Organic & Biomolecular Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/obc

ARTICLE



Synthesis and anti-tubercular activity of N^2 arylbenzo[g]isoquinolin-5,10-dione-3-iminium bromides

Nguyen,^d K. Huygen,^c L. Maes,^b P. Cos^b and K. Abbaspour Tehrani^a*

Received 00th January 20xx, Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/

Tuberculosis has remained a challenge for medicinal chemists worldwide. In the framework of a collaborative program to identify and evaluate novel antitubercular candidate compounds, the biological properties of benzo[g]isoquinoline-5,10diones have been found to be very promising. In this paper we have further expanded the library by incorporation of an amidinium molety into the benzo[g]isoquinoline-5,10-dione scaffold. The presence of this functional group also increased the solubility of the quinones in polar solvents. To that purpose N^2 -arylbenzo[g]isoquinolin-5,10-dione-3-iminium bromides were synthesized in a straightforward way by means of a reaction of anilines with 2-(bromomethyl)-3-(cyanomethyl)-1,4-dimethoxynaphthalene. Following the biological evaluation, N^2 -(4-chlorophenyl)-1,4-dihydro-5,10dimethoxybenzo[q]isoquinoline-3(2H)-iminium bromide (MIC = 0.59 μ M, CC₅₀ = 11.25 μ M, SI = 19.07), was selected as the most promising representative. Apart from nano-molar anti-mycobacterial activity, the compound was able to target intracellular residing Mycobacterium tuberculosis and the susceptibility of a multi-drug-resistant strain towards the

G. Rotthier,^{a,‡} D. Cappoen,^{b,c‡} Quang Trung Nguyen,^d Tuyet Anh Dang Thi,^d V. Mathys,^e Van Tuyen

Introduction

Tuberculosis (TB) is an infectious pulmonary disease caused by the weakly Gram-positive Mycobacterium tuberculosis (Mtb). Transmitted through aerosol, Mtb creates more victims than any other single infectious agent, second only to HIV. A high intrinsic tolerance towards most conventional chemotherapeutics and the increasing incidence of acquired resistance to first and second line TB drugs cause an ongoing demand for novel compounds for treatment. Further fuelled by the HIV pandemic, the search for a cure of multi-drug- and extensively-drug-resistant strains is of paramount importance. In our program to identify and evaluate candidate molecules for

compound was confirmed.

- *‡* Both authors contributed equally to the paper.
- *Corresponding author: Kourosch Abbaspour Tehrani. Tel. 32 03 265 32 26, fax: 32 03 265 32 33, E-mail: <u>Kourosch.AbbaspourTehrani@uantwerpen.be</u> Electronic Supplementary Information (ESI) available: ¹H and ¹³C NMR spectra for **7**,

the treatment of TB, it was recently discovered that benzo[*j*]phenanthridine-7,12-diones showed remarkable potency against Mtb.1 This fact corresponds with the observation that structurally related angular phenanthridine dione derivatives such as the naturally occurring phenanthroviridin² and the calothrixins³ also possess several attractive biological activities. In addition, the biological properties of this compound class could be tailored by varying the substitution pattern and the incorporation of functional groups. Until now, the focus of modifying the tetracyclic scaffold shown in figure 1 has been restricted to the A and D rings.1

Our research has since long been focused on the synthesis of 2aza-anthraquinones and analogs.⁴ Since the previously described 2-azaanthraquinones suffer from poor solubility in polar solvents like DMSO and water, the strategy was to increase compound solubility and to study its impact on the bioactivity of the compounds. In an attempt to further increase the biological activity of the 2-aza-anthraquinone scaffold, a straightforward synthesis was envisaged to modify the C-ring. As a strategy, it was chosen to generate a small library in which a functional amidine group was incorporated in the quinone



Figure 1 General benzophenanthridine-7-12-dione structure used as the scaffold in our program and the target N-arylbenzo[g]isoquinolin-5,10-dione-3-iminium bromides.

^{a.} Organic Synthesis, Faculty of Sciences, University of Antwerp, Groenenborgerlaan 171, B-2020 Antwerp Belgium.

^{b.} Laboratory of Microbiology, Parasitology and Hygiene (LMPH), S7, Faculty of Pharmaceutical, Biomedical and Veterinary Sciences, University of Antwerp, Universiteitsplein 1, B-2610 Wilrijk, Belgium.

Scientific Service Immunology, O.D. Communicable & Infectious Diseases. Scientific Institute of Public Health (Site Ukkel), Engelandstraat 642, B-1180 Ukkel, Belaium.

^d Institute of Chemistry, Vietnam Academy of Sciences and Technology, 18-Hoang Quoc Viet, Cau Giav, Ha Noi, Vietnam.

^{e.} Program Tuberculosis and Mycobacteria, Service Bacterial Diseases, O.D. Communicable & Infectious Diseases, Scientific Institute of Public Health (Site Ukkel), Engelandstraat 642, B-1180 Ukkel, Belgium.

^{8, 9, 11}a-k, 12a-j, 13i, j and 14. In addition the technical details of the materials and methods for the biological evaluation are included. See DOI: 10.1039/x0xx00000x

ARTICLE

scaffold. The choice of this group is not surprising, considering its occurrence in the nucleobases and their presence as a fundamental entity in medicinal chemistry.⁵

While retaining the A, B, C ring structure, the D ring was not present in this compound library in order to conveniently generate the amidinium salts and increase the solubility. The basic amidine functionality is present in many active pharmaceutical ingredients, such as the antiprotozoal/ antipneumocystic pentamidine (1), the widely used veterinary anthelminthic pyrantel (2), and the anticancer compounds gefitinib (3) and dasatinib (4) (Figure 2).



Figure 2 Examples of biologically active amidines.

Therefore we were interested in incorporating the amidine moiety into this promising class of bioactive compounds, and evaluating its effect on the anti-mycobacterial activity.

Secondary amidines have been synthesized by various methods.⁶ The oldest, though still commonly used method to prepare amidines is the Pinner synthesis.⁷ In this reaction, amidines are formed by the acid-catalyzed reaction of an alcohol with a nitrile, forming an N-protonated imidate or "Pinner salt", followed by substitution of the alkoxy group with an amine. Several intramolecular variations of the Pinner reaction have been reported for the synthesis of 2-iminopiperidines⁸ and their annulated analogs.⁹



Scheme 1 Retrosynthetic analysis for *N*-arylbenzo[*g*]isoquinolin-5,10-dione-3-iminium bromides.

Therefore, it was envisaged that a suitable aminonitrile would form the N^2 -arylbenzo[g]isoquinolin-5,10-dione-3-iminium bromides after intramolecular Pinner-type reaction.

This aminonitrile could be prepared conveniently from a 2-(bromomethyl)-3-(cyanomethyl)naphthalene derivative, made in turn from 2-methylnaphthoquinone (5) (Scheme 1). The heterocyclization of the simple (2-bromomethylphenyl)acetonitrile has been effectuated in low yields (12-68%) by means of a reaction with aliphatic and aromatic amines.¹⁰

Results and Discussion

Following a procedure described by Ito et al., 2-methyl-1,4naphthoquinone (vitamin K3) was reduced to the corresponding which was then methylated hydroguinone. using dimethylsulfate in the presence of sodium methoxide.¹¹ Bromomethylation at the 3-position using paraformaldehyde in a concentrated hydrobromic acid solution yielded the substitution product 7 in good yield.¹² This benzylic bromide was reacted with an excess of KCN in acetonitrile/water in order to obtain the corresponding nitrile 8. Benzylic bromination of the latter, by means of N-bromosuccinimide (NBS) in the presence of 0.05 equivalents of benzoyl peroxide (BPO), yielded the benzylic bromide 9 in 80% yield (Scheme 2). Compound 9 is a new ω -halonitrile synthon, which upon tandem addition/cyclisation reaction with organometallic reagents could result in cyclic imine 10,¹³ which may be further oxidized to give 3-substituted 2-azaanthraquinones. The reaction of 9 with MeLi or MeMgCl at -78°C or 0°C, however, only led to complex reaction mixtures.



Scheme 2 Synthesis of 2-(bromomethyl)-3-(cyanomethyl)-1,4-dimethoxynaphthalene (9).

The reaction of the bifunctional compound 9 with aniline in isopropanol under reflux directly leads to the cyclic amidine salt 11a after simple evaporation of the solvent. The disappearance of the nitrile absorption in the IR spectrum (2247 cm⁻¹) was diagnostic for the ring closing reaction. Attempts to convert the cyclic amidinium salt to the corresponding free base, by treating it with aqueous NaOH (1 M) and subsequent extraction with organic solvents, all failed, indicating the high lability of these compounds. In principle hydrolysis of the C=N functionality in 11a could lead to N-phenyl-5,10-dimethoxy-1,4-dihydrobenzo[g]-isoquinoline-3(2H)-one.¹⁴ Upon treatment of the HBr salt 11a with aqueous HCl (2 M) at room temperature, besides degradation products, no trace of this compound was observed. Fortunately, compound 11a rather conveniently crystallized from the reaction mixture upon cooling to room temperature. Because it was difficult to remove eventual traces of unreacted aniline, this reagent was chosen as the limiting reagent (0.95 equivalents). Traces of unreacted bromonitrile could be removed by means of a diethyl ether washing step. Having the conditions at hand for the synthesis and the purification of amidinium salt 11a, a range of anilines was reacted with 9. The latter reacts with both electron- rich and -poor anilines to give the corresponding cyclic amidinium salts 11a-k in moderate to good yields (Table 1). Interestingly, for the amidinium salts 11h-j, derived from ortho-substituted anilines, the ¹H NMR signals for the methylene protons are seen as doublets. The diastereotopic nature of these protons indicates that the N-aryl substituent and the amidine fragment are non-coplanar due to the non-symmetrical substitution of the aryl group. By recording the ¹H NMR of **11h** in DMSO at 110 °C no coalescence of these signals could be induced.

 Table 1
 Synthesis of 5,10-dimethoxy- N^2 -aryl-1,2-dihydrobenzo[g]isoquinolin-3(4H)-iminium bromides



In order to obtain the corresponding quinones from **11a-k** (Table 1), some classical approaches were attempted, such as silver oxide oxidation in dioxane with nitric acid¹⁵ (Table 2, entry 1). Unfortunately only the unchanged starting material was observed. Krapcho-conditions were also attempted by using lithium bromide in DMF as the demethylating agent but no reaction at all was observed in this case and starting material was unaltered (Table 2, entry 2).



Scheme 3 Formation of O,O'-diacetyl esters.

Heating the starting material 11b with LiBr in DMF under an oxygen atmosphere resulted in complete degradation of the product (Table 2, entry 2). Aryl ether cleavage was accomplished by heating the starting material 11b overnight in 11% HBr in acetic acid, and initially afforded the desired quinone, but when this method was repeated it proved not to be reproducible, and a lot of side product formation was observed together with the demethylation/oxidation product (Table 2, entry 6). Since these amidinium salts 12 cannot be purified by any straightforward method, the search for a reproducible method, affording pure quinones was continued. At first variations in the reaction temperature were evaluated, but this had no particular effect on the reaction outcome (Table 2, entries 4, 5). As varying amounts of water in the acetic acid were suspected to be the source of the irreproducible results, it was attempted to use dry acetic acid (Table 2, entries 6-8). In order to obtain dry acetic acid, various equivalents of acetic anhydride were added to the reaction mixture. With high levels anhydride (60 equivalents), the O,O'of acetic diacetylhydroquinones 13i,j were obtained (Scheme 3) (yield not determined). At lower levels of acetic anhydride, no change in the reaction outcome could be observed (Table 2, entry 9). When 5% water was added to the reaction mixture of 9b and 11% HBr in acetic acid, side product formation was almost completely suppressed, and only the corresponding quinones 12 were isolated (Table 2, entry 9). This method then proved to be applicable to all analogs 11a-j, and compounds 12a-j were obtained in good yields without an extra purification step (Table 3). The 4-methoxyphenyl analog 11k, under these conditions, gave a mixture of partially demethylated quinones which could not be separated.

ARTICLE



^a Yield after workup

 Table 3 Oxidative demethylation of 11





^a Yield after crystallization from isopropanol. ^b The ethoxycarbonyl group was hydrolyzed to a carboxylic acid under the reaction conditions used.

Mechanism

Since the reaction of **9** with aniline proceeds in isopropanol, a Pinner type mechanism could be expected, which would imply the formation of the imidate **15**. Subsequent intramolecular addition of the aniline nitrogen atom across the imidate C=N of **15** would lead to the amidine **11a**. Interestingly, no trace of the imidate **15** was observed during LC-MS monitoring of the reaction mixture, which may suggest a different mechanism. The fact that the conversion of **9** into **11a** also proceeds in 1,2dichloroethane and DMSO, albeit in a lower yield, also indicates that a different mechanism is in play. The most likely intermediate is the secondary aniline **14**, which was independently prepared in 94% yield by reaction of **9** with an excess of aniline in dichloromethane at room temperature.



Scheme 4 Plausible reaction mechanism for the cyclic amidine formation.

Journal Name

This journal is © The Royal Society of Chemistry 20xx

Treatment of this compound **14** with 1 equivalent of HBr in acetic acid at room temperature furnished the ring closed amidinium salt **11a** after 20 minutes, in 63% yield. This fact suggests a mechanism in which the secondary amine in **14** adds across an *in situ* formed imidoyl bromide **16**, as was suggested by Ogonor, thus achieving ring closure and amidinium salt formation (Scheme 4).¹⁶ The addition of hydrogen bromide to a nitrile is a classical reaction for the preparation of imidoyl bromides or iminium bromides.¹⁷ Since never such an imidoyl bromide or the derived imidate **15** was isolated, the direct intramolecular addition of the amine across the protonated nitrile can be considered as well.

Biology

The series of amidinium salts 11a-j and 12a-j were tested for their antimycobacterial activity against Mycobacterium tuberculosis as shown in Table 4. A model based on a luminescent Mtb H37Rv strain (H37Rvlux) was used. As reported previously in the literature, this technique is a sensitive and reproducible tool, able to replace fastidious plating.^{1b,18} Antitubercular properties of the synthesized compounds are reported by a reduction of luminescence emitted by a culture exposed to the compound, compared with a negative control culture. After six days of exposure of Mtb H37Rvlux to serial dilutions of the compounds, the potency was calculated as the Minimal Inhibitory Concentration (MIC) at which the mycobacterial growth is reduced by 90%. In parallel, the acute toxicity of the analogs against eukaryotic J774 A.1 cells, a murine macrophage-like monocyte cell line, was studied in a neutral uptake assay. The macrophage model was chosen since macrophages function as the most frequent host cell for Mtb in a tuberculosis infection. The neutral red uptake assay relies on the ability of viable cells to bind and incorporate the dye neutral red.¹⁹ The acute toxic concentration (CC₅₀) of a compound is defined as the concentration at which the uptake of the neutral red dye by the cells is reduced by 50%. By dividing the IC_{50} by the MIC the selectivity index (SI) could be derived.

As shown in Table 4, quinones 12a-j were more active than their corresponding dimethoxylated precursors 11a-j. This confirms earlier findings on the necessity of the presence of a quinone moiety in the structure for its activity. In the less active compounds from this series it can be observed that the difference in toxicity between the quinones 12 and their precursors 11 is directly proportional to the anti-mycobacterial activity. This is illustrated by comparing the unsubstituted derivative 12a and 12g carrying a carboxyl substituent and its dimethoxylated precursor 11g. In view of the similar Selectivity Index (SI) for compounds 12a, 12f, 12g and their precursors 11a, 11f, 11g no advantage can be ascribed to the use of these quinones. On the other hand, the more active members of the library, like the halogenated quinone derivatives 12c, 12d and 12j, have stronger anti-mycobacterial activities than the nonhalogenated quinones 12a, 12b, 12e and 12f. Further, one may notice that the bioactivity is not only influenced by the nature of the N-phenyl substituents but also by their position on the Nphenyl moiety. To this extent, *para* substituted analogs (11c,

11d, **12c**, **12d**) were compared with *ortho* substituted compounds (**11h**, **11i**, **12i**, **12j**) from which it can be seen that the former is favored in terms of bioactivity. Most remarkable was the disproportional difference in SI between the halogenated quinones **12c**, **12d** and **12j** and their respective dimethoxylated precursors **11c**, **11d** and **11j**. While the CC_{50} concentration remains of the same order, the MIC concentrations are lowered to nanomolar for **12d** and **12j**. These observations confirm the importance of the quinone functionality for the anti-mycobacterial properties. In the same time the importance of the compound substitution pattern as a basis for selectivity and for directing the quinone functionality to its target is highlighted.

 Table 4 Growth inhibition of *Mtb* and acute toxicity against macrophages by the *N*-arylbenzo[g]isoquinolin-5,10-dione-3-iminium bromides **12a-j** and their dimethoxylated precursors **11a-j**.

		$MIC^{a}\left(\mu M\right)$	$CC_{50}{}^{b}(\mu M)$	SI ^c
11a	Н	21.49	56.55	2.63
11b	4-Me	21.49	23.25	1.08
11c	4-Cl	10.4	24.12	2.32
11d	4-Br	10.13	21.01	2.07
11e	4-NO ₂	15.75	49.23	3.13
11f	4-EtOC(O)	12.43	124.21	9.99
11g	3-EtOC(O)	23.41	98.25	4.20
11h	2-Cl	15.92	21.14	1.33
11i	2-Br	14.83	19.41	1.31
11j	2-Br-4-Me	10.84	23.45	2.16
12a	Н	11.21	31.25	2.79
12b	4-Me	2.68	18.15	6.77
12c	4-Cl	1.16	28.51	24.58
12d	4-Br	0.65	15.34	23.60
12e	4-NO ₂	2.79	19.48	6.98
12f	4-COOH	23.6	145.12	6.15
12g	3-COOH	38.11	153.24	4.02
12h	2-Cl	3,00	29.12	9.71
12i	2-Br	1.85	31.24	16.89
12j	2-Br-4-Me	0.59	11.25	19.07
\mathbf{INH}^{d}		0.13	>100	
RIF ^e		0.18	>100	

^a The Minimal Inhibitory Concentration (MIC) at which 99% growth inhibition of the *Mtb* H37Rv^{lux} lab strain was observed as calculated from triplicate cultures (SD values < 10%). ^b The Cytotoxic Concentration (CC₅₀) at which viability of the J774 A.1 macrophages was reduced by 50% as calculated from triplicate cultures (SD values < 10%). ^c The selectivity index (*SI*), calculated as CC₅₀/MIC. ^d INH, isoniazid used as a positive control. ^e RIF, Rifampicin, used as a positive control.

To study the potential of the compound class for the treatment of drug-resistant tuberculosis, the susceptibility to the compounds of a multi-drug-resistant *Mtb* LAM-1 strain was tested, as shown in Figure 3. The LAM-1 strain was spoligotyped and resistance against isoniazid, rifampicin, rifabutine and prothionamide was confirmed by the National Reference Laboratory of Belgium. The critical concentration at which the mycobacterial growth is reduced by 99% was

ARTICLE

determined with fluorometric BACTEC MGIT 960TM Mycobacteria Growth Indicator Tubes.²⁰ The growth of a culture, 100 times diluted upon inoculation (C/100), is also measured. By comparing the Growth Index (GI) of this culture with the cultures exposed to the compound, the critical concentration can be determined. At day 6, the C/100 has reached a GI of 1232 units. For 4-chlorophenyl derivative **12c**, the GI at 0.50 μ M was 2132 while at 2.50 μ M the GI was 723 units, placing the critical concentration of **12c** between these two concentrations and confirming the susceptibility of the





Figure 3 Susceptibility of an MDR *Mtb* LAM-1 strain to the compounds **12c**, **12d** and **12j**. Growth of MDR *Mtb* was monitored in BACTEC MGIT 960TM. ^{a,b} Growth was compared to an untreated bacterial suspension and an untreated bacterial suspension diluted 100 times (C/100) to study the minimal inhibitory concentration MIC. ^c Resistance was confirmed with isoniazid (INH). ^d Mfx: moxifloxacin, a second line TB drug included as control at 1.14 μ M.

Figure 4 Growth inhibition of intracellular replicating *Mtb* by derivatives **12c**, **12d** and **12j**. Growth inhibition of *Mtb* H37Rv inside J774 A.1 macrophages. Results are presented as mean + SEM Relative Light Units (RLU) of triplicate cultures. Treatment of the infected cells started 24 hours post infection and lasted for 3 days. The first line drug INH was used as a positive control at 0.1 μ M.

This journal is © The Royal Society of Chemistry 20xx

MDR strain towards the derivative. Susceptibility of the MDR LAM-1 strain for 4-bromo derivative **12d** was similar with a GI of 1852 units at 0.50 μ M and 652 units at 2.50 μ M. For compound **12j**, the GI measured 1342 units after 6 days' exposure while the GI at 0.50 μ M was 354 units.

As macrophages are an important host for Mtb, a macrophage infection model as described earlier²¹ was used to study if the selected derivatives **12c**, **12d** and **12j** are able to reach their target through the membrane structure of the macrophages. In the macrophage infection assay setup, the murine J774 A.1 macrophage like monocyte cell line were first infected with Mtb H37Rv^{lux}. After 24 hours of infection, the infected macrophages were washed and treated with a serial dilution of the compounds for 4 days. Afterwards, the monolayer was washed, lysed and the mycobacterial load estimated by luminometry.

At the four concentrations tested (0.25 μ M, 0.50 μ M, 2.50 μ M and 5.00 μ M), derivatives **12c**, **12d** and **12j** showed potency to reduce the bacterial load inside monocytes. For compound **12c**, a one log10 reduction, considered a MIC, was only achieved at 5 μ M with a 91.4 \pm 2.5% reduction in bacterial load. This reduction fell back to 74.4 \pm 2.4% at 2.5 μ M for derivative **12c**. Similarly for compound **12d**, the bacterial replication inside macrophage was reduced by 92.5 \pm 1.18% at 5.0 μ M while at 2.5 μ M the growth inhibition dropped to 83.5 \pm 4.1%. For compound **12j**, a MIC was achieved at both 5.0 μ M and 2.5 μ M with 95.0 \pm 0.6% and 92.2 \pm 1.2%. At 0.5 μ M a reduction of 82.6 \pm 3.6% was achieved for compound **12j** (Figure 4).

Conclusion

In summary, a new class of benzo[g]isoquinoline-5,10-diones carrying an amidine functionality was synthesized by reaction of anilines with 2-(bromomethyl)-3-(cyanomethyl)-1,4dimethoxynaphthalene, which was in turn synthesized efficiently from 2-methylnaphthoquinone (vitamin K3). After oxidative dealkylation of the dimethoxylated compounds 11a-j with hydrobromic acid in acetic acid, the corresponding quinones 12a-j were obtained in good yields. From the biological evaluation it can be concluded that the quinone 12a-j derivatives were much more active than their dimethoxylated precursors 11a-j. Compounds carrying a halogen substituent on the phenyl ring **12c** (MIC = 1.16 μ M, CC₅₀ = 28.51 μ M, SI = 24.58), **12d** (MIC = 0.65μ M, CC₅₀ = 15.34μ M, SI = 23.60) and **12j** (MIC = 0.59μ M, CC₅₀ = 11.25μ M, SI = 19.07) showed an improved selectivity index compared to derivatives carrying alternative substituents. This information confirms the necessity of the quinone functionality in the structures for their activity and it can be argued that the selectivity for the target is tailored by the substitution pattern. The activity against intracellular replicating Mtb and multi-drug-resistant Mtb confirmed these compounds as a promising scaffold. In addition, the straightforward synthesis of these amidine-quinones will allow the generation of more complex quinone derivatives as potential anti-tubercular compounds.

General experimental

¹H (¹³C) NMR spectra were recorded at 400 (100) MHz on a Bruker spectrometer using CDCl₃ as solvent and TMS as internal standard, or DMSO with the residual solvent peak as a standard in the case of the amidinium salts. The ¹³C chemical shifts were referenced to residual solvent signals at δ_C 77.00 (CDCl₃) or δ_C 39.52 (DMSO). *J* values are given in Hertz (Hz) and chemical shifts are given in ppm. Melting points were determined on a Büchi melting point apparatus B-540 and are uncorrected. UPLC analyses were obtained with an Acquity (Waters) with a reversed phase C-18 column (Halo, 2.1 × 30 mm, 2.7 µm). The mobile phase (water/acetonitrile) contained 0.1% formic acid. The standard gradient consisted of a 1.5 min run from 5% to 95% acetonitrile at a flow of 0.6ml/min with diode array UV detection.

TLC-analysis was performed on glass backed plates (Merck) coated with 0.2 mm silica $60F_{254}$. Solvents used in extraction and purification were distilled prior to use. Flash column chromatography was performed by means of an automated chromatography system with Silica Flash Cartridges (Grace®). For high resolution mass spectrometric analysis, samples were dissolved in CH₃CN and diluted to a concentration of approximately 10^{-5} mol/L. Compound **6** was prepared according to literature procedures.¹¹

Synthesis of 3-(bromomethyl)-1,4-dimethoxy-2methylnaphthalene (7)

This compound was synthesized according to a modified literature procedure.¹¹ To 1,4-dimethoxy-2-methylnaphthalene (4) (15.89 g, 79 mmol) was added a 33% w/w solution of hydrobromic acid in acetic acid (14.5 g, 59 mmol HBr) and the mixture was stirred for 10 min at room temperature. After this time, paraformaldehyde (7.08 g, 79 mmol, 1 Eq.) was added along with another 30 g of 33 % HBr solution in acetic acid (122 mmol HBr). The mixture was stirred at room temperature for 4 hours. After this time, water (200 ml) was added to the mixture, and the aqueous phase was extracted with CH_2Cl_2 (3 × 200 ml). The combined organic layers were washed with saturated NaHCO3 solution until visible gas evolution had stopped, washed with brine (80 ml) and dried over anhydrous MgSO₄. After filtration and removal of the solvent in vacuo, a brown oil was obtained which was purified by flash chromatography (100% heptane) giving a white powder. Yield 16.00 g (69%), mp 79.2-79.9 °C (lit.11, 82°C). Since the spectral data are completely different from those reported in literature,¹² full data are reported here.

¹H NMR $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.52 (s, 3H, ArCH₃), 3.88 (3 H, s, OCH₃), 4.06 (3 H, s, OCH₃), 4.83 (2 H, s, CH₂Br), 7.46-7.54 (2 H, m, Ar*H*), 8.05-8.08 (2 H, m, Ar*H*). ¹³C NMR $\delta_{\rm C}$ (100 MHz, CDCl₃) 11.76, 26.88, 61.59, 62.63, 122.49, 122.98, 125.93, 126.23, 126.84, 127.03, 127.20, 129.34, 150.65, 151.58. IR (ATR) $\nu_{\rm max}/{\rm cm}^{-1}$ 1351, 1054, 964, 771, 718.

Synthesis of 3-(cyanomethyl)-1,4-dimethoxy-2-methylnaphthalene (8)

Experimental

solution of 3-(bromomethyl)-1,4-dimethoxy-2-To а methylnaphthalene (7) (13.12 g, 44.4 mmol) in a mixture of acetonitrile/water (233 ml/80 ml) was added KCN (14.47 g, 222 mmol). After refluxing for 6 h the volatiles were removed in vacuo and 180 ml of water was added to the residu. The aqueous phase was extracted with of CH_2Cl_2 (3 × 120 ml). The combined organic layers were dried (MgSO₄) and evaporated. In most cases, no further purification was required. $R_f = 0.32$ (EtOAc/cHex : 3/17). The compound was obtained as brown crystals. Yield 9.86 g (92%), mp 107.3-107.5°C (lit.²²: 105°C). Since no NMR spectra have been reported full data are given here. ¹H NMR $\delta_{\rm H}$ (400 MHz, CDCl₃) 2.51 (3 H, s, ArCH₃), 3.89 (3 H, s, OCH₃), 3.91 (2 H, s, ArCH₂CN), 3.99 (3 H, s, OCH₃), 7.49-7.56 (2 H, m, ArH), 8.04-8.10 (2 H, m, ArH). ¹³C NMR δ_C (63 MHz, CDCl₃) 12.4 (CH₃), 15.3 (CH₂CN), 61.5 and 62.6 (OCH₃), 117.8 and 119.7 (Cq), 122.3 and 122.4 (C-5 and C-8), 125.4 (C_a), 126.0 and 126.7 (C-6 and C-7), 126.9 and 128.7 (C_a), 150.6 (2 × COMe). IR (KBr) v_{max}/cm^{-1} 2932, 2248 (CN), 1676, 1587, 1357, 1270.

Synthesis of 2-(bromomethyl)-3-(cyanomethyl)-1,4dimethoxynaphthalene (9)

ARTICLE

In a dry 500 ml flask equipped with a reflux condenser, 1,4dimethoxy-2-methyl-3-(cyanomethyl)naphthalene (8) (5 g, 20.72 mmol) was dissolved in CCl₄ (100 ml). To this solution there was added *N*-bromosuccinimide (4.06 g, 22.79 mmol, 1.1 Eq.) and benzoyl peroxide (0.502 g, 2.072 mmol, 0.1 Eq.). The mixture was heated at reflux temperature (85°C) for 3 hours. After this time, the reaction mixture was cooled and filtered through a celite pad. The residue was washed with CH₂Cl₂ (100 ml). Then, after drying over MgSO₄ the volatiles were removed under reduced pressure and the residue was purified by flash chromatography over silica gel applying a heptane-ethyl acetate gradient (from 100% heptane to 100% ethyl acetate in 25 minutes, 40 ml/min). R_f = 0.29 (EtOAc/cHex : 3/17). Yield 5.31 g (80 %), mp 121.0-122.0°C.

¹H NMR $\delta_{\rm H}$ (400 MHz, CDCl₃) 4.02 (3 H, s, OCH₃), 4.06 (3 H, s, OCH₃), 4.12 (2 H, s, ArCH₂CN), 4.91 (2 H, s, ArCH₂Br), 7.58-7.62 (2 H, m, ArH), 8.06-8.14 (2 H, m, ArH). ¹³C NMR $\delta_{\rm C}$ (63 MHz, CDCl₃) 14.1, 25.2, 62.70, 62.74, 117.8, 118.9, 122.8, 123.2, 125.1, 127.3, 127.6, 128.6, 128.8, 151.6, 152.1. IR (KBr) $\nu_{\rm max}/{\rm cm}^{-1}$ 2934, 2247 (CN), 1587, 1357, 1270. HRMS (ESI): *m*/*z* calcd for C₁₅H₁₄BrNO₂ + H⁺: 320.0281; found 320.0293.

Synthesis of 5,10-dimethoxy-1,2-dihydrobenzo[g]isoquinoline-3(4H)-iminium bromides 11

General procedure. To a solution of 2-(bromomethyl)-3-(cyanomethyl)-1,4-dimethoxy-naphthalene (9) (200 mg, 0.63 mmol) in isopropanol (10 ml) was added the aniline (0.95 eq.) and the mixture was heated for 16 h at reflux temperature (82° C). The solvent was removed *in vacuo* and the residue was washed with dry diethyl ether (3×10 ml), which yielded the pure amidine **11**. In some cases (**11b**, **11d**, **11f**, **11g**, **11i**, **11j**) an extra purification step was required using active carbon. To a solution of 100 mg of compound **11** in a 1/1 MeCN/H₂O mixture (10 ml) was added 100 mg of active carbon and the mixture was stirred for 10 min. Then the suspension was filtered over a celite pad and the filter cake washed with another 10 ml of 1/1 MeCN/H₂O mixture. The solvents were removed *in vacuo* to yield the pure product.

N²-Phenyl-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2H)-iminium bromide

(11a). Yield 191 mg (74%). ¹H NMR $\delta_{\rm H}$ (250 MHz, DMSO) 3.83 (3 H, s, OCH₃), 3.95 (3 H, s, OCH₃), 4.39 (2 H, s, ArCH₂), 5.06 (2 H, s, NCH₂), 7.55 -7.70 (7 H, m, arom. H), 8.05-8.15 (2 H, m, arom. H), 8.41 (1 H, br s, N(H)H⁺), 9.70 (1 H, br s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (63 MHz, DMSO) 28.1, 50.8, 62.2, 62.5, 118.5, 120.7, 122.1, 122.3, 126.2, 126.9, 127.0, 127.1, 127.7, 130.0, 130.9, 139.2, 147.2, 148.2, 164.4. IR (ATR) $\nu_{\rm max}/{\rm cm}^{-1}$ 3300, 2991, 2845, 1655, 1357, 1066. HRMS (ESI): m/z calcd for C₂₁H₂₁N₂O₂⁺: 333.1598; found 333.1580.

N²-(4-Methylphenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2H)-iminium bromide (11b). Yield 190 mg (71%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.41 (3 H, s, 4-CH₃Ph), 3.83 (3 H, s, OCH₃), 3.95 (3 H, s, OCH₃), 4.34 (2 H, s, ArCH₂), 5.05 (2 H, s, NCH₂), 7.42 (2 H, d, J = 8.5 Hz, 4-MePh), 7.46 (2 H, d, J = 8.5 Hz, 4-MePh), 7.64-7.68 (2 H, m, arom. H), 8.08-8.13 (2 H, m, arom. H), 8.35 (1 H, s, N(H)H⁺), 9.62 (1 H, s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 20.8, 28.1, 50.9, 62.2, 62.5, 118.4, 120.7, 122.0, 122.2, 125.9, 126.8, 127.0, 127.1, 127.7, 131.3, 136.7, 139.6, 147.2, 148.2, 164.4. HRMS (ESI): m/z calcd for C₂₂H₂₃N₂O₂⁺: 347.1754; found 347.1774.

N²-(4-Chlorophenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11c). Yield 123 mg (44%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.83 (3 H, s, OCH₃), 3.94 (3 H, s, OCH₃), 4.34 (2 H, s, ArCH₂), 5.03 (2 H, s, NCH₂), 7.63 (2 H, br d, J = 8.0 Hz, 4-ClPh), 7.64-7.67 (2 H, m, arom. H), 7.73 (2 H, br d, J = 8.0 Hz, 4-ClPh), 8.06-8.14 (2 H, m, arom. H), 8.52 (1 H, br s, N(H)H⁺), 9.77 (1 H, br s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 28.2, 50.7, 62.0, 62.3, 118.1, 120.4, 121.9, 122.0, 126.6, 126.7, 127.0, 127.6, 128.3, 130.7, 134.4, 137.9, 147.1, 148.1, 164.7. HRMS (ESI): m/z calcd for C₂₁H₂₀N₂O₂Cl⁺: 367.1208; found 367.1230.

N²-(4-Bromophenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11d). Yield 145 mg (47%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.84 (3 H, s, OCH₃), 3.95 (3 H, s, OCH₃), 4.38 (2 H, s, ArC*H*₂), 5.05 (2 H, s, NCH₂), 7.56 (2 H, br d, *J* = 8.7 Hz, 4-BrPh), 7.62-7.68 (2 H, m, arom. H), 7.87 (2 H, br d, *J* = 8.7 Hz, 4-BrPh), 8.08-8.13 (2 H, m, arom. H), 8.53 (1 H, s, N(H)*H*⁺), 9.75 (1 H, s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 28.1, 50.7, 62.1, 62.5, 118.3, 120.5, 122.0, 122.2, 123.2, 126.8, 127.0, 127.1, 127.7, 128.7, 133.8, 138.5, 147.2, 148.2, 164.6. HRMS (ESI): *m/z* calcd for C₂₁H₂₀BrN₂O₂⁺: 405.0703; found 411.0708.

N²-(4-Nitrophenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11e). Yield 129 mg (45%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.85 (3 H, s, OCH₃), 3.96 (3 H, s, OCH₃), 4.42 (2 H, s, ArCH₂), 5.12 (2 H, s, NCH₂), 7.64-7.67 (2 H, m, arom. H), 7.92 (2 H, d, J = 9.0 Hz, 4-NO₂Ph), 8.08-8.14 (2 H, m, arom. H), 8.50 (2 H,

d, J = 9.0 Hz, 4-NO₂Ph), 8.71 (1 H, s, N(*H*)H⁺), 9.90 (1 H, s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 28.3, 50.5, 62.0, 62.3, 118.0, 120.2, 121.9, 122.0, 125.9, 126.6, 126.7, 127.0, 127.6, 128.4, 144.4, 147.1, 147.8, 148.1, 164.9. HRMS (ESI): m/z calcd for C₂₁H₂₀N₃O₄⁺: 378.1448; found 378.1447.

N²-(4-(Ethoxycarbonyl)phenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2H)-iminium bromide (11f). Yield 103 mg (34%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO)

1.35 (3 H, t, J = 7.0 Hz, OCH₂CH₃), 3.84 (3 H, s, OCH₃), 3.96 (s, 3 H, OCH₃), 4.38 (2 H, q, OCH₂CH₃ overlaps with 4.43 (2 H, s, ArCH₂), 5.10 (2 H, s, NCH₂), 7.64-7.68 (2 H, m, arom. H), 7.75 (2 H, br d, J = 8.7 Hz, 4-CO₂EtPh), 8.08-8.12 (2 H, m, arom. H), 8.20 (2 H, br d, J = 8.7 Hz, 4-EtO₂CPh), 8.57 (1 H, s, N(H)H⁺), 9.84 (1 H, s, N(H)H⁺). ¹³C NMR δ_{C} (100 MHz, DMSO) 14.1, 28.3, 50.5, 61.2, 62.2, 62.5, 118.3, 120.5, 122.1, 122.2, 126.8, 126.95, 127.00, 127.1, 127.7, 131.1, 131.6, 143.0, 147.3, 148.2, 164.7, 164.9. HRMS (ESI): m/z calcd for C₂₄H₂₅N₂O₄⁺: 405.1809; found 405.1819.

N²-(3-(Ethoxycarbonyl)phenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11g). Yield 180 mg (59%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 1.34 (3 H, t, J = 7.0 Hz, OCH₂CH₃), 3.84 (3 H, s, OCH₃), 3.96 (3 H, s, OCH₃), 4.37 (2 H, s, ArCH₂), 4.37 (2 H, q, J = 7.0 Hz, OCH₂CH₃), 5.08 (2 H, s, NCH₂), 7.64.-7.67 (2 H, m, arom. H), 7.81-7.86 (2 H, m, arom. H), 8.08-8.16 (4 H, m, 3-EtO₂CCPh), 8.48 (1 H, s, N(H)H⁺), 9.71 (1 H, s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 14.1, 28.2, 50.7, 61.2, 62.2, 62.5, 118.4, 120.6, 122.0, 122.2, 126.8, 127.0, 127.1, 127.4, 127.7, 130.5, 131.3, 131.5, 132.3, 139.5, 147.2, 148.2, 164.77, 164.83. HRMS (ESI): *m/z* calcd for C₂₄H₂₅N₂O₄⁺: 405.1809; found 405.1815.

N²-(2-Chlorophenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11h). Yield 137 mg (49%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.86 (3 H, s, OCH₃), 3.97 (3 H, s, OCH₃), 4.36 (1 H, d, *J* = 18.4 Hz, ArC(H)*H*), 4.50 (1 H, d, *J* = 18.4 Hz, ArC(*H*)H), 4.96 (1 H, d, *J* = 15.6 Hz, NC(H)*H*), 5.08 (1 H, d, *J* = 15.6 Hz, NC(*H*)H), 7.63-7.70 (4 H, m, 2-CIPh), 7.76-7.80 (2 H, m, arom. H), 8.09-8.14 (2 H, m, arom. H), 8.75 (1 H, s, N(H)*H*⁺), 9.92 (1 H, s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 28.2, 50.0, 62.4, 62.5, 118.2, 120.8, 122.1, 122.3, 126.8, 127.0, 127.1, 127.8, 129.2, 129.8, 130.1, 131.1, 132.1, 135.9, 147.3, 148.2, 165.0. HRMS (ESI): m/z calcd for C₂₁H₂₀N₂O₂Cl⁺: 367.1208; found 367.1204.

N²-(2-Bromophenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11i). Yield 145 mg (39%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.86 (3 H, s, OCH₃), 3.95 (3 H, s, OCH₃), 4.37 (1 H, d, *J* = 18.5 Hz, ArC(H)*H*), 4.49 (1 H, d, *J* = 18.5 Hz, ArC(H)*H*), 4.92 (1 H, d, *J* = 16 Hz, NC(H)*H*), 5.10 (1 H, d, *J* = 16 Hz, NC(*H*)H), 7.56 (1 H, td, *J* = 7.8, 1.6 Hz, 2-BrPh), 7.63-7.71 (3 H, m, 2-BrPh and arom. H), 7.76 (1 H, dd, *J* = 8.0, 1.6 Hz, 2-BrPh), 8.10-8.14 (2 H, m, arom. H), 8.71 (1 H, s, N(H)*H*⁺), 9.90 (1 H, s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 28.2, 50.0, 62.4, 62.5, 118.1, 120.4, 120.8, 122.1, 122.3, 126.9, 127.0, 127.2, 127.8, 129.3, 130.4, 132.3, 134.3, 137.5, 147.3, 148.2, 164.8. HRMS (ESI): *m/z* calcd for C₂₁H₂₀N₂O₂Br⁺: 411.0703; found 411.0726.

N²-(2-Bromo-4-methylphenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11j). Yield 177 mg (56%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.42 (3 H, s, 4-*CH*₃Ar), 3.86 (3 H, s, OCH₃), 3.95 (3 H, s, OCH₃), 4.34 (1 H, d, *J* = 18.0 Hz, ArC(H)*H*), 4.47 (1H, d, *J* = 18.0 Hz, ArC(*H*)H), 4.88 (1 H, d, *J* = 15.0 Hz, NC(H)*H*), 5.07 (1 H, d, *J* = 15.0 Hz, NC(*H*)H), 7.50 (1 H, d, *J* = 8.0 Hz, 2-Br-4-MePh), 7.62 (1 H, br d, *J* = 8.0 Hz, 2-Br-4-MePh), 7.61-7.68 (2 H, m, arom. H), 7.78 (1 H, br s, 2-Br-4-MePh), 8.10-8.14 (2 H, m, arom. H), 8.67 (1 H, s, N(H)*H*⁺), 9.85 (1 H, s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (101 MHz, DMSO) 20.4, 28.1, 50.1, 62.4, 62.5, 118.1, 119.9, 120.8, 122.1, 122.3, 126.9, 127.0, 127.2, 127.8, 128.7, 130.9, 134.4, 135.0, 142.5, 147.3, 148.2, 164.8. HRMS (ESI): m/z calcd for C₂₂H₂₂N₂O₂Br⁺: 425.0859; found 425.0883.

N²-(4-Methoxyphenyl)-1,4-dihydro-5,10-

dimethoxybenzo[g]isoquinoline-3(2*H*)-iminium bromide (11k). Yield 136 mg (49%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 3.85 (2 × 3 H, 2 × s, 2 × OCH₃), 3.95 (3 H, s, OCH₃), 4.34 (2 H, s, ArCH₂), 5.01 (2 H, s, NCH₂), 7.18 (2 H, d, *J* = 8.9 Hz, 4-MeOPh), 7.46 (2 H, d, *J* = 8.9 Hz, 4-MeOPh), 7.60-7.68 (2 H, m, arom. H), 8.07-8.14 (2 H, m, arom. H), 8.28 (1 H, br s, N(H)*H*⁺), 9.55 (1 H, br s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (101 MHz, DMSO) 28.0, 51.0, 55.5, 62.0, 62.2, 115.8, 118.2, 120.6, 121.9, 122.0, 126.5, 126.7, 126.9, 127.3, 127.6, 131.6, 147.0, 148.1, 159.8, 164.6. HRMS (ESI): m/z calcd for C₂₂H₂₃N₂O₃⁺: 363.1703; found 363.1710.

Synthesis of 5,10-dioxobenzo[g]isoquinoline-3(2H)-iminium bromides 12

General procedure. To a 10 ml microwave vial there was added 50 mg of compound 11. Then, acetic acid (2 ml) was added along with of a 33 % w/w solution of hydrobromic acid in acetic acid (1 ml). Finally water (0.15 ml) was added and the tube was sealed. The mixture was heated at 115 °C for 16 h. After this time the volatiles were removed in vacuo and the residue was sonicated with dry diethyl ether (10 ml). The ether was decanted, which yielded the pure crystalline product. In some cases (12b, 12d, 12e, 12h, 12i, 12j) an extra purification step was performed by using column chromatography on C18 silica. Typically 50 mg of crude product was separated on a 16 ml column and eluted with MeCN/H₂O (6/4). In some of the 1 H NMR spectra of compounds 12 a triplet at 7.09 ppm (J = 51.1Hz) can be observed (see ESI). This signal is characteristic for the presence of the ammonium cation.²³ The origin of the ammonium salt could be ascribed to some degradation during the HBr treatment of compounds 11. The presence of water, which was required to make the oxidative demethylation work, could give rise to the hydrolysis of the $C=NH_2^+$ moiety in 11 or 12, thus giving rise to NH_4Br formation.

N²-Phenyl-5,10-dioxobenzo[g]isoquinoline-3(2H)-iminium

bromide (12a). Yield 37 mg (74%). ¹H NMR $\delta_{\rm H}$ (250 MHz, DMSO) 7.66-7.75 (5 H, m, Ph), 7.78 (1 H, s, 4-H), 7.99-8.08 (2 H, m, arom. H), 8.24-8.31 (2 H, m, arom. H), 8.56 (1 H, br s, N(H)H⁺), 8.77 (1 H, s, 1-H), 10.04 (1 H, br s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (63 MHz, DMSO) 112.7, 117.7, 126.4, 127.1, 127.5,

ARTICLE

130.9, 131.5, 133.4, 133.6, 135.1, 135.8, 137.3, 141.2, 143.3, 156.8, 178.6, 179.8. IR (ATR) υ_{max} /cm⁻¹ 3656, 2981, 2888, 1632, 1385, 1282, 954. HRMS (ESI): m/z calcd for $C_{19}H_{13}N_2O_2^+$: 301.0972; found 301.0961.

*N*²-(4-Methylphenyl)-5,10-dioxobenzo[*g*]isoquinoline-3(2*H*)iminium bromide (12b). Yield 29.6 mg (64%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.47 (3 H, s, PhCH₃), 7.56 (2 H, d, *J* = 8.0 Hz, 4-MePh), 7.60 (2 H, d, *J* = 8.0 Hz, 4-MePh), 7.78 (1 H, s, 4-H), 8.00-8.07 (2 H, m, arom. H), 8.26-8.30 (2 H, m, arom. H), 8.51 (1 H, br s, N(H)*H*⁺), 8.71 (1 H, s, 1-H), 9.97 (1 H, br s, N(*H*)*H*⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 20.9, 112.6, 117.7, 126.1, 127.0, 127.4, 131.3, 133.4, 133.6, 134.8, 135.1, 135.8, 141.2, 141.3, 143.3, 156.9, 178.6, 179.7. HRMS (ESI): m/z calcd for C₂₀H₁₅N₂O₂⁺: 315.1128; found 315.1134.

*N*²-(4-Chlorophenyl)-5,10-dioxobenzo[*g*]isoquinoline-3(*2H*)iminium bromide (12c). Yield 11.6 mg (25%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.78 (2 H, d, *J* = 8.9 Hz, 4-ClPh), 7.81 (1 H, s, 4-H), 7.83 (2 H, d, *J* = 8.9 Hz, 4-ClPh), 8.00-8.07 (2 H, m, arom. H), 8.26-8.31 (2 H, m, arom. H), 8.65 (1 H, br s, N(H)*H*⁺), 8.83 (1 H, s, 1-H), 10.05 (1 H, br s, N(*H*)*H*⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.8, 117.5, 127.0, 127.4, 128.7, 130.9, 133.2, 133.5, 135.1, 135.8, 136.0, 136.1, 140.9, 143.2, 156.9, 178.5, 179.6. HRMS (ESI): m/z calcd for C₁₉H₁₂N₂O₂Cl⁺: 335.0582; found 335.0588.

*N*²-(4-Bromophenyl)-5,10-dioxobenzo[*g*]isoquinoline-3(*2H*)iminium bromide (12d). Yield 42.1 mg (90%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.70 (2 H, d, *J* = 8.8 Hz, 4-BrPh), 7.76 (1 H, s, 4-H), 7.97 (2 H, d, *J* = 8.8 Hz, 4-BrPh), 8.00-8.07 (2 H, m, arom. H), 8.26-8.30 (2 H, m, arom. H), 8.64 (1 H, br s, N(*H*)H⁺), 8.84 (1 H, s, 1-H), 9.99 (1 H, br s, N(H)*H*⁺). 13C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.7, 117.6, 125.0, 127.0, 127.5, 128.9, 133.3, 133.6, 133.9, 135.1, 135.8, 136.4, 141.2, 143.3, 156.8, 178.6, 179.7. HRMS (ESI): m/z calcd for C₁₉H₁₂N₂O₂Br⁺: 379.0077; found 379.0084.

N^2 -(4-Nitrophenyl)-5,10-dioxobenzo[g]isoquinoline-3(2H)-

iminium bromide (12e). Yield 46.5 mg (100%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.80 (1 H, s, 4-H), 8.03-8.07 (2 H, m, arom. H), 8.07 (2 H, d, J = 9.0 Hz, 4-NO₂Ph), 8.27-8.31 (2 H, m, arom. H), 8.60 (2 H, d, J = 9.0 Hz, 4-NO₂Ph), 8.74 (1 H, br s, N(H) H^+), 8.94 (1 H, s, 1-H), 10.09 (1 H, br s, N(H) H^+). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 113.1, 117.6, 126.1, 127.1, 127.5, 129.1, 133.3, 133.6, 135.2, 135.8, 141.2, 142.1, 143.0, 149.1, 156.8, 178.6, 179.7. HRMS (ESI): m/z calcd for C₁₉H₁₂N₃O₄⁺: 346.0822; found 346.0828.

```
N^2-(Phenyl-4-carboxylic
```

```
acid)-5,10-
```

dioxobenzo[g]isoquinoline-3(2H)-iminium bromide (12f). Yield 38.6 mg (83%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.78 (1 H, s, 4-H), 7.86 (2 H, d, J = 8.7 Hz, 4-CO₂HPh), 8.00-8.07 (2 H, m, arom. H), 8.28 (2 H, d, J = 8.7 Hz, 4-CO₂HPh), 8.25-8.31 (2 H, m, arom. H), 8.64 (1 H, br s, N(H) H^{+}), 8.86 (1 H, s, 1-H), 10.00 (1 H, br s, N(H) H^{+}). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.8, 117.8, 127.1, 127.1, 127.5, 131.8, 133.3, 133.5, 133.6, 135.1, 135.8, 140.5, 141.2, 143.0, 156.7, 166.3, 178.6, 179.7. HRMS (ESI): m/z calcd for C₂₀H₁₃N₂O₄⁺: 345.0870; found: 345.0888.

N^2 -(Phenyl-3-carboxylic acid)-5,10dioxobenzo[g]isoquinoline-3(2H)-iminium bromide (12g).

Journal Name

Page 10 of 12

Yield 21.4 mg (46%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.78 (1 H, s, 4-H), 7.87 (1 H, t, J = 7.8 Hz, 3-CO₂HPh), 7.95-7.97 (1 H, m, 3-CO₂HPh), 8.01-8.08 (2 H, m, arom. H), 8.25-8.31 (4 H, m, 3-CO₂HPh and arom. H), 8.61 (1 H, br s, N(H)H⁺), 8.88 (1 H s, 1-H), 9.97 (1 H, br s, N(H)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.9, 117.6, 127.1, 127.5, 127.8, 130.0, 130.9, 131.3, 132.0, 133.3, 133.6, 135.1, 135.8, 137.4, 141.1, 143.5, 156.9, 166.1, 178.6, 179.8. HRMS (ESI): m/z calcd for C₂₀H₁₃N₂O₄⁺: 345.0870; found 345.0880.

*N*²-(2-Chlorophenyl)-5,10-dioxobenzo[*g*]isoquinoline-3(2*H*)iminium bromide (12h). Yield 11.6 mg (25%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.73 (1 H, ddd, *J* = 7.8, 7.8, 1.4 Hz, 2-ClPh), 7.79 (1 H, ddd, *J* = 7.8, 7.8, 1.7 Hz, 2-ClPh), 7.85 (1 H, s, 4-H), 7.88 (1 H, dd, *J* = 7.8, 1.7 Hz, 2-ClPh), 7.89 (1 H, dd, *J* = 7.8, 1.4 Hz, 2-ClPh), 7.99-8.06 (2 H, m, arom. H), 8.26-8.30 (2 H, m, arom. H), 8.87 (1 H, br s, N(H)*H*⁺), 8.96 (1 H, s, 1-H), 10.18 (1 H, br s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.5, 118.4, 126.8, 127.2, 129.1, 129.5, 129.8, 131.2, 133.2, 133.5 (2 carbons), 134.0, 134.9, 135.5, 141.5, 142.7, 156.2, 178.3, 179.5. HRMS (ESI): m/z calcd for C₁₉H₁₂N₂O₂Cl⁺: 335.0582; found 335.0569.

*N*²-(2-Bromophenyl)-5,10-dioxobenzo[*g*]isoquinoline-3(2*H*)iminium bromide (12i). Yield 36.5 mg (78%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 7.70 (1 H, td, *J* = 8.0 Hz, 1.5 Hz, 2-BrPh), 7.76 (1 H, td, *J* = 8.0 Hz, *J* = 1.5 Hz, 2-BrPh), 7.83 (1 H, s, 4-H), 7.87 (1 H, dd, *J* = 8 Hz, *J* = 1.5 Hz, 2-BrPh), 7.99-8.06 (2 H, m, arom. H), 8.25-8.29 (2 H, m, arom. H), 8.81 (1 H, br s, N(H)*H*⁺), 8.96 (1 H, s, 1-H), 10.11 (1 H, br s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 112.5, 118.4, 119.8, 127.0, 127.4, 129.2, 130.4, 133.5, 133.6, 133.7, 134.5, 135.1, 135.6, 135.8, 142.0, 143.1, 156.2, 178.6, 179.5. HRMS (ESI): m/z calcd for C₁₉H₁₂N₂O₂Br⁺: 379.0077; found 379.0074.

N²-(2-Bromo-4-methylphenyl)-5,10-

dioxobenzo[g]isoquinoline-3(2*H*)-iminium bromide (12j). Yield 21.5 mg (43%). ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.47 (3 H, s, Me), 7.56 (1 H, d×m, J = 8.1 Hz, 2-Br-4-MePh), 7.73 (1H, d, J = 8.1 Hz, 2-Br-4-MePh), 7.80 (1 H, d, J = 0.4 Hz, 4-H), 7.89-7.90 (1 H, m, 2-Br-4-MePh), 7.99-8.06 (2 H, m, arom. H), 8.25-8.29 (2 H, m, arom. H), 8.76 (1 H, br s, N(H)*H*⁺), 8.89 (1 H, d, J = 0.4 Hz, 1-H), 10.08 (1 H, br s, N(*H*)*H*⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 20.5, 112.4, 118.4, 119.4, 127.0, 127.4, 128.7, 130.9, 133.3, 133.5, 133.7, 134.6, 135.1, 135.6, 142.0, 143.2, 144.0, 156.3, 178.6, 179.5. HRMS (ESI): m/z calcd for C₂₀H₁₄N₂O₂Br⁺: 393.0233; found 393.0232.

Synthesis of 5,10-diacetoxybenzo[g]isoquinoline-3(2H)-iminium bromides 13

General Procedure. To a dry MW vial there was added 44 mg of compound **11**. This was dried in vacuum for 2 h at 60°C and sealed under air. In another MW vial, HOAc (2 ml), Ac₂O (0.5 ml) and a 33% HBr solution in acetic acid (1 ml) was heated at 125°C for 2 hours, and then cooled. The content of the second vial was transferred via a syringe to the first vial, which was sealed. The mixture was heated overnight (16 h) at 115°C. After this time the volatiles were removed *in vacuo*. The residue was sonicated with dry diethyl ether, which afforded the crystalline compound.

N²-(2-Bromophenyl)-5,10-diacetoxy-1,4-

dihydrobenzo[g]isoquinoline-3(2*H*)-iminium bromide (13i). Yield not determined. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.59 (3 H, s, MeCO), the signal for the other MeCO overlaps with the solvent peak, 4.23 (1 H, d, J = 19.0 Hz), 4.31 (1 H, d, J = 19.0Hz), 4.86 (1 H, d, J = 15.9 Hz), 4.99 (1 H, d, J = 15.9 Hz), 7.54-7.59 (1 H, m), 7.67-7.74 (4 H, m + d overlap), 7.92 (1 H, d, J = 7.9 Hz, 2BrPh), 7.98-8.05 (2 H, m, arom. H), 8.71 (1 H, br s, N(H)*H*⁺), 9.81 (1 H, br s, N(*H*)H⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 20.7, 21.1, 28.5, 49.9, 118.8, 120.6, 121.5, 121.8, 122.0, 126.3, 126.8, 128.0, 128.1, 129.3, 130.5, 132.4, 134.4, 137.2, 139.4, 140.5, 164.2, 169.2, 169.3. No HRMS could be recorded due to the lability of this compound.

N²-(2-Bromo-4-methylphenyl)-5,10-diacetoxy-1,4-

dihydrobenzo[g]isoquinoline-3(2*H*)-iminium bromide (13j). Yield not determined. ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO) 2.42 (3 H, s), 2.49 (3 H, s), 2.60 (3 H, s), 4.23 (1 H, d, J = 18.4 Hz), 4.32 (1 H, d, J = 18.4 Hz), 4.82 (1 H, d, J = 16.1 Hz), 4.97 (1 H, d, J = 16.1 Hz), 7.48 (1 H, d, J = 8.0 Hz, 2-Br-4-MePh), 7.57 (1 H, d, J = 8.0 Hz, 2-Br-4-MePh), 7.68-7.74 (2 H, m, arom. H), 7.77 (1 H, s, 2-Br-4-MePh), 7.98-8.04 (2 H, m, arom. H), 8.69 (1 H, br s, N(H)*H*⁺), 9.82 (1 H, br s, N(*H*)*H*⁺). ¹³C NMR $\delta_{\rm C}$ (100 MHz, DMSO) 20.46, 20.54, 20.57, 28.4, 49.9, 118.8, 120.1, 121.5, 121.8, 122.0, 126.2, 126.8, 127.9, 128.0, 128.7, 130.9, 134.4, 134.6, 139.3, 140.4, 142.6, 164.2, 169.1, 169.2. No HRMS could be recorded due to the lability of this compound.

Synthesis of 2-(1,4-dimethoxy-3-

((phenylamino)methyl)naphthalen-2-yl)acetonitrile (14)

To a solution of 2-(bromomethyl)-3-(cyanomethyl)-1,4dimethoxynaphthalene (9) (100 mg, 0.31 mmol) in dichloromethane (10 ml) was added aniline (73 mg, 0.78 mmol) at room temperature. After stirring for 4 h, the reaction mixture was poured in water (15 ml) and extracted with dichloromethane (3×15 ml). The combined extracts were dried and evaporated in vacuo. Chromatography over silica gel furnished yellow crystals. $R_f = 0.41$ (EtOAc/cHex : 1/3). Yield 97 mg (94%), mp 164.0-165.0 °C. ¹H NMR $\delta_{\rm H}$ (250 MHz, CDCl₃) 3.64 (1 H, br s, NH), 3.95 (3 H, s, OMe), 4.04 (3 H, s, OMe), 4.06 (2 H, s, CH₂CN), 4.56 (2 H, s, CH₂N), 6.77-6.84 (3 H, m, Ph), 7.22-7.29 (2 H, m, Ph), 7.56-7.62 (2 H, m, arom. H), 8.06-8.17 (2 H, m, arom. H). ¹³C NMR $\delta_{\rm C}$ (63 MHz, CDCl₃) 14.6, 40.3, 62.7, 63.6, 113.3, 118.5, 118.6, 120.1, 122.7, 123.0, 125.8, 127.1, 127.2, 128.3, 128.9, 129.4, 147.8, 151.5, 151.9. IR (KBr) v_{max}/cm⁻¹ 3361 (NH), 2937, 2841, 2249 (CN), 1602, 1507, 1360, 1055, 755. HRMS (ESI): m/z calcd for C₂₁H₂₀N₂O₂ + H⁺: 333.1603; found 333.1621.

Acknowledgements

We gratefully acknowledge the Research Foundation–Flanders (FWO-Vlaanderen) for a mandate for DC under grant 12N5915N, the Bilateral Scientific Research Cooperation Projects between FWO (Flanders, G005514N)) and NAFOSTED (Vietnam, FWO.104.2013.12) and the IWT for financial support of this research. We are thankful to Ing. Heidi

Seykens and Ing. Norbert Hancke for recording NMR spectra and HRMS. We thank Dr. Thomas Ronson for the careful editing of the text.

References

- (a) D. Cappoen, J. Jacobs, T. N. Van, S. Claessens, G. Diels, R. Anthonissen, T. Einarsdottir, M. Fauville, L. Verschaeve, K. Huygen and N. De Kimpe, *Eur. J. Med. Chem.*, 2012, **48**, 57.
 (b) P. Claes, D. Cappoen, B. M. Mbala, J. Jacobs, B. Mertens, V. Mathys, L. Verschaeve, K. Huygen and N. De Kimpe, *Eur. J. Med. Chem.*, 2013, **67**, 98. (c) D. Cappoen, P. Claes, J. Jacobs, R. Anthonissen, V. Mathys, L. Verschaeve, K. Huygen and N. De Kimpe, *J. Med. Chem.*, 2014, **57**, 2895. (d) P. Claes, D. Cappoen, C. Uythethofken, J. Jacobs, B. Mertens, V. Mathys, L. Verschaeve, K. Huygen and N. De Kimpe, *Eur. J. Med. Chem.*, 2014, **77**, 409.
- 2 M. P. Gore, S. J. Gould and D. D. Weller, *J. Org. Chem.*, 1991, 56, 2289.
- 3 D. Mal, J. Roy, K. Biradha, Org. Biomol. Chem., 2014, 12, 8196.
- 4 (a) S. Claessens, J. Jacobs, S. Van Aeken, K. Abbaspour Tehrani and N. De Kimpe, J. Org. Chem. 2008, 73, 7555. (b) J. Jacobs, J. Deblander, B. Kesteleyn, K. Abbaspour Tehrani and N. De Kimpe, Tetrahedron, 2008, 64, 5345. (c) J. Jacobs, S. Claessens, K. Huygen, K. Abbaspour Tehrani and N. De Kimpe, Pure Appl. Chem., 2011, 83, 1651.
- 5 J. V. Greenhill and P. Lue, Prog. Med. Chem., 1993, 30, 203.
- 6 T. R. M. Rauws and B. U. W. Maes, *Chem. Soc. Rev.* 2012, **41**, 2463 and references cited therein.
- 7 T. Brückl, F. Klepper, K. Gutsmiedl and T. Carell, Org. Biomol. Chem., 2007, 5, 3821.
- 8 R. Shapiro, R. DiCosimo, S. M. Hennessey, B. Stieglitz, O. Campopiano and G. C. Chiang, Org. Proc. Res. Dev., 2001, 5, 593.
- 9 (a) G. Kalaus, N. Malkieh, I. Katona, M. Kajtár-Peredy, T. Koritsánszky, A. Kálmán, L. Szabó and C. Szántay, J. Org. Chem., 1985, 50, 3760. (b) M. Lounasmaa, D. Din Belle and A. Tolvanen, Heterocycles, 1999, 51, 1125. (c) T. Murata, M. Shimada, S. Sakakibara, T. Yoshino, T. Masuda, T. Shintani, H. Sato, Y. Koriyama, K. Fukushima, N. Nunami, M. Yamauchi, K. Fuchikami, H. Komura, A. Watanabe, K. B. Ziegelbauer, K. B. Bacon and T. B. Lowinger, *Bioorg. Med. Chem. Lett.*, 2004, 14, 4019. (d) Jahangir, M. A. Brook, D. B. Maclean and H. L. Holland, *Tetrahedron*, 1987, 43, 5761.
- L. M. Potikha and V. A. Kovtunenko, *Rus. Chem. Rev.*, 2009, 78, 513 and references cited therein.
- 11 T. Ito, T. Ikemoto, T. Yamano, Y. Mizuno and K. Tomimatsu, *Tetrahedron: Asymmetry*, 2003, **14**, 3525.
- 12 L. Syper, J. Młochowski and K. Kloc, *Tetrahedron*, 1983, **39**, 781.
- 13 D. F. Fry, C. B. Fowler and R. K. Dieter, Synlett, 1994, 836.
- 14 J. Jacobs, S. Claessens, B. Kesteleyn, K. Huygen and N. De Kimpe, *Tetrahedron*, 2007, **63**, 2503.
- 15 C. D. Snyder and H. Rapoport, J. Am. Chem. Soc., 1972, 94, 227.
- 16 J. I. Ogonor, Tetrahedron, 1981, 37, 2909.
- 17 H. Ulrich, The Chemistry of Imidoyl Halides, Plenum Press, New York, 1968.
- (a) V. A. Snewin, M.-P. Gares, P. Ó Gaora, Z. Hasan, I. N. Brown and D. B. Young, *Infect. Immun.* 1999, **67**, 4586. (b) D. Forge, D. Cappoen, J. Laurent, D. Stanicki, A. Mayence, T. L. Huang, L. Verschaeve, K. Huygen and J. J. Vanden Eynde, *Eur. J. Med. Chem.*, 2012, **49**, 95. (c) D. Cappoen, D. Forge, F. Vercammen, V. Mathys, M. Kiass, V. Roupie, R. Anthonissen, L. Verschaeve, J. J. Vanden Eynde and K. Huygen, *Eur. J. Med. Chem.*, 2013, **63**, 731.

This journal is © The Royal Society of Chemistry 20xx

- 19 G. Repetto, A. del Peso and J. L. Zurita, *Nature Protocols*, 2008, **3**, 1125.
- 20 B. A. Hanna, A. Ebrahimzadeh, L. B. Elliott, M. A. Morgan, S. M. Novak, S. Rusch-Gerdes, M. Acio, D. F. Dunbar, T. M. Holmes, C. H. Rexer, C. Savthyakumar and A. M. Vannier, J. Clin. Microb., 1999, **37**, 748.
- 21 D. Schnappinger, S. Ehrt, M. I. Voskuil, Y. Liu, J. A. Mangan, I. M. Monahan, G. Dolganov, B. Efron, P. D. Butcher, C. Nathan and G. K. Schoolnik, *J. Exp. Med.*, 2003, **198**, 693.
- 22 W. E. Bondinell, S. J. DiMari, B. Frydman, K. Matsumoto and H. Rapoport, J. Org. Chem., 1968, **33**, 4351.
- 23 (a) S. A. Richards and J. C. Hollerton, Essential practical NMR for organic chemistry, Wiley, 2011. (b) A. Savini, P. Belanzoni, G. Bellachioma, C. Zuccaccia, D. Zuccaccia and A. Macchioni, *Green Chem.*, 2011, 13, 3360.