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Determination of tryptophan and tyrosine by chemiluminescence based on luminol-N-bromosuccinimide-ZnS quantum dots system Shifeng Li*, Min Xing, Haiyan Wang, Li Zhang, Yiman Zhong, Lei Chen

Abstract

In this work, water-soluble ZnS quantum dots (QDs) modified with 3-Mercaptopropionic acid (MPA), L-Cysteine (L-Cys) and Thioglycolic acid (TGA) were synthesized. The nanostructure and optical properties were characterized by X-ray powder diffraction (XRD), Transmission electron microscope 10 (TEM), UV-Vis absorption spectrum and Fluorescence spectrum. Through the study of ZnS QDs effects on the chemiluminescence (CL) system of luminol, we found that ZnS QDs could obviously enhance the CL of luminol-N-bromosuccinimide (NBS) system in alkaline medium. At the optimal experimental conditions, we 15 researched the impacts of 17 kinds of amino acids on this system. It was found that L-tryptophan (Trp) and L-tyrosine (Tyr) had significantly inhibitory effects on luminol-NBS-ZnS QDs CL. Based on the inhibitory effects, a novel CL method with wider linear range and lower detection limit for determination of Trp ₂₀ and Tyr was developed. The detection limits were 1.5×10⁻¹¹ g/mL and 2.0×10⁻¹⁰ g/mL, respectively. The possible CL reaction mechanism was also discussed briefly.

Introduction

Tryptophan(2S-2-amino-3-(1H-indol-3-yl))propanoic acid)
25 and tyrosine (4-hydroxyphenylalanine) are two aromatic amino
acids which are important precursors for synthetic peptide
hormones, antibiotics, dopamine and some drugs [1-3].
Meanwhile, Trp and Tyr play crucial roles on synthesizing
melanin and melanocytes in the blood. Many means for detecting
30 Trp and Tyr have been reported including electrochemistry [4,5],
high performance liquid chromatography [6,7],
spectrofluorometry [8], mass spectrometry [9] and capillary
electrophoresis [10]. However, these methods often suffer from
complicated pretreatment, poor reproducibility and low

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35 sensitivity defect. Therefore, it is necessary to design a rapid and sensitive method for detection of Trp and Tyr.

Flow injection analysis with low power consumption, wide liner range, low detection limit advantage has been widely used in biological immune analysis [11-13]. Luminol as an ordinary 40 CL reagent has been extensively studied by researchers, including luminol-ferricyanide [14], luminol-potassium permanganate [15-17], luminol-hydrogen peroxide system [18,19] and so on [20, 21]. Many of these oxidants have color interfere with analysis, therefore, finding more stable and no color oxidants become our 45 primary problem. HBrO as a strong oxidant could be applied in the field of CL. The biggest drawback is the reagent poor stability which limited its application scopes. In order to solve this problem, we choose NBS as oxidant. NBS can be hydrolyzed in aqueous solution for producing HBrO and the chemical properties 50 are stable than HBrO. In addition, this agent can both dissolved in the polar and non-polar solvent, which has a greatly potential application in the organic phase CL. Many researchers have used different methods to enhance luminol-NBS system CL, including Au nanoparticles [22], some drugs[23].

Recently, QDs or colloidal semiconductor nanocrystals have stimulated great interest in many fields due to their unique electronic and optical properties [24-28]. ZnS QDs is direct broadband semiconductor, its fluorescence wavelength can cover ultraviolet to infrared, high fluorescence efficiency, narrow and continuous excitation spectrum etc advantages, which make it has a wide application prospect in optoelectronic aspect. For example, as a light emitting diode, quantum dots laser, biological fluorescent probes and optical luminescence catalyst [29-36].

In this paper, we studied the presence of ZnS QDs in the CL reaction of luminol-NBS system, it was found that ZnS QDs could catalyze luminol-NBS CL. Through researching the 17 kinds of amino acids effects on this system, Trp and Tyr could significantly inhibit luminol-NBS-ZnS QDs system CL intensity. Based on this inhibition, we have developed a flow injection method for measuring Trp and Tyr. This method has a wider liner range and lower sensitivity advantages for the samples analysis with satisfactory results.

Experimental Section

75 Reagents and chemicals

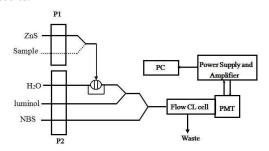
All the chemicals were of analytical-reagent grade and used without further purification, and prepared in double-distilled water. 3-Aminophthal-hydrazide (luminol) was purchased from Alfa Aesar. (Britain). N-bromosuccinimide (NBS), Zinc chloride (ZnCl₂) and Ethanol were purchased from Sinopharm Chemical Reagent Co. (Shanghai, China). Sodium sulfide (Na₂S) was purchased from Wuxi Chemical Co. (Jiangsu, China). MPA and TGA were purchased from Aladdin Chemistry Co. (Shanghai, China). Amino acids were purchased from Huixing Chemical Reagent Co. (Shanghai, China).

Apparatus

All the CL measurements were carried out on the ultra weak luminescence analyzer (IFFM-E, Remex Electronic Institute Limited Co., Xi'an China). The CL signals were monitored by the PMT adjacent to the flow CL cell. When the CL system was used for investigating the effects of amino acids on the CL system, the sample solution and ZnS QDs were injected simultaneously and mixed with each other before further reaction with luminol-NBS solutions. Amino acids were injected after the optimized experimental concentrations.

Procedure

The FI-CL system procedure is shown in Scheme 1. The solutions of luminol, NBS and carrier water were delivered into the flow cell by peristaltic pumps at 3.0 mL min⁻¹ flow rate, respectively . The ZnS QDs colloid solution was injected by a valve injector with 100 μ L sample loop. The light output from the CL reaction was detected and amplified by the PMT and luminometer. The signal was imported to the computer for date acquisition. The determination of certain analytes of interest was based on changes in CL intensity $\Delta I = I_s - I_o$, where I_s and I_o were the CL intensity of sample and blank solutions, respectively, which was used for quantitative analysis of amino acid molecules.



Scheme 1 Schematic diagram of the FI-CL system

Synthesis of ZnS QDs capped with MPA (TGA or L-Cys)

As it was cited in the literature[37-39], 0.3408 g of zinc chloride (ZnCl₂) and 0.21 mL MPA (0.21mL TGA or 0.2908 g L-Cys) as a stabilizing regent were dissolved in 265 mL of deionized water then stirred for 10 min to get a homogeneous mixture. The pH of the resulting solution was adjusted to 10.2 by adding 1 mol/L NaOH solution then transferred to the water bath with 70 °C for preheating 10 min, after that dropwise added a to certain concentration of Sodium sulphide solution for reacting 2 h. Then the resultant MPA capped ZnS QDs was centrifuged and washed with absolute ethanol three times. Finally, ZnS QDs was dried in 60 °C for 6 h. The sample obtained in preparation for the subsequent experiments.

125 Results and discussion

Characterization of ZnS QDs

As shown in Fig.1(A), the size of the as-prepared MPA/ZnS QDs was characterized by TEM and statistically analyzed as 5 ± 1 nm in diameter. TEM images of TGA or L-Cys modified ZnS QDs were also investigated (see Supplementary material Fig. S1). A histogram of approximate distribution of MPA/ZnS QDs as shown in Fig.1(B) based on TEM observations. Fig.1(C) was 70°C ZnS X-ray power diffraction pattern, the peak location is consistent with cubic ZnS standard card 05-0556. Fig.1(D) was different modifier modified ZnS QDs UV-Vis absorption and fluorescence spectra, the ultraviolet absorption peak appear at 285 nm, the emission peak of fluorescence was appeared at 438 nm.

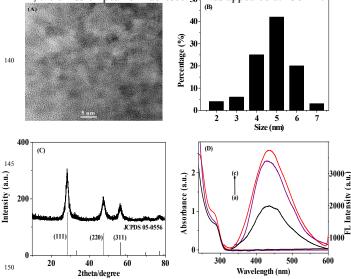


Fig. 1. (A) TEM of the MPA/ZnS QDs at 70 °C, (B) the size distribution histogram, (C) XRD image of MPA/ZnS QDs and (D) UV-Vis absorption spectrum and Fluorescence spectrum ZnS QDs(a-c: TGA/ZnS, L-155 Cys/ZnS, MPA/ZnS, respectively).

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Kinetic curves of CL reaction

In our experiments, we had synthesized and characterized three different modifiers capped ZnS QDs. All of the size were about 5 nm, which meant that the effect of particle size on catalytic properties could basically be eliminated. As it shown in Fig. 2 insert, On injection of ZnS QDs into the luminol-NBS system (Curve a), CL intensity was substantially increased. Compared with two other modifiers (TGA/L-Cys, Curveb/c), the enhance effect of MPA modified ZnS QDs was the best (Curve

So we selected MPA modified ZnS QDs as the reaction catalyst. Under the optimal condition, through the sensing research of 17 amino acids, the results showed that Trp and Tyr significantly inhibit the CL intensity (Fig. 2).

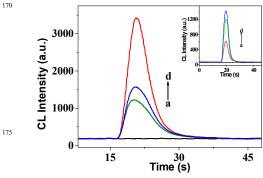


Fig. 2 Kinetic curves of CL reaction (Insert a-d): luminol-NBS, luminol-NBS-TGA/ZnS, luminol-NBS-L-Cys/ZnS, luminol-NBS-MPA/ZnS. (luminol: 3.5×10^{-7} mol/L, NaOH: 0.02mol/L, NBS: 2×10^{-5} mol/L, ZnS: 2×10^{-5} mol/L) and (a-d): luminol-NBS, luminol-NBS-MPA/ZnS-Trp, luminol-NBS-MPA/ZnS-Tyr, luminol-NBS-MPA/ZnS. (luminol: 3.5×10^{-7} mol/L NaOH: 0.025 mol/L, NBS: 2×10^{-5} mol/L, ZnS: 4×10^{-5} mol/L, Trp and Tyr: 1×10^{-6} g/mL).

Optimization of the experimental conditions

Effect of NaOH Concentration

Considering luminol in the alkaline medium has a strong CL signal, we chose the same concentration of NaOH, Na₂CO₃, and NaHCO₃ to add into the luminol solution. NaOH was selected as a reactive medium because of the strongest signal intensity. We chose 0.025mol/L NaOH (Fig. 3a) as the reaction medium for consequent research.

195 Effect of luminol Concentration

The effect of luminol concentration on the CL intensity was examined from 7.5×10^{-8} to 1×10^{-6} mol/L. The results showed that the CL intensity increased with luminol added (Fig. 3b). Considering too large concentration was unstable and difficultly test, 3.5×10^{-7} mol/L was chosen as the optimum concentration of luminol for further research.

Effect of NBS Concentration

The effect of the NBS concentration ranging from 8.0×10⁻⁶ to 9×10⁻⁵ mol/L on the CL signal was investigated. It was obvious found 2.0×10⁻⁵ mol/L NBS solution had a maximum CL intensity (Fig. 3c). Therefore, 2.0×10⁻⁵ mol/L NBS was chosen for the subsequent experiment.

Effect of ZnS QDs Concentration

The effect of ZnS QDs concentration on the changes in relative CL intensity was studied at different concentrations from 6.0×10^{-6} to 7×10^{-5} mol/L. The maximum CL intensity appeared at the concentration of 4×10^{-5} mol/L, so 4×10^{-5} mol/L ZnS (Fig. 3d) was selected for further research.

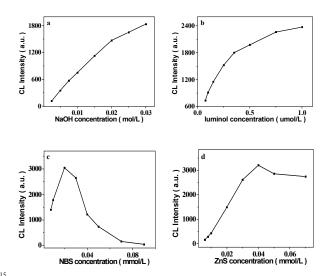


Fig. 3 Effects of reaction on the CL intensity: (a) NaOH concentration, luminol, 2.5×10^{-7} mol/L; NBS, 2.0×10^{-5} mol/L; MPA/ZnS, 5×10^{-5} mol/L. (b) luminol concentration (0.025 mol/L NaOH); NBS, 2.0×10^{-5} mol/L; MPA/ZnS, 5×10^{-5} mol/L. (c) NBS concentration, luminol, 3.5×10^{-7} mol/L (d) MPA/ZnS concentration, luminol, 3.5×10^{-7} mol/L (d) MPA/ZnS concentration, luminol, 3.5×10^{-7} mol/L(0.025 mol/L NaOH), NBS, 2.0×10^{-5} mol/L.

Mechanism discussion

The CL spectra of the reactions is obtained by using the modified F-4500 spectrofluorimeter, whose light entrance slot was shut. We mixed each substance and found the peak appeared at 425nm, as shown in Fig. 4a, indicating the same maximal peak as some literature reported [18-20]. Thus, we explored that the main emitters of luminol-NBS-ZnS QDs CL system were related to the excited state of 3-aminophthalate molecule with characteristic maximum emissions of around 425 nm [40, 41]. The enhanced CL signals are thus ascribed to the possible catalysis from ZnS QDs [20].

We also used UV-Vis absorption spectrum to explore the possible mechanism, shown as Fig. 4b. We could clearly see the

absorption peak of luminol-NBS system appear at 302 nm and 352 nm. Merging with ZnS, the CL system absorption peak location was no change. The light absorption of the mixed system was approximately equal to the sum of the light absorption of the 240 two individual systems. Therefore, we concluded that ZnS QDs might be used as the catalyst for this reaction.

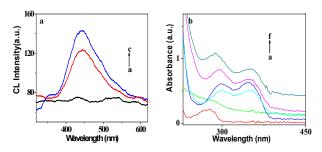


Fig. 4a CL spectra of CL system: (a):luminol, (b):luminol-NBS, (c):(b)-ZnS; 4b UV-vis absorption spectra of CL system: (a):Trp, (b):ZnS, 245 (c):luminol, (d):luminol-NBS, (e):(d)-ZnS, (f):(e)-Trp.

Using the flow injection analyzer, we could not found signal when the lower concentration of luminol and NBS mixed. When ZnS QDs added into the system, it could be observed a 250 significant CL signal. A lot of reactive oxygen and hydroxyl radical could be produced. Moreover, ZnS QDs and reactive oxygen could catalyze luminol and NBS reaction intermediates reaction, which might greatly improve the speed of light-emitting and signal strength. Therefore, we explored this CL system 255 mechanism as following in Scheme 2.

Scheme 2 Possible mechanism for the luminol-NBS-ZnS system

Analytic application

Under the optimum conditions, we researched the behavior of amino acids with luminol-NBS-ZnS QDs CL. It was found that Trp and Tyr both had stronger inhibition for the above CL, as shown in Table 1. The CL inhibitory ratios of amino acids on this 265 system were shown in Fig. 5. Based on this inhibition effects, a novel CL method for detection of Trp and Tyr was developed. Compared with other conventional method, this mean has a greatly lower detection limit and wide linear range advantage for samples analysis (Fig. 6). We obtained that Trp with a linear 270 range from 4.0×10^{-11} to 2.0×10^{-6} g/mL, Tyr with a linear range from 6.0×10⁻¹⁰ to 1.0×10⁻⁶ g/mL. The detection limits were 1.5×10⁻¹¹ g/mL, 2.0×10⁻¹⁰ g/mL of Trp and Tyr, respectively. The relative standard deviation (R.S.D., n=11) for Trp in water samples at 4.0×10^{-7} g/mL was 0.92 % (Fig. 7). Moreover, 275 recovery tests of standard addition in Trp samples showed the recoveries between 98.6% and 103.8%, which were at confidence interval of 100 ± 5 % (Table 2). The results showed that this flow injection CL was a sensitive and accurate method for the Trp and Tyr detection.

Table 1 Effects of amino acids on luminol-NBS-ZnS CLsystem

Table 1 Effects of anniho acids on fullillior-NDS-2115 CESystem				
Amino acids	ΔI			
L-Tryptophan	-1380			
L-Tyrosine	-1180			
L-Cysteine	-130			
L-Valine	+62			
L-Hitsidine	+95			
L-Aspartic acid	+105			
L-Threonine	+105			
L-Proline	+125			
L-phenylalanine	+125			
L-Cystine	+145			
L-Asparagine	+215			
L-Glycine	+255			
L-Isoleucine	+270			
L-Methionine	+285			
L-Leucine	+290			
L-Arginine	+320			
L-Serine	+335			

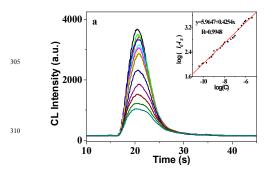
Concentration of amino acids: 1×10^{-6} g/mL, ΔI : net CL Intensity. CL conditions: luminol, 3.5×10^{-7} mol/L (0.02mol/L NaOH); NBS, 2.0×10^{-5} mol/L; ZnS QDs, 4.0×10^{-5} mol/L; Flow rate: 3.0 mL/min, negative high voltage: 450V. I_0 obtained by luminol-NBS-ZnS QDs CL system CL single intensity was 2160.

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Fig. 5 The Chemiuminescence inhibitory ratios of amino acids on this system.



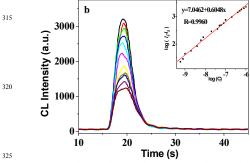


Fig. 6a Analysis characteristic curve of Trp (Illustration is linear relationship) and 6b Analysis characteristic curve of Tyr (Illustration is linear relationship).

Table 2. The recoveries of the determination of Trp in sample.

	Sample	Initially	Added	Found	Recovery
		present	(g/mL)	(g/mL)	(%)
		(g/mL)			
335	1	4.72×10 ⁻⁷	2.0×10 ⁻⁷	6.97×10 ⁻⁷	103.7
			3.0×10^{-7}	8.01×10^{-7}	103.8
	2	6.46×10 ⁻⁷	1.0×10 ⁻⁷	7.48×10^{-7}	100.3
			3.0×10^{-7}	9.41×10^{-7}	99.5
340	3	6.98×10^{-7}	1.0×10 ⁻⁷	8.05×10^{-7}	100.9
			2.0×10 ⁻⁷	8.85×10 ⁻⁷	98.6

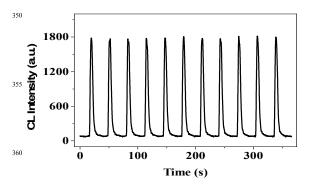


Fig. 7 Parallel determination (n=11) for Trp at 4.0×10^{-7} g mL⁻¹

365 Conclusion

In this paper, we successfully synthesized MPA functionalized ZnS QDs which could catalyze the luminol-NBS CL system. Through consulting relevant articles, UV-vis and CL spectra experiments, we explored the possible reaction and analysis, we found that Trp and Tyr could significantly inhibited the CL intensity. Based on this phenomenon, we could design a sensitive and rapid detection of Trp and Tyr sensor.

Acknowledgements

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References

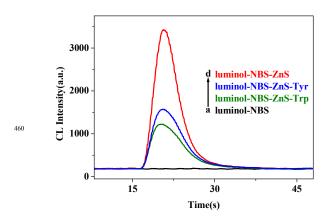
- J. S. Meyer, K. M. Welch, V. D. Deshmukh, F. I. Perez, R. H. Jacob, D.
 B. Haufrect, N. T. Mathew, R. M. Morrell, J. Am. Geriatr. Soc, 1977, 25, 289
 - 2 A. J. Gelenberg, C. J. Gisbon, J. D. Wojcik, *Psychopharmacol, Bull*, 1982, 18, 7.
- 3 Y. Zhu, Y. Yang, G. Zhou, M. Li, C. Jiang, C. Zhang, S. Chen, Food. 385 Chem, 2010, 118, 159.
 - 4 Q. Wang, A. Vasilescu, P. Subramanian, A. Vezeanu, V. Andrei, Y. Coffinier, M. Li, R. Boukherroub, S. Szunerits, *Electrochem. Commun*, 2013, **35**, 84.
- 5 M. Y. Wan, W. F. Li, Y. M. Long, Y. F. Tu, *Anal. Methods*, 2012, **4**, 90 2860.
- 6 M. Sa, L. Ying, A. G. Tang, L. D. Xiao, Y. P. Ren, *Clinica. Chimica. Acta*, 2012, **413**, 973.
- 7 J. Yang, S. Wang, J. Liu, A. Raghani, *J. Chromatogr. A*, 2007, **1156**, 174.
- 395 8 K. Lotte, R. Plessow, A. Brockhinke, *Photochem. Photobiol. Sci*, 2004, 3, 348.
 - 9 T. Miron, M. Wilchek, Anal. Biochem, 2013, 440, 12.

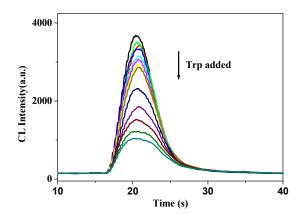
www.rsc.org/[JOURNAL] | [JOURNAL NAME HERE]

- 10 B. Arvidsson, N. Johannesson, A. Citterio, P. G. Righetti, J. Bergquist, J. Chromatogr. A, 2007, 1159, 154.
- 400 11 Y. Chi, J. Duan, S. Lin, G. Chen, Anal. Chem, 2006, 78, 1568.
 - 12 D. Du, J. Wang, J. N. Smith, C. Timchalk, Y. Lin, *Anal. Chem*, 2009, **81**, 9314.
 - 13 Z. P. Wang, J. Li, B. Liu, J. Q. Hu, X. Yao, J. H. Li, *J. Phys. Chem. B*, 2005, **109**, 23304.
- 405 14 C. Guo, H. J. Zeng, X. J. Ding, D. He, J. J. Li, R. Yang, L.B. Qu, J. Luminescence, 2013, 134, 888.
 - 15 X. Chen, C. Wang, X. Tan, J. Wang, Anal. Chim. Acta, 2011, 689, 92.
 - 16 B. Du, T. Wang, S. Han, X. Cao, T. Qu, F. Zhao, H. Yao. *Spectrochim. Acta. A*, 2015, **136**, 149.
- 410 17 Z. Y. Yan, A. Xiao, H. Lu, Z. Liu, J. Q. Chen. New. Carbon. Mater, 2014, 29, 216.
 - 18 X. Zhong, Y. Q. Chai, R. Yuan, Talanta, 2014, 128, 9.
 - 19 M. Deng, S. J. Xu, F. N. Chen, Anal. Methods, 2014, 6, 3117.
 - 20 S. F. Li, X. Z. Li, Y. Q. Zhang, F. Huang, F. F. Wang, X. W. Wei,
- 415 Microchim. Acta, 2009, 167, 103.
 - 21 Z. H. Song, S. Hou, J. Pharm. Biomed. Anal, 2002, 28, 683.
 - 22 J. X. Du, J. Quan, Y. D. Wang, *Talanta*, 2012, 90, 117.
 - 23 Z. P. Wang, Z. J. Zhang, Z. F. Fu, W. F. Luo, X. Zhang, *Talanta*, 2004, **62**, 611.
- 420 24 N. Wang, R. J. Zou, H. H. Chen, H. H. Chen, Y. G. Sun, J. H. Wu, Q. W. Tian, J. M. Yang, Z. G. Chen, J. G. Hu, *CrystEngComm*, 2011, 13, 1518.
 - 25 A. Khataee, A. Hasanzadeh, M. Iranifam, S. W. Joo. *Sensors. Actuat. B-Chem*, 2015, **215**: 272.
- 425 26 I. Costas-Mora, V. Romero, I. Lavilla, C. Bendicho. *Anal. Chim. Acta*, 2013, 788, 114.
 - 27 P. T. Shao, Q. H. Zhang, Y. G. Li, H. Z. Wang, J. Mater. Chem, 2011, 21, 151.
 - 28 M. Hosseini, M. R.Ganjali, Z. Vaezi, F. Faridbod, B. Arabsorkhi, M.
- 430 H. Sheikhha. Spectrochim. Acta. A, 2014, 121, 116.
 - 29 S. K. Panda, S. G. Hickey, H. V. Demir, A. Eychmüller, *Angew. Chem. Int. Ed*, 2011, **50**, 4432.
 - 30 J. L. Zhang, B. X. Li. Spectrochim. Acta. A, 2014, 125, 228.
 - 31 D. I. Son, H. H. Kim, D. K. Hwang, S. Kwon, W. K. Choi, J. Mater.
- 435 Chem. C, 2014, 2, 510.
 - 32 P. Wu, T. Zhao, S. L. Wang, X. D. Hou, Nanoscale, 2014, 6, 43.
 - 33 A. Mandal, A. Dandapat, G. De, Analyst, 2012, 137, 765.
 - 34 J. Kang, X. Li, J. Geng, L. Han, J. Tang, Y. Jin, Y. Zhang. *Food. Chem*, 2012, **134**, 2383.
- 440 35 X. Y. Cao , F. Shen , M. W. Zhang, J. X. Bie, X. Liu, Y. L. Luo, J. J. Guo, C. Y. Sun, RSC. Adv, 2014, 4, 16597.
 - 36 P. Wu, X. P. Yan, Chem. Soc. Rev, 2013, 42, 5489.
 - 37 W. Vogel, P. H. Borse, N. Deshmukh, S. K. Kulkarni, *Langmuir*, 2000, **16**, 2032.
- 445 38 B. Gilbert, F. Huang, Z. Lin, C. Goodell, H. Zhang, J. F. Banfield, *Nano. Lett.* 2006, 6, 605.

- L. Shen, X. Cui, H. Qi, C. Zhang, J. Phys. Chem. C, 2007, 111, 8172.
 S. F. Li, X. M. Zhang, W.X. Du, Y.H. Ni, X. W. Wei, J. Phys. Chem. C, 2009, 113, 1046.
- 450 41 G. Merenyi, J. Lind, T. E. Eriksen, *J. Biolumin. Chemilumin*, 1990, 5, 53.

455 Graphic Abstract:





ZnS QDs as a catalyst can catalyze luminol-NBS system CL, based on Trp and Tyr can inhibit this system CL intensity, we were designed a rapid and sensitive sensor for determination of Trp and Tyr.