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1 **Removal of methyl violet dye by adsorption onto *N*-benzyltriazole**
2 **derivatized dextran**

3

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22 **Abstract**

23

24 In this work, *N*-benzyltriazole derivatized dextran was evaluated for its potential as a novel carbohydrate-
25 based adsorbent for the removal of methyl violet dye from water. The modified dextran was synthesized by a
26 click reaction of pentynyl dextran and benzyl azide, and the structure was characterized with nuclear magnetic
27 resonance spectroscopy, elemental analysis, and scanning electron microscopy. Dextran was substituted with a
28 triazole-linked benzyl group. For decolorization of dyeing effluent, adsorption is a very effective treatment; here,
29 the driving force is based on hydrogen bonding, pi stacking, and electrostatic interaction between the methyl
30 violet dye and the *N*-benzyltriazole derivatized dextran. Batch experiments were carried out to investigate the
31 required contact time and the effects of pH, initial dye concentrations, and temperature. The experimental data
32 were analyzed with equilibrium isotherms including the Langmuir, Freundlich, and Temkin models. Based on
33 the Langmuir isotherm, the maximum adsorption capacity was determined to be 95.24 mg of dye per gram of
34 the adsorbent. The adsorption obeyed pseudo-second order kinetics, and a negative ΔG° value indicated
35 spontaneous adsorption in nature.

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44 **Keywords**

45 Methyl violet; Dye removal; Adsorption; *N*-benzyltriazole derivatized dextran; Click reaction.

46 1.Introduction

47 Dyes are widely used in textile and paper industries; however, their effluents cause environmental
48 pollution¹. Since dyes partially block transmission of sunlight into water, they can inhibit the photosynthesis of
49 aquatic plants and the growth of bacteria². Therefore, color is considered to be one obvious indicator of water
50 pollution. Due to their high solubility in water, the removal of dyes is difficult to achieve by conventional
51 physicochemical and biological treatment methods. Methyl violet is a member of a group of basic dyes, are
52 highly visible even at very low concentrations. Since it is also mutagen and mitotic poison, much attention has
53 been paid to developing remediation procedures for methyl violet pollution^{3,4}. Although various dye removal
54 methods have been reported in the literature, such as chemical- and photo-oxidation⁵, biological treatment⁶,
55 coagulation⁷, and adsorption, adsorption has particular advantages: no remaining toxic fragments are left behind,
56 it has flexibility and simplicity of design, and it is a highly effective, easy, and low-cost method. Activated
57 carbon is the most common material used to remove dye by adsorption but it is expensive and requires
58 reactivation of the carbon^{8,9}; inorganic compounds such as alum, polyaluminum chloride, and silica gel present
59 their own environmental problems¹⁰. Therefore, the development of eco-friendly polysaccharide-based
60 adsorbents would be of great importance. Although chitosan and its derivatives have been investigated for use in
61 the removal of several acidic dyes^{11,12}, there has been no report on the use of modified dextran for the removal
62 of toxic dyes from water.

63 Dextran is an extracellular polysaccharide of varying lengths (M_r 3-2000000), synthesized from
64 sucrose by *Leuconostoc mesenteroides*¹³. The general structure of dextran consists of a linear backbone of
65 repeating α -1,6-linked D -glucopyranosyl units branched with α -1,2, α -1,3, and α -1,4 linkages, and is
66 viscoelastic in water¹⁴. Since dextran is non-toxic, biocompatible, and inert in physiological conditions, they
67 have been targeted for use in some industrial fields, such as food, cosmetics, and pharmaceuticals^{15,16}. To
68 further the potential applications of dextran, it is possible to modify their hydroxyl groups with specific
69 substituents; for instance, cross-linked and carboxymethyl- and diethylaminoethyl-derivatized dextran have
70 been utilized as chromatographic separation resins¹⁷.

71 In the present study, dextran was modified with a benzyltriazole moiety and explored as functional
72 biopolymers for the methyl violet dye removal. In biomedical science, click chemistry has been used for DNA
73 conjugation and the synthesis of microcapsules and glycopolymers¹⁸. In this case, the triazole formed is stable

74 against reduction, hydrolysis, and oxidation. Given the physicochemical properties of the hydrophobic-
75 functionalized dextran, we investigated it for use in the adsorption of methyl violet. The driving force of the
76 adsorption process is considered as non-covalent bonds including hydrogen bonds, hydrophobic effects, π -
77 stacking interactions, and electrostatic interactions as well as the backbone structure of the polymer.
78 Furthermore, the effects of the contact time, initial dye concentration, initial solution pH, and reaction
79 temperature are presented and discussed. Equilibrium isotherms were analyzed according to the Langmuir,
80 Freundlich, and Temkin equations. A pseudo-second order kinetic model and thermodynamic parameters have
81 been determined.

82

83 **2. Experimental**

84 **2.1. Chemicals**

85 Methyl violet 2B and dextran ($M_r \sim 500000$, from *Leuconostoc* sp.) were purchased from Fluka
86 analytical and Sigma–Aldrich Co, respectively. Diethyl ether (99.5%) from J.T.Baker and dimethyl sulfoxide
87 (DMSO, 99.9%) from Sigma–Aldrich Co. were used as organic solvents. All other chemicals used were of
88 analytical reagent grade.

89

90 **2.2. Synthesis of dimsyl lithium**

91 A basic agent, dimsyl lithium ($\text{CH}_3\text{SOCH}_2^-\text{Li}^+$) introduced by Hakomori¹⁹, was prepared as follows.
92 In a round bottom flask dried by heating under nitrogen, 1.6 M methyllithium in diethyl ether was added to an
93 equal volume of DMSO. The mixture was stirred under nitrogen for 90 min, and the freshly prepared dimsyl
94 lithium was used immediately.

95

96 **2.3. Synthesis of *O*-(*N*-benzyl-[1,2,3]-triazoyl)-propyl dextran via a click reaction of pentynyl dextran with 97 benzyl azide**

98 Pentynyl dextran was synthesized using dimsyl lithium, 5-chloro-1-pentyne, and dextran as previously
99 described²⁰. Pentynyl dextran (5 g) was dissolved in DMSO/H₂O (4:1 v/v, 150 mL). After dissolution, benzyl
100 azide (2 equiv. per alkynyl group) was added, followed by a freshly prepared 1 M aqueous solution of sodium L-
101 ascorbate (20 mol % per alkynyl group) and CuSO₄·5H₂O (5 mol % per alkynyl group). The reaction mixture

102 was stirred at room temperature for 96 h²¹. The product was dialyzed (molecular weight cutoff (MWCO) >
103 10000) against deionized water and the final solution was lyophilized.

104

105 **2.4. Preparation of succinoglycan and N-benzyltriazole derivatized succinoglycan**

106 As an acidic polysaccharide, succinoglycan was isolated and purified from *Sinorhizobium meliloti*²².

107 N-benzyltriazole derivatized succinoglycan was synthesized by the same protocol described in section 2.3²³.

108

109 **2.5. Nuclear magnetic resonance (NMR) spectroscopy**

110 A Bruker Avance 500 spectrometer was used to record ¹H-NMR spectra. The samples were dissolved

111 in d₆-DMSO at room temperature.

112

113 **2.6. Scanning electron microscopy (SEM)**

114 To fix the samples on a brass stub, double-sided adhesive carbon tape was used. The samples were

115 coated with a thin gold layer at 30 W for 30 s in a vacuum. SEM images were acquired using a 20 kV

116 accelerating voltage on a JSM-6380 (Jeol, Japan) scanning electron microscope.

117

118 **2.7 Fourier-transform Infrared (FT-IR) Spectroscopy**

119 FT-IR Spectra were obtained in potassium bromide matrix by using Bruker IFS-66 spectrometer

120 (AMX, Germany). The spectra were recorded in the scanning range was 400–4000 cm⁻¹.

121

122 **2.8. Intraparticle diffusion study**

123 The possibility of intra-particle diffusion resistance affecting adsorption was explored by using the

124 intraparticle diffusion model as:

$$125 \quad q_t = k_{id} t^{1/2} + C \quad (1)$$

126 where k_{id} (mg/g·min^{1/2}) is the intraparticle diffusion rate constant and C (mg/g) is the boundary layer thickness

127 ²⁴.

128

129 **2.9. Activation energy and thermodynamics of adsorption**

130 Using an Arrhenius plot²⁵, the Arrhenius energy of activation (E_a) can be calculated as follows:

$$131 \ln k_2 = \ln A - E_a/RT \quad (2)$$

132 where A and R refer to the Arrhenius frequency factor and the gas constant respectively.

133 Using Eyring's equation (3), the enthalpy (ΔH^\ddagger) and entropy (ΔS^\ddagger) of activation were calculated from
134 the slope and intercept, respectively, of a plot of $\ln(k_2/T)$ versus $1/T$:

$$135 \ln k_2/T = \ln(k_B/h) + (\Delta S^\ddagger/R) - (\Delta H^\ddagger/RT) \quad (3)$$

136 where k_B and h are the Boltzmann and Planck constants, respectively.

137 The free energy of activation (ΔG^\ddagger) was obtained as follows:

$$138 \Delta G^\ddagger = \Delta H^\ddagger - T\Delta S^\ddagger \quad (4)$$

139 The spontaneity of the adsorption process was evaluated by measuring changes in the thermodynamic
140 parameters, namely the free energy change (ΔG^0 , kJ/mol), the enthalpy change (ΔH^0 , kJ/mol), and the entropy
141 change (ΔS^0 , J/mol·K). The values of ΔG^0 , ΔH^0 , and ΔS^0 were calculated by equations 5 and 6. A plot of $\ln K_c$ vs.
142 $1/T$ yields a straight line with $-\Delta H^0/R$ and $\Delta S^0/R$ as its slope and intercept, respectively.

$$143 \Delta G^0 = -RT \ln K_c \quad (5)$$

$$144 \ln K_c = \Delta S^0/R - \Delta H^0/RT \quad (6)$$

145 where K_c , the distribution coefficient of the adsorbate, is equal to q_e/C_e .

146

147 3. Results and discussion

148 3.1. *N*-benzyltriazole derivatized dextran and its structural analysis

149 A structural analysis of polysaccharides and information about substituent distribution in their
150 derivatives may offer the most fundamental understanding about the functions and properties of polysaccharides.
151 Dextran and its derivatives have been investigated as the critical polysaccharides for medical and industrial
152 applications²⁶. In the present study, we evaluated the prepared *O*-(*N*-benzyl-[1,2,3]-triazoyl)-propyl dextran as a
153 carbohydrate-based adsorbent for the dye removal. *N*-benzyltriazole derivatized dextran was synthesized via a
154 click reaction of pentynyl dextran and benzyl azide. The primary dextran derivative, pentynyl dextran, allowed
155 the introduction of aromaticity in the second reaction step (a copper-catalyzed azide-alkyne cycloaddition). The

156 resulting *N*-benzyltriazole derivatized dextran was evaluated for the adsorption of methyl violet which has
157 aromatic ring structures with heteroatoms.

158 *N*-benzyltriazole derivatized dextran was characterized by NMR, elemental analysis (EA), and SEM.
159 As shown in the ¹H NMR spectrum (Fig. 1A), aromatic protons and protons at the 2'-position of the benzyl
160 group had peaks located at 7.29 and 5.49 ppm, respectively. Glucose ring protons appear in the range of 3.00-
161 4.00 ppm, and the hydroxyl protons are visible at 4.46 ppm (OH-2), 4.83 ppm (OH-3), and 4.90 ppm (OH-4).
162 Some signals between 1.00 and 3.00 ppm are due to the resonance from residual pentynyl groups²³. The EA
163 results of *N*-benzyltriazole derivatized dextran and the parent unsubstituted dextran are summarized in the
164 supporting material, in Table S1. Chemical modifications led to a significant increase in the carbon and nitrogen
165 contents of *N*-benzyltriazole derivatized dextran compared with those of parent dextran. Based on the nitrogen
166 contents, the degree of substitution (DS) value was determined to be 0.56. The spherical morphology of the
167 parent dextran was also changed into an uneven state after modification (Fig. 2A and Fig. S2). These structural
168 analyses indicated that the novel adsorbent of dye, *N*-benzyltriazole derivatized dextran was successfully
169 synthesized.

170

171 **3.2. Adsorption isotherm analysis**

172 Table 1 summarizes the Langmuir, Freundlich, and Temkin isotherm constants for the adsorption of
173 methyl violet 2B onto *N*-benzyltriazole derivatized dextran. The applicability of the isotherm equation was
174 evaluated through the correlation coefficients, r^2 (Table 1 and Fig. S3). The best fit, with an r^2 value of 0.9815,
175 was obtained from the Langmuir isotherm model. Langmuir adsorption quantitatively describes the formation of
176 an adsorbate monolayer on the outer surface of the adsorbent, after which no further adsorption takes place²⁷.
177 The maximum amount of the dye adsorbed from the aqueous solution, q_m , was 95.24 mg/g; the constant
178 associated with the binding energy of the sorption system, K_L , was 0.16 L/mg. Other natural adsorbents such as
179 bagasse fly ash, sugarcane dust, and sunflower seed hull have shown q_m values in the range of 25-93 mg/g for
180 methyl violet adsorption (Table S2)^{3, 4, 28}. These agricultural materials are localized, and their structural
181 information in the literature is deficient. In these respects, modified polysaccharides can be developed with
182 advantage, and applied for the conventional methods of dye removal from water. The separation factor, R_L ,
183 calculated from the Langmuir constant, indicates the nature of the isotherm. The values of R_L in the present

184 investigation were found to range from 0.286 to 0.032 for initial dye concentrations ranging from 10 to 200 ppm
185 (Fig. S4). Higher R_L values at lower dye concentrations showed that the adsorption was more favorable at lower
186 initial adsorbate concentrations; however, all of the R_L values indicate favorable adsorptions, as they all lie in
187 the range of $0 < R_L < 1$. This favorable adsorption may be attributed to van der Waals forces and hydrogen
188 bonding between methyl violet 2B and *N*-benzyltriazole derivatized dextran, as well as π - π interactions between
189 the aromatic residues of the dye and the adsorbent.

190 Freundlich adsorption isotherms are commonly used to describe the adsorption characteristics of
191 heterogeneous surfaces. The extent of adsorption varies directly depending on the pressure until the saturation
192 pressure is reached. The constant K_f and n are the empirical parameters which must be determined by data
193 fitting²⁹. The constant K_f is a rough indicator of the adsorption capacity, and $1/n$ is the adsorption intensity.
194 Chemical adsorption is expected if $1/n < 1$, while if $n = 1$, the partition between the two phases is independent of
195 the concentration, and $1/n > 1$ indicates that the adsorption is a favorable physical process³⁰. Therefore, the $1/n$
196 value of 2.77 suggests that the adsorption of methyl violet 2B onto *N*-benzyltriazole derivatized dextran is a
197 favorable physical process.

198 Temkin's adsorption isotherm model takes into account the adsorbate/adsorbent interaction, and
199 assumes that the heat of adsorption of all molecules in the layer will decrease linearly with increased surface
200 coverage³¹. It predicts a uniform distribution of binding energies over the whole population of surface binding
201 sites. Theoretically, such a uniform distribution of binding energies would arise from a truly random
202 arrangement of surface binding sites. According to the Temkin plot of adsorption data collected in this
203 experiment ($r^2 = 0.9433$), the constants were calculated as $A_T = 1.37$ L/g and $B = 22.73$ J/mol. Since the heat of
204 sorption lower than 20 kJ/mol is characteristics for physisorption³².

205

206 **3.3. Effect of initial dye concentration on methyl violet 2B removal**

207 The effect of the initial dye concentration on its subsequent removal was studied (Fig. 3). The
208 experiments were performed at different initial dye concentrations (12.5, 25, and 50 ppm) with *N*-benzyltriazole
209 derivatized dextran acting as the adsorbent (1 g/L). In the case of the initial concentration of 12.5 ppm, 89% of
210 the dye was removed from water at 25 °C within 30 min. At a 50 ppm initial dye concentration, 57% was
211 removed after 2 h. Using the graph shown in Fig. 3B, the rate constants of the pseudo-second order, k_2 (g/mg

212 min), and amounts of dye adsorbed at equilibrium, q_e (mg/g), were determined to be: $k_2 = 0.0515$ (12.5 ppm),
213 0.0271 (25 ppm), and 0.0042 (50 ppm); $q_e = 11.05$ (12.5 ppm), 23.47 (25 ppm), 30.21 (50 ppm). At a low initial
214 concentration, the adsorption of dye by the adsorbent is very intense and reaches equilibrium very quickly. For
215 the diffusion mechanism, the kinetic results were further analyzed by the intraparticle diffusion model. Fig. 3C
216 shows plots of q_t vs. $t^{1/2}$ at different initial dye concentration, and they are presented as at least two linear graphs.
217 The results indicate that two or more steps simultaneously operate the adsorption process³³. In addition, the
218 presence of intercept implies that the intraparticle diffusion is not the only rate-limiting step³⁴. The first portion
219 represents boundary layer diffusion through the bulk liquid to the external surface of the adsorbent. The second
220 linear phase is a gradual adsorption stage through the intraparticle diffusion. The intraparticle diffusion rate
221 constants were calculated as: K_{id} (mg/g·min^{1/2}) = 0.1274 (12.5 ppm), 0.1372 (25 ppm), and 1.0389 (50 ppm).
222 Therefore, we suggest that both surface adsorption and intraparticle diffusion affect the present adsorption
223 mechanism.

224

225 3.4. Comparison with *N*-benzyltriazole derivatized succinoglycan

226 Considering that methyl violet 2B is a basic dye (Fig. 4A), it is meaningful to investigate the
227 contribution of acidic groups to its adsorption. Since succinoglycan is an anionic glucan containing succinyl and
228 pyruvyl substituents²², *N*-benzyltriazole derivatized succinoglycan was prepared via a click reaction for the
229 comparison with *N*-benzyltriazole derivatized dextran. Figs. 4B and 4C show the unmistakable effectiveness of
230 *N*-benzyltriazole derivatized dextran as an adsorbent for the removal of methyl violet, with residual color less
231 than 5%. Parent dextran showed a better result than the unmodified succinoglycan, and *N*-benzyltriazole
232 derivatized succinoglycan gave a removal improvement of 30% over unmodified succinoglycan. In a previous
233 study, the removal of another class of organic contaminant, polycyclic aromatic hydrocarbons (PAH), was
234 attributed to the exposed aromatic cores in the modified pine bark³⁵. The aromatic cores resulted in a stronger
235 specific π - π interaction between the PAH and the adsorbent. Based on the EA of the tested polysaccharides
236 (Table S1), the values of (N+O)/C indicating polarity, decreased after modification from 0.96 and 0.92 to 0.50
237 and 0.50 for succinoglycan and dextran, respectively. The aliphatic character (H/C) of each also decreased from
238 2.06 and 1.98 to 1.46 and 1.37, respectively. Thus, aromaticity is not the only deciding factor for adsorption; the
239 polysaccharide backbone structure is also important.

240 As another key, the surface morphology of *N*-benzyltriazole derivatized dextran, with its roughness,
241 supports the effective adsorption of dye (Fig. 2A). The craterous surface structure of *N*-benzyltriazole
242 derivatized dextran may allow methyl violet 2B to be trapped and adsorbed into its holes³⁶. Conversely, *N*-
243 benzyltriazole derivatized succinoglycan shows a laminar sheet-like surface²³, and parent dextran has a
244 spherical structure with the average diameter of 20.03 μm (Fig. S2). In reality, the uneven surface of the *N*-
245 benzyltriazole derivatized dextran was filled after the adsorption of dye, as shown in Fig. 2B. Morphological
246 changes were also observed for the adsorption by other adsorbents^{4,37}. From the plot profiles of the selected 50
247 μm line in the SEM data (according to Fiji image processing package), the 8 potholes of 2-5 μm width and the
248 flattened surface are described in blue and red line, respectively (Fig. 2C). The gray scale can be considered as
249 an underestimation due to the image treatment software effect, however the degree of surface roughness is
250 clearly differentiated. These results indicate that various non-covalent bonds including π - π interaction and its
251 uneven surface work at the present dye adsorption.

252

253 **3.5. FT-IR spectra of *N*-benzyltriazole derivatized dextran/methyl violet 2B composite**

254 After adsorption methyl violet 2B onto *N*-benzyltriazole derivatized dextran, the precipitated
255 composite was analyzed with FT-IR spectroscopy. Fig. 5 shows shifts or changes of IR absorption peaks,
256 indicating interactions of the adsorbent and methyl violet. All the observed changes are summarized in Table S3,
257 and the characteristic change is present at $\text{C}\equiv\text{C}$ and aromatic $\text{C}=\text{C}$ stretching vibration. A disappearance in the
258 $\text{C}\equiv\text{C}$ stretch may be attributed to the electrostatic interaction of tertiary ammonium ion of methyl violet with the
259 terminal alkyne group of *N*-benzyltriazole derivatized dextran (Fig. 1). A novel peak (1587 cm^{-1}) belong to
260 methyl violet 2B was formed, and band shifts took place from 1643 to 1637 cm^{-1} . This result demonstrates the
261 additional function of residual alkyne group in *N*-benzyltriazole derivatized dextran for electrostatic interaction
262 with methyl violet. Similar observations have also been reported in methylene blue/exfoliated graphene oxide
263 composite³⁸.

264

265 **3.6. Effect of pH on methyl violet 2B removal**

266 Adsorption processes are affected by solution pH, as it influences not only the surface charge of the
267 adsorbent but also the chemistry of dye in solution³⁹. We depict the effect of pH on dye removal by *N*-

268 benzyltriazole derivatized dextran (Fig. 6); we found that the dye adsorption decreased as the pH increased.
269 Under alkaline condition, the hydrogen bonds stabilizing the adsorbate-adsorbent interaction can be destroyed,
270 and the *N*-benzyltriazole derivatized dextran may adopt another conformation because of charge repulsion.
271 Whereas the opposite trend was observed in the adsorption of basic dye onto sunflower seed hull and bagasse
272 fly ash^{3,4}.

273

274 **3.7. Effect of temperature, activation energy, and a thermodynamic study**

275 The adsorption studies were carried out at different temperatures, ranging from 15 to 75 °C (Table 2
276 and Fig. S5). The adsorption capacity and the rate constant were found to increase with increasing temperature,
277 indicating that the adsorption is an endothermic process. This may be due to an increase in the mobility of the
278 dye or a change of active sites within the internal structure of the *N*-benzyltriazole derivatized dextran as the
279 temperature increases. The present dye adsorption follows the pseudo-second order model with the best
280 correlation coefficient ($r^2 > 0.99$); using the rate constant for pseudo-second order adsorption (k_2) at several
281 temperatures, the energy of activation (E_a) was calculated to be 33.96 kJ/mol from an Arrhenius plot (Fig. S6A
282 and Table 2). In general, activation energies less than 40 kJ/mol indicate a physisorption process, while those
283 higher than 40 kJ/mol signify chemisorption⁴⁰. Thus, the present adsorption of methyl violet 2B by *N*-
284 benzyltriazole derivatized dextran can be regarded as predominantly physisorption.

285 Table 3 shows thermodynamic parameters as obtained from the intercept and slope of a plot of $\ln K_c$
286 vs. $1/T$ (Fig. S7). The positive ΔH° value, 16.47 kJ/mol, also indicates that the adsorption is an endothermic
287 process; furthermore, the magnitude of this value provides information about the type of sorption. In principle,
288 the heat evolved during physical adsorption is of the same order of magnitude as the heat of condensation, i.e.,
289 2.1-20.9 kJ/mol, while the heat of chemisorption generally falls in the range of 80-200 kJ/mol⁴¹. Although the
290 low value of ΔS° implies that no remarkable change in entropy occurred during adsorption, the positive value
291 may be attributed to an increased randomness at the solid-solution interface during adsorption. The negative
292 values of ΔG° (-3.90, -5.61, and -6.71 kJ/mol) at the given temperatures (288, 308, and 328 K) indicate that the
293 adsorption processes at all three recorded temperatures are spontaneous.

294

295 **4. Conclusion**

296 In the present study, *N*-benzyltriazole derivatized dextran was successfully synthesized by a click
297 reaction. It was then evaluated for its effectiveness as a polysaccharide adsorbent for the removal of methyl
298 violet dye from water. Equilibrium and kinetic studies were carried out for the adsorption of methyl violet onto
299 *N*-benzyltriazole derivatized dextran. The monolayer adsorption capacity was determined to be 95.24 mg of dye
300 per gram of *N*-benzyltriazole derivatized dextran. The adsorption efficiency was found to be dependent on the
301 initial dye concentration, solution pH, adsorbate contact time, and temperature. The adsorption kinetics followed
302 a pseudo-second order model. A thermodynamic study supported the theory that the adsorption of methyl violet
303 2B onto *N*-benzyltriazole derivatized dextran is a spontaneous and endothermic process at the studied
304 temperatures. We suggest that methyl violet removal by *N*-benzyltriazole derivatized dextran is caused by
305 physical adsorption based on hydrogen bonding, van der Waals interactions, and π -stacking interactions, and
306 electrostatic interactions. The present study indicates great potential for the removal of cationic dyes from
307 aqueous solutions using natural biomaterials. Furthermore, the practical use of tailor made
308 polysaccharide-derivative adsorbents to remove various dye and organic pollutants as target chemicals
309 could be expected in the environmental field.

310

311

312 **Acknowledgements**

313 This paper was supported by the KU Research Professor Program of Konkuk University. This research was also
314 supported by the National Research Foundation of Korea Grant, funded by the Korean Government (NRF-
315 2013R1A1A2012568 and NRF-2011-619-E0002), and by the Priority Research Centers Program through the
316 National Research Foundation of Korea (NRF), funded by the Ministry of Education, Science, and Technology
317 (2012-0006686). SDG.

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320 **References**

- 321 1. Y. Anjaneyulu, N. S. Chary and D. S. S. Raj, *Reviews in Environmental Science and Bio/Technology*,
322 2005, **4**, 245-273.
- 323 2. N. Koprivanac and H. Kusic, *Hazardous organic pollutants in colored wastewaters*, Nova Science
324 Publishers Hauppauge, 2009.
- 325 3. I. D. Mall, V. C. Srivastava and N. K. Agarwal, *Dyes and pigments*, 2006, **69**, 210-223.
- 326 4. B. Hameed, *Journal of hazardous materials*, 2008, **154**, 204-212.
- 327 5. M. S. Lucas and J. A. Peres, *Dyes and Pigments*, 2006, **71**, 236-244.
- 328 6. H. S. Rai, M. S. Bhattacharyya, J. Singh, T. Bansal, P. Vats and U. Banerjee, *Critical Reviews in*
329 *Environmental Science and Technology*, 2005, **35**, 219-238.
- 330 7. B. Shi, G. Li, D. Wang, C. Feng and H. Tang, *Journal of Hazardous Materials*, 2007, **143**, 567-574.
- 331 8. Y. S. Al-Degs, M. I. El-Barghouthi, A. H. El-Sheikh and G. M. Walker, *Dyes and Pigments*, 2008, **77**,
332 16-23.
- 333 9. G. Walker and L. Weatherley, *Water Research*, 1997, **31**, 2093-2101.
- 334 10. S. Lambert, N. Graham, C. Sollars and G. Fowler, *Water science and technology*, 1997, **36**, 173-180.
- 335 11. V. Dhanapal and K. Subramanian, *Carbohydrate polymers*, 2014.
- 336 12. G. Crini and P.-M. Badot, *Progress in polymer science*, 2008, **33**, 399-447.
- 337 13. A. Jeanes, C. A. Wilham and J. C. Miers, *The Journal of biological chemistry*, 1948, **176**, 603-615.
- 338 14. A. Misaki, M. Torii, T. Sawai and I. J. Goldstein, *Carbohydrate research*, 1980, **84**, 273-285.
- 339 15. M. Naessens, A. Cerdobbel, W. Soetaert and E. J. Vandamme, *Journal of Chemical Technology and*
340 *Biotechnology*, 2005, **80**, 845-860.
- 341 16. R. Mehvar, *Journal of controlled release*, 2000, **69**, 1-25.
- 342 17. B. Porsch and L.-O. Sundelöf, *Journal of Chromatography A*, 1994, **669**, 21-30.
- 343 18. K. Nwe and M. W. Brechbiel, *Cancer Biotherapy and Radiopharmaceuticals*, 2009, **24**, 289-302.
- 344 19. S.-I. HAKOMORI, *Journal of Biochemistry*, 1964, **55**, 205-208.
- 345 20. M. N. Tahir, C. Bork, A. Risberg, J. C. Horst, C. Komoß, A. Vollmer and P. Mischnick,
346 *Macromolecular Chemistry and Physics*, 2010, **211**, 1648-1662.

- 347 21. P. F. Tankam, P. Mischnick, H. Hopf and P. G. Jones, *Carbohydrate research*, 2007, **342**, 2031-2048.
- 348 22. L.-X. Wang, Y. Wang, B. Pellock and G. C. Walker, *Journal of bacteriology*, 1999, **181**, 6788-6796.
- 349 23. E. Cho, K. Kim, M. N. Tahir, J. Y. Lee and S. Jung, *Notes*, 2014, **35**, 2589.
- 350 24. W. Weber and J. Morris, *J. Sanit. Eng. Div. Am. Soc. Civ. Eng.*, 1963, **89**, 31-60.
- 351 25. J. W. Grissom, T. L. Calkins, H. A. McMillen and Y. Jiang, *The Journal of organic chemistry*, 1994, **59**,
- 352 5833-5835.
- 353 26. D. de Belder, *Polysaccharides in Medicinal Applications. New York, NY: Marcel Dekker*, 1996, 505-
- 354 524.
- 355 27. I. Langmuir, *Journal of the American Chemical Society*, 1918, **40**, 1361-1403.
- 356 28. Y.-S. Ho, W.-T. Chiu and C.-C. Wang, *Bioresource technology*, 2005, **96**, 1285-1291.
- 357 29. A. Akgerman and M. Zardkoobi, *Journal of Chemical & Engineering Data*, 1996, **41**, 185-187.
- 358 30. S. V. Mohan and J. Karthikeyan, *Environmental pollution*, 1997, **97**, 183-187.
- 359 31. R. D. Johnson and F. H. Arnold, *Biochimica et Biophysica Acta (BBA)-Protein Structure and Molecular*
- 360 *Enzymology*, 1995, **1247**, 293-297.
- 361 32. P. Atkins, Oxford University Press, 1998.
- 362 33. L. Ai, M. Li and L. Li, *Journal of Chemical & Engineering Data*, 2011, **56**, 3475-3483.
- 363 34. S. Ghorai, A. Sarkar, M. Raoufi, A. B. Panda, H. Schönherr and S. Pal, *ACS applied materials &*
- 364 *interfaces*, 2014, **6**, 4766-4777.
- 365 35. Y. Li, B. Chen and L. Zhu, *Bioresource technology*, 2010, **101**, 7307-7313.
- 366 36. H. L. Parker, A. J. Hunt, V. L. Budarin, P. S. Shuttleworth, K. L. Miller and J. H. Clark, *RSC Advances*,
- 367 2012, **2**, 8992-8997.
- 368 37. S. Dawood and T. K. Sen, *Water research*, 2012, **46**, 1933-1946.
- 369 38. G. Ramesha, A. V. Kumara, H. Muralidhara and S. Sampath, *Journal of colloid and interface science*,
- 370 2011, **361**, 270-277.
- 371 39. N. M. Mahmoodi, B. Hayati, M. Arami and C. Lan, *Desalination*, 2011, **268**, 117-125.
- 372 40. T. Anirudhan and P. Radhakrishnan, *The Journal of Chemical Thermodynamics*, 2008, **40**, 702-709.
- 373 41. Y. Liu and Y.-J. Liu, *Separation and Purification Technology*, 2008, **61**, 229-242.

374

375 **Figure Legends**

376

377 **Fig. 1.** ¹H-NMR spectra of (A) *O*-(*N*-benzyl-[1,2,3]-triazoyl)-propyl dextran and (B) dextran. The left insets
378 show the chemical structures of *N*-benzyltriazole derivatized dextran and dextran.

379

380 **Fig. 2.** SEM images of (A) *N*-benzyltriazole derivatized dextran and (B) *N*-benzyltriazole derivatized dextran
381 with dye adsorbed. (C) Plot profiles of the selected yellow line (blue line : *N*-benzyltriazole derivatized dextran;
382 red line : *N*-benzyltriazole derivatized dextran with dye adsorbed). Black arrows indicate the gap width.

383

384 **Fig. 3.** (A) The effect of initial dye concentration. (B) Pseudo-second order kinetic plots on the removal of
385 methyl violet 2B by adsorption onto *N*-benzyltriazole derivatized dextran. (C) Intraparticle diffusion plots for
386 the adsorption of methyl violet onto *N*-benzyltriazole derivatized dextran. (25 °C; 1g/L adsorbent)

387

388 **Fig. 4.** (A) The chemical structure of methyl violet 2B. (B) Color changes and (C) degree of residual color after
389 reaching adsorption equilibrium. Polysaccharide adsorbents: a, no addition; b, succinoglycan; c, *N*-
390 benzyltriazole derivatized succinoglycan; d, dextran; e, *N*-benzyltriazole derivatized dextran. (10 g/L adsorbent)

391

392 **Fig. 5.** FT-IR spectra of *N*-benzyltriazole derivatized dextran before (in red) and after adsorption (in blue).

393

394 **Fig. 6.** Effect of pH on dye removal by *N*-benzyltriazole derivatized dextran. (100 ppm dye, 10 g/L adsorbent;
395 buffers used: pH 2, HCl-KCl buffer; pH 4, citrate buffer; pH 7, phosphate buffer; pH 9, borate buffer; pH 10,
396 NaHCO₃·NaOH buffer; pH 11, NaHCO₃·NaOH buffer)

397

398 **Tables and Figures**

399

400 **Table 1.** Isotherm constants for methyl violet adsorption onto *N*-benzyltriazole derivatized dextran at 25 °C.

401 (Adsorbent dose: 5g/L)

Langmuir isotherm constant		
K_L (L/mg)	q_m (mg/g)	r^2
0.16	95.24	0.9815
Freundlich isotherm constant		
K_f (mg/g)	n	r^2
16.04	0.36	0.9787
Temkin isotherm constant		
B (J/mol)	A_T (L/g)	r^2
22.73	1.37	0.9433

402

403 **Table 2.** Activation parameters for dye adsorption onto *N*-benzyltriazole derivatized dextran. (25 ppm dye; 1g/L
404 adsorbent)

Temperature (°C)	k_2 (mg/g min)	E_a (kJ/mol)	r^2	ΔH^\ddagger (kJ/mol)	ΔS^\ddagger (J/mol K)	ΔG^\ddagger (kJ/mol)	r^2
15	0.0031					84.19	
35	0.0068					87.86	
45	0.0148	33.96	0.9602	31.33	-183.548	89.70	0.9531
55	0.0221					91.53	
75	0.0310					95.20	

405

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410

411 **Table 3.** Thermodynamic parameters of dye adsorption onto *N*-benzyltriazole derivatized dextran. (25 ppm dye;
412 1 g/L adsorbent)

413

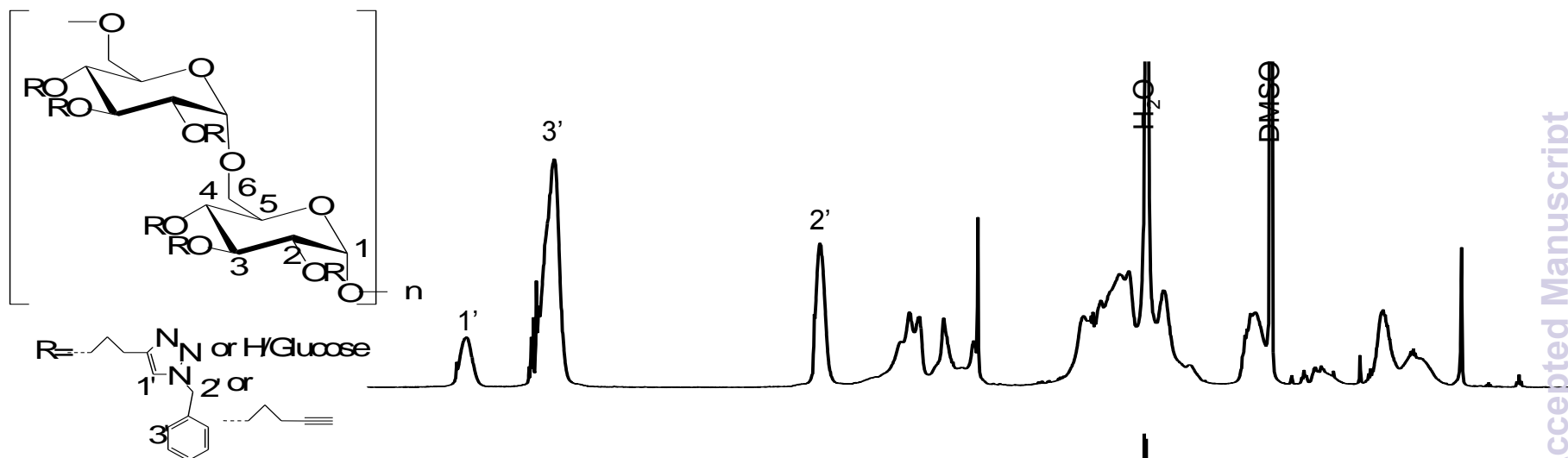
Temperature (K)	K_c	ΔG° (kJ/mol)	ΔH° (kJ/mol)	ΔS° (J/mol·K)	r^2
288	1.6293	-3.8993			
308	2.1905	-5.6067	16.4677	71.0308	0.9746
328	2.4629	-6.7130			

414

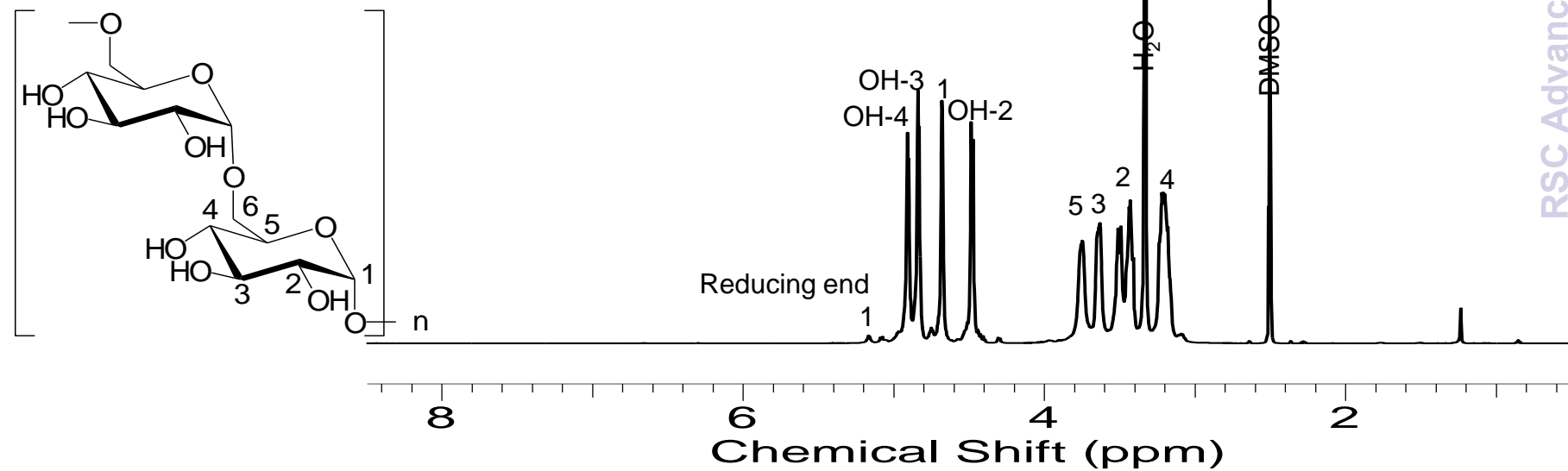
415

Fig. 1.

(A)



(B)



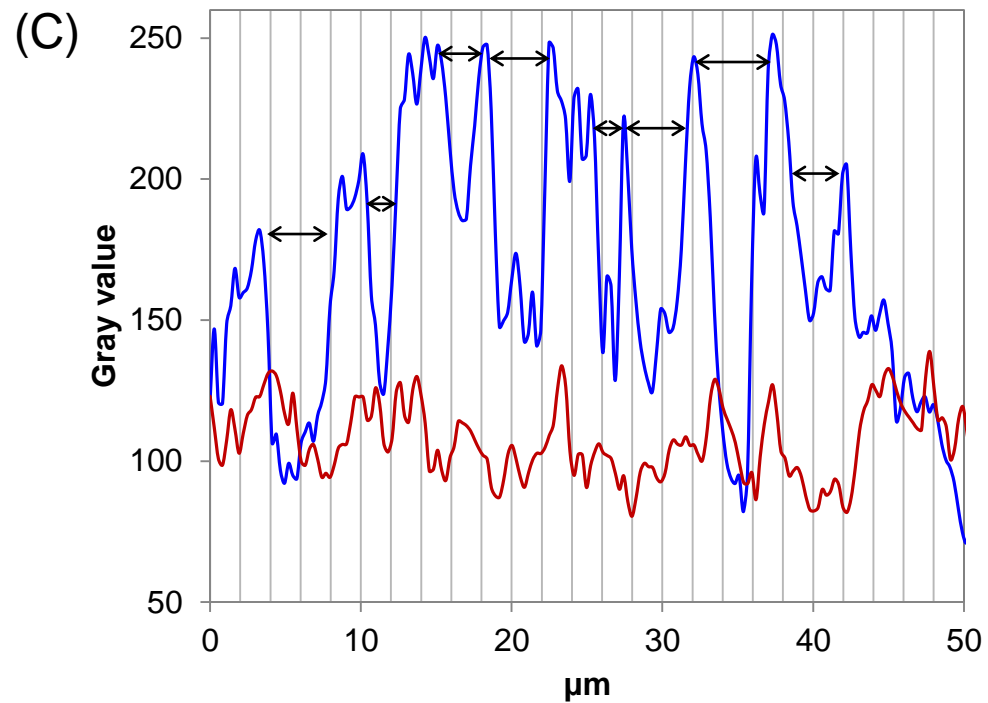
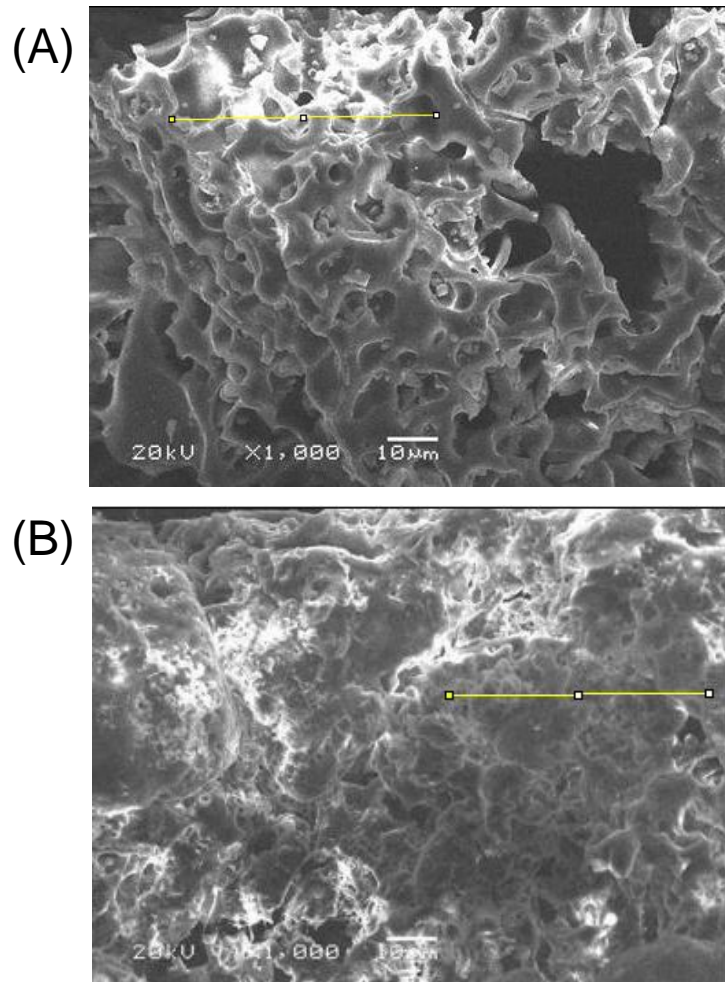
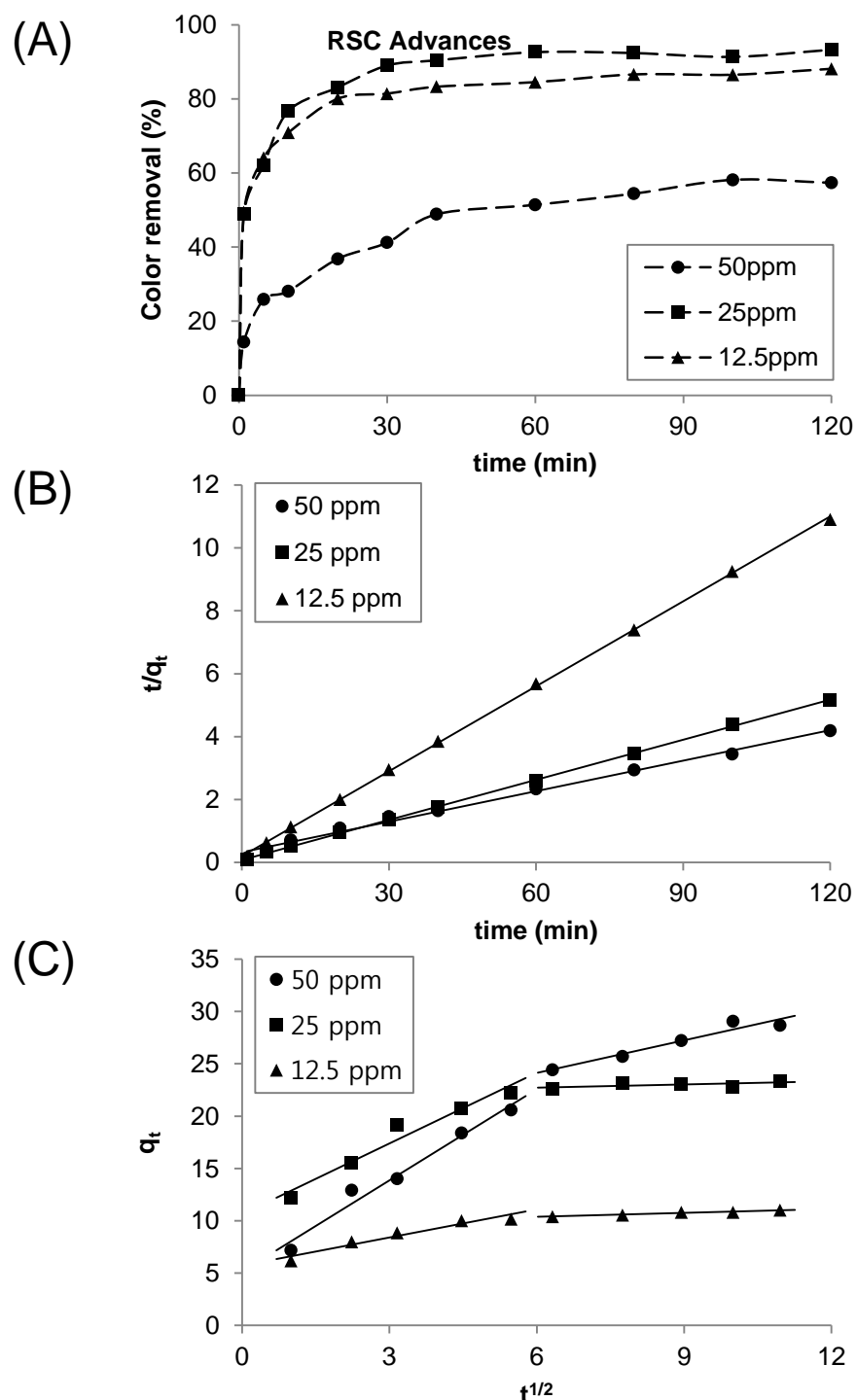
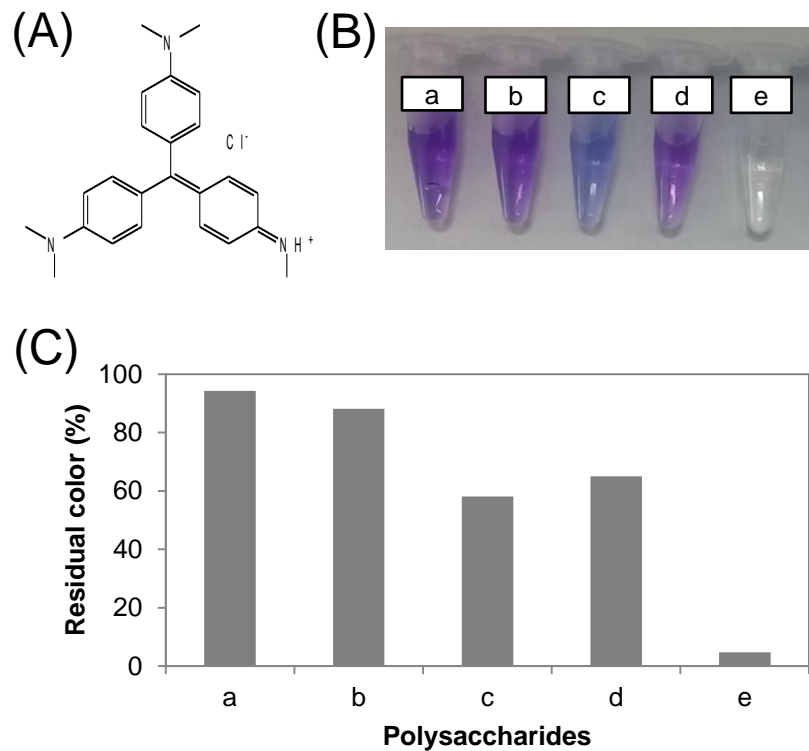


Fig. 3.





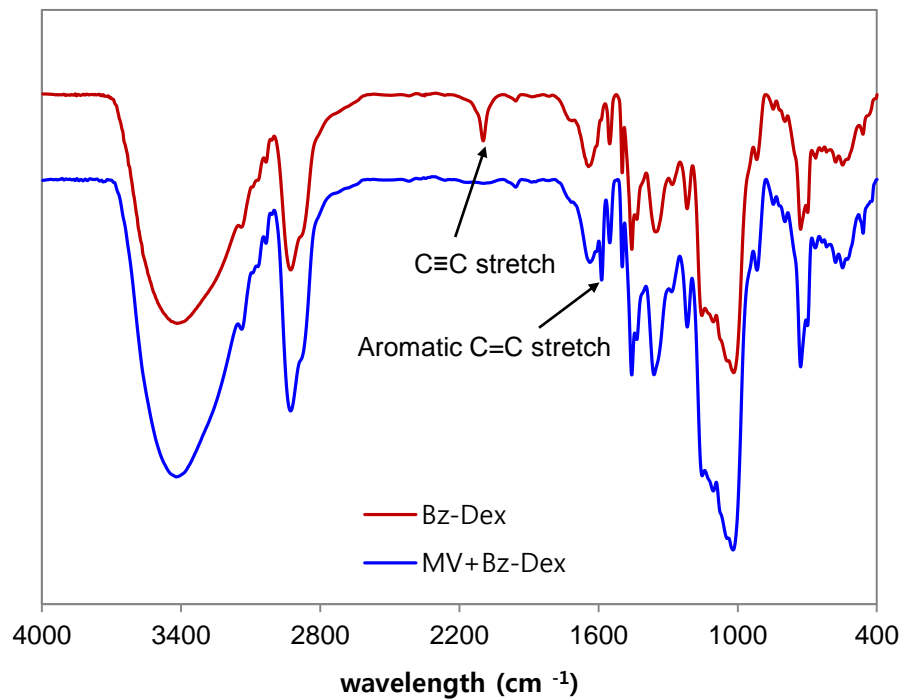


Fig. 6.

