Organic & Biomolecular Chemistry

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/obc

Journal Name

ARTICLE



OPG

An organocatalytic strategy for the stereoselective synthesis of Cgalactosides with fluorine at the pseudoanomeric carbon Ahmad S. Altiti, ^a S. Bachan, ^a W. Alrowhani ^a and D. R. Mootoo ^a as enantiomerically pure catalyst gave intractable mixtures of products. These results are in line with the results for simple achiral aldehydes, and suggest that stereochemistry in the reactions of these chiral, highly substituted, carbohydratederived aldehydes are controlled primarily by the chirality in the catalyst. OPG "F+" organocatalvst сно OH HO C (CH₂)₂₄CH₃ HN OH ΗÒ

Received 00th January 20xx. Accepted 00th January 20xx

DOI: 10.1039/x0xx00000x

www.rsc.org/

The α -fluorination of α - and β - C-ethanals of galactose using Jørgensen catalysts and NFSI was investigated. The crude reaction products were transformed to their primary alcohol or methylenated derivatives, which are versatile precursors to biologically interesting fluorinated glycomimetics. The α -C-glycoside substrate gave moderate to high yields of fluorinated α -C-glycosides with minor amounts of β -C-glycoside analogues. The reactions on the β -C-glycoside were lower yielding but gave exclusively fluorinated β -C-glycosides. For both α - and β - C-glycoside substrates (R) and (S) catalyst showed complementary stereoselectivity. The preparation of difluorinated materials required the use of racemic catalyst

Introduction

Fluorination of bioactive materials is an active area of research in medicinal chemistry because the electronic and conformational effects of fluorine and the stability of the C-F bond may enhance binding in receptor sites and improve drug absorption, distribution, metabolism and excretion (ADME).^{1,2,3} Accordingly a variety of strategies for introduction of fluorine into biomolecules have been developed.4,5,6 However, while these methods have met much success in simple substrates, their application to more highly functionalized substrates like carbohydrates is often problematic.^{7,8,9} In this context, fluorinated C-glycosides like 2 and versatile derivatives thereof that can be transformed to complex analogues like 3 and 4, are 1).^{10,11,12,13,14,1516,17,18,19,2021,22,23,24} interest (Fig. of The organocatalytic α -fluorination of simple C-glycoside aldehydes 1 is an appealing route to such precursors because of the mild reaction conditions, operational simplicity and easy access to reaction substrates.^{25,26,27} Herein, we describe the application of this strategy to C-glycosides of galactose. While the proline catalyzed amination of related C-glycoside aldehydes has been studied, to the best of our knowledge the mechanistically similar α -fluorination reaction has not been reported.²⁸

Results and Discussion

The substrates for this study were the $\alpha\text{-}$ and $\beta\text{-}$ C-linked ethanals 5a and 5b, which were obtained from their readily available C-allyl derivatives.^{20,21,29} Initial experiments were performed on 5a (Scheme 1). This material was treated with two molar equivalents of NFSI in THF in the presence of 2.5 mol % of (S) or (R) Jørgensen catalyst.²⁵ Because of the instability of the fluorinated aldehyde products, the crude with NaBH₄ aldehyde was treated or methylenetriphenylphosphorane to give the primary alcohol or terminal alkene derivative.³⁰ NaBH₄ reduction of the product from the fluorination with the (S) catalyst afforded α -Cmonofluorinated glycoside **6**, and a β -C-glycoside **7** of undetermined configuration at the fluorinated carbon, in 75 and 10% yield respectively from 5a. The fluorination/reduction

^{a.} Department of Chemistry, Hunter College, 695 Park Avenue New York, NY 10065, USA. E-mail: dmootoo@hunter.cuny.edu; Fax.: +1 212 772 5332; Tel.: +1 212 772 4356;

^{*}Electronic Supplementary Information (ESI) available: Copies of 'H, 13 C, 2D NMR spectra and tabulated peak assignments for 0 and 00 – 00, X-ray data for 00,. See DOI: 10.1039/x0xx00000x

СНО X/Y = H/F; F/H; F/FHO OH HO HO HO х CH₂)₁₃CH₃ нс ŌН 3 4 Figure 1 Fluorinated precursors for complex C-glycosides

ARTICLE

Journal Name

sequence with the (R) catalyst produced the α -C-fluorohydrin **8** as the major product, together with **6**, the β -C-fluorohydrin **9** that was epimeric to **7** at the fluorinated carbon, and the α -Cdifluorinated product 10 in 68, 6, 3 and 20% yield respectively.



Scheme 1 Fluorination-reduction on α-C-ethanal 5a

Wittig methylenation on the crude fluorination product with the (S) catalyst gave the α -C-alkene **11** and the β -C-product **12** in 73 and 9% yields, respectively (Scheme 2). The fluorination/methylenation sequence with the (R) catalyst afforded 13 (53%), the α -C-glycoside that was epimeric to 11 at the fluorinated carbon, and an intractable mixture of other fluorinated alkenes (31%).



Towards more stereoselective syntheses of the β -Cfluorohydrins 7 and 9, the fluorination-reduction protocol on the β -C-ethanal **5b**, was next investigated (Scheme 3). For optimal results, these reactions were performed at higher catalyst loading and shorter reaction times, compared to the reactions on 5a. With 30 mol% catalyst 7 and 9 were obtained as the only observed mono-fluorinated products with the (S) and (R) catalysts, in 44 and 50 % yields, respectively.



Scheme 3. Fluorination-reduction on β -C-ethanal 5b

The synthesis of difluorinated derivatives was next examined more carefully (Scheme 4). Reactions on 5a using excess NFSI, extended reaction times and increased loading of the (R) or (S) catalyst, gave complex mixtures with low yields of fluorinated materials and other unidentified products. Experiments with pyrrolidine instead of the chiral catalyst gave similar results. However when the reaction was performed with a 20 mol% of a racemic mixture of the catalyst, and the crude product treated with NaBH₄, or subjected to methylenation, the difluorinated products 10 and 14 were obtained in 83 and 40% yield, respectively. The alkene 14 was also prepared by oxidation of a purified sample of 10 with Dess-Martin periodinane followed by Wittig olefination on the crude aldehyde. The overall yield of 14 from 10 following this procedure was 66%. As for the mono-fluorination reactions, the difluorination-reduction protocol on 5b was lower yielding compared to case of 5a, giving a 16% yield of the difluorinated β -C-glycoside **15** with 30 mol% of the racemic catalyst.



Scheme 4 Synthesis of difluorinated derivatives

Journal Name

Stereochemical assignment of the fluorinated compounds rests on X-ray analysis of alkene 11, $J_{\text{H},\text{H}}$ values and chemical correlation between alkene and alcohol derivatives (Fig. 2, Supporting Information, Scheme 2). The X-ray structure of 11 indicated the α -C1 orientation and the R configuration at the fluorinated carbon. Interestingly, the molecule adopted the ${}^{1}C_{4}$ chair conformation. A similar result has been reported for a related mono-fluorinated α -C-galactoside. A $J_{1,2}$ of 2.4 Hz also supported the $\alpha\text{-C1}$ orientation, but is ambiguous in relation to assignment of ring conformation. Similarly J_{1.2} values between 0 - 4.5 and close to 10 Hz were consistent with the α -C1 in 6, 8, 10, 13, 14, and a $\beta\text{-C1}$ in 7, 9, 12 and 15 respectively. The configuration at the fluorinated carbon in the α -C-glycosides **6**, 8, 11 and 13 rests on correlation to 11. Thus, alkene 13 was deduced as S on the basis of its epimeric relationship to 11. The α -C- fluorohydrins 6 and 8 were assigned by correlating them to 11 and 13 respectively, through an ozonolysis/aldehyde reduction sequence. The configurations at the fluorinated carbons in the $\beta\text{-C-glycosides}$ 7, 9 and 12 were tentatively assigned using the stereochemical model proposed for the Jørgensen catalyst (vide infra). That the (S) v.s. (R) catalyzed reactions selectively gave epimeric fluorides, 6 vs. 8, 7 vs. 9, 11 vs. 13 as the major products provides a measure of self-consistency to this argument.



The stereoselectivity in the fluorination of α -C-ethanal **5a** is in agreement with the model proposed for the Jørgensen catalysts (Scheme 5). Thus, the (*S*) and (*R*) catalyst lead to *E* enamines **16** and **20**, which should favour fluorination on the *Re* and *Si* face respectively, which is in line with the experimental result, stereoselective formation of **6** and **8**. Similarly high stereoselectivities have been observed in the proline catalysed amination on related carbohydrate

ARTICLE

aldehydes. The apparently better performance of the (*S*) catalyst compared to the (*R*), on the α -C-glycosoide substrate may represent a "matched/mistmatched" scenario. Thus **16** and **20** are expected to adopt the "exo-anomeric" conformation with respect to the pseudoglyconic torsion (i.e. the aminated carbon of the enamine is gauche to O5 and H1 of the sugar ring).³¹ The facial selectivity imposed by the (*R*) catalyst (i.e. on **20**), is opposed by steric hindrance from *O*5 of the sugar. For **16**, the enamine from the (S) catalyst, the facial selectivity of the catalyst is matched to that imposed by the sugar residue. Therefore, fluorination on **20** may be less stereoselective and side reactions more likely compared to **16**.

The results for the difluorination reactions also appear to be consistent with the Jørgensen model. Thus, for a given enantiomer of the catalyst, α -deprotonation on the monofluorination iminium ion (i.e. 17, 21), is sterically hindered by the bis-arylcarbinol side chain in the catalyst residue.²⁵ A racemic mixture of catalyst allows for amine exchange on 17 and **21** to give diastereomeric iminum ions in which α deprotonation is not sterically blocked, thereby facilitating formation of the di-fluorinated product. As in the initial fluorination step, conformational effects may also impact on difluorination and explain why the (S) catalyst apparently shows a lesser propensity for difluorination compared to the (R) catalyst. Thus, "exo-anomeric" conformations for 17 and 21, suggests that deprotonation in 17 (the favored monofluorinated iminium ion with the (S) catalyst), is hindered by both the sugar ring and the catalyst moiety, whereas the hindrance to deprotonation on 21, the corresponding species with the (R) catalyst, comes from the catalyst moiety only.

Without definitive assignment of the configuration at the fluorinated carbons in the β -C-glycosides, it is impossible to tell whether the Jørgensen model extends to the fluorination of **5b**. However, the finding that enantiomeric catalysts exhibit opposite stereoselectivity in the fluorination of **5b**, supports this notion. This observation is also relevant to the mechanism by which the fluorinated β -glycosides **7** and **12** are formed from the α -C-glycoside **5a**. A pathway in which **5a** or **16** first epimerizes to **5b** or the β -C-glycoside of **16** respectively (via an retro-Michael-Michael mechanism), followed by fluorination, seems more likely than epimerization of a first formed α -C-fluorinated species.³² In the latter scenario, the individual reactions of **5a** with the enantiomeric catalysts might have been expected to give mixtures of the β -C-glycosides **7** and **12**.

Conclusions

The α -fluorination of C-linked galactosyl ethanals using the Jørgensen catalyst and NFSI is a promising route to versatile precursors for complex fluorinated glycomimetics. Enantiomeric catalysts influenced complementary facial selectivity in line with the results for simple achiral aldehydes. The high diastereoselectivity observed for these stereochemically complex substrates is similar to that observed for the organocatalyzed amination on related

ARTICLE

substrates, and point to the powerful stereodirecting effects of these proline-based catalysts in enamine reactions. These findings provide insight on the effects of conformation and remote substituents in such complex substrates, and provide the basis for the wider application of this chemistry in carbohydrate synthesis.



Experimental section

Solvents were purified by standard procedures or used from commercial sources as appropriate. Petroleum ether refers to the fraction of petroleum ether boiling between 40 and 60 °C. Ether refers to diethyl ether. Unless otherwise stated thin layer chromatography (TLC) was done on 0.25 mm thick precoated silica gel 60 (HF-254, Whatman) aluminium sheets and flash column chromatography (FCC) was performed using Kieselgel 60 (32-63 mesh, Scientific Adsorbents). Elution for FCC usually employed a stepwise solvent polarity gradient, correlated with TLC mobility. Chromatograms were observed under UV (short and long wavelength) light, and/or were visualized by heating plates that were dipped in a solution of ammonium (VI) molybdate tetrahydrate (12.5 g) and cerium (IV) sulfate tetrahydrate (5.0 g) in 10% aqueous sulphuric acid (500 mL), or a solution of 20% sulfuric acid in ethanol. ¹H and ¹³C and ¹⁹F NMR spectra were recorded using Bruker Ultra Shield 400 MHz, Bruker Ultra Shield 500 MHz and Bruker Ultra Shield Plus 600 MHz instruments, in CDCl₃ or C₆D₆ solutions with residual CHCl₃ or C_6H_6 as internal standard (δ_H 7.27, 7.16 and δ_C 77.2, 128.4 ppm). Chemical shifts are quoted in ppm relative to tetramethysilane ($\delta_{\rm H}$ 0.00) and coupling constants (J) are given in Hertz. First order approximations are employed throughout. High resolution mass spectrometry was performed on an Ultima Micromass Q-TOF or Waters Micromass LCT Premier mass spectrometers.

General procedure for fluorination-reduction sequence on aldehydes: 2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2-*R*-fluoroethanol (6) and 2-(2', 3', 4', 6'-Tetra-O-benzyl- β -Dgalactopyranosyl)-2-*R*-fluoroethanol (7)

To a solution of aldehyde 5a (250 mg, 0.44 mmol) in THF (2.2 mL) was added (S)-Jørgensen catalyst (7.0 mg, 0.012 mmol). The mixture was stirred at rt for 15 min at which time NFSI (275 mg, 0.87 mmol) was added, and stirring continued at rt for 6 h, or after NMR indicated disappearance of the starting material. The reaction mixture was then concentrated at rt under reduced pressure and the residue filtered over a short column of silica gel. The filtrate was evaporated in vacuo at rt, the residue taken up in methanol (10 mL) and NaBH₄ (250 mg, 6.8 mmol) slowly added at 0 °C to the mixture. The reaction was stirred for 30 min, then pH adjusted to 6 with 1M HCl, and the volatiles removed under reduced pressure at rt. The residue was extracted with EtOAc and the organic phase washed with saturated aqueous NaHCO3, and brine, dried (Na₂SO₄), and evaporated in vacuo. FCC of the crude product gave 6 (195 mg, 75%) and 7 (27 mg, 10%).

For **6**: white amorphous solid, m.p = 79-83 °C; R_f 0.50 (60% hexanes: 40% EtOAc); $[\alpha]_D^{25}$ + 40 (c 1.6, CH₂Cl₂); ¹HNMR (500 MHz, CDCl₃) δ 7.38-7.16 (m, 20H), 4.65 (ddt, *J* = 2.8, 8.9, 48 Hz,

Journal Name

Journal Name

1H), 4.58-4.42 (m, 8H), 4.43 (m, partially buried, 1H), 4.33 (m, 1H), 4.20 (t, J = 8.1 Hz, 1H), 3.98 (dd, J = 3.2, 6.3 Hz, 1H), 3.95-3.78 (m, 2H), 3.75 (t, J = 3.5 Hz, 1H), 3.69 (m, 1H), 3.60 (dd, J = 3.1, 11.6 Hz, 1H), 2.91 (br s, 1H); ¹³CNMR (125 MHz, CDCl₃) δ 138.3, 138.2, 138.0, 137.8, 128.7, 128.6, 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 90.4 (d, $J_{C-F} = 169$ Hz), 74.2, 73.7, 73.5, 73.4 (two peaks), 73.3, 73.2, 71.7, 65.5, 65.2 (d, $J_{C-F} = 32$ Hz), 63.1 (d, $J_{C-F} = 21$ Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -198.5 (s); HRMS (ESI, MNa⁺) m/z calc for C₃₆H₃₉FNaO₆ 609.2628, found 609.2618.

For **7**: colorless oil; $R_f = 0.28$ (60% hexanes: 40% EtOAc); $[\alpha]_D^{25}$ +24 (c 0.7, CH_2Cl_2); ¹HNMR (600 MHz, CDCl₃) δ 7.32-7.22 (m, 20H), 5.00 (m, 2H), 4.91 (dt, J = 3.8, 51.7 Hz, 1H), 4.78 (m, 1H), 4.82-4.65 (m, 4H), 4.45 (ABq, J = 11.6 Hz, $\Delta \delta = 0.06$ ppm, 2H), 4.19 (t, J = 9.6 Hz, 1H), 4.05 (ddd, J = 6.2, 12.6, 19.6 Hz, 1H), 3.97 (bs, 1H), 3.85 (m, 1H), 3.66 (dd, J = 2.2, 9.5 Hz, 1H), 3.61 (dd, J = 6.3, 8.7 Hz, 1H), 3.58 (t, J = 6.1 Hz, 1H), 3.48 (dd, J = 5.8, 8.8 Hz, 1H), 3.43 (dd, J = 9.2, 28.4 Hz, 1H), 2.64 (m, 1H); ¹³CNMR (150 Mz, CDCl₃) δ 138.5, 138.2, 138.1, 137.7, 128.7, 128.4, 128.3, 128.2, 128.1, 127.9, 127.8, 75.8, 74.4, 74.3 (d, $J_{C-F} = 4.4$ Hz), 73.8, 72.9, 72.4, 69.3, 62.9 (d, $J_{C-F} = 23$ Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -206.9 (s); HRMS (ESI, MNa⁺) m/z calc for C₃₆H₃₉FNaO₆ 609.2628, found 609.2625

2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2-S-fluoroethanol (8), 2-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-2-*R*-fluoroethanol (9) and 2-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-2,2-difluoroethanol (10)

Aldehyde **5a** (150 mg, 0.265 mmol) was subjected to the standard fluorination-reduction procedure with (R)-Jørgensen catalyst, as described for the synthesis of **6** and **7**. FCC of the product after the reduction step afforded **6** (5 mg, 3%), **8** (105 mg, 68%), **9** (9 mg, 6%) and **10** (31 mg, 20%).

For **8**: colorless oil; $R_f = 0.38$ (40% EtOAc/60% hexanes); $[\alpha]_D^{25}$ + 39 (c 1.3, CH₂Cl₂); ¹HNMR (500 MHz, CDCl₃) δ 7.27-7.18 (m, 20H), 4.87 (dq, *J* = 4.3, 46.7 Hz, 1H), 4.76 (app d, J = 12.0 Hz, 1H), 4.71 (app d, J = 10.5 Hz, 1H), 4.67 (s, 2H), 4.53 (app d, J = 11.5 Hz, 1H), 4.48 (app d, J = 11.5 Hz, 1H), 4.40 (ABq, J = 12.0 Hz, $\Delta\delta$ = 0.08 ppm, 2H), 4.14 (m, 1H), 4.05 (br t, J = 7.1 Hz, 1H), 3.97 (ddd, *J* = 3.0, 5.9, 34.1 Hz, 1H), 3.96 (dd, *J* = 2.0, 9.5 Hz 1H), 3.84 (t, *J* = 2.6 Hz, 1H), 3.81-3.76 (m, 2H), 3.59 (t, *J* = 9.5 Hz, 1H), 3.31 (dd, *J* = 4.4, 10.2 Hz, 1H), 2.43 (br s, 1H); ¹³CNMR (125 MHz, CDCl₃) δ 138.7, 138.6, 138.3, 138.2, 128.7, 128.6, 128.5, 128.3, 128.1, 128.0, 127.8, 93.5 (d, *J*_{C-F} = 35 Hz), 69.2, 63.1 (d, *J*_{C-F} = 25 Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -198.5 (s); HRMS (ESI, MNa⁺) *m*/*z* calc for C₃₆H₃₉FNaO₆ 609.2628, found 609.2618.

For **9**: colorless oil; $R_f = 0.48$ (40% EtOAc/60% hexanes); $[\alpha]_{D}^{25}$ + 32 (c 0.1, CH₂Cl₂); ¹HNMR (500 MHz, C₆D₆) δ 7.03-7.38 (m, 20H), 4.94 (m, 2H), 4.88 (dt, J = 4.5, 45 Hz, 1H partially buried under m at 4.94 ppm), 4.55 (m, 2H), 4.30 (ABq, *J* = 11.8 Hz, $\Delta\delta$ = 0.06 ppm, 2H), 4.24 (ABq, *J* = 11.8 Hz, $\Delta\delta$ = 0.05, 2H), 4.08 (t, *J* = 9.6 Hz, 1H), 3.85 (m, 3H), 3.70 (ddd, *J* = 1.8, 10.0, 11.8 Hz, 1H), 3.63 (t, *J* = 8.1 Hz, 1H), 3.52 (dd, *J* = 5.4, 8.9 Hz, 1H), 3.36

ARTICLE

(m, 1H), 3.33 (dd, J = 2.7, 8.2 Hz, 1H); ¹³CNMR (125 MHz, C₆D₆) δ 139.7, 138.2, 139.2, 139.1, 129.0 (two peaks), 128.9, 128.5 128.3 (two peaks), 128.1, 128.0, 93.4 (d, *J*_{C-F} = 174 Hz), 85.7, 80.5 (d, *J*_{C-F} = 21 Hz), 77.5, 75.5 (d, *J*_{C-F} = 8 Hz), 75.4, 75.3, 74.1, 73.9, 72.4, 69.1, 62.3 (d, *J*_{C-F} = 25 Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -196.5 (s); HRMS (ESI, MNa⁺) *m/z* calc for C₃₆H₃₉FNaO₆ 609.2628, found 609.2626.

For **10**: colorless oil; $R_f = 0.63$ (40% EtOAc/60% hexanes); $[\alpha]_D^{26}$ +44 (c 0.6, CH₂Cl₂); ¹HNMR (500 MHz, C₆D₆) δ 7.22-7.07 (m, 20H), 4.62 (ddd, J = 2.1, 8.9, 18.0 Hz, 1H), 4.52 (apparent d , J = 11.5 Hz, 1H), 4.47 (m, 1H), 4.38 (br m, 5H), 4.18 (ABq, *J* = 11.8 Hz, $\Delta\delta$ = .05 ppm, 1H), 4.10 (br m, 1H), 4.05 (dd, *J* = 2.2, 5.0 Hz, 1H), 3.99 (dd, *J* = 3.1, 5.3 Hz, 1H), 3.93 (bm, 1H), 3.80 (dd, *J* = 3.1, 5.0 Hz, 1H), 3.71 (dd, *J* = 2.9, 11.2 Hz, 1H), 3.27 (t, *J* = 7.5 Hz, 1H); ¹³CNMR (125 MHz, C₆D₆) δ 139.0, 138.8, 138.7, 132.4, 132.2, 131.9, 131.7, 128.8, 128.1, 127.9, 127.8, 127.7, 127.6, 127.5, 127.4, 127.2, 125.6, 125.2, 123.6 (t, *J_{C-F}* = 243 Hz), 75.6, 75.0, 74.9, 74.3, 73.9, 73.8, 73.7, 72.6, 68.3 (dd, *J_{C-F}* = 25, 34 Hz), 66.6, 64.1 (dd, *J_{C-F}* = 28, 34 Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -114.6 (ABq, *J* = 256 Hz, $\Delta\delta$ = 9.8 ppm, 2F); HRMS (ESI, MNa⁺) *m/z* calc for C₃₆H₃₈F₂NaO₆ 627.2534, found 627.2523.

Synthesis of 7 from 5b

Aldehyde **5b** (46 mg, 0.081 mmol) was subjected to the standard fluorination procedure with NFSI (51 mg, 0.16 mmol) and (*S*)-Jørgensen catalyst (15 mg, 0.024 mmol) for 1.5 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **7** (21 mg, 44%).

Synthesis of 9 from 5b

Aldehyde **5b** (46 mg, 0.081 mmol) was subjected to the standard fluorination procedure with NFSI (51 mg, 0.16 mmol) and (*R*)-Jørgensen catalyst (15 mg, 0.024 mmol) for 1.5 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **9** (24 mg, 50%).

Synthesis of 10 using racemic catalyst

Aldehyde **5a** (330 mg, 0.58 mmol) was subjected to the fluorination procedure described for **6**, with NFSI (730 mg, 2.30 mmol), (*R*)-Jørgensen catalyst (35 mg, 0.058 mmol) and and (*S*)-Jørgensen catalyst (35 mg, 0.058 mmol) for 18 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded **10** (290 mg, 83%).

General procedure for fluorination-Wittig methylenation sequence on aldehydes: 3-(2', 3', 4', 6'-Tetra-O-benzyl- α -Dgalactopyranosyl)-3-*R*-fluoropropene (11) and 3-(2', 3', 4', 6'-Tetra-O-benzyl- β -D-galactopyranosyl)-3-*R*-fluoropropene (12)

To a solution of aldehyde **5a** (490 mg, 0.87 mmol) in THF (5.0 mL) was added (*S*)-Jørgensen catalyst (25 mg, 0.022 mmol). The mixture was stirred at rt for 15 min at which time NFSI (540 mg, 1.7 mmol) was added, and stirring continued at rt for

ARTICLE

6 h. The reaction was monitored and processed as described for the synthesis of **6**. The crude fluoro-aldehyde was used directly in the next step. To a solution of Ph_3PCH_3Br (932 mg, 2.6 mmol) in anhydrous THF (10 mL) was added 2M NaHMDS in THF (1.3 mL, 2.6 mmol) at -78 °C. The resulting bright orange solution was stirred for 1 h at this temperature and then a solution of the product from the previous step in THF (10 mL) was added. The reaction mixture was warmed to rt over 2 h, then poured into saturated aqueous NH₄Cl and extracted with EtOAc. The organic phase was washed with brine, dried (Na₂SO₄), filtered, and evaporated under reduced pressure. FCC of the residue afforded **11** (423 mg, 73%) and **12** (51 mg, 9%).

For **11**: White needles (ether:methanol); m.p = 97-98 $^{\circ}$ C; R_f = 0.35 (20 % ethyl ether: 80% hexanes); $[\alpha]_{D}^{25}$ +53 (c 0.5, CH₂Cl₂); ¹HNMR (500 MHz, CDCl₃) δ 7.27-7.10 (m, 20H), 6.05 (m, 1H), 5.43 (br d, J = 17.5 Hz, 1H), 5.28 (d, J = 11.0 Hz, 1H), 5.02 (ddd, *J* = 5.5, 7.0, 46.8 Hz, 1H), 4.40-4.59 (m, 8H), 4.33 (t, *J* = 7.2 Hz, 1H), 4.07 (dd, *J* = 8.5, 11.5 Hz, 1H), 3.99 (dd, *J* = 2.4, 7.0 Hz, 1H), 3.72 (m, 3H); ¹³CNMR (125 MHz, CDCl₃) δ 138.8, 138.5, 138.3, 138.0, 134.8 (d, *J*_{C-F} = 17 Hz), 128.6, 128.5, 128.4, 128.2, 127.9, 127.8, 127.7, 118.0 (d, *J*_{C-F} = 13 Hz), 90.2 (d, *J*_{C-F} = 168 Hz), 75.3, 74.7, 74.0, 73.6, 73.5, 73.3, 73.2, 71.8, 69.6 (d, *J*_{C-F} = 29 Hz), 65.8; ¹⁹FNMR (377 MHz, CDCl₃) δ -190.5 (s); HRMS (ESI, MNa⁺) *m/z* calc for C₃₇H₃₉FNaO₆ 605.2679, found 605.2668.

For **12**: colorless oil; $R_f = 0.27$ (20 % ethyl ether: 80% hexanes); $[\alpha]_D^{26+}$ 30 (c 1.1, CH₂Cl₂); (500 MHz, CDCl₃) δ 7.30-7.15 (m, 20H), 6.01 (m, 1H), 5.32 (ddd, J = 1.0, 2.5, 17.0 Hz, 1H), 5.21 (d, J = 10.5 Hz, 1H), 5.09 (dd, J = 8.0, 47.0 Hz, 1H), 4.91 (app d, J = 10.5, 1H), 4.87 (app d, J = 11.5, 1H), 4.70-4.55 (m, 4H), 4.37 (ABq, J = 11.5, $\Delta\delta$ = 0.05 ppm, 2H), 4.08 (t, J = 9.5 Hz, 1H), 3.90 (d, J = 2.0 Hz, 1H), 3.53 (m, 3H), 3.46 (t, J = 6.3 Hz, 1H), 3.20 (ddd, J = 1.7, 9.7, 27.7 Hz, 1H); ¹³CNMR (125 MHz, CDCl₃) δ 138.9, 138.5 (two peaks), 138.2, 133.6 (d, $J_{C-F} = 20$ Hz), 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 118.9 (d, $J_{C-F} = 12$ Hz), 90.7 (d, $J_{C-F} = 174$ Hz), 85.0, 80.9 (d, $J_{C-F} = 20$ Hz), 78.0, 75.7, 74.9 (d, $J_{C-F} = 4$ Hz), 74.6, 73.8, 73.5, 72.5, 69.2; ¹⁹FNMR (377 MHz, CDCl₃) δ -193.7 (s); HRMS (ESI, MNa⁺) m/z calc for C₃₇H₃₉FNaO₆ 605.2679, found 605.2679.

3-(2', 3', 4', 6'-Tetra-O-benzyl-α-D-galactopyranosyl)-3-(S)fluoropropene (13)

Aldehyde **5a** (540 mg, 0.95 mmol) was subjected to the standard fluorination-Wittig methylenation procedure with (R)-Jørgensen catalyst, as described for the synthesis of **11** and **12**. FCC of the product after the methylenation step afforded **13** (309 mg, 53 %) and an unseparated mixture of fluorinated products (180 mg, ca 31%).

For **13**: colorless oil; $R_f = 0.06 (CHCl_3); [\alpha]_D^{25} + 19$ (c 5.5, CH_2Cl_2); ¹HNMR (500 MHz, CDCl_3) δ 7.25-7.17 (m, 20H), 5.82 (m, 1H), 5.30 (dt, J = 1.6, 18.6 Hz, 1H), 5.20 (d, J = 10.7 Hz, 1H), 5.19 (dt, J = 5.0, 47.8 Hz, 1H), 4.69-4.40 (m, 8H), 4.19 (m, 1H), 3.92 (m, 3H), 3.82 (dt, J = 4.5, 29.0 Hz, 1H), 3.70 (dd, J = 7.2, 10.3 Hz, 1H), 3.52 (dd, J = 5.1, 10.3 Hz, 1H); ¹³CNMR (125 MHz, CDCl₃) δ 138.8, 138.7, 138.6, 138.4, 133.2 (d, $J_{C-F} = 19$ Hz),

128.7, 128.6, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 119.0 (d, $J_{C-F} = 11$ Hz), 94.1 (d, $J_{C-F} = 174$ Hz), 77.9, 76.2, 74.9, 74.5, 73.8, 73.6, 73.5, 73.4, 73.3 (d, $J_{C-F} = 14$ Hz), 73.2, 67.9; ¹⁹FNMR (377 MHz, CDCl₃) δ -184.7 (s); HRMS (ESI, MNa⁺) m/z calc for C₃₇H₃₉FNaO₆ 605.2679, found 605.2671.

3-(2', 3', 4', 6'-Tetra-O-benzyl- α -D-galactopyranosyl)-3,3difluoropropene (14)

To a solution of aldehyde 5a (5.30 g, 9.35 mmol) in THF (50 mL) was added (S)-Jørgensen (250 mg, 0.47 mmol) and (R)-Jørgensen catalyst (250 mg, 0.47 mmol). The mixture was stirred at rt for 15 min at which time NFSI (7.00 g, 22.4 mmol) was added, and stirring continued at rt for 12 h. The reaction mixture was concentrated at rt under reduced pressure and the residue filtered over a short column of silica gel. The filtrate was evaporated in vacuo at rt, and the crude aldehyde was used in the next step without further purification. To a solution of Ph₃PCH₃Br (15.0 g, 37.4 mmol) in anhydrous THF (90 mL) was added 2M NaHMDS in THF (23 mL, 46.0 mmol). The resulting bright orange solution was stirred for 1 h at this temperature at which time a solution of the crude aldehyde in THF (30 mL) was added. The reaction mixture was warmed to rt over 2 h, then processed following the general procedure for the Wittig reaction, as described for the synthesis of 11 and 12. FCC of the crude product afforded 14 (2.23 g, 40%): colorless oil; $R_f = 0.17$ (CHCl₃); $[\alpha]_D^{25} + 37$ (c 0.4, CH₂Cl₂); ¹HNMR (600 MHz, C_6D_6) δ 7.04-7.38 (m, 20H), 6.36 (m, 1H), 5.71 (bd, J = 17.5 Hz, 1H), 5.10 (d, J = 11.2 Hz, 1H), 4.57 (m, 1H), 4.35-4.50 (m, 6H), 4.31 (ABq, J = 11.0 Hz, $\Delta\delta$ = 0.03 ppm, 2H), 4.22 (app d, J = 11.5 Hz, 1H), 4.20 (dd, J = 7.9, 11.3 Hz, 1H), 4.06 (dd, J = 2.4, 5.3 Hz, 1H), 4.00 (m, 1H), 3.97 (dd, J = 3.5, 11.3 Hz, 1H), 3.82 (dd, J = 2.8, 4.4 Hz, 1H); ¹³CNMR (150 MHz, C₆D₆) δ 139.7, 139.2 (two peaks), 138.8, 132.9 (t, J_{C-F} = 24 Hz), 129.3, 129.2, 129.1, 129.0, 128.9, 128.8, 128.7, 128.5, 128.1, 128.0, 127.9, 127.8, 120.9 (t, $J_{C-F} = 242$ Hz), 119.3 (t, $J_{C-F} = 10$ Hz), 76.1 (two peaks), 76.0, 75.9, 74.5, 73.7 (two peaks), 72.9, 72.3 (t, J_{CF} = 31 Hz), 67.0; ¹⁹FNMR (377 MHz, CDCl₃) δ -103.2 (ABq, $J = 264 \text{ Hz}, \Delta \delta = 7.3 \text{ ppm}, 2\text{F}$); HRMS (ESI, MNa⁺) m/zcalc for C₃₇H₃₈F₂NaO₆ 623.2585, found 623.2571.

Synthesis of difluoroalkene 14 from alcohol 10

To a solution of alcohol **10** (40 mg, 0.066 mmol) in dry DCM (1 mL) was added Dess-Martin periodinane solution (0.1 mL of a 15% wt/vol in DCM, 0.035 mmol). The reaction mixture was stirred at rt for 20 min, then concentrated at rt under reduced pressure. Next, the residue was filtered over a short column of silica gel, and the filtrate evaporated in *vacuo* at rt. The crude aldehyde product was used in the next step without further purification. To a solution of Ph₃PCH₃Br (400 mg, 1.1 mmol) in anhydrous THF (1 mL) was added 2M NaHMDS in THF (0.7 mL, 1.4 mmol). The resulting bright orange solution of the crude aldehyde in THF (1.0 mL) was added. The reaction mixture was warmed to rt over 2 h, then processed following the standard procedure for Wittig reactions. FCC of the crude product afforded **14** (32 mg, 80%).

Unemistry Accepted Manuscrip

& **Biomolecular**

ganic

$2\mathchar`-(2', 3', 4', 6'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 3', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathchar`-(2', 5'-Tetra-O-benzyl-\beta-D-galactopyranosyl)\mathcha$

Aldehyde 5b (65 mg, 0.115 mmol) was subjected to the fluorination procedure described for 6, with NFSI (181 mg, 0.570 mmol), (R)-Jorgensen catalyst (10 mg, 0.017 mmol) and and (S)-Jorgensen catalyst (10 mg, 0.017 mmol) for 18 h. Reduction of the crude product with NaBH₄ following the standard procedure and FCC of the product afforded 15 (11 mg, 16%): colorless oil; R_f = 0.60 (5% acetone: 95% CH₂Cl₂); $[\alpha]_{D}^{20}$ +20 (c 0.1, CHCl₃); ¹HNMR (600 MHz, C₆D₆) δ 7.40-7.05 (m, 20H), 4.84 (ABq, J = 10.7 Hz, $\Delta\delta$ = 0.14 ppm, 2H), 4.74 (ABq, J = 11.6 Hz, $\Delta\delta$ = 0.40 ppm, 2H), 4.45 (t, J = 9.4 Hz, 1H), 4.41 (ABq, J = 11.8 Hz, $\Delta\delta$ = 0.02 ppm, 2H), 4.38 (br m, 5H), 4.22 (ABq, J = 11.8 Hz, $\Delta\delta$ = 0.10 ppm, 2H), 3.91 (br m, 1H), 3.78 (br m, 2H), 3.70 (d, J = 1.90 Hz, 1H), 3.52 (dd, J = 6.3, 9.2 1H), 3.47 (dd, J = 6.6, 9.2 Hz, 1H), 3.34 (dd, J = 2.8, 9.3 Hz, 1H), 3.30 (t, J = 6.5 Hz, 1H), 2.15 (t, J = 6.5 Hz, 1H); 13 CNMR (150 MHz, C₆D₆) δ 139.6, 139.5, 139.2, 138.9, 130.0-127.0 (several lines buried under C₆D₆ triplet), 121.9 (t, J_{C-F} = 245 Hz), 85.2, 78.2 (t, J_{C-F} = 28 Hz), 77.9, 75.5, 75.3, 75.1 (d, J_{C-F} = 3 Hz), 74.3, 74.0, 73.0, 69.5, 64.2 (t, J_{C-F} = 32 Hz); ¹⁹FNMR (377 MHz, CDCl₃) δ -114.3 (ABq, $J = 264 \text{ Hz}, \Delta \delta = 6.6 \text{ ppm}, 2\text{F}$); HRMS (ESI, MNa⁺) m/zcalc for $C_{36}H_{38}F_2NaO_6$ 627.2534, found 627.2521.

General procedure for correlation of allylic fluorides and fluorohydrin derivatives: Transformation of 11 to 6

A stream of O_3/O_2 was bubbled through a solution of **11** (50 mg, 0.09 mmol) in DCM 20 mL at -78 °C, for 5 min. PPh₃ (50 mg, 0.19 mol) was then added to the mixture and stirring continued at rt for an additional 2 h. The volatiles were evaporated in *vacuo*, and NaBH₄ (30 mg, 0.8 mmol) was then slowly added at 0 °C to a solution of the residue in MeOH (2 mL). The reaction mixture was processed following the standard procedure for NaBH₄ reduction. FCC of the crude product provided **6** (31 mg, 62%).

Acknowledgements

This work was supported by the National Science Foundation (#1301330). A Research Centers in Minority Institutions Program grant from the National Institute of Health Disparities (MD007599) of the National Institutes of Health (NIH), which supports the infrastructure at Hunter College, is also acknowledged. We thank Dr. Matthew Devany for help with NMR experiments.

Notes and references



- 2 D. O'Hagan, Chem. Soc. Rev., 2008, 37, 308-319.
- 3 S. Purser, P. R. Moore, S. Swallow and V. Gouverneur, *Chem. Soc. Rev.*, 2008, **37**, 320–330
- 4 P. A. Champagne, J. Desroches, J.-D. Hamel, M. Vandamme and J.-F. Paquin, *Chem. Rev.*, 2015, **115**, ASAP, doi: 1021/cr500706a.
- 5 X. Yang, T. Wu, R. J. Phipps and D. F. Toste, *Chem. Rev.*, 2015, **115**, 826-870.
- 6 G. Valero, X. Companyó and R. Rios, *Chem. Eur. J.* 2011, **17**, 2018-2037.
- 7 D. Schedler and D. C. Baker, Carbohydr. Res., 2004, 339, 1585-1595.
- 8 Y. Cheng, A.-L. Guo and D.-S. Guo, *Cur. Org. Chem.*, 2010, **14**, 977-999.
- 9 E. Leclerc, X. Pannecoucke, M. Ethève-Queiquejeu and M. Sollogoub, Chem. Soc. Rev. 2013, 42, 4270-4283.
- 10 R. Plantier-Royon and C. Portella, *Carbohydr. Res.*, 2000, **327**, 119-146.
- 11 J. Jiménez-Barbero, R. Demange, K. Schenk and P. Vogel, J. Org. Chem., 2001, 66, 5132–5138.
- 12 S, Marcotte, F. D'Hooge, S. Ramadas, C. Feasson, X. Pannecoucke and J.-C. Quirion, *Tetrahedron Lett.*, 2001, 42, 5879-5882.
- 13 J. Picard, J., N. Lubin-Germain, J. Uziei and J. Augé, *Synthesis*, 2006, 979-982.
- K. A. Tony, R. W. Denton, A. Dilhas, J. Jiménez-Barbero and D. R. Mootoo, *Org. Lett.*, 2007, 9, 1441-1444.
- 15 J. Pérez-Castells, J. J. Hernández-Gay, R. W. Denton, K. A. Tony, D. R. Mootoo, D. R. and J. Jiménez-Barbero, Org. Biomol. Chem., 2007, 5, 1087-1092.
- 16 F. Poulain, A.-L. Serre, J. Lalot, E. Leclerc and J.-C. Quirion, *J. Org. Chem.*, 2008, **73**, 2435-2438.
- 17 S. Colombel, N. Van Hijfte, T. Poisson, E. Leclerc and X. Pannecoucke, *Chem. Eur. J.*, 2013, **19**, 12778-12787.
- 18 S. Colombel, N. Van Hijfte, T. Poisson, X. Pannecoucke, F. Monneaux and E. Leclerc, J. Fluorine Chem. 2015, 173, 84-91.
- 19 For examples of general strategies for transforming Cglycoside alkenes and aldehydes to more complex Cglycosides, T. Gustafsson, M. Saxin, and J. Kihlberg, *J. Org. Chem.*, 2003, **68**, 2506-2509, and ref. 19 – 23.
- 20 P. Wipf and J. G. Pierce, Org. Lett., 2006, 8, 3375-3378.
- 21 J. Pu and R. W. Franck, Tetrahedron, 2008, 64, 8618-8629.
- 22 V. N. Thota, J. Gervay-Hague and S. S. Kulkarni, Org. Biomol. Chem., 2012, **10**, 8132-8139.
- 23 V. N. Thota, M. Brahmaiah and S. S., Kulkarni, J. Org. Chem., 2013, 78, 12082-12089.
- 24 A. S. Altiti and D. R. Mootoo, Org. Lett., 2014, 16, 1466-1469.
- 25 M. Marigo, D. Fielenbach, A. Braunton, A. Kjærsgaard and K. A. Jørgensen, Angew. Chem. Int. Ed., 2005, 44, 3703 –3706.
- 26 D. D. Šteiner, N. Mase and C. F. Barbas, III, Angew. Chem. Int. Ed., 2005, 44, 3706-3710.
- 27 T. D. Beeson and D. W. C. MacMillan, J. Am. Chem. Soc., 2005, 127, 8826-8828.

Organic & Biomolecular Chemistry Accepted Manuscript

- 28 A. Nuzzi, A. Massi and A. Dondoni, *Org. Lett.*, 2008, **10**, 4485-4488.
- 29 E. G. Nolen, M. M. Watts and D. J. Fowler, *Org. Lett.*, 2002, 4, 3963-3965.
- 30 For related one-pot reactions on simple aldehydes using the Jørgensen catalyst, H. Jiang, A. Falcicchio, K. L. Jensen, M. W. Paixão, S. Bertelsen and K. A. Jørgensen, J. Am. Chem. Soc., 2009, **131**, 7153-7157.
- 31 J. Jimenez-Barbero, J. F. Espinosa, J. L. Asensio, F. J. Cañada, and A. Poveda, *Adv. Carbohydr. Chem. Biochem.*, 2001, **56**, 235-284.
- 32 A. Massi, A. Nuzzi and A. Dondoni, *J. Org. Chem.*, 2007, **72**, 10279-10282.