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Synthesize of Ag-HNTs-MnO₂ Nanocomposites and Its Application for Nonenzymatic Hydrogen Peroxide Electrochemical Sensing

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Abstract: Natural halloysite nanotubes (HNTs) were attached to the flower-like MnO₂ and HNTs-MnO₂ composites were obtained, then silver nanoparticles were successfully deposited on the surface of HNTs-MnO₂ to produce Ag-HNTs-MnO₂ nanocomposites and they were used for fabricating nonenzymatic hydrogen peroxide (H₂O₂) sensor. Scanning electron microscopic, energy-dispersive X-ray spectrum, transmission electron microscope and Fourier transform infrared spectrum were applied to investigate the structures and morphologies of the resultant samples. The Ag-HNTs-MnO₂ composite-based modified electrode exhibited high electrocatalytic activity to the reduction of H₂O₂ with a linear range of 2.0 μM to 4.71 mM, a detection limit of 0.7 μM (S/N = 3) and a sensitivity of 11.9 μA mM⁻¹ cm⁻². In addition, high specific surface area, low cost and good biocompatibility made the modified electrode have a bright perspective in biosensors and biocatalysis.

Keywords: Nonenzymatic sensor; MnO₂; Halloysite nanotubes; Hydrogen peroxide

1. Introduction

Accurate detection of hydrogen peroxide (H₂O₂) became increasingly important because it can not only serve as an oxidizing agent in a general industrial process, but also have a great significance for food, industrial, pharmaceutical, clinical and environmental analysis [1-4]. Among the techniques for the detection of H₂O₂, including spectrophotometry [5], titrimetry [6], chromatography [7], electrochemistry [8], and chemiluminescence [9, 10], electrochemical technique based on a simple and low cost electrode has been extensively applied for the detection of H₂O₂ [11-13]. Traditionally, the electrochemical sensor includes enzyme sensors and non-enzyme sensors. Although, many enzymatic H₂O₂ assays possess good sensitivity and selectivity, they are environmentally unstable and comparatively expensive [14].

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1 Compared with enzymatic H_2O_2 assays, nonenzymatic assays that employ metal
2 oxides and their composites are more stable, very easy to synthesize, quite cost
3 effective, even at high temperature and needs very less maintenance [15, 16]. In the
4 light of these characteristics, how to develop enzyme-free H_2O_2 sensors with low
5 detection limit and wide responding range have been paid increasing attention.

6 Nowadays, with the development of nanotechnology, metal nanoparticles (NPs)
7 have been widely used to fabricate enzyme-free H_2O_2 sensors due to their unique
8 properties of biocompatibility, catalysis and low toxicity. As a typical nanomaterial,
9 silver nanoparticle (Ag NP) exhibits excellent physicochemical properties and shows
10 good catalytic activity toward the reduction of H_2O_2 [17, 18]. For these reasons, many
11 researchers have been synthesized silver nanoparticles to fabricate H_2O_2 sensors. Ag
12 NPs and multiwalled carbon nanotubes were combined by Li, and the obtained
13 functionalized composites were applied to fabricate a novel nonenzymatic H_2O_2
14 sensor [19]. Lu et al. synthesized multilayer films of polyelectrolyte/Ag NPs through
15 the method of layer-by-layer self-assembling for enzymeless H_2O_2 sensing [20]. Wang
16 et al. prepared nonenzymatic hydrogen peroxide sensor based on the electrodeposition
17 of silver nanoparticles on poly (ionic liquid)-stabilized graphene sheets [21].

18 In view of the above researches, it can be seen that homogeneously dispersed silver
19 nanoparticles are extremely significant in fabrication of H_2O_2 sensors and some
20 available substrates are necessary to prevent the aggregation of Ag NPs. Commonly
21 used materials include carbon nanomaterials, metallic oxide and polymeric membrane
22 et al. [19-21]. Nowadays, metallic oxides (TiO_2 [22], SiO_2 [23], CuO [24], Fe_3O_4 [25],
23 etc.) which display a lot of advantages such as low cost, simple synthesis, unique
24 electrochemical and optic properties have been attracted much attentions. Among
25 them, manganese dioxide (MnO_2) is considered as one of the candidates on account of
26 its low cost, high energy density, environmental pollution-free and nature abundance.
27 Several kinds of MnO_2 nanomaterials have been utilized in fabrication of
28 electrochemical sensors [26-28]. As we known, due to the basic unit MnO_6 octahedral
29 is linked in different ways of MnO_2 [29], it have some crystallographic forms in
30 nature (such as α , β , γ , and δ). Different from (1D) MnO_2 nanostructures, it is not easy
31 to obtain two- and three-dimensional (2D and 3D) ordered nanostructured MnO_2

1 semiconducting materials though they are urgent need for advanced optoelectronic,
2 information storage and nanoscale electronic applications [30]. Through our tireless
3 efforts, a facile one-step solution phase shape-controlled of 3D hierarchical
4 nanostructures of MnO₂ synthetic approach at room temperature have been researched
5 and it could overcome this problem primely. It is obviously to see that the method is
6 environmentally friendly, which could be regard as a promising green chemical
7 synthesis in widespread practical applications. However, the application of 3D MnO₂
8 was quite few in contrast with 1D MnO₂ and 2D MnO₂ and few researchers have been
9 paid attention to the application to electrochemistry of 3D MnO₂. We think it may
10 have a good exhibition in the areas of sensor for its high specific surface of
11 hierarchical nanostructures. In this work, we employ (3D) MnO₂ as a catalyst support.
12 The specific flower-like structure is formed by soft silk films with wrinkle which
13 looks like the structure of monolayer grapheme and the high surface-to-volume ratio
14 of flower-like MnO₂ can provide large interspaces for the immobilization of Ag NPs
15 [31], thus preventing the aggregation of silver nanoparticles effectively and obtaining
16 highly dispersed silver nanoparticles, achieving faster electron transfer, promoting the
17 property of constructed H₂O₂ sensors. In view of the above mentioned points, the
18 research on (3D) MnO₂ nanomaterials seems to be a hot pot for their alternation of
19 other noble metals in the area of electrocatalysis. To further increase the amount of
20 adsorbed metal particles, making the surface of flower-like MnO₂ rougher so that the
21 nanocomposites could play the greatest potential to improve the detection
22 performance of the sensor towards H₂O₂, the activity of Ag-MnO₂ for H₂O₂ reduction
23 needs to be further enhanced.

24 Recently, Halloysite nanotubes (HNTs) have been aroused much attention as an
25 immobilization matrix for biosensors and biocatalysis [32]. They are naturally
26 occurring alumi-nosilicates (Al₂Si₂O₅(OH)₄· nH₂O) with a regular nanotubular bulk
27 structure, morphology, rich mesopores and nanopores. The size of halloysite
28 nanotubes varies from 50 to 70 nm in external diameter, 15 nm diameter lumen and
29 1±0.5 μm length. Due to the siloxane and a few hydroxyl groups occupied the outer
30 surfaces of HNTs, HNTs could disperse in solution more uniformly than other natural

1 silicates (such as kaolinite and montmorillonite) and possess the unique property to
2 form hydrogen bonding [33]. Different from other layered silicates, the reason of
3 easily dispersion on halloysites was that their infirm secondary interactions among the
4 nanotubes via Vander Waals forces and hydrogen bonds [34]. In addition, the naturally
5 occurring HNTs are much cheaper and easily available. Compared to carbon
6 nanotubes (CNTs), HNTs were selected as reliable substrates in many scopes due to
7 their unique characteristics, such as different outside and inside chemistry and
8 adequate hydroxyl groups on the surface of HNTs. [35]. Considering its special
9 performance, we would disperse it on the flower-like MnO_2 . In terms of the
10 nanocomposites, not only the MnO_2 own the ability to catalyze hydrogen peroxide,
11 but also the HNTs nanotubes could increase the surface area of flower-like MnO_2 ,
12 creating favorable conditions for a large number of silver nanoparticles adsorption.

13 The aim of the present work is to synthesize Ag-HNTs- MnO_2 nanocomposites by
14 employing reduction reaction and ultrasonic agitation, fabricating a novel
15 non-enzymatic sensor of H_2O_2 based on the unique material by simple cast method
16 and used for sensitively detect H_2O_2 .

17 **Experimental**

18 1.1 Materials

19 Halloysite nanotubes were purchased from Natural Nano. Inc. Chitosan (CS,
20 MW5- 6×10^5 , >90% deacetylation) was got from Shanghai Yuanju Biotechnology Co,
21 Ltd (shanghai, China). H_2O_2 (30%, v/v aqueous solution) was purchased from Tianjin
22 Tianli Chemistry Reagent Co., Ltd (Tianjin, China). 0.1 M phosphate buffered saline
23 (PBS, pH 7.2). All other chemicals and reagents were of analytical reagent grade and
24 deionized water was used in experiments.

25 1.2 Apparatus and electrochemical measurements

26 Scanning electron microscope (SEM) and energy dispersive spectroscopy (EDS)
27 were got with a JSM-6390F scanning electron microscope (JEOL, Japan).
28 Transmission electron microscopy (TEM) patterns were done on a JEM-2100
29 scanning electron microscope (JEOL, Japan). All of the electrochemical
30 measurements were obtained on a CHI 660 electrochemical analyzer (Shanghai
31 Chenhua Instrument Co. Ltd., China). A conventional three-electrode cell was used,
32 including a platinum wire as counter electrode, a saturated calomel electrode (SCE) as

1 reference electrode, and the modified glassy carbon electrodes (GCE, 3mm in
2 diameter) as working electrode. The analytical solutions were purged with highly
3 purified nitrogen for at least 30 minutes before electrochemical experiments and
4 maintained under nitrogen atmosphere during the experiments. All the measurements
5 were conducted at room temperature (25 ± 2 °C).

6 2.3. Preparation of the sensor

7 2.3.1. Synthesis of HNTs-MnO₂

8 In a typical synthesis, 200 mL of manganous chloride (MnCl₂) solution (20 mM)
9 was mixed with ethylenediaminetetraacetic acid disodium salt (EDTA) solution. 80
10 mg HNTs we dispersed in the above solution under sonication. After sonicating for 1
11 h, 200 mL of sodium hydroxide (250 mM) was added to the system. Then, 200 mL of
12 potassium persulfate (K₂S₂O₈) was added to initiate the redox reaction. After standing
13 at 30°C for overnight, the solid product was collected by filtration, washed with
14 doubly distilled water and finally with ethanol, followed by vacuum drying at 80 °C.
15 The obtained sample was denoted as HNTs-MnO₂.

16 2.3.2. Synthesis of Ag-HNTs-MnO₂

17 HNTs-MnO₂ powder (10 mg) was dispersed in 50 mL ethanol-water (1:1, v/v ratio)
18 solution, ultrasonically mixed with 2.0 mL of 0.1 mM AgNO₃ solution, and
19 subsequently added excess NaBH₄ solution in a dropwise manner under stirring
20 condition. The reductive reaction was performed under room temperature for 2 h with
21 continuous magnetic stirring, after which, the composite products were separated
22 from the solution in a centrifuge, ultrafiltration and thoroughly washed with doubly
23 distilled water. The obtained black powder was dried in a vacuum oven at 70°C for 12
24 h. For comparison, Ag-MnO₂ and HNTs-MnO₂ were prepared by the same process.

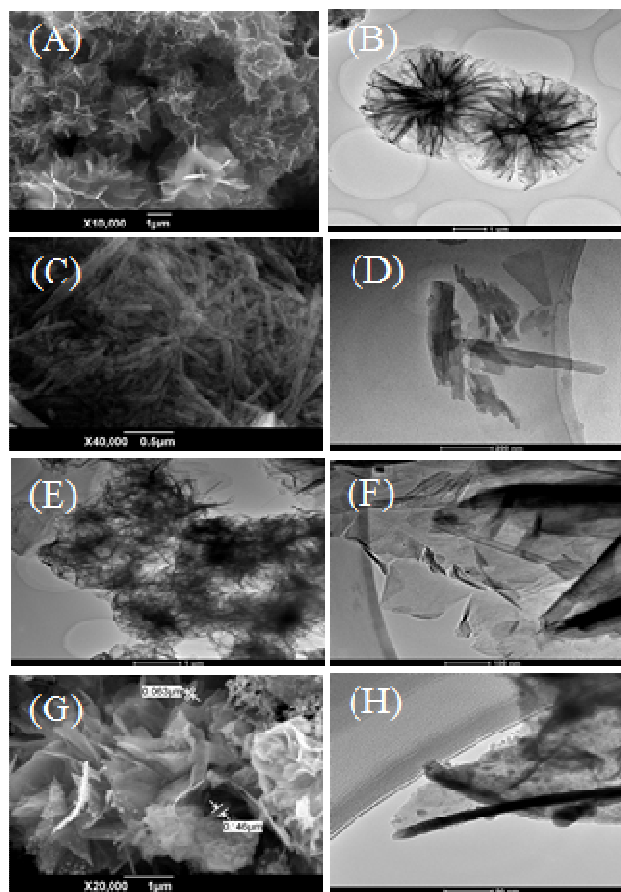
25 2.3.3. Electrode modification

26 The glass carbon electrode (GCE) was prepared by a simple casting method. Prior
27 to use, the GCE was polished with 1.0 and 0.3 μm alumina powder to obtain mirror
28 like surface, respectively, and rinsed with doubly distilled water, followed by
29 sonication in ethanol solution and doubly distilled water successively. Then, the GCE
30 was allowed to dry in a stream of nitrogen. The composites (5 mg) were dispersed
31 into chitosan (5 mL, 0.5 %) and sonicated for 30 minutes; suspension (5 μL) was cast
32 onto the GCE and then dried in air at room temperature. The resulted electrode was
33 denoted as Ag-HNTs-MnO₂/GCE.

34 2. Results and Discussion

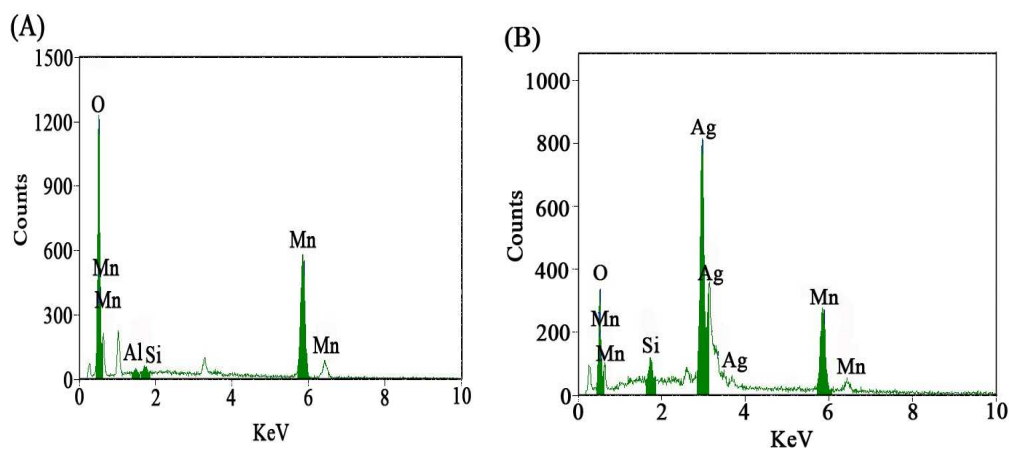
1 **2.1 Characterization of Ag-HNTs-MnO₂ nanocomposites**

2 The morphologies and structure of the MnO₂, HNTs, MnO₂-HNTs and
3 Ag-HNTs-MnO₂ were characterized by SEM and TEM as shown in Fig. 1. An overall
4 view in Fig. 1A indicates many flower-like nanoarchitectures. The high-magnification
5 image in Fig. 1B shows that a double flowery nanostructure is made of tiny nanopetals
6 growing outside from different sites. An obvious phenomenon was worth mentioning
7 that although suffer from long-time sonication, the new-get nanostructure cannot be
8 destroyed into discrete petals, suggesting that the nanocomposites are actually
9 integrated tightly rather than delicate aggregates. From Fig. 1C and Fig. 1D, it can be
10 seen that HNTs are the tubular structures of hollow and open-ended in the
11 submicrometer range. Commonly, the size of HNTs varies from 1200 to 500 nm in
12 length, with the internal diameter of ~15 nm and the outer diameter of ~100 nm. Fig.
13 1E showed that the vast majority of HNTs were coated with flower-like MnO₂, and
14 almost no parts of the HNTs were naked, implying the strong binding between HNTs
15 and MnO₂. It can be seen clearly from Fig. 1F that HNTs nanotubes loaded onto the
16 MnO₂ sheet thus could further enhance the active surface area of MnO₂ and adsorb
17 more silver nanoparticles. Fig. 1G and Fig. 1H showed that granular nanosilver
18 attached on MnO₂ surfaces without any Ag NPs aggregation.



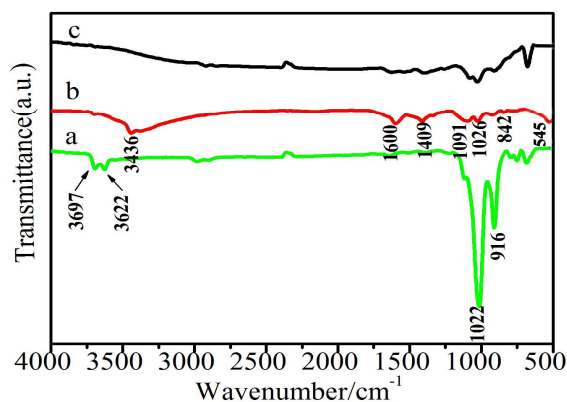
1
2 Fig. 1 TEM and SEM images of nanocomposites: (A, B) MnO₂, (C, D) HNTs, (E, F)
3 HNTs-MnO₂ and (G, H) Ag-HNTs-MnO₂

4 Fig. 2 showed the EDS patterns of HNTs-MnO₂ and Ag-HNTs-MnO₂. From Fig.
5 2A and 2B, the EDS patterns revealed that the nanocomposites were composed of O,
6 Al, Mn and Ag elements, suggesting that HNTs-MnO₂ and Ag-HNTs-MnO₂
7 nanocomposites had been synthesized successfully.



8
9 Fig. 2 EDS spectrum of (A) HNTs-MnO₂ and (B) Ag-HNTs-MnO₂ nanocomposites

1 FTIR spectra were helpful to further understand the formation of nanocomposites.
2 Fig. 3a showed that four peaks present in the FTIR spectrum of HNTs. Double peaks
3 at 3697 and 3622 cm^{-1} appeared on the spectrum of HNTs (curve a), which were due
4 to the stretching vibrations of hydroxyl groups at the surface of HNTs. The peaks
5 about 1000 cm^{-1} were assigned to Si-O groups in HNTs. In addition, a single Al_2OH
6 bending band at 916 cm^{-1} , and a band at 1022 cm^{-1} attributed to Si-O-Si stretching
7 vibrations. Compared with HNTs, the other two peaks of HNTs- MnO_2 (curve b) were
8 observed at 1600 cm^{-1} and 545 cm^{-1} . The peak at 1600 cm^{-1} was related to water -OH
9 bending and the peak at 545 cm^{-1} should be ascribed to the Mn-O and Mn-O-Mn
10 vibrations in $[\text{MnO}_6]$ octahedral. After metallic Ag (curve c) particles were loaded, the
11 final nanocomposites exhibit low absorption-peak intensity of the functional group;
12 the main reason was that once Ag nanoparticles existencing, the absorption peak of
13 MnO_2 was covered.

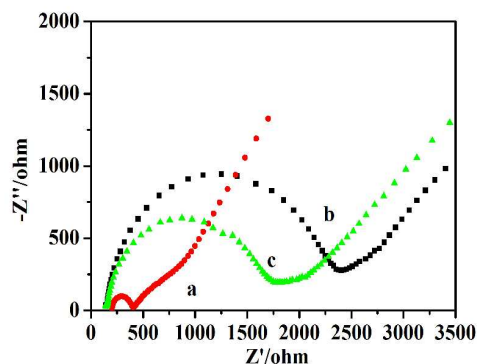


14
15 Fig. 3 FTIR spectra of (a) HNTs, (b) HNTs- MnO_2 and (c) Ag-HNTs- MnO_2
16 nanocomposites

17 3.3. Electrochemical properties of Ag-HNTs- MnO_2

18 Electrochemical impedance spectroscopy (EIS) can study the interfacial properties
19 of surface-modified electrodes usefully. As we known, the semicircle diameter
20 equaled to the electron transfer resistance (R_{ct}). We can seen from Fig. 4, the value of
21 R_{ct} is increased from 1750 Ω (curve c) to 2300 (curve b) after introducing HNTs onto
22 the MnO_2 modified electrode while the value is decreased to 500 Ω after introducing
23 AgNPs (curve a). These results suggest that after MnO_2 was modified on GCE,
24 electron transfer between the solution and the electrode is less efficient which is
25 ascribed to the semiconductive of MnO_2 nanoflowers. When HNTs is immobilized

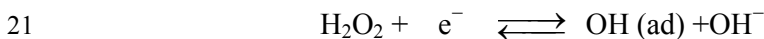
1 onto electrode, a remarkable increase in the semicircle diameter was observed due to
 2 the poor electric conductivity of HNTs, after Ag is immobilized onto electrode, the
 3 electron transfer resistance value is reduced owing to the good conductivity of Ag NPs
 4 that decreased the impedance of the electrode. The results were indicating that the
 5 Ag-HNTs-MnO₂ could efficiently enhance the electron transfer.



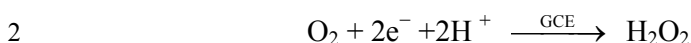
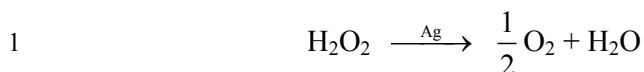
6
 7 Fig. 4 EIS of (a) Ag-HNTs-MnO₂/GCE, (b) HNTs-MnO₂/GCE, (c) MnO₂/GCE
 8 HNTs/GCE in 5.0 mM [Fe(CN)₆]^{4-/3-} containing 0.1 M KCl from 10⁵ to 10⁻² Hz at
 9 amplitude of 5 mV

10 Fig. 5 recorded cyclic voltammograms (CVs) in N₂-saturated 0.1 M PBS (pH 7.2)
 11 of the bare GCE (a, b), HNTs-MnO₂/GCE (c, d) and the Ag-HNTs-MnO₂/GCE (e, f)
 12 at a scan rate of 100 mVs⁻¹ in 0.1 M PBS. Bare GCE (a), HNTs-MnO₂/GCE (c) and
 13 the Ag-HNTs-MnO₂/GCE (e) showed weak electrochemical response in the absence
 14 of H₂O₂. After adding 1.0 mM H₂O₂, the electrochemical responses were increased
 15 accordingly. However, compared with bare GCE (b), HNTs-MnO₂/GCE (d),
 16 Ag-HNTs-MnO₂/GCE (curve f) exhibited greatly current response about 8.5 μA in
 17 intensity at -0.65 V, indicating that the Ag NPs exhibited excellent catalytic
 18 performance toward H₂O₂.

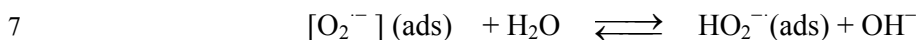
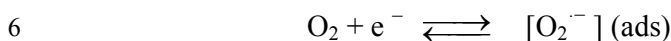
19 According to the literature [36], the catalytic mechanism proposed for the
 20 reduction of H₂O₂ was as follows:



24 When the Ag NPs were deposited on the electrode, the reaction became more
 25 irreversible [36]:



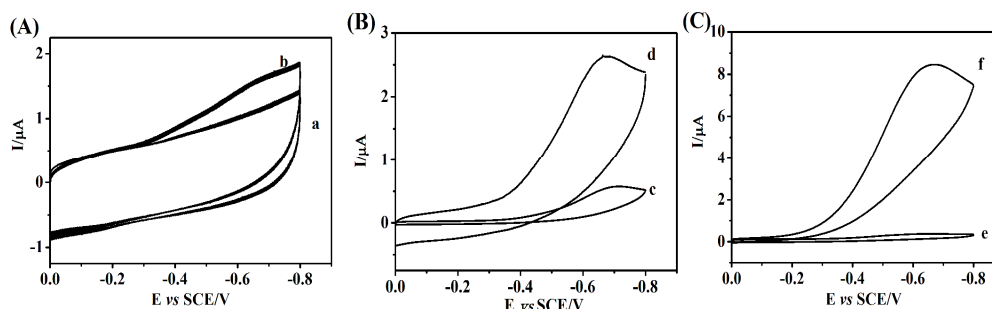
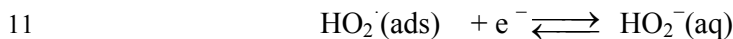
3 Then the O_2 generated in the action above would turn into the detection signal on
4 electrode. It had been proposed [37] that the electroreduction of oxygen on electrode
5 occurred via the mechanism shown below [38]:



8 Then



10 Or

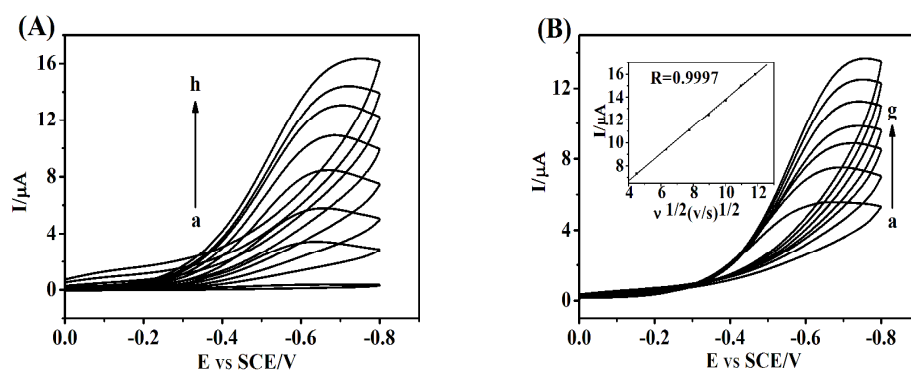


12
13 Fig. 5 CVs of bare GCE (A), HNTs-MnO₂/GCE (B), Ag-HNTs-MnO₂/GCE (C) in
14 N₂-saturated 0.1 M PBS in the absence (a, c and e) and presence (b, d and f) of 1.0
15 mM H₂O₂

16 With bring Ag NPs modified onto the HNTs-MnO₂ electrode, the detection signal
17 of H₂O₂ was amplified. There are some rational reasons could be considered: Firstly,
18 flower-like MnO₂ layer can provide roomy space for Ag NPs adsorption, thus obtain
19 more electroactive sites. Secondly, in terms of MnO₂ smooth surface, the rough HNTs
20 could further support high surface area for nanoparticles loading to keep the high
21 catalytic activity. Thirdly, HNTs lives as a natural holder, increase the effective
22 surface area of unique morphology MnO₂, with homogeneous Ag NPs, they all have
23 the good ability to catalyze H₂O₂, once integrate there own unique properties, a new
24 designed nanocomposites sensor will present excellent performance towards
25 catalyzing.

26 The catalytic activity of Ag-HNTs-MnO₂ nanocomposites by changing the

1 concentration of H_2O_2 was shown in Fig. 6A. It can be seen that no characteristic peak
 2 was shown when no H_2O_2 was introduced into the system. After injecting H_2O_2 into
 3 the N_2 -saturated 0.1 M PBS (pH 7.2), a gradually increased reduction current
 4 appeared, indicating the final nanocomposite own the excellent response and
 5 electrocatalytic activity towards H_2O_2 . It was notable that with the increment of H_2O_2
 6 concentration, the reduction current gradually increased. From Fig. 6B we can get the
 7 effect of potential scan rate on peak current of Ag-HNTs- MnO_2 . The cathodic peak
 8 current increased in a linear relationship with the square root of with increasing the
 9 scan rates from 20 to 140 $\text{mV}\cdot\text{s}^{-1}$, indicating that the reactions occurring on the
 10 modified electrode were irreversible and this process was diffusion-controlled.

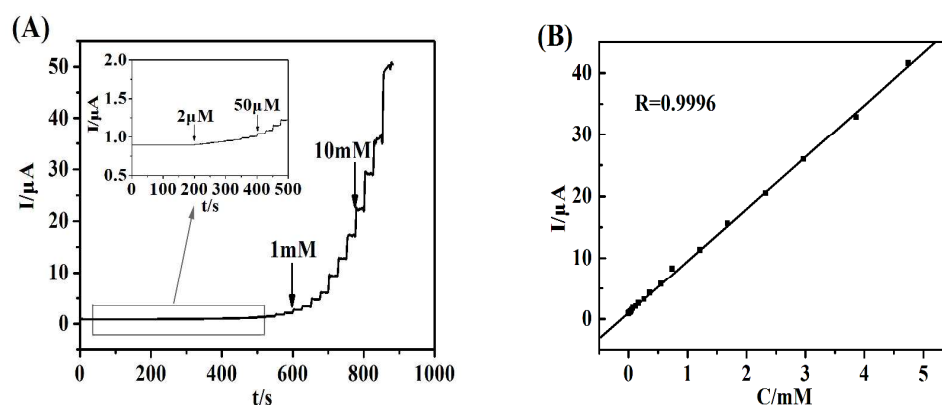


11
 12 Fig. 6 (A) CVs of the Ag-HNTs- MnO_2/GCE in N_2 -saturated 0.1M PBS (pH 7.2) in the
 13 absence and presence of H_2O_2 with different concentrations (from a to g: 0, 1, 2, 3, 4,
 14 5, 6 and 7 mM at the scan rate of 100 mV/s). (B) CVs of the Ag-HNTs- MnO_2/GCE
 15 N_2 -saturated 0.1M PBS (pH 7.2) containing 5.0 mM H_2O_2 at different scan rates
 16 (from a to g: 20, 40, 60, 80, 100, 120 and 140 mV/s). Inset: plot of electrocatalytic
 17 peak current of H_2O_2 versus $v^{1/2}$

18 Fig. 7A showed a typical amperometric response curve of H_2O_2 at
 19 Ag-HNTs- MnO_2/GCE in N_2 -saturated 0.1 M PBS (pH 7.2) for the different
 20 concentrations of H_2O_2 . We can observe a stable, well-defined and fast amperometric
 21 response under successive step additions of H_2O_2 . Although the modified electrode
 22 exhibited biggest catalytic activity at -0.65 V, the background was much too high to
 23 interfere the detection. So detect H_2O_2 was carried out at -0.3 V to ensure a low
 24 applied potential, good signal-to-noise ratio and less interference of other
 25 electroactive species in the solution. It was clear that the response current of the
 26 modified electrode increased to steady-state values less than 2 s upon the addition of
 27 H_2O_2 , indicating a fast amperometric response behavior. Fig. 7B showed the

1 calibration curve for the H_2O_2 . With increasing add H_2O_2 , the working electrode gave
 2 a linear dependence in the H_2O_2 concentration range of 2.0 μM to 4.71 mM and the
 3 linear regression equation was expressed as $I_p (\mu\text{A}) = 1.03 + 8.45 \cdot C (\text{mM})$ with a
 4 correlation coefficient of 0.9996, a sensitivity of $11.9 \mu\text{A mM}^{-1} \text{cm}^{-2}$ and a detection
 5 limit of 0.7 μM at a signal-to-noise ratio of 3. These results indicated that the
 6 Ag-HNTs- MnO_2/GCE can be used for the preparation of an amperometric sensor for
 7 H_2O_2 with quick response and wide linear range, deeply illustrate this new sensor own
 8 excellent property.

9 As shown in Table 1, several typical non-enzymatic and enzymatic H_2O_2 sensors
 10 reported previously have been compared. We can observe that the wide linear range,
 11 the low detection limit and the short response time due to the high surface-to-volume
 12 ratio and larger surface area for H_2O_2 molecules adsorb and react. The unique
 13 morphology of MnO_2 provides larger surface and effective surface area for the
 14 attachment of Ag NPs. In addition, Ag NPs and flower-like MnO_2 are also acts the
 15 role of the electron transfer promoter. As a result, the new construct nanocomposites
 16 can be used for detecting H_2O_2 efficiently.



17
 18 Fig. 7 (A) Typical amperometric response of the Ag-HNTs- MnO_2/GCE on successive
 19 injection of H_2O_2 into the stirring N_2 -saturated 0.1 M PBS (pH 7.2), applied potential:
 20 -0.3 V . (B) Calibration curve of H_2O_2 versus its concentration

21 **Table. 1** Comparison of several typical nonenzymatic H_2O_2 sensors

| Sensors | Linear range (mM) | Sensitivity ($\mu\text{A mM}^{-1} \text{cm}^{-2}$) | Detection limit (μM) | Literature |
|-------------------------------------|----------------------|---|--------------------------------------|------------|
| $\text{MnO}_2/\text{GO}/\text{GCE}$ | 0.005–0.6 | - | 0.8 | [39] |
| $\text{MnO}_2/\text{carbon fiber}$ | 0.01-0.26 | 10.6 | 5.4 | [40] |

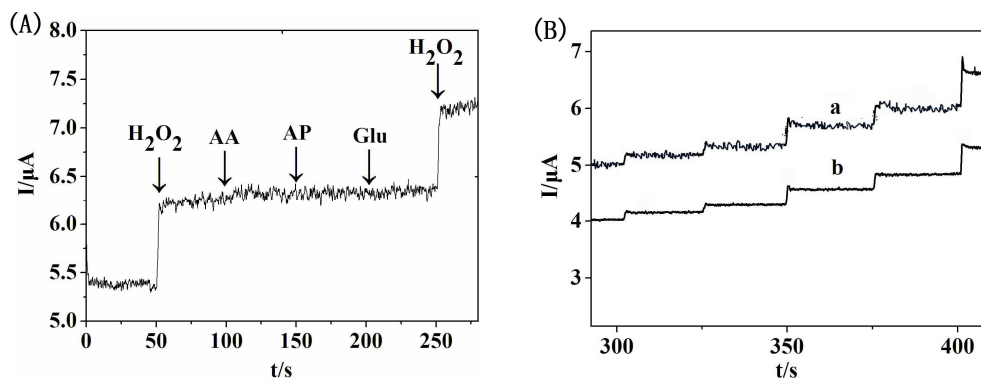
| | | | | |
|-----------------------------------|------------|------|------|-----------|
| (DNA-AgNCs)/GE | 0.02-23 | - | 3 | [41] |
| Platinum hierarchical nanoflowers | 0.01-4.0 | 1.39 | 1.05 | [42] |
| AgNP/SnO ₂ | 0.01-3.5 | - | 5.0 | [43] |
| PtAu/G-CNTs | 0.002-8.6 | - | 0.6 | [44] |
| Ag-HNTs-MnO ₂ | 0.002-4.71 | 11.9 | 0.7 | This work |

1

2 **3.4. Interference study**

3 In terms of non-enzyme H₂O₂ sensor, good selectivity is very important. Under the
 4 optimized experimental conditions, some potential interfering species such as ascorbic
 5 acid (AA), uric acid (UA), glucose, and acetaminophen (AP) have been investigated.
 6 As shown in Fig. 8 (A), H₂O₂ solution was firstly injected into the N₂-saturated 0.1 M
 7 PBS (pH 7.2) at a working potential of -0.3 V, and followed by the addition of
 8 interferences (0.01 mM, respectively). It can be seen that an obvious amperometric
 9 response appeared at once 0.1 mM H₂O₂ were injected while Ascorbic acid (AA),
 10 acetaminophen (AP), glucose (Glu) did not cause any further amperometric changes.
 11 The result indicates that the modified electrode exhibited good ability of
 12 anti-interference to electroactive species, which attribute to the relatively lower
 13 potential at -0.3 V. The interfering species of oxygen were also investigated by testing
 14 the amperometric responses of H₂O₂ at the same potential. As seen from Fig. 8 (B), a
 15 stable response current and good signal-to-noise ratio can be observed in N₂-saturated
 16 0.1 M PBS at -0.3 V (curve b). In O₂-saturated 0.1 M PBS at -0.3 V (curve a),
 17 although the background noise increased due to the electro-reduction of O₂, the
 18 response currents almost remained unchanged, meaning the resulting electrode
 19 exhibited good ability of anti-interference to O₂.

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21

1 Fig. 8 Amperometric response of the Ag-HNTs-MnO₂/GCE to (A) successive addition
2 of H₂O₂, AA, AP, Glu (0.05 mM, respectively) in N₂-saturated 0.1 M PBS (pH 7.2) at
3 -0.3 V and (B) successive addition of H₂O₂ (0.05 mM) in (a) O₂-saturated and (b)
4 N₂-saturated 0.1 M PBS (pH 7.2) at -0.3 V

5 **3.5. Repeatability and stability**

6 The repeatability and stability of the resulted H₂O₂ sensors were also investigated.
7 There are four Ag-HNTs-MnO₂ modified electrodes were investigated at the same
8 condition to compare their amperometric current responses. The result suggests that
9 the relative standard deviation for current determination of H₂O₂ was 3.5%,
10 confirming that the Ag-HNTs-MnO₂/GCE nanocomposite modified electrode was
11 highly reproducible. The stability of the modified GCE was also estimated every one
12 week, founding the modified electrode remained 90% of its initial current response.
13 Thus, the modified GCE processed acceptable repeatability and stability.

14 **4. Conclusion**

15 In summary, Ag-HNTs-MnO₂ nanocomposites had been synthesized successfully
16 by a facile simple strategy and a novel non-enzymatic H₂O₂ sensor based on the
17 nanocomposites was fabricated. The novel sensor exhibits good electrocatalytic
18 activities toward H₂O₂ reduction, low detection limit, long-time stability and high
19 response sensitivity, indicating its prominent electrochemical behavior towards
20 electroactive biomolecules. Extraordinary, flower-like MnO₂ provides more binding
21 sites for Ag NPs to enhance the catalyst ability, making the performance of the
22 proposed modified electrode more effectively. Finally, we believe that the type of
23 high-performance nanostructured sensor, combined with a low-cost and scalable
24 technique could be used as promising platform for the construction of various
25 nonenzymatic electrochemical sensors for further study.

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