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A rheological study on non-rubber component network in natural rubber

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Non-rubber components, mainly indicating phospholipid and protein, were separately removed from natural rubber to individually study their effect on structure and properties of the rubber. Fourier Transform Infrared Spectroscopy (FTIR) and ¹H nuclear magnetic resonance (¹H NMR) were applied to characterize the chemical structure and the non-rubber component residual. Rheology study and stress relaxation measurement were adopted to study the role non-rubbers played in natural networks. Rheological study of natural rubber (NR) and deproteinized natural rubber (DPNR) exhibited similar dynamic modulus at 170°C. The lack of superposition in van Gurp Palmen (vGP) curves at different temperature for NR and DPNR, together with the shape of vGP curves proved that long chain branching was mainly constructed by phospholipid. Stress relaxation measurement at room temperature was fitted with Maxwell model and showed that NR relaxation curve underwent a quick decrease and then come to 58% equilibrium stress retention, about 3 times higher than that of DPNR, indicating that protein in NR contributed to the network structure at room temperature. Combined the chemical with molecule dynamic study, the interaction between protein and phospholipid in non-rubber component network was proposed.

Introduction

Natural rubber (NR) has been particularly interesting and attractive materials since their first vulcanization by C. Goodyear in 1839. And the natural rubber (NR) latex, collected from the Hevea trees as a colloidal suspension [1], is the main commercial feedstock for rubber and latex dipping industries. The field of tire industry is one for which the application of natural rubber is highly attractive, and obvious superiorities in mechanical and universal properties in a wide temperature range could allow such materials to be employed as innovative solutions to satisfy the demanding requirements that a modern-day tire must fulfill, such as wet grip, mechanical and thermal resistance, low cost, recycling, etc.[2, 3] Nowadays, natural rubber has become increasingly important for applications spanning many fields of chemistry, the hygienic and medical sectors, and it is further involved in a broad spectrum of consumer products in our daily life [4-7].

As a natural material, the yield of natural rubber which is restricted seriously by land resources and natural conditions, has become the fatal bottlenecks for extensive application. Under this circumstance, synthetic analogue polyisoprene with high expectations to replace natural rubber is modified mainly by various molecule design [8-13], while the comprehensive properties were disappointing.

It is reported that the non-rubbers [14], especially proteins and lipids, confer to the natural rubber latex excellent properties unsurpassed by any synthetic latex [15]. As a consequence, great industrial and scientific attention is paid to non-rubbers structure and their primary importance on natural rubber structure to construct the magical mechanical performance [16], such as high strength of raw rubber (green strength), shape retention, high tensile strength, high crack growth resistance and minimal heat built up in the vulcanized state. Several attempts have also been made to study the non-rubber components and investigate the corresponding structure in natural rubber. Tanaka, Y [17, 18] first proposed that cis-polyisoprenes obtained from H. brasiliensis and Parthenium ardentatum consist of more than 5000 isoprene units and that there are trans-isoprene units per rubber chain. And the mainstream view now is that the NR molecules comprise of 2 trans-isoprene units connected to a long-chain of cis-isoprene units. Two terminal groups, referred to as α and ω, have been postulated to link with mono- and di-phosphate groups associated with phospholipids by H-bonding at the α-terminal, whereas the ω-terminal is postulated to be a modified dimethylallyl group linked to protein by H-bonding [19]. Then the non-rubbers can connect the linear polyisoprene chains in NR through functional terminals and generate the branching or net-working topology, which intitules the “naturally occurring
network”. [20] But since Goodyear medal [19] in 2001, no clear evidence of structure of end-linking network is reported yet. Although the real cross-linking structure and role of non-rubbers are not fully understood, it was suggested that the cross-linking of NR formed by non-rubbers can be eliminated by deproteination and transesterification. After the removal of protein and phospholipid, linear rubber chain can be obtained. [21] 

The effect of proteins and phospholipids on strain induced crystallization and the correlation with the tensile strength of NR have been extensively studied by X-ray diffraction and microscopic methods [22, 23]. While spectroscopy method is static and mostly accompanied by solvent, and X-ray characterization is indirect in “natural occurring” networking characterization. Rheological property is highly sensitive to long chain branching because even a low degree of long-chain branching has a significant effect on the viscoelasticity in polyethylene [23-28] or polypropylene [29]. Van Gurp Palmen curve, which was first proposed to judge the feasibility of Time-Temperature Superposition (TTS) principles and then evolved to branched structure study independent of chemical components of the macromolecule chain, is also very sensitive to the long chain branching of the polyisoprene melt in unvulcanized NR [30]. Therefore, the rheological behavior of unvulcanized NR can be characterized by a dynamic spectrum, related to molecular structure in melt, which is strongly impacted by branching or networking. However, to our knowledge, there is a lack of systematic investigation on how the non-rubber components, affect the rheological behavior of rubber chains in melt and inversively testify role of non-rubber components from rheology, especially effect of the long chain branching on rheological property. While few rheological studies were conducted in NR, among which most focused on blended NR with plastic [31], silica [32, 33], carbon black [34] or other natural ingredient [35] in vulcanized regime, and hardly any was on unvulcanized rubber. While in unvulcanized NR, the studies mainly emphasized on processability [36] or nonlinear viscoelastic behavior [37]. In this study, we conducted rheological study on the unvulcanized NR melt, and introduced the widely used vGP curve into unvulcanized NR to study the effect of non-rubber component.

Additionally, the stress relaxation behavior is also of important guiding sense in probing the network, additionally the equilibrium relaxation modulus $G_e$ is proportional to crosslinking density. [28] The stress relaxation behavior of natural rubber is often investigated in vulcanized rubber [29-31], while scare reports in unvulcanized regime is reviewed if there were. In this study, to investigate the network structure constructed by non-rubbers in unvulcanized NR, we introduced stress relaxation test and analyze the equilibrium relaxation modulus to evaluate the crosslinking density and constraint to relaxation units.

In this work, an attempt was made to clarify the “naturally occurring network” by molecule dynamic study, combining the chemical structure analysis. Fourier Transform Infrared Spectroscopy (FTIR), Rheology and stress relaxation were applied to analyze the composition and structure of the natural network. To individually study the effect of non-rubber component, three models were prepared to individually study the effect of non-rubber component and listed as follows, NR containing all the non-rubber component, deproteined natural rubber (DPNR) containing phospholipid and fatty acid, transesterified deproteined natural rubber (TEDPNR) containing only rubber chain.

**Experimental**

**Materials**

Natural rubber latex used in the present work was commercial high ammonia natural rubber (HANR) latex purchased from Dongfeng company (China). Total solid content and dry rubber content of the HANR latex, determined according to ASTM D 1076, were 61.3% and 60.8 w/w%, respectively. Triton X-100 and protease (P-5380) was provided from Sigma-Adrich. Other reagents used were analytical grade.

**Preparation of specimens**

Three models were prepared to individually study the effect of non-rubber component and listed as follows, natural rubber containing all the non-rubber component, deproteined natural rubber (DPNR) containing phospholipid and fatty acid, transesterified deproteined natural rubber (TEDPNR) containing only rubber chain.

**Natural rubber**

HANR latex was centrifugation once for the removal of dash, and then spread into a thin layer to dry in under reduced pressure at ambient temperature until constant weight in vacuum oven.

**Deproteinization of natural rubber**

Deproteinization of the HANR latex was carried out by incubation with 0.08 wt% protease in the presence of 0.15% v/v Triton X-100 and 1 wt% aquea ammonia at 37°C for 12 h and the dry rubber content was 30%. The cream fraction was redispersed in the surfactant, in which dry rubber content was adjusted to 30 wt% with deionized water. It was washed twice or three times by centrifugation. The rubber was recovered by centrifugation followed by coagulation with methanol and dried under reduced pressure at ambient temperature until constant weight.

**Measurement of nitrogen content of the deproteined natural rubber (DPNR)** was made by nitrogen elemental analyzer (CARLO ERBA 1106, Italian), and the nitrogen content was reduced from 0.5% to 0.07% after deproteinization.

**Transesterification of the deproteinized rubber**

Transesterification of the rubber from DPNR was carried out in 2% w/v toluene by reaction with freshly prepared sodium methoxide and stirring at room temperature for 8 h. The resulting transesterified deproteinized natural rubber (TEDPNR) was purified by precipitation of the rubber solution using excess of methanol and then dried in vacuum oven.
Measurement of phosphorus content of TEDPNR was made by phosphorus elemental analyzer (IRIS 1000 ICP-AES, Thermo Electron Co. USA), and the phosphorus content was reduced from 224ppm to 65ppm after transesterification.

The molecular weight of all samples was determined by gel permeation chromatography (HLC-8320, Waters), and the number averaged molecule weight was about 150-160kg mol⁻¹ for NR and DPNR, 134kg mol⁻¹ for TEDPNR, while MW of commercial linear isoprene rubber (IR) was 317 kg mol⁻¹.

**Characterization**

**FTIR.** The rubber samples for FTIR analysis were prepared by casting 1% w/v solutions of the rubber in tetrahydrofuran (THF) on a KBr disk, conducted with a Nicolet-560 infrared spectrometer (USA) using a potassium bromide (KBr) pellet. FTIR spectra were recorded in the spectruml range of 4000-400 cm⁻¹ with a 2 cm⁻¹ resolution and 32 scans.

**¹H-NMR.** ¹H NMR spectra was recorded on a spectrometer (Varian INOVA-400) at 25°C using CDCl₃ as the solvent and TMS as the external standard.

**Rheology Analysis.** Dynamic mechanical measurements were performed by means of the HAAKE PXR800 rheometer. Shear deformation was applied under conditions of controlled deformation amplitude, always remaining in the range of linear viscoelastic response. Frequency dependencies of the storage (G') and loss (G'') shear moduli were measured in the frequency range 0.01-100 Hz at a deformation amplitude ranging from 2 to 4% depending on temperature and at various temperatures between 160°C and 190°C. The geometry of parallel plates was employed with plate diameters of 20 mm and a sample thickness of 1.2 mm. Temperature control (±0.1°C) was achieved via water circulation system. Before measurements, the samples were processed into even thickness plate by heatressing at 150°C for 5 min in a mold and then isothermal for 12h in vacuum at 40°C.

**Stress relaxation.** Stress relaxation was conducted in INSTRONG universal tensile test machine at a proper strain of 20%. The total length was 15mm and elongated to target strain at 500mm min⁻¹ tension rate and then keep the strain for 1200s. Before measurements, the samples were processed into even thickness plate by heatressing at 150°C for 5 min in a mold and then isothermal for 12h in vacuum at 40°C.

**Results and discussion**

**Chemical structure characterization of NR samples**

Infrared spectroscopy was used as a method of chemical structure analysis, and also to study intermolecular interactions caused by hydrogen bonding, because the vibrational modes of the donor and acceptor groups are sensitive to this interaction leading to a change in a vibrational characteristic [32]. And the FTIR spectrums of NR, DPNR and TEDPNR at room temperature were plotted in Fig.1.

First, a wide H-bond peak at around 3400cm⁻¹ was absent in TEDPNR. And in NR the wide peak comprised a relatively sharp peak at about 3300cm⁻¹ related to the N-H symmetric stretch in H-bond, and the wide H-bond peak is mainly ascribed to O-H symmetric stretch in H-bond in non-rubbers, overlapping with small amount of environment water absorbed by non-rubbers.

Second, to pick out the non-rubbers, the rubber chain peaks assignment are listed below. vmax/cm⁻¹ 836 (Trisubstituted olefin out-of-plane =CH wag), 1129 (-CH₂ rock), 1300 (-CH₂ wag), 1376 (-CH₃ symmetric deformation), 1450 (-CH₂ symmetric and -CH₃ asymmetric deformation), 1664 (C=C stretch), 2720 (Overtone of -CH₂ umbrella), 2850 (-CH₂ and -CH₁ symmetric stretch), 2920 (-CH₂ asymmetric stretch), 2962 (-CH₃ asymmetric stretch), 3030 (Olefin =CH- stretch) [33, 34].

Then, the recession of characteristic peaks of protein at 1663 and 1546cm⁻¹ [35] is obviously observed in DPNR compared with NR, and the peaks of C=O in phospholipid at 1738cm⁻¹ [36] in TEDPNR. The FTIR results, combining with the elemental analysis in section 2, proved that the intended samples were successfully prepared. As for the different intensity of C=C stretch peak in normalized infrared spectrogram, it can be interpreted by various infrared activity with different non-rubber environment.

The shoulder peak at 1260cm⁻¹ in NR shifted to 1248cm⁻¹ and turn sharp after protein removal, continually shifting to about 1243cm⁻¹ and receding rapidly after phospholipid deprivation. It has been reported that the asymmetric O-P-O vibrational band is extremely sensitive to hydration, and the O-P-O asymmetric stretching of the hydrated phospholipid bilayer is usually observed at 1230 cm⁻¹, whereas dried or anhydrous lipid always appears at 30 cm⁻¹ higher frequency. [37] The shift of O-P-O asymmetric stretching peak strongly suggests that phospholipids in NR interacted with protein via H-bond, and in DPNR rubber aggregate or link together via H-bond. [38]
The ¹H-NMR spectra of NR, DPNR and TEDPNR were shown in Fig. 2, and a small amount of residual chloroform peak was detected at 7.28ppm. All the spectrums were normalized by the =C-H at the backbone triple (J=6.5Hz) peak at 5.15ppm. A small multiplet signals resonating at 3.91ppm was expected to be derived from the nonequivalent methylene protons linked to a phosphate group, -CH₂OP-. The multiple peaks at 2.365ppm, 2.375ppm and 2.395ppm were assignable to -CH₂- of long-chain fat, the multiple peak at 2.06ppm was assignable to -CH₃- in the backbone, the single peak at 1.7ppm was assignable to -CH₃ in the backbone, the multiple peak at 1.43ppm was assigned to -CH₃- of protein, the multiple peak at 1.13ppm was assigned to -CH₃ of protein. From the extended spectrums we can see that protein was removed after deproteinization, long-chain fat was eliminated after transesterification, while phospholipid was partly removed in TEDPNR. This can be interpreted that some phospholipid was connected with the rubber chain terminal. [16] Combining ¹H-NMR and FTIR results, we can conclude that the intended samples were successfully prepared.

Branching molecular structure in unvulcanized nature rubber

To further study the effect of non-rubbers on the main chain structure, rheological behavior was analyzed in this section. All measurements were conducted at a fixed strain, ranging from 2% to 4% at 170°C where all samples display the linear
viscoelastic performance respectively, and the storage modulus $G'$ as functions of frequency is shown in Fig. 3. At so high temperature, most H-bonding or other non-bonding interaction was destroyed at this temperature.

Fig.3 Dynamic spectrum of NR, DPNR and TEDPNR at 170°C.

NR and DPNR show a higher modulus at low frequency than TEDPNR, indicating long-chain branching, crosslinking or phase-separation existence. [40, 41] However, the modulus of NR is a little higher than that of DPNR. DPNR displayed no apparent slope change at low frequency, while the modulus shifted up as much as 2 magnitudes contrasted with TEDPNR.

To further investigate the modulus shift, dynamic spectrum in different temperatures for all samples were obtained, and van Gurp-Palmen plots were drawn from dynamic shear modulus at three different temperatures: 160, 170, and 190°C, displayed in Fig.3. Generally, fine superposition of vGP curves was observed for classical viscoelastic polymers, and little amount of long chain can significantly distort the vGP curve and thus cause a misalign. [42, 43] The rubber models were confirmed to be thermo-rheology melt from the discrepancy of vGP curves for NR and DPNR in Fig.4, possibly with long-chain branching or phase-separation.

Fig.4 vGP curve of (a) NR, (b) DPNR and (c) TEDPNR at various temperatures.

To judge the reason of vGP curves discrepancy, temperature sweep from 100°C to 200°C in a frequency of 1 Hz at 1% strain was conducted. As is known, phase-separation can cause a sharp uprush in the modulus of temperature sweep. [44, 45] But now, the result shows no uprush in dynamic modulus in Fig.5, indicating that no phase separation occurred in the testing temperature region. As is shown in section 2, all samples were unvulcanized, so we can excluded the crosslinking. Now after excluding phase-separation, the modulus shift of NR and DPNR in Fig.3 is attributed to long chain branching, and a small amount of protein involved in forming long-chain branch.

Fig.5 Temperature sweep in a frequency of 1 Hz at 1% strain from 100°C to 200°C.

In contrast to NR and DPNR, the overlap of vGP curves among the various temperature is good for TEDPNR, indicating that
TEDPNR is simple fluid. And from the shape of curve—only one minimum, spread monotonously to 90° in the phase angle—we can conclude that TEDPNR was probably linear. Combining the good linearity of Han map [46] in Fig.6, we can conclude that TEDPNR was linear at high temperature, in accordance with the dynamic spectrum.

![Fig.6 Han map of DPNR and TEDPNR at 170°C.](image)

The above rheological results showed that long-chain branching existed only in NR and DPNR melt, indicating that phospholipid and a small amount of protein acted as the branching point in the long-chain branching construction. Furthermore, we can conclude, phospholipid and a small amount of protein connected with rubber terminal point by chemical bond or very strong interaction (survive at 170°C), then the non-rubbers coagulate by micellar interaction [47], and possibly formed star [48] in melt.

**Network structure in unvulcanized nature rubber**

Rheological demonstration on structure of three models at high temperature confirms the stable topology in NR, which proves in turn that the natural network is further constructed mainly by thermal unstable H-bonding or non-bond interaction between non-rubber component and rubber chain end, in accordance with the FTIR result. However, does the natural network exist in melt at room temperature? To answer this question, stress relaxation test was conducted. As is known, stress relaxation analysis is an effective method to study the network, and the equilibrium relaxation modulus $G_\infty$ is proportional to crosslinking density. [44]

In this study, stress relaxation curves of NR, DPNR and TEDPNR at 25°C is displayed in Fig.7, where all samples were quickly subject to a proper strain of 20% to avoid the influence of strain induced crystallinity (SIC) [49], and then hold the strain for 1200s. For the sake of contrast, a relaxation curve of fairly linear IR with a 317kg mol⁻¹ number average molecule weight (MW) was added. Generally, the stress of linear viscoelastic polymer quickly relaxed to a very small value, and even to zero.

![Fig.7 Stress relaxation curves of NR, DPNR, TEDPNR and IR at 25°C.](image)

To illustrate the effect of non-rubbers on the stress relaxation tendency, all the instant stress was normalized by the corresponding initial stress, as is displayed in Fig.7. Relaxation curves of NR tended to form a plateau after about 300s and the retention is more than 50%, performed the largest stress retention, indicating that protein mainly contributed to the natural network at room temperature. Unexpectedly, DPNR performed nearly the same behaviors as that of IR, but relaxed slightly slower than that of linear IR. The TEDPNR relaxed to zero during the testing period. It can be confirmed that relaxation units in NR, were obviously confined by chemical or strong physical network. While in DPNR, the network constraint was scarcely observed in contrast to NR. However, comparing with linear IR, we can conclude that the restriction derived from long chain branching. TEDPNR displayed typical relaxation behavior of linear molecular chains with fully relaxed stress.

To further confirm the above assumption, we adopted Maxwell model [41, 50, 51] to describe the stress relaxation of rubber, read in equation 1:

$$\sigma(t) = \sigma_e + \sigma_i e^{-\tau_i}$$  \hspace{1cm} \text{equ.1}

In equation 1, $\sigma(t)$ represented the stress in the relaxation, $\sigma_e$ was equilibrium stress, $\sigma_i$ is coefficient of the $i$th Maxwell model and $\tau_i$ was relaxation time of the $i$th Maxwell model. Elastomer relaxation followed a 7-element Maxwell model [52], read in equation 2:

$$\sigma(t) = \sigma_e + \sigma_1 e^{-\tau_1} + \sigma_2 e^{-\tau_2} + \sigma_3 e^{-\tau_3}$$  \hspace{1cm} \text{equ.2}

Divided by the initial stress $\sigma_0$ in both sides, equation 2 may then be inverted to read:

$$\frac{\sigma(t)}{\sigma_0} = \frac{\sigma_e}{\sigma_0} + \frac{\sigma_1}{\sigma_0} e^{-\frac{t}{\tau_1}} + \frac{\sigma_2}{\sigma_0} e^{-\frac{t}{\tau_2}} + \frac{\sigma_3}{\sigma_0} e^{-\frac{t}{\tau_3}}$$  \hspace{1cm} \text{equ.3}

Let

$$A(t) = \frac{\sigma(t)}{\sigma_0}$$  \hspace{1cm} \text{equ.4}

Then we fit the experimental data by equation 3 and the result was listed in Table 1.
From equation 4, we can see that $A_i$ represent the contribution of $t_i$, and $t_i$ corresponds to the different relaxation unit in the samples. The $A_1$, $A_2$, $A_3$ increased in the same samples with the increasing magnitude of the relaxation time $t_1$, $t_2$, $t_3$. The consistency of $A_i$ and $t_i$ proved that the larger relaxation unit contributed larger stress relaxation.

<table>
<thead>
<tr>
<th>Sample</th>
<th>$A_0$</th>
<th>$A_1$</th>
<th>$t_1$</th>
<th>$A_2$</th>
<th>$t_2$</th>
<th>$A_3$</th>
<th>$t_3$</th>
<th>$R^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td>NR</td>
<td>0.58</td>
<td>0.06</td>
<td>9.877</td>
<td>0.137</td>
<td>57.85</td>
<td>0.187</td>
<td>364.7</td>
<td>0.987</td>
</tr>
<tr>
<td>DPNR</td>
<td>0.15</td>
<td>0.09</td>
<td>12.62</td>
<td>0.202</td>
<td>82.65</td>
<td>0.515</td>
<td>544.9</td>
<td>0.995</td>
</tr>
<tr>
<td>IR</td>
<td>0.06</td>
<td>0.13</td>
<td>17.76</td>
<td>0.274</td>
<td>96.31</td>
<td>0.501</td>
<td>561.9</td>
<td>0.996</td>
</tr>
<tr>
<td>TEDPNR</td>
<td>0.09</td>
<td>0.21</td>
<td>16.88</td>
<td>0.336</td>
<td>115.2</td>
<td>0.487</td>
<td>740.3</td>
<td>0.982</td>
</tr>
</tbody>
</table>

Second, the crosslinking density was discuss through the $A_0$. As is show in table 1, NR displays the largest $A_0$, quadruple of DPNR, while $A_0$ of DPNR is also higher than that in linear IR, and TEDPNR shows a meaningless negative $A_0$.

![Fig.8 Maxwell relaxation time histogram of NR, DPNR, TEDPNR and IR.](image)

While in TEDPNR, the linear topology and lower MW results in the absence of strong entanglement network.

Then the relaxation time was also considered. And the relaxation time of all samples was displayed in the histogram in Fig.8.

As is shown in Fig.8, the relaxation time decrease in this order: NR>DPNR<IR<TEDPNR. In vulcanized rubber, the relaxation time decreases with increasing crosslinking density, which can be interpreted as the crosslinking point restriction. [38] Similarly, in the unvulcanized rubber, the relaxation units were also constrained by the natural network, and then displayed a decrease in relaxation time. The result confirmed that the crosslinking density increased in the order: NR>DPNR>IR>TEDPNR, which agreed with the previous study in $A_0$.

![Scheme 1 Assumed network structure in (a) NR and (b) DPNR.](image)

From the above results, we proposed the non-rubber in the structure of NR and DPNR in Scheme 1. In NR, the protein is connected in $\omega$-terminal and then coagulated with each other by H-bond, part of which can also interact with phospholipid by H-bond. Moreover, in $\alpha$-terminal, mono- or di-phosphate is bonded with the rubber chain, and then connected by micelle cluster. While in DPNR, the phospholipid or protein fragment after protease treatment can also interact with the $\omega$-terminal by polar-polar interaction or H-bond, together with the $\alpha$-terminal connected to phospholipid to form the network, whose density is much lower than that in NR.

**Conclusions**

The long chain branching point formed by phosphate, and crosslinking point consisted of protein and phospholipid are confirmed by the rheological and stress relaxation change of the three models. The modulus decreased at low frequency, and equilibrium stress retention decreased by 75% after deproteinization, suggesting that the network was broken with removing protein. The van Gurp Palmen curves showed a fine overlap and the stress relaxed to zero after transesterification, indicating the decomposition of long chain branching point by phosphatase. The FTIR study of NR and DPNR further reveals that the H-bonding between protein and phosphatase connected with the terminal phosphatide mainly contributed to the physical crosslinking in NR. While in DPNR, part of phospholipid and protein fragments after protease treatment...
may also connected with α-terminal by H-bond to form the weak network, and H-bond between phosphatase and α-terminal phosphatide participated together in the formation of long chain branching point.

Acknowledgements
We acknowledge funding from the national natural science foundation of China to carry out this work (contract grant number 51333003). Authors must particularly acknowledge GS Huang for her forceful guidance and generous assistance.

Notes and references

2. Mitarai, K., Kneading carbon black and silica to a discharging temperature of 130 degrees c. or higher; nonhardening on storage, 2004, Google Patents.


Protein can also interact with phospholipid, while in DPNR, phospholipid or protein fragment after can also interact with the ω-terminal.