoptosis in renal cells through activation of caspase-3 leads to nephrotoxicity [47-48]. About three decades before Baltimore invent one more ROS mediated cellular signalling (NF κ B). It has been found NF κ B signalling leads to atherosclerosis, schizophrenia and cancer generation [49-51]. NF κ B has five distinct transcription factors (p65, p50, p52, c-Rel and RelB) which react with DNA through homology domain and may stimulate transcription of various genes. Furthermore, seven individual proteins of IkB family (inhibitory proteins) can modulate NFkB signalling cascade [52-54]. Schreck and his colleagues primarily revealed the important role of oxidative stress in stimulating NFkB and -associated transcription factors [55]. Oxidative stress mediated NF κ B signalling activation may lead to cell inflammation, proliferation, and apoptosis [56-57]. ROS generated oxidative stress initiate glomerular, tubular injuries and may initiate NF- κ B. NF- κ B further alters the transcription of the intercellular adhesion molecule-1 and tumor necrosis factor (TNF- α) which cause renal injury [47-48, 58] [Figure 2]. ER stress associated production involved in wide range of pathologic condition such as cancer, neurodegenerative diseases, cardiac diseases, diabetes mellitus and muscle degeneration diseases [29-30]. Thus, mentioned all above evidences suggest that ROS generated oxidative stress may cause serious pathological condition. However, pathological role of ROS is only associated with its elevated level.

Physiological role of ROS

ROS including superoxide anions (O^{2-}) and hydrogen peroxide (H_2O_2), are highly reactive molecules which may leads to organ pathogenesis. However, it is recognized that low level of ROS act as vital signalling molecules for modulating the normal cellular processes [59]. ROS are salient mediators and signal modifiers during

numerous biological processes and cascades such as G protein-coupled receptors (GPCR), Notch [60], Wnt- $\beta\beta$ -catenin [61], MAPK, JAK-STAT, NF- κ B, and PI3K/AKT [62].

Physiological role of ROS associated with myocardium

Recent evidence suggests that ROS could serve as paracrine signalling mediators upon pathological stimulation [63]. Paracrine signalling in the epicardium and endocardium is associated with fibroblast growth factor and retinoic acid dependent signalling. These vital signalling pathways carry the proper growth and differentiation of the myocardium [64-65].

Physiological role of ROS associated with insulin sensitivity

ROS promotes tyrosine phosphorylation-dependent signaling (protein tyrosine phosphatises, PTP) which leads to phosphorylation of PTP1B and dephosphorylation of tyrosyl phosphorylated substrates and PTEN enzyme. PTP is regulated by cysteines which inactivate its nucleophilic properties and increase insulin sensitivity and glucose homeostasis. ROS also improve the insulin sensitivity by terminating PI3K signalling via dephosphorylation of phosphatidylinositol-3,4,5-triphosphate (PIP3) [66]. ROS play an important role in the regulation of insulin receptor kinase activity by auto phosphorylation of the insulin receptor kinase at Tyr-1158, Tyr-1162, and Tyr-1163 sites [67]. In general, insulin controls several physiological functions such as glucose metabolism, lipid metabolism and synthesis of protein. Loh et al. (2009) also revealed supportive evidence on the basis of *in vivo* study conducted by him suggesting the enhancement of insulin signalling by ROS [66].

Physiological role of ROS associated with autophagy

ROS regulate the process of autophagy which in turn implicated with various physiological processes and maintain the cell homeostasis. Autophagy is a process of engulfing or breaking of intracellular proteins and organelles of cells by lysosome and further repurposes the constituents for new biosynthesis. In-addition autophagy plays a central role in maintaining the homeostasis during starvation and prevention of pathogenic infection [37, 68]. Starvation activates PI3K signalling followed by mitochondrial ROS generation which inactivate the cysteine protease Atg4 and promote autophagy.

Physiological role of ROS associated with apoptosis

Mitochondrial ROS significantly leads to apoptosis which have physiologic functions during embryogenesis. Interestingly, mitochondrial cytochrome-C which leads to caspase signalling activation intensely arbitrated by direct or indirect ROS action and that ultimately causes cell death. In-spite of that, ROS have their own anti-apoptotic actions also [69]. Apoptosis has an imperative biological function in the improvement and homeostasis of cell masses [70]. Apoptosis also involved in certain physiologic action including involution of breast [71], endometrium detachment during menstruation [72], removal of T cells during thymus development [73], castration-induced prostate atrophy [74] and epidermal cell death [75]. Recent study revealed apoptosis-dependent cavitation is an essential step for the formation of embryogenesis and growth of mammary gland, lung, and kidney by epithelial lumen creation route [76].

Physiological role of ROS associated with immunity

The physiological role of ROS also includes their potential against pathogens in favour to immune response. Superoxide generation may kill the growth of bacteria

and resist the host from infection. Additionally, evidence supports that ROS derived NADPH oxidase also plays a crucial role in host protection [77-79]. ROS initiate the immunity of cells by causing oxidative burst via NADPH oxidase to kill the pathogens in cells. Uncoupling proteins 1–3 (UCPs1-3; mitochondrial anion carrier proteins) are supposed to play an important role in minimizing ROS discharge from the electron transport chain [80]. However, UCP2 in macrophages increase mitochondrial ROS for MAPK activation and oxidative burst agumentation in favour of pathogen elimination [81-82]. More recent studies have demonstrated that mitochondrial ROS are significantly initiated by Toll-like receptors (TLR) and associated pathways. The stimulation of cell-surface TLRs (TLR1, TLR2, and TLR4), but not endosomal TLRs (TLR3, TLR7, TLR8, and TLR9) leads to an increase in mitochondrial ROS production through ECSIT (evolutionarily conserved signalling intermediate in Toll pathways) and TRAF6 (tumour necrosis factor receptorassociated factor 6 signalling). Collectively, the study concluded that, TLRs which are prominently obtained from bacteria can produce immunity against bacterial infection by inducing ROS [83].

Physiological role of ROS associated with capacitation

ROS have another crucial liability in early events of capacitation. The capacitation is a term used to define the complex and not well-characterized process that allows spermatozoa to complete their preparations to fertilize oocytes. Some of the early events of capacitation include a calcium influx, rise in pH [84], increase in intracellular cyclic adenosine monophosphate (cAMP) [85] and the generation of low and organize level of superoxide anion [86]. Another important aspect of capacitation, reported more than ten years before is that the proteins from the fibrous sheath are subjected to tyrosine phosphorylation (P-Tyr) [87]. Moreover, post translational

modifications of these proteins become very important for sperm function, because spermatozoa are unable to synthesize proteins. Studies revealed that this potential role of protein P-Tyr is dependent on ROS [88-89]. Ford, (2004) publish a report which revealed the increment of capacitation by the addition of ROS and vice versa. This study also support ROS-induced cAMP and tyrosine kinase activation through protein tyrosine phosphorylation as signalling cascade for regulation of sperm function [90]. Conversely, excessive ROS generation may devastate its protective action in capacitation. The posivtive and negative influence of ROS generation on fertilizing ability but the threshold level of ROS-associated with sperm cell maturation is still unclear. Thus further revisit on this aspects is needed for the contemporary treatment of infertility [90-92].

Physiological role of ROS associated with signalling cascade

Nrf2 is another prominent regulator of cellular resistance to oxidants. Keap-1 (Kelchlike ECH-associated protein1) dependent and independent activation of Nrf2 plays a pivotal role in regulating anti-oxidant and anti-inflammatory defence mechanism through various physiological cellular signalling. It also revealed that low level of ROS can significantly initiate the activation of Nrf2 signalling [93-94].

Albeit, H_2O_2 is an inevitable by-product of cellular metabolism. The production of H_2O_2 is most commonly associated in neutrophils which play important defensive role in host. Moreover, some studies reveal that H_2O_2 act as novel signalling messenger at low level. Several prosurvival pathways regulated by H_2O_2 including kinase, oxidation of cysteines and transcriptional factors such as p53, NF-kappa B and AP-1 [95].

Thus, the diverse biological activities of oxygen radicals and related species are likely rooted in the oxygen-dependent evolution of complex life forms. These critical evidences collectively suggest the physiological role of ROS may offer a precious therapeutic option and that may be indispensable in cellular pathology.

Conclusion

For a long time, evidences pursuit the stochastic action of ROS but the devoted role of ROS has soared in both physiology and pathology. ROS is a double-edged sword, with protective and toxic capabilities. All pathological conditions instigate involvement of high level of ROS. However, low levels of ROS are not only suspicious against diseases, but also have the additional property to regulate physiological cellular signalling. Both these facts widely asserted and acknowledged, but the mystery regarding the level of ROS conversion from physiological action to pathological action is still unclear. Ample numbers of important cellular pathways, where ROS are involved. For instance, MAPK [96-97], 'Guardian of the genome' p53 [98-99], Hypoxia-inducible factor 1-α (HIF1-α) [100], SP-1 transcription factor [101-102], AP-1 transcription factor [101, 103], caspase regulation [104], cytokine [105], platelet derived growth factor [106-107] and fibroblast-derived growth factor [108], are well known remnant key signalling cascades which have been altered by ROS. Thus it is further important to clarify that what extant of ROS is protective and another one is aggressive. Concept becomes more worthy for the individuals getting anti-oxidant treatment in life-threatening diseases like cancer. Moreover, potent antioxidant may cause developmental deformity due to crucial reduction of ROS level as compared to its threshold level needed for therapeutic effect. In light of this view point we suggest there should be a precise border line between patho-phyisological actions with statistical level of ROS. This statistical approach may offer new

perspectives for better and in-time treatment of ROS -associated diseases with specific amount of anti-oxidant agents.

Conflict of interest

No conflict of interest.

Acknowledgment

We express our gratefulness to Dr. Pitchai Balakumar, Associate Professor and Head, Pharmacology Unit, Faculty of Pharmacy, Asian Institute of Medicine, Science and Technology (AIMST) University, Semeling, 08100 Bedong, Kedah Darul Aman, Malaysia for the expert suggestion, inspiration and review.

Reference

[1] G. Siciliano, C. Simoncini, A. Lo Gerfo, D. Orsucci, G. Ricci, M. Mancuso, Effects of aerobic training on exercise-related oxidative stress in mitochondrial myopathies, Neuromuscul Disord. 22 (2012) 172-177.

[2] Y. Liu, G. Fiskum, D. Schubert, Generation of reactive oxygen species by the mitochondrial electron transport chain, J Neurochem. 80 (2002) 780-787.

[3] T.P. Devasagayam, J.C. Tilak, K.K. Boloor, K.S. Sane, S.S. Ghaskadbi, R.D. Lele, Free radicals and antioxidants in human health: current status and future prospects, J Assoc Physicians India. 52 (2004) 794-804.

[4] P. Perez-Matute, M.A. Zulet, J.A. Martínez, Reactive species and diabetes:
 counteracting oxidative stress to improve health, Curr Opin Pharmacol. 9 (2009) 771 779.

[5] E. Barbieri, P. Sestili, L. Vallorani, M. Guescini, C. Calcabrini, A.M. Gioacchini, G. Annibalini, F. Lucertini, G. Piccoli, V. Stocchi, Mitohormesis in muscle cells: a morphological, molecular, and proteomic approach, Muscles Ligaments Tendons J. 3 (2014) 254-266.

[6] C. Kunsch, R.M. Medford, Oxidative stress as a regulator of gene expression in the vasculature. Circ Res.85 (1999) 753-766.

[7] W.H. Lee, C.Y. Loo, M. Bebawy, F. Luk, R.S. Mason, R. Rohanizadeh, Curcumin and its derivatives: their application in neuropharmacology and neuroscience in the 21st century, Curr Neuropharmacol. 11 (2013) 338-378.

[8] Z.A. Ma, Z. Zhao, J. Turk, Mitochondrial dysfunction and β-cell failure in type 2 diabetes mellitus, Exp Diabetes Res. 2012 (2012) 703538.

[9] W. Zhu, Q. Jia, Y. Wang, Y. Zhang, M. Xia, The anthocyanin cyanidin-3-O-βglucoside, a flavonoid, increases hepatic glutathione synthesis and protects hepatocytes against reactive oxygen species during hyperglycemia: Involvement of a cAMP-PKA-dependent signalling pathway, Free Radic Biol Med. 52 (2012) 314-327.

[10] T.V. Fiorentino, A. Prioletta, P. Zuo, F. Folli, Hyperglycemia-induced oxidative stress and its role in diabetes mellitus related cardiovascular diseases, Curr Pharm Des. 19 (2013) 5695-5703.

[11] T. Ishibashi, Molecular hydrogen: new antioxidant and anti-inflammatory therapy for rheumatoid arthritis and related diseases, Curr Pharm Des. 19 (2013) 6375-6381.

[12] S. Fulda, A.M. Gorman, O. Hori, A. Samali, Cellular stress responses: cell survival and cell death, Int J Cell Biol. 2010 (2010) 214074.

[13] X. Chen, B.T. Andresen, M. Hill, J. Zhang, F. Booth, C. Zhang, Role of Reactive Oxygen Species in Tumor Necrosis Factor-alpha Induced Endothelial Dysfunction, Curr Hypertens Rev. 4 (2008) 245-255.

[14] A.A. Sablina, A.V. Budanov, G.V. Ilyinskaya, L.S. Agapova, J.E. Kravchenko,P.M. Chumakov, The antioxidant function of the p53 tumor suppressor, Nat Med. 11(2005) 1306-1313.

[15] A.K. Sharma, D. Khanna, Diabetes mellitus associated cardiovascular signalling alteration: a need for the revisit, Cell Signal. 25 (2013) 1149-1155.

[16] J.C. Campos, K.M. Gomes, J.C. Ferreira, Impact of exercise training on redox signalling in cardiovascular diseases, Food Chem Toxicol. 62 (2013) 107-119.

[17] L. Ernster, G. Schatz, Mitochondria: a historical review, J Cell Biol. 91 (1981)227-255.

[18] M.L. Circu, T.Y. Aw, Reactive oxygen species, cellular redox systems, and apoptosis, Free Radic Biol Med. 48 (2010) 749-762.

[19] L.A. MacMillan-Crow, J.P. Crow, J.A. Thompson, Peroxynitrite-mediated inactivation of manganese superoxide dismutase involves nitration and oxidation of critical tyrosine residues, Biochemistry. 37 (1998) 1613–1622.

[20] C.A. Piantadosi, H.B. Suliman, Redox regulation of mitochondrial biogenesis,Free Radic, Biol. Med. 53 (2012) 2043–2053.

[21] M. Bourens, F. Fontanesi, I.C. Soto, J. Liu, A. Barrientos, Redox and reactive oxygen species regulation of mitochondrial cytochrome c oxidase biogenesis, Antioxid Redox Signal. 19 (2013) 1940-1952.

[22] M.D. Brand, The sites and topology of mitochondrial superoxide production,Exp. Gerontol. 45 (2010) 466–472.

[23] F.L. Muller, Y. Liu, H. Van Remmen, Complex III releases superoxide to both sides of the inner mitochondrial membrane, J. Biol. Chem. 279 (2004) 49064–49073.

[24] R. Fato, C. Bergamini, S. Leoni, P. Strocchi, G. Lenaz, Generation of reactive oxygen species by mitochondrial complex I: Implications in neurodegeneration, Neurochem. Res. 33 (2008) 2487–2501.

[25] M.P. Murphy, How mitochondria produce reactive oxygen species, Biochem. J.417 (2009) 1–13.

[26] Y. Kushnareva, A.N. Murphy, A. Andreyev, Complex I-mediated reactive oxygen species generation: Modulation by cytochrome c and NAD(P)+ oxidation-reduction state, Biochem. J. 368 (2002) 545–553.

[27] M. Taura, K. Miyano, R. Minakami, S. Kamakura, R. Takeya, H. Sumimoto, A region N-terminal to the tandem SH3 domain of p47phox plays a crucial role in activation of the phagocyte NADPH oxidase, Biochem. J. 419 (2009) 329–338.

[28] P. Balakumar, M.K. Arora, J. Reddy, M.B. Anand-Srivastava, Pathophysiology of diabetic nephropathy: involvement of multifaceted signalling mechanism, J. Cardiovasc. Pharmacol. 54 (2009) 129–138.

[29] S. Xu, S. Sankar, N. Neamati, Protein disulfide isomerase: a promising target for cancer therapy. Drug Discov Today. 19 (2014) 222-40.

[30] S. Xu, A.N. Butkevich, R. Yamada, Y. Zhou, B. Debnath, R. Duncan, E. Zandi, N.A. Petasis, N. Neamati, Discovery of an orally active small-molecule irreversible

inhibitor of protein disulfide isomerase for ovarian cancer treatment. Proc Natl Acad Sci U S A. 109 (2012) 16348-53.

[31] D.W. Bak, E. Weerapana, Cysteine-mediated redox signalling in the mitochondria. Mol Biosyst. 11 (2015) 678-97.

[32] P. Balakumar, S. Kathuria, G. Taneja, S. Kalra, N. Mahadevan, Is targeting eNOS a key mechanistic insight of cardiovascular defensive potentials of statins? J Mol Cell Cardiol. 52 (2012) 83-92.

[33] W.A. Pryor, G.L. Squadrito, The chemistry of peroxynitrite: a product from the reaction of nitric oxide with superoxide, Am J Physiol 268 (1995) 699-722.

[34] N. Kuzkaya, N. Weissmann, D.G. Harrison, S. Dikalov, Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: implications for uncoupling endothelial nitric-oxide synthase, J Biol Chem. 278 (2003) 22546-22554.

[35] V.B. O'Donnell, B.A. Freeman, Interactions between nitric oxide and lipid oxidation pathways: implications for vascular disease, Circ Res. 88 (2001) 12-21.

[36] A. K. Sharma, S. Haidarali, U. Nagaich, Apoptosis: A Potential target site for natural bioactive agents during myocardial infarction, J. Adv. Pharm. Edu. & Res. 4 (2014) 264-284.

[37] B. Levine, N. Mizushima, H.W. Virgin, Autophagy in immunity and inflammation, Nature. 469 (2011) 323–335.

[38] Z.Y. Li, Y. Yang, M. Ming, B. Liu, Mitochondrial ROS generation for regulation of autophagic pathways in cancer, Biochem. Biophys. Res. Commun. 414 (2011) 5–8.

[39] G. Ishdorj, L. Li, S.B. Gibson, Regulation of autophagy in hematological malignancies: Role of reactive oxygen species, Leuk Lymphoma. 53 (2012) 26–33.

[40] S. Loft, H.E. Poulsen, Cancer risk and oxidative DNA damage in man, J Mol Med. 74 (1996) 297-312.

[41] S. N. Goyal, A. K. Sharma, S. Haiderali, N. Reddy, D. S. Arya and C. R. Patil, Prediabetes: grounds of pitfall signalling alteration for cardiovascular disease, RSC Adv 5 (2015) 1619.

[42] T.M. Paravicini, R.M. Touyz, NADPH oxidases, reactive oxygen species, and hypertension: clinical implications and therapeutic possibilities, Diabetes Care. 2 (2008) 170-180.

[43] F. Desjardins, J.L. Balligand, Nitric oxide-dependent endothelial function and cardiovascular disease, Acta Clin. Belg. 61 (2006) 326–334.

[44] A. K. Sharma, D. Khanna, P. Balakumar, Low-dose dipyridamole treatment partially prevents diabetes mellitus-induced vascular endothelial and renal abnormalities in rats, Int J Cardiol. 172 (2014) 530-532.

[45] D. Tousoulis, N. Papageorgiou, A. Briasoulis, E. Androulakis, M. Charakida, E. Tsiamis, C. Stefanadis, Conflicting effects of nitric oxide and oxidative stress in chronic heart failure: potential therapeutic strategies, Heart Fail Rev. 17 (2012) 65-79.

[46] U. Forstermann, H. Li, Therapeutic effect of enhancing endothelial nitric oxide synthase (eNOS) expression and preventing eNOS uncoupling, Br J Pharmacol. 164 (2011) 213-223.

[47] S. Jaiman, A. K. Sharma, K. Singh, D. Khanna, Signalling mechanisms involved in renal pathological changes during cisplatin-induced nephropathy, Eur J Clin Pharmacol 69 (2013) 1863–1874.

[48] M.M. Said, The protective effect of eugenol against gentamicin-induced nephrotoxicity and oxidative damage in rat kidney, Fundam Clin Pharmacol. 25 (2011) 708–716.

[49] R. Sen, D. Baltimore, Multiple nuclear factors interact with the immunoglobulin enhancer sequences, Cell. 46 (1986) 705–716.

[50] Monaco C, Andreakos E, Kiriakidis S, Mauri C, Bicknell C, Foxwell B, Cheshire N, Paleolog E, Feldmann M, Canonical pathway of nuclear factor kappa B activation selectively regulates proinflammatory and prothrombotic posses in human atherosclerosis, Proc. Natl. Acad. Sci. USA. 101(2004) 5634–5639.

[51] X.Q. Song, L.X. Lv, W.Q. Li, Y.H. Hao, J.P. Zhao, The interaction of nuclear factor-kappa B and cytokines is associated with schizophrenia, Biol. Psychiatry. 65 (2009) 481–488.

[52] S. Ghosh, M.J. May, E.B. Kopp, NF-kappa B and Rel proteins: Evolutionarily conserved mediators of immune responses, Annu. Rev. Immunol. 16 (1998) 225–260.

[53] S.K. Hansen, L. Guerrini, F. Blasi, Differential DNA sequence specificity and regulation of HIV-1 enhancer activity by cRel-RelA transcription factor, J. Biol. Chem. 269 (1994) 22230–22237.

[54] A.M. Brown, M.W. Linhoff, B. Stein, K.L. Wright, A.S. Baldwin, P.V. Basta, J.P. Ting, Function of NF-kappa B/Rel binding sites in the major histocompatibility

complex class II invariant chain promoter is dependent on cell-specific binding of different NF-kappa B/Rel subunits, Mol. Cell Biol. 14 (1994) 2926–2935.

[55] Schreck R, Rieber P, Baeuerle PA, Reactive oxygen intermediates as apparently widely used messengers in the activation of the NF-kappa B transcription factor and HIV-1, EMBO J. 10(1921) 2247–2258.

[56] D. Fouad, E. Siendones, G. Costan, J. Muntane, Role of NF-kappaB activation and nitric oxide expression during PGE protection against d-galactosamine-induced cell death in cultured rat hepatocytes, Liver Int. 24 (2004) 227–236.

[57] K.M. Lee, B.S. Kang, H.L. Lee, S.J. Son, S.H. Hwang, D.S. Kim, J.S. Park, H.J. Cho, Spinal NF-kB activation induces COX-2 upregulation and contributes to inflammatory pain hypersensitivity, Eur. J. NeuroSci. 19 (2004) 3375–3381.

[58] N. Sakai, T. Wada, K. Furuichi, Y. Iwata, K. Yoshimoto, K. Kitagawa, S. Kokubo, M. Kobayashi, A. Hara, J. Yamahana, T. Okumura, K. Takasawa, S. Takeda, M. Yoshimura, H. Kida, H. Yokoyama, Involvement of extracellular signal-regulated kinase and p38 in human diabetic nephropathy, Am J Kidney Dis. 45 (2005) 54–65.

[59] L. Covarrubias, D. Hernandez-Garcia, D. Schnabel, E. Salas-Vidal, S. Castro-Obregon, Function of reactive oxygen species during animal development: passive or active? Dev. Biol. 320 (2008) 1–11.

[60] N. Coant, S. Ben Mkaddem, E. Pedruzzi, C. Guichard, X. Tréton, R. Ducroc, J.N.
Freund, D. Cazals-Hatem, Y. Bouhnik, P.L. Woerther, D. Skurnik, A. Grodet, M. Fay,
D. Biard, T. Lesuffleur, C. Deffert, R. Moreau, A. Groyer, K.H. Krause, F. Daniel, E.
Ogier-Denis, NADPH oxidase 1 modulates WNT and NOTCH1 signalling to control

the fate of proliferative progenitor cells in the colon, Mol Cell Biol. 30 (2010) 2636-2650.

[61] S. Kajla, A.S. Mondol, A. Nagasawa, Y. Zhang, M. Kato, K. Matsuno, C. Yabe-Nishimura, T. Kamata, A crucial role for Nox 1 in redox-dependent regulation of Wnt-β-catenin signalling, FASEB J. 26 (2012) 2049-2059.

[62] J. Abe, M. Kusuhara, R.J. Ulevitch, B.C. Berk, J.D. Lee, Big mitogen-activated protein kinase 1 (BMK1) is a redox-sensitive kinase, J Biol Chem. 271 (1996) 16586-16590.

[63] N.R. Love, Y. Chen, S. Ishibashi, P. Kritsiligkou, R. Lea, Y. Koh, J.L. Gallop, K. Dorey, E. Amaya, Amputation-induced reactive oxygen species are required for successful Xenopus tadpole tail regeneration, Nat. Cell Biol. 15 (2013) 222–228.

[64] E. Merki, M. Zamora, A. Raya, Y. Kawakami, J. Wang, X. Zhang, J. Burch, S.W. Kubalak, P. Kaliman, J.C. Izpisua Belmonte, K.R. Chien, P. Ruiz-Lozano, Epicardial retinoid X receptor alpha is required for myocardial growth and coronary artery formation, Proc. Natl. Acad. Sci. USA. 102 (2005) 18455–18460.

[65] T. Brade, S. Kumar, T.J. Cunningham, C. Chatzi, X. Zhao, S. Cavallero, P. Li, H.M. Sucov, P. Ruiz-Lozano, G. Duester, Retinoic acid stimulates myocardial expansion by induction of hepatic erythropoietin which activates epicardial Igf2, Development. 138 (2011) 139–148.

[66] K. Loh, H. Deng, A. Fukushima, X. Cai, B. Boivin, S. Galic, C. Bruce, B.J. Shields, B. Skiba, L.M. Ooms, N. Stepto, B. Wu, C.A. Mitchell, N.K. Tonks, M.J. Watt, M.A. Febbraio, P.J. Crack, S. Andrikopoulos, T. Tiganis, Reactive oxygen species enhance insulin sensitivity, Cell Metab. 10 (2009) 260–272.

[67] J.R. Flores-Riveros, E. Sibley, T. Kastelic, M.D. Lane, Substrate phosphorylation catalyzed by the insulin receptor tyrosine kinase. Kinetic correlation to autophosphorylation of specific sites in the beta subunit, J Biol Chem. 264 (1989) 21557–21572.

[68] G. Kroemer, G. Marino, B. Levine, Autophagy and the integrated stress response, Mol. Cell. 40 (2010) 280–293.

[69] H.U. Simon, A. Haj-Yehia, F. Levi-Schaffer. Role of reactive oxygen species(ROS) in apoptosis induction, Apoptosis. 5 (2000) 415-418.

[70] C. Fleury, B. Mignotte, J.L. Vayssière, Mitochondrial reactive oxygen species in cell death signalling, Biochimie. 84 (2002) 131–41.

[71] R. Strange, R.R. Friis, L.T. Bemis, F.J. Geske, Programmed cell death during mammary gland involution, Meth Cell Biol. 46 (1995) 355–368.

[72] K. Kokawa, T. Shikone, R. Nakano, Apoptosis in the human uterine endometrium during the menstrual cycle, J Clin Endocrinol Metab. 81 (1996) 4144–4147.

[73] L.B. King, J.D. Ashwell, Thymocyte and T cell apoptosis: is all death created equal? Thymus. 23 (1994) 209–230.

[74] J.F. Kerr, J. Searle, Deletion of cells by apoptosis during castrationinduced involution of the rat prostate, Virchows Arch B Cell Pathol. 13 (1973) 87–102.

[75] M. Weil, M.C. Raff, V.M. Braga, Caspase activation in the terminal differentiation of human epidermal keratinocytes, Curr Biol. 9 (1999) 361–364.

[76] Qi Y, Tian X, Liu J, Han Y, Graham AM, Simon MC, Penninger JM, Carmeliet P, Li S, Bnip3 and AIF cooperate to induce apoptosis and cavitation during epithelial morphogenesis, J. Cell Biol. 198 (2012) 103–114.

[77] D.E. Morgenstern, D.A. Gifford, L.L. Li, C.M. Doerschuk, M.C. Dinauer, Absence of respiratory burst in X-linked chronic granulomatous disease mice leads to abnormalities in both host defense and inflammatory response to Aspergillus fumigates, J. Exp. Med. 185 (1997) 207–218.

[78] J.D. Lambeth, NOX enzymes and the biology of reactive oxygen, Nat. Rev. Immunol. 4 (2004) 181–189.

[79] A. Kanayama, Y. Miyamoto, Apoptosis triggered by phagocytosis-related oxidative stress through FLIPS down-regulation and JNK activation, J. Leukoc. Biol. 82 (2007) 1344–1352.

[80] R.J. Mailloux, M.E. Harper, Uncoupling proteins and the control of mitochondrial reactive oxygen species production, Free Radic. Biol. Med. 51 (2011) 1106–1115.

[81] T. Kizaki, K. Suzuki, Y. Hitomi, N. Taniguchi, D. Saitoh, K. Watanabe, K. Onoe, N.K. Day, R.A. Good, H. Ohno, Uncoupling protein 2 plays an important role in nitric oxide production of lipopolysaccharide-stimulated macrophages, Proc. Natl. Acad. Sci. USA. 99 (2002) 9392–9397.

[82] Y. Emre, C. Hurtaud, T. Nubel, F. Criscuolo, D. Ricquier, A.M. Cassard-Doulcier, Mitochondria contribute to LPS-induced MAPK activation via uncoupling protein UCP2 in macrophages, Biochem. J. 402 (2007) 271–278.

[83] A.P. West, I.E. Brodsky, C. Rahner, D.K. Woo, H. Erdjument-Bromage, P .Tempst, M.C. Walsh, Y. Choi, G.S. Shadel, S. Ghosh, TLR signalling augments macrophage bactericidal activity through mitochondrial ROS, Nature. 472 (2011) 476–480.

[84] L.R. Fraser, Ionic control of sperm function, Reprod. Fertil. Dev. 7 (1995) 905– 925.

[85] J. Parinaud, P. Milhet, Progesterone induces Ca-dependent 3',5'-cyclic adenosine monophosphate increase in human sperm, J. Clin. Endocrinol. Metab. 81 (1996) 1357–1360.

[86] E. de Lamirande, C. Gagnon, Capacitation-associated production of superoxide anion by human spermatozoa, Free Radic. Biol. Med. 18 (1995) 487–495.

[87] P. Leclerc, E. de Lamirande, C. Gagnon, Cyclic adenosine 3',5' monophosphatedependent regulation of protein tyrosine phosphorylation in relation to human sperm capacitation and motility, Biol. Reprod. 55 (1996) 684–692.

[88] B.J. Awda, M.M. Buhr, Extracellular signal-regulated kinases (ERKs) pathway and reactive oxygen species regulate tyrosine phosphorylation in capacitating boar spermatozoa, Biol Reprod. 83 (2010) 750–758.

[89] G. Donà, I. Kožuh, A.M. Brunati, A. Andrisani, G. Ambrosini, G. Bonanni, E. Ragazzi, D. Armanini, G. Clari, L. Bordin, Effect of astaxanthin on human sperm capacitation, Mar Drugs. 11 (2013) 1909–1919.

[90] W.C. Ford, Regulation of sperm function by reactive oxygen species. Hum Reprod Update. 10 (2004) 387-99.

[91] D. Sanocka, M. Kurpisz, Reactive oxygen species and sperm cells. Reprod Biol Endocrinol. 2 (2004) 12.

[92] S.S. Du Plessis, A. Agarwal, J. Halabi, E. Tvrda, Contemporary evidence on the physiological role of reactive oxygen species in human sperm function. J Assist Reprod Genet. 32 (2015) 509-20.

[93] H. Motohashi, M. Yamamoto, Nrf2-Keap1 defines a physiologically important stress response mechanism, Trends Mol Med. 10 (2004) 549–557.

[94] D. Trachootham, W. Lu, M.A. Ogasawara, R.D. Nilsa, P. Huang, Redox regulation of cell survival. Antioxid Redox Signal. 10 (2008) 1343-74.

[95] G. Groeger, C. Quiney, T.G. Cotter, Hydrogen peroxide as a cell-survival signaling molecule. Antioxid Redox Signal 11 (2009) 2655-71.

[96] H. Kamata, T. Manabe, J. Kakuta, S. Oka, H. Hirata, Multiple redox regulation of the cellular signalling system linked to AP-1and NFkappaB: Effects of Nacetylcysteine and H2O2on the receptor tyrosine kinases, the MAP kinase cascade, and IkappaB kinases, Ann. N. Y. Acad. Sci. 973 (2002) 419–422.

[97] P.K. Vayalil, K.E. Iles, J. Choi, A.K. Yi, E.M. Postlethwait, R.M. Liu, Glutathione suppresses TGF-beta-induced PAI-1 expression by inhibiting p38 and JNK MAPK and the binding of AP-1, SP-1, and Smad to the PAI-1 promoter, Am. J. Physiol, Lung Cell Mol. Physiol. 293 (2007) 1281–1292.

[98] J. Li, L.M. Fan, V.T. George, G. Brooks, Nox2 regulates endothelial cell cycle arrest and apoptosis via p21cip1 and p53, Free Radic. Biol. Med. 43 (2007) 976–986.

[99] B. Liu, Y. Chen, D.K. St Clair, ROS and p53: A versatile partnership, Free Radic. Biol. Med. 44 (2008) 1529–1535.

[100] V.A. Shatrov, V.V. Sumbayev, J. Zhou, B. Brune, Oxidized low-density lipoprotein (oxLDL) triggers hypoxia-inducible factor-1alpha (HIF-1alpha) accumulation via redox-dependent mechanisms, Blood. 101 (2003) 4847–4849.

[101] P.K. Vayalil, K.E. Iles, J. Choi, A.K. Yi, E.M. Postlethwait, R.M. Liu, Glutathione suppresses TGF-beta-induced PAI-1 expression by inhibiting p38 and JNK MAPK and the binding of AP-1, SP-1, and Smad to the PAI-1 promoter, Am. J. Physiol. Lung Cell Mol. Physiol. 293 (2007) 1281–1292.

[102] S. Akiba, M. Chiba, Y. Mukaida, T. Sato, Involvement of reactive oxygen species and SP-1 in fibronectin production by oxidized LDL, Biochem. Biophys. Res. Commun. 310 (2003) 491–497.

[103] Y. Sun, L.W. Oberley, Redox regulation of transcriptional activators, Free Radic. Biol. Med. 21 (1996) 335–348.

[104] J. Moungjaroen, U. Nimmannit, P.S. Callery, L. Wang, N. Azad, V. Lipipun, P. Chanvorachote, Y. Rojanasakul, Reactive oxygen species mediate caspase activation and apoptosis induced by lipoic acid in human lung epithelial cancer cells through Bcl-2 down-regulation, J. Pharmacol. Exp. Ther. 319 (2006) 1062–1069.

[105] M. Kurdi, G.W. Booz, Evidence that IL-6-type cytokine signalling in cardiomyocytes is inhibited by oxidative stress: Parthenolide targets JAK1 activation by generating ROS, J. Cell Physiol. 212 (2007) 424–431.

[106] M. Sundaresan, Z.X. Yu, V.J. Ferrans, K. Irani, T. Finkel, Requirement for generation of H2O2for platelet-derived growthfactor signal transduction, Science. 270 (1995) 296–299.

[107] M.L. Circu, T.Y. Aw, Reactive oxygen species, cellular redox systems, and apoptosis, Free Radic. Biol. Med. 48 (2010) 749–762.

[108] N. Maulik, D.K. Das, Redox signalling in vascular angiogenesis, Free Radic.Biol Med. 33 (2002) 1047–1060.

Legends for Figures:

Figure 1: Figure portrays the involvement of various sites in generation of ROS. NADPH-oxidase indicates nicotinamide adenine dinucleotide phosphate-oxidase; eNOS indicates endothelial nitric oxide synthase; NO indicates nitric oxide; RNS indicates reactive nitrogen species; ROS indicates reactive oxygen species.

Figure 2: Figure interpret the pre-diabetic-associated cardiovascular signalling alterations and their possible contribution to cardiovascular disease pathology. ROS indicates reactive oxygen species; TNF α indicates tumor necrosis factor- α ; NF κ B indicates nuclear factor kappa-B cells; mTOR indicates mammalian target of PI3K rapamycin; JNK indicates c-Jun N-terminal indicates kinases; phosphatidylinositide 3-kinase; PKB/Akt indicates protein kinase B; GPCR indicates G protein-coupled receptors; Nrf2 indicates nuclear factor erythroid 2-related factor 2; cAMP indicates cyclic adenosine monophosphate; MAPK indicates mitogen-activated protein kinase; TLR indicates toll-like receptors; HIF1- α indicates hypoxia-inducible factor $1-\alpha$.





