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Enantioselective Organocatalytic Michael Addition of Isorhodanines to α , β -Unsaturated Aldehydes⁺

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Miao,^a Jianzhong Yao,^a Wei Wang^{*b,c} Wannian Zhang,^{*a} and Chunquan Sheng^{*} Michael reaction of substituted isorhodanines with α , β -unsaturated aldehydes in the presence of catalytic amount of a

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chiral secondary amine is presented. This transformation proceeds in good to high yields furnishing the corresponding 4,5disubstituted isorhodanine adducts in good to excellent enantioselectivities.

Introduction

Asymmetric organocatalysis is a powerful and fruitful research area of organic chemistry by providing efficient and environmental friendly access to enantiomerically pure compounds in the synthesis of biologically active compounds and natural products.¹ In particular, asymmetric Michael reaction is a viable strategy for stereoselective formation of carbon-carbon bonds. In recent years, organocatalytic enantioselective Michael reactions has attracted wide interests and numerous synthetic approaches have been developed.² It is noteworthy that a variety of enolizable nucleophiles have been explored for this purpose. In these processes, generally highly active pre-enolized or readily enolizable nucleophilic species such as silyl enol ethers,³ enamides,⁴ nitroalkanes,⁵ 1,3-dicarbonyls,⁶ bis-sulfones,⁷ active thioesters,⁸ and others⁹ are used.¹⁰

Given a broad spectrum of biological activities, such as antitrypanosomal, antitumor, antimycobacterial, and antiexudative activities and so on of isorhodanine related molecules (Figure 1),¹¹ synthesis of chiral isorhodanines is highly valuable for medicinal chemistry and drug discovery. In our previous work^{10j}, we reported a new approach to create a divergent cascade organocatalysis strategy for the facile construction of new "privileged" substructure-based DOS

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(pDOS) library. Diverse molecular architectures were produced available facilelv from readily simple synthons thiazolidinedione and its analogues and α , β -unsaturated aldehydes. Reactions of isorhodanine via a Michael-cyclization cascade lead to structurally different fused thiopyranoid scaffolds.^{10j} Inspired these results, we envision that the tautomerization of isorhodanines 2 to active enol form 2' as new Michael donors could participate in an organocatalytic enantioselective conjugate addition reactions with enals (Scheme 1).









Antiexudative activity

Antitumor and Antitrypanosomal activity antimycobacterial activity

Figure 1. Representative bioactive molecules containing the isorhodanine unit.

Previous work



Scheme 1. Tautomerization of substituted-isorhodanines.

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Results and discussion

To probe the feasibility, we initiated the study by the reaction of 4-(benzylthio)thiazol-2(3H)-one (2a) with transcinnamaldehyde (1a) in toluene catalyzed by I (10 mol %) at room temperature (rt) (Table 1). To our delight, the desired product 3a was obtained in 82% yield with excellent enantioselectivity (ee = 93%) (entry 1). Inspired by the outcome, catalysts II-V were then probed (entries 2-6). In the presence of catalyst III, the reaction also proceeded smoothly with high yield (83%) and excellent enantioselectivity (94% ee) (entry 3). However, catalysts II, IV, V and VI were not suitable for this reaction, giving lower ee values (9-91% ee). Moreover, larger catalyst load (20 mol% catalyst III) did produce better yield and enantioselectivity (entry 7). No reaction was observed between substrates 1a and 2a without the catalyst, suggesting the activation of the enal via an iminium ion is necessary (entry 8).

On the basis of catalysts I and III, various solvents (entries 1-14) were screened accordingly. They delivered the similar yields, but the reaction with catalyst III offered higher ee value than that of I (*e.g.* entries 9 vs 17, entries 10 vs 18, and entries 11 vs 19). Lower yields (33-78% yield) and enantioselectivity (80-94% ee) were observed in the presence of an additive (entries 23-28). These studies led to establishing the optimal reaction condition by using toluene as reaction medium and III as promoter (entry 3). The absolute configuration was determined by the X-ray crystal structure analysis of the product (**3a**) to be as (*S*), when *S*-chiral secondary amine (III) was used as catalyst (Figure 2).¹²

Table 1. Exploration and optimization of organocatalytic Michael-reduction cascade reaction of isorhodanine **2a** with *trans*-cinnamaldehyde **1a**.^a

Ph CHO +	O H S Ph	1. cat. (10 mol %) additive solvent 2. NaBH ₄ , MeOH	O HN Ph S Ph
1a	2a		3a
Ar I: Ar Ar II: Ar H OR V: Ar V: Ar	$\begin{array}{l} \text{additive} \\ \textbf{A1} = PhCO_2H \\ \textbf{A2} = AcOH \\ \textbf{A3} = K_2CO_3 \\ \textbf{A4} = Li_2CO_3 \\ \textbf{A5} = HCOOH \\ \textbf{A6} = 4-NO_2PhCO_2H \end{array}$		

Entry	Cat.	Additive	Solvent	t (h)	Yield [%] ^b	ee [%] ^c
1	I	-	toluene	11	82	93
2	II	-	toluene	11	80	80
3	Ш	-	toluene	12	83	94
4	IV	-	toluene	13	75	58
5	v	-	toluene	13	76	91
6	VI	-	toluene	12	72	9

7 ^d	ш	-	toluene	13	78	95
8	-	-	toluene	>15	trace	nd ^e
9	I	-	DCM	13	77	9
10	I	-	CI(CH ₂) ₂ CI	13	71	72
11	I	-	MeOH	12	77	8
12	I	-	(CH ₃ CH ₂) ₂ O	11	71	60
13	I	-	THF	15	63	26
14	I	-	MeCN	12	71	69
15	ш	-	EtOH	12	74	36
16	ш	-	CCl ₄	13	61	84
17	111	-	DCM	13	70	83
18	ш	-	CI(CH ₂) ₂ CI	13	79	75
19	ш	-	MeOH	12	78	7
20	ш	-	(CH ₃ CH ₂) ₂ O	11	77	80
21	ш	-	THF	15	60	39
22	ш	-	MeCN	12	76	64
23	ш	A1	toluene	15	78	84
24	ш	A2	toluene	15	61	86
25	ш	A3	toluene	18	33	87
26	ш	A4	toluene	16	71	84
27	ш	A5	toluene	15	66	94
28	ш	A6	toluene	15	74	80
29 ^f	111	-	toluene	15	76	90

^a Reaction conditions, unless specified, **1a** (0.16mmol, 1.2 equiv), **2a** (0.13 mmol, 1.0 equiv), and catalyst **III** (10 mol %) in solvent (1.0 mL) at rt, then methanol (1.0 mL), NaBH₄(0.13 mmol, 1.0 equiv) to give **3a**. ^b Yield of isolated product after column chromatography. ^c Determined by chiral HPLC (see the Supporting Information). ^d cat (20 mol %) was used. ^e Not determined. ^f **1a** (0.26 mmol, 3.0 equiv)

With the optimal reaction conditions in hand, the scope of the Michael reaction of isorhodanine **2a** with an array of α , β -unsaturated aldehydes **1** was probed. As shown in Table 2, a number of α , β -unsaturated aldehydes bearing electron-withdrawing, -neutral, -donating substituents and different substituted position on the benzene ring were tolerated (entries 1-9 and 13). It is found that the reaction is sensitive to

electronic effect and the position of substituted groups. It is hard to rationalize these results. Generally good to excellent enantioselectivities were obtained for enals bearing 3substituted or 4-substituted groups (60-91% ee) (entries 3-9), and the ee value was decreased slightly. However, 4-nitrosubstituted (91% ee, entry 3) substrates with strong electronwithdrawing substitutions gave the best results. In contrast, the enantioselectivities were dropped dramatically for substrates with 2-substituted groups (43% and 27% ee, entries 2 and 13). The process is also applicable for alkyl-substituted enals with good enantioselectivities (71-83% ee, entries 10-12). It is observed that the enals bearing *othro*-substituents gave poor ee (entries 2 and 13), we believed the steric effect was significantly detrimental to the enantioselectivity.

Table 2. Scope of the Michael-reduction cascade reaction of substituted isorhodanine **2a** with α , β -unsaturated aldehydes^a.

	S-	ı		– S	ОН
R ¹ CH	0 ₊ 0 − N	$S \sim Ph = \frac{1.111, to}{2.11, to}$	oulene	→ Ph	
1	H 2	2. NaB a	H ₄ , MeOI	<u>-</u> S 3	R'
Entry	Product	R ¹	t (h)	Yield	ee
				[%] ^b	[%] ^c
1	3a	Ph	12	83	94
2	3b	$2-NO_2C_6H_4$	16	79	43
3	3c	$4-NO_2C_6H_4$	12	70	91
4	3d	$4-CH_3OC_6H_4$	12	68	78
5	3e	$3-CH_3C_6H_4$	12	71	65
6	3f	$4-CH_3C_6H_4$	12	72	63
7	3g	$4-CF_3C_6H_4$	12	77	79
8	3h	3-CIC ₆ H ₄	12	71	70
9	3i	$4-BrC_6H_4$	12	75	70
10	Зј	CH ₃	13	55	71
11	3k	CH ₂ CH ₃	13	51	83
12	31	CH(CH ₃) ₂	13	56	74
13	3m	2-CH ₃ OC ₆ H ₄	12	70	27

^a Reaction conditions, unless specified, **1** (0.16mmol, 1.2 equiv), **2a** (0.13 mmol, 1.0 equiv), and catalyst **III** (10 mol %) in solvent (1.0 mL) at rt, then methanol (1.0 mL), NaBH₄ (0.13 mmol, 1.0 equiv) to give **3a-m**. ^b Yield of isolated product after column chromatography. ^c Determined by chiral HPLC (see the Supporting Information).





Figure 2 X-ray crystal structure of the product 3a

Furthermore, we explored the substrate scope by the variation of isorhodanines 2 (Table 3). To our delight, under the same conditions, the processes between different substituted isorhodanines $\boldsymbol{2}$ and $\boldsymbol{\alpha},$ $\boldsymbol{\beta}\text{-unsaturated}$ aldehyde $\boldsymbol{1a}$ proceeded smoothly with good enantioelectivity (61-> 99% ee) and yields (50-83%). Excellent entioselectivity was obtained for the methyl- (> 99% ee, entry 2) and ethyl-substituted (92% ee, entry 3) isorhodanines, whereas their yields were relatively low (50% and 56%, respectively). For 2-naphthylmethylenesubstituted isorhodanines, the enantioselectivity was decreased (88% ee) but with high yields (83% yield, entry 4). It is suggested that the size of substituted group was one of the crucial factors to improve the enantioselectivity. The substituted benzyl isorhodanines (entries 5-11) gave high yields (68-81% yield) and good to excellent ee values (61-97% ee). For the 3-substituted or 4-substituted substrate, the enantioselectivity was decreased (entries 5, 7, and 9-11), however, 3-substituted or 4-substituted substrates bearing strong electron-withdrawing groups benefit the enantioselectivity (such as 93% ee, entry 9). For the 2substituted substrates, excellent enantioselectivities maintained (97% and 94% ee, entries 6 and 8). For the substrate bearing acetonitrile substitution (entry 12), the yield was good (71% yield), but the enantioselectivity was decreased (76% ee). It appears that steric demanding is beneficial to the enantioselectivty (entries 4, 6 and 8), while less hindered ones deliver relatively poor outcomes (entries 5, 7 and 12). When the N-substitued substrates were reacted with compound 1a, no desired products were obtained (entry 13). It is suggested that the enol form in isorhodanines is critical for the reaction process (Scheme 1). It is difficult for N-substituted isorhodanines to produce the active species via tautomerization (Scheme 2).

 Table 3. Scope of the Michael-reduction cascade reaction of substituted-isorhodanines 2 with *trans*-cinnamaldehyde 1a^a.

Ph C	CHO ₊ R		1. III, toulene 2. NaBH ₄ , MeOH	R ² -N R ^{3-S} 4	Ph
Entry	Product	R ² , R ³	t (h)	Yield	ee
				[%]	[%]
1	3a	H, Ph	12	83	94
2	4a	H, CH₃	13	50	>99
3	4b	H, CH₃C⊦	l ₂ 13	56	92

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4	4c	H, 2-naphthylCH ₂	13	83	88
5	4d	H, 3-FC ₆ H ₄ CH ₂	12	71	78
6	4e	H, 2-CH ₃ C ₆ H ₄ CH ₂	12	76	97
7	4f	H, 4-CH ₃ C ₆ H ₄ CH ₂	12	81	61
8	4g	H, 2-NO ₂ C ₆ H ₄ CH ₂	12	70	94
9	4h	H, 4-NO ₂ C ₆ H ₄ CH ₂	12	68	93
10	4i	H, 4-CNC ₆ H ₄ CH ₂	12	80	83
11	4j	H,4- <i>tert</i> - butylC ₆ H ₄ CH ₂	12	78	83
12	4k	H, CH ₂ CN	18	71	76
13	41	CH ₃ , CH ₃	36	NR^d	-

^a Reaction conditions, unless specified, **1** (0.16mmol, 1.2 equiv), 2a (0.13 mmol, 1.0 equiv), and catalyst III (10 mol %) in toluene (1.0 mL) at rt, then methanol (1.0 mL), NaBH₄ (0.13 mmol, 1.0 equiv) to give 4a-l. ^b Yield of isolated product after column chromatography. ^c Determined by chiral HPLC (see the Supporting Information). ^d No reaction.



Scheme2. Proposed mechanism for the designed Michael addition of isorhodanines to α , β -unsaturated aldehydes.

Conclusions

In summary, we have developed an efficient organocatalytic enantioselective Michael addition of isorhodanines derivatives to α,β -unsaturated aldehydes. Under the optimal reaction conditions, good yields and good to excellent Page 4 of 8

enantioselectivities are achieved with broad substrate scope. The synthetic application of the resulting 4,5-disubstituted isorhodanines as versatile building blocks for the rapid synthesis of chiral isorhodanines is demonstrated. Moreover, due to diverse bioactivity of isorhodanines, the new derivatives reported herein will inspire medicinal chemists to investigate their pharmacological activity and discover drug leads. Such chemical and medicinal investigations are currently ongoing in our laboratories.

Experimental

General Information

Commercial reagents were used as received, unless otherwise stated. ¹H and ¹³C NMR spectra were recorded on Bruker AVANCE II 300, AVANCE II 500 or AVANCE II 600 MHz nuclear magnetic resonance spectrometer (CDCl₃ or DMSO- d_6 as solvent) and teramethylsilane (TMS) was used as reference. Multiplicities were given as: s (singlet), d (doublet), t (triplet), dd (double of doublet) or m (multiplets). Coupling constants were reported in Hertz (Hz). High resolution mass spectrometry (HRMS) was recorded on an Agilent 6538 UHD Accurate-Mass Q-TOF LC/MS spectrometer. Analytical thin layer chromatography (TLC) was performed on silica gel GF254 (Qingdao Haiyang Chemical China) and compounds were visualized with a UV light at 254 nm. Flash chromatography was performed on silica gel 100-200 mesh with freshly distilled solvents. Enantioselectivities were determined by High performance liquid chromatography (HPLC) analysis employing a Daicel Chiralpak AS-H, OZ-H, AD-H. Optical rotations were measured in CH₂Cl₂ or Ethyl Acetate (EtOAc) on a SGW-1 automatic polarimeter (Shanghai Precision Scientific Instrument CO., LTD) with a 0.8 mL cell (c given in g/100 mL). Absolute configuration of the products was determined by Xray.

General reaction procedure and characterization for 3

Catalyst III (22.0 mg, 0.06 mmol, 0.1 equiv) was added to a solution of α , β -unsaturated aldehyde **1** (21.3 mg, 0.16 mmol, 1.2 equiv) and S-benzyl-substituted isorhodanine 2a (30.0 mg, 0.13 mmol, 1.0 equiv) in toluene (1.0 mL). The mixture was stirred for 12 h at room temperature. Then, methanol (1.0 mL) and NaBH₄ (5.1 mg, 0.13 mmol, 1.0 equiv) were added. After stirring for 0.5 h, the solvent was evaporated and the crude product was purified though column chromatography over silica gel with hexane/EtOAc (5:1-3:1) as the eluent to yield 3a (38.5 mg, 83% yield) as a white solid. Compounds 3b-m were prepared with the same procedure.

(S)-4-(Benzylthio)-5-(3-hydroxy-1-phenylpropyl)thiazol-2(3H)-o**ne (3a).** ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.84 (s, 1H), 7.31-7.36 (m, 4H), 7.29-7.30 (m, 1H), 7.21-7.25 (m, 3H), 7.13-7.16 (m, 2H), 4.41 (t, J = 8.1 Hz, 1H), 3.87-3.98 (m, 2H), 3.38-3.53 (m, 2H), 2.05-2.16 (m, 1H), 1.80-1.92 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.92, 141.93, 137.23, 131.14, 128.96, 128.84, 128.78, 127.65, 127.32, 127.03, 118.78, 60.15, 40.25, 39.45, 37.93, 29.71; HRMS (ESI) calcd for C₁₉H₁₈NO₂S₂ [M-H]⁻ =

356.4897, found 356.2246; HPLC (Chiralpak AS-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 30/70, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 12.8 min, t_{major} = 30.6 min, ee = 94.1%; [α]²⁵_D = -12.5 (*c* = 0.32 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(2-nitrophenyl)propyl)thiazol-2(3*H*)-one (3b). ¹H NMR (600 MHz, CDCl₃, TMS): $\delta = 8.99$ (s, 1H), 7.77 (d, *J* = 8.1 Hz, 1H), 7.55 (t, *J* = 7.6 Hz, 1H), 7.45 (d, *J* = 7.6 Hz, 2H), 7.37 (t, *J* = 7.6 Hz, 3H), 7.29 (t, *J* = 7.6 Hz, 1H), 7.23-7.24 (m, 1H), 7.17 (d, *J* = 7.6 Hz, 2H), 4.94 (t, *J* = 7.3 Hz, 1H), 3.83 (d, *J* = 12.9 Hz, 1H), 3.78 (d, *J* = 12.9 Hz, 1H), 3.41-3.50 (m, 2H), 2.04-2.10 (m, 1H), 1.83-1.89 (m, 1H); ¹³C NMR (150 MHz, DMSO-*d*₆, TMS): δ = 171.11, 149.09, 136.29, 135.99, 132.54, 128.43, 128.37, 127.39, 127.31, 124.13, 120.16, 59.47, 39.11, 38.40, 34.07; HRMS (ESI) calcd for C₁₉H₁₇N₂O₄S₂ [M-H]⁻ = 401.4872, found 401.2043; HPLC (Chiralpak AS-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*propanol/hexane = 25/75, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 50.3 min, t_{major} = 81.7 min, ee = 43.1%; [α]²⁵_D = 2.9 (*c* = 0.19 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(4-nitrophenyl)propyl)thiazol-2(3*H*)-one (3c). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.87 (s, 1H), 8.08 (d, *J* = 7.7 Hz, 2H), 7.29-7.30 (m, 2H), 7.25-7.26 (m, 1H), 7.19 (s, 1H), 7.16-7.18 (m, 3H), 4.47 (t, *J* = 7.6 Hz, 1H), 3.93 (s, 2H), 3.36-3.46 (m, 2H), 2.01-2.06 (m, 1H), 1.82-1.87 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.02, 148.83, 146.37, 136.61, 128.43, 128.40, 128.25, 127.70, 127.32, 123.47, 119.56, 59.08, 39.45, 38.82, 37.04; HRMS (ESI) calcd for C₁₉H₁₇N₂O₄S₂ [M-H]⁻ = 401.4872, found 401.2130; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 38.6 min, t_{major} = 43.8 min, ee = 91.1%; [α]²⁵_D = -40.0 (*c* = 0.20 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(4-methoxyphenyl)propyl) thiazol-2(3*H*)-one (3d). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 8.72 (s,1H), 7.26-7.31 (m, 3H), 7.18 (d, *J* = 7.4, 2H), 7.04 (d, *J* = 8.5 Hz, 2H), 6.80 (d, *J* = 8.5 Hz, 2H), 4.36 (t, *J* = 7.8 Hz, 1H), 3.86 (s, 2H), 3.77 (s, 3H), 3.40-3.49 (m, 2H), 2.01-2.07 (m, 1H), 1.80-1.86 (m, 1H); ¹³C NMR (125 MHz, CDCl₃, TMS): δ = 171.46, 158.06, 136.75, 133.51, 131.39, 128.40, 127.82, 127.24, 117.55, 113.67, 59.73, 54.76, 39.25, 39.08, 37.66; HRMS (ESI) calcd for C₂₀H₂₀NO₃S₂ [M-H] = 386.5156, found 386.4143; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 14.1 min, t_{major} = 23.6 min, ee = 77.5%; [α]²⁵_D = 46.7 (*c* = 0.09 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(m-tolyl)propyl)thiazol-2(3*H*)-one (3e). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.71 (s, 1H), 7.31 (m, 2H), 7.14-7.29 (m, 4H), 7.05-7.12 (m, 1H), 6.96-7.00 (m, 1H), 4.37 (t, *J* = 7.8 Hz, 1H), 3.96 (d, *J* = 13.4 Hz, 1H), 3.88 (d, *J* = 13.0 Hz, 1H), 3.37-3.52 (m, 2H), 2.35 (s, 3H), 2.20-2.14 (m, 1H), 1.78-1.89 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.82, 141.87, 138.43, 137.23, 131.30, 128.05, 128.94, 128.81, 128.67, 128.06, 127.81, 127.65, 124.32, 118.61, 60.21, 40.24, 39.47, 38.08, 29.70, 21.53; HRMS (ESI) calcd for C₂₀H₂₂NO₂S₂ [M+H]⁺ = 372.5162, found 372.1102; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, ARTICLE

particle size: 5 µm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 37.4 min, t_{major} = 46.8 min, ee = 65.3%; $[\alpha]^{25}_{ D}$ = 31.2 (*c* = 0.16 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(p-tolyl)propyl)thiazol-2(3*H*)-one (3f). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.53 (s, 1H), 7.26-7.30 (m, 3H), 7.17-7.18 (m, 2H), 7.06-7.09 (m, 2H), 6.99-7.00 (m, 2H), 4.34 (t, *J* = 7.6 Hz, 1H), 3.90 (d, *J* = 13.0 Hz, 1H), 3.85 (d, *J* = 13.0 Hz, 1H), 3.37-3.46 (m, 1H), 2.29 (s, 3H), 2.01-2.07 (m, 1H), 1.78-1.83 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.02, 148.83, 146.37, 136.61, 128.43, 128.40, 12825, 127.70, 127.32, 123.47, 119.56, 59.08, 39.45, 38.82, 37.04; HRMS (ESI) calcd for C₂₀H₂₂NO₂S₂ [M+H]⁺ = 372.5162, found 372.2172; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 9.8 min, t_{major} = 16.7 min, ee = 63.3%; [α]²⁵_D = 32.5 (*c* = 0.16 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(4-(trifluoromethyl)phenyl)propyl)thiazol-2(3*H*)-one (3g). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.51 (s, 1H), 7.51-7.52 (m, 1H), 7.47 (s, 1H), 7.41-7.44 (m, 1H), 7.32-7.34 (m, 2H), 7.30-7.32 (m, 2H), 7.20-7.23 (m, 2H), 4.49-4.51 (m, 1H), 3.93 (dd, *J* = 13.1, 23.0 Hz, 2H), 3.47-3.51 (m, 1H), 3.41-3.44 (m, 1H), 2.06-2.12 (m, 1H), 1.86-1.92 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.40, 143.06, 137.07, 130.97, 130.79, 129.91, 129.82, 129.29, 129.14, 128.88, 127.79, 124.03, 124.01, 123.95, 119.47, 59.77, 39.99, 39.47, 37.90, 29.70; HRMS (ESI) calcd for C₂₀H₁₇F₃NO₂S₂ [M-H]⁻ = 424.4876, found 424.4721; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 24.8 min, t_{major} = 30.3 min, ee = 79.3%; [α]²⁵_D = 26.9 (*c* = 0.23 in EtOAc).

(*S*)-4-(Benzylthio)-5-(1-(3-chlorophenyl)-3-hydroxypropyl)thiazol-2(3*H*)-one (3h). ¹H NMR (600 MHz, DMSO-*d*₆, TMS): δ =11.45 (s, 1H), 7.27-7.29 (m, 2H), 7.22-7.25 (m, 3H), 7.19-7.20 (m, 2H), 7.04 (s, 1H), 6.91 (d, *J* = 7.1 Hz, 1H), 4.45 (t, *J* = 5.1 Hz, 1H), 4.24 (t, *J* = 7.6 Hz, 1H), 4.00-4.05 (m, 2H), 3.12-3.16 (m, 2H), 1.85-1.90 (m, 1H), 1.65-1.71 (m, 1H); ¹³C NMR (150 MHz, DMSO-*d*₆, TMS): δ = 170.04, 144.89, 137.03, 133.07, 130.40, 128.72, 128.48, 127.44, 126.83, 126.61, 125.87, 119.31, 58.20, 40.07, 37.87, 37.60; HRMS (ESI) calcd for C₁₉H₁₉CINO₂S₂ [M+H]⁺ = 392.9347, found 392.4198; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 11.2 min, t_{major} = 14.2 min, ee = 70.3%; [α]²⁵_D = -135.5 (*c* = 0.09 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(1-(4-bromophenyl)-3-hydroxypropyl)thiazol-2(3*H*)-one (3i). ¹H NMR (600 MHz, DMSO- d_6 , TMS): δ = 11.44 (s, 1H), 7.37-7.39 (m, 2H), 7.26-7.30 (m, 3H), 7.19-7.20 (m, 2H), 6.86-6.88 (m, 2H), 4.44 (t, *J* = 4.6 Hz, 1H), 4.20 (t, *J* = 7.7 Hz, 1H), 4.06 (d, *J* = 12.9 Hz, 1H), 4.00 (d, *J* = 12.9 Hz, 1H), 3.13-3.17 (m, 2H), 1.84-1.89 (m, 1H), 1.64-1.70 (m, 1H); ¹³C NMR (150 MHz, DMSO- d_6 , TMS): δ = 170.08, 141.79, 137.16, 131.30, 129.28, 128.77, 128.54, 127.73, 127.29, 119.55, 119.09, 58.24, 39.12, 37.84, 37.41; HRMS (ESI) calcd for C₁₉H₁₇BrNO₂S₂ [M-H]⁻ = 435.3857, found 434.9761; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle

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size: 5 μm, 25 °C, *i*-propanol/hexane = 10/90, flow rate: 0.8 mL/min, λ = 254 nm): t_{minor} = 13.6 min, t_{major} = 16.6 min, ee = 70.5%; [α]²⁵_D = 36.6 (*c* = 0.18 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(4-hydroxybutan-2-yl)thiazol-2(3*H*)-one (3j). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.46 (s, 1H), 7.29-7.32 (m, 2H), 7.24-7.25 (m, 1H), 7.20-7.21 (m, 2H), 3.92 (dd, *J* = 13.2, 19.4 Hz, 2H), 3.45-3.48 (m, 1H), 3.34-3.38 (m, 1H), 3.09-3.15 (m, 1H), 1.70 (s, 1H), 1.60-1.66 (m, 1H), 1.37-1.43 (m, 1H), 0.92 (d, *J* = 6.8 Hz, 3H); HRMS (ESI) calcd for C₁₄H₁₆NO₂S₂ [M-H]⁻ = 294.4203, found 294.1168; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{major} = 10.0 min, t_{minor} = 12.3 min, ee = 71.3%; [α]²⁵_D = 44.3 (*c* = 0.11 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(1-hydroxypentan-3-yl)thiazol-2(3*H*)-one (3*k*). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.45 (s, 1H), 7.30-7.32 (m, 2H), 7.26-7.27 (m, 1H), 7.22-7.25 (m, 2H), 3.94 (dd, *J* = 13.0, 23.0 Hz, 2H), 3.44-3.48 (m, 1H), 3.28-3.33 (m, 1H), 2.95-3.00 (m, 1H), 1.71-1.77 (m, 1H), 1.35-1.54 (m, 3H), 0.75 (t, *J* = 0.75 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.99, 137.28, 131.44, 128.92, 128.58, 127.70, 119.08, 60.20, 39.75, 38.50, 37.01, 29.32, 11.83; HRMS (ESI) calcd for C₁₅H₁₈NO₂S₂ [M-H]⁻ = 308.4469, found 3708.4225; HPLC (Chiralpak AD-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 15/85, flow rate: 0.8 mL/min, λ = 254 nm): t_{major} = 17.0 min, t_{minor} = 21.0 min, ee = 82.9%; [α]²⁵_D = 25.5 (*c* = 0.18 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(1-hydroxy-4-methylpentan-3-yl)thiazol-2(3*H*)-one (3l). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.67 (s, 1H), 7.27-7.34 (m, 5H), 4.03 (d, *J* = 12.8 Hz, 1H), 3.95 (d, *J* = 13.2 Hz, 1H), 4.43 (m, 1H), 3.20 (m, 1H), 2.89 (m, 1H), 1.90 (m, 1H), 1.59 (m, 1H), 1.40 (m, 1H), 0.94 (d, *J* = 6.2 Hz, 3H), 0.79 (d, *J* = 6.2 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.59, 136.77, 129.12, 128.42, 128.34, 127.19, 119.38, 59.95, 41.31, 39.00, 35.14, 32.81, 20.08, 19.88; HRMS (ESI) calcd for C₁₆H₂₀NO₂S₂ [M-H]⁻ = 322.4734, found 322.4475; HPLC (Chiralpak AD-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*-propanol/hexane = 15/85, flow rate: 0.8 mL/min, λ = 254 nm): t_{major} = 14.1 min, t_{minor} = 22.2 min, ee = 73.5%; [α]²⁵_D = 28.7 (*c* = 0.16 in CH₂Cl₂).

(*S*)-4-(Benzylthio)-5-(3-hydroxy-1-(2-methoxyphenyl)propyl) thiazol-2(3*H*)-one (3m). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.24 (s, 1H), 7.28-7.31 (m, 2H), 7.18-7.26 (m, 5H), 6.93-6.95 (m, 1H), 6.85-6.88 (m, 1H), 4.80 (t, *J* = 8.3 Hz, 1H), 3.92 (d, *J* = 13.0 Hz, 1H), 3.85 (s, 3H), 3.42 (t, *J* = 6.3 Hz, 2H), 2.08-2.03 (m, 1H), 1.87-1.93 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 182.53, 155384, 137.30, 130.54, 130.07, 128.87, 128.82, 128.27, 128.14, 127.68, 121.02, 118.96, 110.91, 60.32, 55.39, 39.59, 37.56, 34.67; HRMS (ESI) calcd for C₂₀H₂₀NO₃S₂ [M-H]⁻ = 386.5156, found 386.4575; HPLC (Chiralpak AD-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 23.2 min, t_{major} = 35.3 min, ee = 25.7%; [α]²⁵_D = 144.4 (*c* = 0.90 in EtOAc).

Reaction procedure and characterization for 4

Catalyst III (7.5 mg, 0.02 mmol, 0.1 equiv) was added to a solution of α , β -unsaturated aldehyde **1a** (32.3 mg, 0.24 mmol,

1.2 equiv) and S-methyl-substituted isorhodanine **2a** (30.0 mg, 0.2 mmol, 1.0 equiv) in toluene (1.0 mL). The resulted solution was stirred for 12 h at room temperature. Then, methanol (1.0 mL) and NaBH₄ (7.7 mg, 0.2 mmol, 1.0 equiv) were added. After stirring for 0.5 h, the solvent was evaporated and the crude product was purified though column chromatography over silica gel with hexane/EtOAc (3:1-1:1) as the eluent to yield **4a** (28.2 mg, 50% yield) as a white solid. Compounds **4b-I** were prepared with the same procedure.

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-(methylthio)thiazol-2 (3*H*)-one (4a). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 10.11 (s, 1H), 7.30-7.32 (m, 2H), 7.27-7.28 (m, 2H), 7.21-7.24 (m, 1H), 4.63 (dd, *J* = 6.8, 9.0 Hz, 1H), 3.58-3.66 (m, 2H), 2.27 (s, 3H), 2.15-2.21 (m, 1H), 2.08-2.14 (m, 2H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 173.46, 142.15, 128.86, 128.77, 127.33, 127.11, 121.18, 60.25, 40.35, 37.94, 18.37; HRMS (ESI) calcd for C₁₃H₁₆NO₂S₂ [M+H]⁺ = 282.3937, found 282.4685; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 12.6 min, t_{major} = 20.0 min, ee > 99%; [α]²⁵_D = -28.4 (*c* = 0.26 in CH₂Cl₂).

(S)-4-(Ethylthio)-5-(3-hydroxy-1-phenylpropyl)thiazol-

2(3*H***)-one (4b).** ¹H NMR (600 MHz, CDCl₃, TMS): δ = 10.07 (s, 1H), 7.30-7.32 (m, 2H), 7.27-7.29 (m, 2H), 4.62 (dd, *J* = 7.1, 8.7 Hz, 2H), 3.59-3.66 (m, 2H), 2.72 (dd, *J* = 7.1, 15.0 Hz, 2H), 2.16-2.21 (m, 1H), 2.08-2.14 (m, 1H), 2.00 (s, 1H), 1.20 (t, *J* = 7.0 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 173.28, 142.20, 129.82, 128.82, 127.37, 127.08, 119.77, 60.27, 40.37, 38.25, 25.43, 14.99; HRMS (ESI) calcd for C₁₄H₁₈NO₂S₂ [M+H]⁺ = 296.4203, found 296.3168; HPLC (Chiralpak OZ-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*-propanol/hexane = 10/90, flow rate: 0.6 mL/min, λ = 254 nm): t_{minor} = 49.6 min, t_{major} = 79.7 min, ee = 92.7%; [α]²⁵_D = -26.6 (*c* = 0.18 in CH₂Cl₂).

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-((naphthalen-2-ylmethyl)thio)thiazol-2(3*H*)-one (4c). ¹H NMR (600 MHz, DMSO- d_6 , TMS): δ = 11.46 (s, 1H), 7.81-7.91 (m, 2H), 7.82-7.84 (m, 1H), 7.66 (s, 1H), 7.48-7.53 (m, 2H), 7.43 (dd, *J* = 1.7, 8.4 Hz, 1H), 7.04 (t, *J* = 7.3 Hz, 1H), 6.93 (t, *J* = 7.8 Hz, 2H), 6.76 (d, *J* = 7.3 Hz, 2H), 4.41 (t, *J* = 4.8 Hz, 1H), 4.17-4.27 (m, 3H), 3.15-3.16 (m, 2H), 1.85-1.92 (m, 1H), 1.64-1.71 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 170.64, 142.71, 135.29, 133.39, 132.71, 129.20, 128.77, 128.70, 128.07, 127.63, 127.53, 127.39, 126.84, 126.77, 126.45, 119.14, 58.83, 38.69, 37.98; HRMS (ESI) calcd for C₂₃H₂₀NO₂S₂[M-H]⁻ = 406.5483, found 406.6723; HPLC (Chiralpak AD-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 29.4 min, t_{major} = 43.2 min, ee = 88.3%; [α]²⁵_D = 46.2 (*c* = 0.16 in EtOAc).

(*S*)-4-((3-Fluorobenzyl)thio)-5-(3-hydroxy-1-phenylpropyl)thiazol-2(3*H*)-one (4d). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 10.03 (s, 1H), 7.26-7.31 (m, 1H), 7.18-7.23 (m, 2H), 7.11-7.14 (m, 2H), 6.90-6.94 (m, 3H), 4.22 (t, *J* = 8.0 Hz, 1H), 3.86 (s, 2H), 3.41-3.54 (m, 2H), 2.05-2.16 (m, 1H), 1.83-1.95 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 173.10, 164.47, 161.20, 141.86, 139.43, 131.30, 130.32, 130.21, 128.81, 127.28, 127.09, 124.63, 124.60, 118.61, 115.96, 115.67, 114.78,

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114.50, 60.14, 40.35, 38.87, 38.85, 38.09; HRMS (ESI) calcd for $C_{19}H_{17}FNO_2S_2$ [M-H]⁻ = 374.4801, found 374.3347; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 0.8 mL/min, λ = 254 nm): t_{minor} = 18.9 min, t_{major} = 22.3 min, ee = 78.7%; [α]²⁵_D = 78.6 (*c* = 0.14 in CH₂Cl₂).

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-((2-methylbenzyl)thio) thiazol-2(3*H*)-one (4e). ¹H NMR (300 MHz, CDCl₃, TMS): δ = 11.45 (s, 1H), 7.19-7.24 (m, 3H), 7.09-7.18 (m, 3H), 7.02-7.04 (m, 1H), 6.93-6.95 (m, 2H), 4.41-4.45 (m, 1H), 4.20 (t, *J* = 7.7 Hz, 1H), 4.05 (s, 2H), 3.13 (s, 2H), 2.36 (s, 3H), 1.83-2.00 (m, 1H), 1.60-1.72 (m, 1H); ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 170.62, 142.89, 136.95, 135.27, 130.93, 130.13, 129.35, 128.97, 128.12, 127.47, 126.98, 126.39, 119.24, 58.89, 38.07, 36.83, 19.14; HRMS (ESI) calcd for C₂₀H₂₀NO₂S₂ [M-H]⁻ = 370.5162, found 370.3602; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 30/70, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 29.4 min, t_{major} = 43.2 min, ee = 97.5%; [α]²⁵_D = 57.8 (*c* = 0.09 in CH₂Cl₂).

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-((4-methylbenzyl)thio) thiazol-2(3*H*)-one (4f). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.66 (s, 1H), 7.28 (s, 1H), 7.25 (s, 1H), 7.19-7.22 (m, 1H), 7.09-7.11 (m, 4H), 7.06-7.07 (m, 2H), 4.38 (t, *J* = 7.9 Hz, 1H), 3.86 (dd, *J* = 9.4, 13.1 Hz, 2H), 3.37-3.49 (m, 2H), 2.31 (s, 3H), 2.05-2.10 (m,1H), 1.81-1.88 (m, 1H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.67, 141.97, 137.53, 134.11, 131.03, 129.50, 128.85, 128.75, 127.35, 137.01, 118.88, 60.20, 40.26, 39.32, 37.99, 21.14; HRMS (ESI) calcd for C₂₀H₂₀NO₂S₂ [M-H]⁻ = 370.5162, found 370.5243; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 30/70, flow rate: 1.0 mL/min, λ = 254 nm): t_{minor} = 7.9 min, t_{major} = 9.4 min, ee = 77.1%; [α]²⁵_D = -50.6 (*c* = 0.17 in CH₂Cl₂).

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-((2-nitrobenzyl)thio)thiazol-2(3*H*)-one (4g). ¹H NMR (300 MHz, CDCl₃, TMS): δ = 9.76 (s, 1H), 8.01 (dd, *J* = 1.1, 7.9 Hz, 1H), 7.47 (td, *J* = 1.1, 7.3, 14.8 Hz, 1H), 7.39 (td, *J* = 1.5, 7.9, 15.9 Hz, 1H), 7.27-7.29 (m, 1H), 7.19-7.23 (m, 2H), 7.13-7.16 (m, 1H), 7.09-7.11 (m, 2H), 4.29-4.35 (m, 2H), 4.18-4.22 (m, 1H), 3.43 (t, *J* = 6.3 Hz, 2H),1.99-2.11 (m, 2H), 1.76-1.88 (m, 1H); ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 172.63, 147.93, 141.72, 133.58, 132.75, 132.19, 132.12, 128.96, 128.85, 127.26, 127.11, 125.74, 118.17, 60.13, 40.46, 38.28, 36.91; HRMS (ESI) calcd for C₁₉H₁₇N₂O₄S₂ [M-H]⁻ = 401.4872, found 401.2156; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 0.8 mL/min, λ = 254 nm): t_{minor} = 29.4 min, t_{major} = 43.2 min, ee = 94.1%; [α]²⁵_D = 25.5 (*c* = 0.22 in CH₂Cl₂).

(*S*)-5-(3-Hydroxy-1-phenylpropyl)-4-((4-nitrobenzyl)thio)thiaz-ol-2(3*H*)-one (4*h*). ¹H NMR (300 MHz, CDCl₃, TMS): δ = 10.26 (s, 1H), 8.05 (d, *J* = 8.7 Hz, 2H), 7.31-7.37 (m, 2H), 7.18-7.23 (m, 3H), 7.04-7.07 (m, 2H), 4.40 (t, *J* = 7.8 Hz, 1H), 3.93 (dd, *J* = 13.1, 31.5 Hz, 2H), 3.47-3.51 (m, 2H), 2.05-2.13 (m, 1H), 2.02 (s, 2H), 1.84-1.96 (m, 1H); ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 173.19, 147.23, 144.42, 141.64, 131.90, 129.71, 128.80, 127.18, 123.98, 118.14, 59.95, 40.32, 38.54, 31.60, 25.35; HRMS (ESI) calcd for $C_{19}H_{17}N_2O_4S_2$ [M-H]⁻ = 401.4872, found 401.3219; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 µm, 25 °C, *i*-propanol/hexane = 30/70, flow rate: 0.8 mL/min, λ = 254 nm): t_{minor} = 20.6 min, t_{major} = 24.9 min, ee = 93.3%; [α]²⁵_D = -46.0 (*c* = 0.20 in CH₂Cl₂).

(*S*)-4-(((*S*-(3-Hydroxy-1-phenylpropyl)-2-oxo-2,3-dihydrothiazol-4-yl)thio)methyl)benzonitrile (4i). ¹H NMR (300 MHz, CDCl₃, TMS): δ = 10.22 (s, 1H), 7.44-7.54 (m, 2H), 7.28-7.31 (m,2H), 7.14-7.22 (m, 3H), 7.04-7.08 (m, 2H), 4.17 (t, *J* = 8.0 Hz, 1H), 3.90 (dd, *J* = 13.0, 29.7 Hz, 2H), 3.44-3.57 (m, 2H), 2.06-2.16 (m, 1H), 1.86-1.97 (m, 1H); ¹³C NMR (75 MHz, CDCl₃, TMS): δ = 173.21, 142.40, 141.70, 132.52, 131.72, 129.63, 128.86, 127.24, 127.21, 118.58, 118.27, 111.37, 59.96, 40.28, 38.88, 38.28; HRMS (ESI) calcd for C₂₀H₁₈N₂O₂S₂ [M-H]⁻ = 381.4991, found 381.5132; HPLC (Chiralpak OZ-H, column size: 0.46 cml.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*propanol/hexane = 25/75, flow rate: 0.7 mL/min, λ = 254 nm): t_{minor} = 33.8 min, t_{major} = 40.7 min, ee = 82.5%; [α]²⁵_D = 35.0 (*c* = 0.12 in EtOAc).

(S)-4-((4-(Tert-butyl)benzyl)thio)-5-(3-hydroxy-1-phenylpropyl)thiazol-2(3H)-one (4j). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 9.78 (s,1H), 7.34-7.35 (m, 2H), 7.28-7.31 (m, 2H), 7.22-7.23 (m, 1H), 7.19-7.21 (m, 2H), 7.14-7.16 (m, 2H), 4.43 (t, J = 7.6 Hz, 1H), 4.14 (dd, J = 7.0, 14.4 Hz, 2H), 3.96 (d, J = 12.9 Hz, 1H), 3.84 (d, J = 12.9 Hz, 1H), 3.43-3.46 (m, 1H), 3.33-3.37 (m, 1H), 2.03-2.10 (m, 1H), 1.78-1.84 (m, 1H), 1.32 (s, 9H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.35, 150.35, 141.56, 133.60, 130.31, 128.29, 128.19, 128.10, 127.58, 126.86, 126.67, 126.52, 125.22, 125.01, 124.95, 118.55, 59.54, 39.71, 38.54, 37.48, 34.09, 30.81; HRMS (ESI) calcd for C₂₃H₂₆NO₂S₂ [M-H]⁻ = 412.5960, found 412.6328; HPLC (Chiralpak OZ-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 μm, 25 °C, ipropanol/hexane = 10/90, flow rate: 0.7 mL/min, λ = 254 nm): $t_{\text{minor}} = 40.6 \text{ min}, t_{\text{major}} = 50.0 \text{ min}, \text{ ee} = 82.5\%; [\alpha]_{D}^{25} = -26.7 (c)$ = 0.15 in EtOAc).

(S)-2-((5-(3-Hydroxy-1-phenylpropyl)-2-oxo-2,3-dihydrothiazol-4-yl)thio)acetonitrile (4k). ¹H NMR (600 MHz, CDCl₃, TMS): δ = 10.32 (s, 1H), 7.28-7.32 (m, 4H), 7.22-7.24 (m, 1H), 4.71 (t, *J* = 7.7 Hz, 1H), 3.64-3.66 (m, 2H), 3.55 (d, *J* = 17.2 Hz, 2H), 3.41 (d, *J* = 17.0 Hz, 2H), 2.12-2.14 (m, 3H); ¹³C NMR (150 MHz, CDCl₃, TMS): δ = 172.32, 141.20, 135.06, 135.06, 128.53, 128.53, 126.82, 115.81, 115.31, 59.22, 39.95, 37.98, 29.20, 19.75; HRMS (ESI) calcd for C₁₄H₁₃N₂O₂S₂ [M-H]⁻ = 305.4032, found 305.3396; HPLC (Chiralpak OZ-H, column size: 0.46 cmI.D. × 25 cmL, particle size: 5 μm, 25 °C, *i*-propanol/hexane = 20/80, flow rate: 1.0 mL/min, λ = 254 nm): *t*_{minor} = 22.4 min, *t*_{major} = 27.6 min, ee = 76.1%; [α]²⁵_D = -24.8 (*c* = 0.29 in EtOAc).

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- 12 The structure of compound derived from molecule **3a** was determined by X-ray crystal analysis. CCDC-1432606 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge via <u>www.ccdc.cam.ac.uk</u>.