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A highly selective and sensitive fluorescent turn-on Al³⁺ chemosensor in aqueous media and living cells: Experimental and theoretical studies

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Abstract

A highly selective and sensitive fluorescent chemosensor (1) based on 2aminobenzoic acid Schiff base has been synthesized in one step. The binding properties of 1 with metal ions were investigated by fluorescence, UV-vis, ¹H NMR and electrospray ionization mass spectrum. This sensor exhibited an enhanced fluorescence in the presence of Al^{3+} in aqueous solution and also in living cells. The sensing mechanism of 1 toward Al^{3+} was explained by DFT/TDDFT calculations. The binding constant was determined to be 3.1 x 10⁸ M⁻¹ which is one of the highest binding affinities among such simple Schiff bases for Al^{3+} in buffer solution. The calibration curve for the analytical performance of Al^{3+} was found to be liner over a concentration range with a very low detection limit of 290 nM (3 σ).

Keywords: aluminum ion, carboxylic group, Schiff base, fluorescent, chemosensor

1. Introduction

The development of selective and sensitive chemosensors for the detection of metal ions has received considerable attention because of their important roles in medicine, living systems and the environment.¹⁻⁷ Among metals, aluminum is the third most prevalent metallic element in the Earth. People are widely exposed to aluminum because of its widespread use in food additives, aluminum-based pharmaceuticals and storage/cooking utensils.⁸⁻¹³ The World Health Organization (WHO) prescribed the average human intake of aluminum as around 3-10 mg day⁻¹ and limited its drinking water concentration to 7.41 µM. Tolerable weekly aluminum intake in the human body is estimated to be 7 mg kg⁻¹ body weight.¹⁴⁻¹⁶ The superfluous ingestion of aluminum influences the absorption of calcium in the bowel, causing softening of the bone, atrophy and even aberrance, and also affects the absorption of iron in blood, causing anemia. In addition, the toxicity of aluminum causes damage to the central nervous system, is suspected to be involved in neurodegenerative diseases such as Alzheimer's and Parkinson's, and is responsible for intoxication in hemodialysis patients.¹⁷⁻¹⁹

In recent years, many efforts have been devoted to design various chemosensors specific for Al³⁺ detection.²⁰⁻⁶¹ However, many of them have suffered from cost of synthesis, the number of synthesis steps, selectivity, binding constant, detection limit, use of harmful organic solvents, and poor water solubility. Therefore, it is necessary to develop new sensors for selectively detecting Al³⁺ in aqueous solution. To increase the water solubility, we used 2-aminobenzoic acid with a carboxylic group acting as a hydrophilic character. To the best of our knowledge, very few studies have been reported on the use of the aminobenzoic acid as a fluorescent chemosensor for the detection of metal ions.^{32,62-65}

Herein, we report an aminobenzoic acid-based Schiff base **1** as a highly selective and sensitive fluorescent chemosensor for Al^{3+} . **1** showed one of the highest binding affinities $(K_a = 3.0 \times 10^8 \text{ M}^{-1})$ among such simple Schiff bases for Al^{3+} in aqueous solution and quite low detection limit $(2.9 \times 10^{-7} \text{ M})$ of a nanomolar concentration level.

2. Experimental section

2.1. Materials and instrumentation

All the solvents and reagents (analytical grade and spectroscopic grade) were obtained from Sigma-Aldrich and used as received. The Live/Dead assay (LIVE/DEAD[®] Viability/Cytotoxicity Kit) kit was purchased from Life Technologies Company. ¹H NMR measurements were performed on a Varian 400 MHz spectrometer and chemical shifts were recorded in ppm. Electrospray ionization mass spectra (ESI-MS) were collected on a Thermo Finnigan (San Jose, CA, USA) LCQTM Advantage MAX quadruple ion trap instrument. Elemental analysis for carbon, nitrogen, and hydrogen was carried out by using a Flash EA 1112 elemental analyzer (thermo) in Organic Chemistry Research Center of Sogang University, Korea. Absorption spectra were recorded at 25 °C using a Perkin Elmer model Lambda 2S UV/Vis spectrometer. Fluorescence measurements were performed on a Perkin Elmer model LS45 fluorescence spectrometer.

2.2. Synthesis of the fluorescence sensor 1

A solution of 2-aminobenzoic acid (0.14 g, 1 mmol) in ethanol was added to a solution containing 4-(diethylamino)-2-hydroxybenzaldehyde (0.19 g, 1 mmol) in ethanol and the mixture was stirred for 5 min. Then, two drops of HCl were added into the reaction solution and it was stirred for 3 h at room temperature. The solvent was evaporated and

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bright yellow product was recrystallization from acetonitrile. The yield: 0.23 g (74.3 %). ¹H NMR (DMSO-*d*₆, 400 MHz) δ : 8.72 (s, 1H), 7.93 (d, 1H, *J* = 7.6 Hz), 7.58 (d, 2H, *J* = 6.8 Hz), 7.44 (d, 1H, *J* = 8.8 Hz), 7.29 (m, 1H, *J* = 7.6 Hz), 6.41 (d, 1H, *J* = 8.4 Hz), 6.39 (s, 1H), 3.45 (q, 4H, *J* = 6.8 Hz), 1.14 (t, 6H, *J* = 7.0 Hz). ESI-mass *m*/*z* [M-H⁺]⁻: calcd, 311.14; found, 311.33. Anal. Calcd for C₁₈H₂₀N₂O₃ (312.15): C, 69.21; H, 6.45; N, 8.97. Found: C, 69.60; H, 6.30; N, 8.70 %.

2.3. Fluorescence and UV-vis measurements

For fluorescence measurements, sensor **1** (1.6 mg, 0.005 mmol) was dissolved in methanol (5 mL) and 30 μ L of the sensor **1** (1 mM) were diluted to 2.97 mL bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)) to make the final concentration of 10 μ M. Al(NO₃)₃ 9H₂O (37.5 mg, 0.1 mmol) was dissolved in methanol (5 mL). 1.5-28.5 μ L of the Al³⁺ solution (20 mM) were transferred to each sensor solution (10 μ M) prepared above. After mixing them for a few seconds, fluorescence spectra were obtained at room temperature.

For UV-vis measurements, all experimental conditions were identical above. 1.5-21 μ L of the Al³⁺ solution (10 mM) were transferred to each sensor solution (10 μ M) prepared above. After mixing them for a few seconds, UV-vis spectra were taken at room temperature.

2.4. Job plot measurement

Sensor **1** (1.6 mg, 0.005 mmol) was dissolved in methanol (5 mL). 30, 27, 24, 21, 18, 15, 12, 9, 6, 3 and 0 μ L of the sensor **1** solution were taken and transferred to vials. Each vial was diluted with bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)) to make a

total volume of 2.97 mL. Al(NO₃)₃·9H₂O (1.9 mg, 0.005 mmol) was dissolved in methanol (5 mL). 0, 3, 6, 9, 12, 15, 18, 21, 24, 27 and 30 μ L of the Al(NO₃)₃ solution were added to each diluted sensor **1** solution. Each vial had a total volume of 3 mL. After shaking the vials for a few seconds, fluorescence spectra were taken at room temperature.

2.5. NMR titration

For ¹H NMR titrations of sensor **1** with Al³⁺, four NMR tubes of sensor **1** (31.2 mg, 0.1 mmol) dissolved in DMSO- d_6 (700 µL) were prepared and then four different concentrations (0, 0.02, 0.05 and 0.1 mmol) of Al(NO₃)₃ 9H₂O dissolved in DMSO were added to each solution of sensor **1**. After shaking them for a minute, ¹H NMR spectra were taken at room temperature.

2.6. Competition with other metal ions

Sensor 1 (1.6 mg, 0.005 mmol) was dissolved in methanol (5 mL) and 30 μ L of the sensor 1 (1 mM) were diluted to 2.97 mL bis-tris buffer (10 mM, water:CH₃OH=1:1 (v/v)) to make the final concentration of 10 μ M. MNO₃ (M = Na, K, 0.1 mmol) or M(NO₃)₂ (M = Mn, Co, Ni, Cu, Zn, Cd, Hg, Mg, Ca, Pb and Cr³⁺, 0.1 mmol) or M(NO₃)₃ (M = Fe, Al, Ga, In, 0.1 mmol) were separately dissolved in methanol (5 mL). 27 μ L of each metal solution (20 mM) were taken and added into 3 mL of each sensor 1 solution (10 μ M) prepared above to make 18 equiv. Then, 27 μ L of Al(NO₃)₃ solution (20 mM) were added into the mixed solution of each metal ion and sensor 1 to make 18 equiv. After mixing them for a few seconds, fluorescence spectra were taken at room temperature.

2.7. Methods for cell imaging and cytotoxicity

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Human dermal fibroblast cells in low passage were cultured in FGM-2 medium (Lonza, Switzerland) supplemented with 10% fetal bovine serum, 1% penicillin/streptomycin in an *in vitro* incubator with 5% CO₂ at 37 °C. Cells were seeded onto a 12 well plate (SPL Lifesciences, Korea) at a density of 2×10^5 cells per well and then incubated at 37 °C for 4 h after the addition of various concentrations (0-200 µM) of Al(NO₃)₃ dissolved in DMSO. After washing with phosphate buffered saline (PBS) two times to remove the remaining Al(NO₃)₃, the cells were incubated with **1** dissolved in medium/DMSO solution (FGM-2-medium : DMSO= 9:1 v/v) at room temperature for 30 min. The cells were observed using a microscope (Olympus, Japan). The fluorescent images of the cells were obtained using a fluorescence microscope (Leica DMLB, Germany) at the excitation wavelength of 410 nm.

To observe cell viability, a live & dead assay was performed for **1** and **1**-Al³⁺ complex. Fibroblasts (P=5) were in vitro cultured to reach 70% confluent. The cells were incubated with **1** (10 μ M) dissolved in medium/DMSO solution (FGM-2-medium : DMSO= 9:1 v/v) for 1 h, 12 h and 24 h, respectively. Reagent (400 μ L) of the live & dead assay was added into each cell culture plate. In case of **1**-Al³⁺ complex, the cells were incubated with Al(NO₃)₃ (150 μ M) dissolved in MeCN. After washing with phosphate buffered saline (PBS) two times to remove the remaining Al(NO₃)₃, the cells were incubated with **1** dissolved in medium/DMSO solution (FGM-2-medium : DMSO= 9:1 v/v) at room temperature for 1 h, 12 h and 24 h, respectively. Reagent (400 μ L) of the live & dead assay was added into each cell culture plate. Both viability and morphological changes of the cells were observed by a fluorescence microscope (Leica DMLB, Leica; Wetzlar, Germany).

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2.8. Theoretical calculations

All theoretical calculations were performed by using the Gaussian 03 suite.⁶⁶ The singlet ground states (S₀) of **1** and Al³⁺-2·**1** complex were optimized by DFT methods with Becke's three parametrized Lee-Yang-Parr (B3LYP) exchange functional^{67,68} with 6-31G** basis set^{69,70}. In vibrational frequency calculations, there was no imaginary frequency for **1** and Al³⁺-2·**1** complex, suggesting that the optimized **1** and Al³⁺-2·**1** complex represented local minima. For all calculations, the solvent effect was considered by using the Cossi and Barone's CPCM (conductor-like polarizable continuum model).^{71,72} To investigate the electronic properties of singlet excited states, time-dependent DFT (TDDFT) was performed in the ground state geometries of **1** and Al³⁺-2·**1** complex. The 20 singlet-singlet excitations were calculated and analyzed. The GaussSum 2.1⁷³ was used to calculate the contributions of molecular orbitals in electronic transitions.

3. Results and discussion

3.1. Spectroscopic measurements of 1 toward Al³⁺

Sensor **1** was obtained by coupling 2-aminobenzoic acid with 4-(diethylamino)-2hydroxybenzaldehyde with 74% yield in ethanol (Scheme 1), and characterized by ¹H NMR, ESI-mass spectrometry, and elemental analysis.



Scheme 1. Synthesis of sensor 1.

We studied the fluorescence response behaviors of **1** towards various metal ions in bistris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)). As shown in Fig. 1, sensor **1** alone displayed a very weak fluorescence emission upon excitation at 410 nm. The addition of Al³⁺ resulted in a prominent fluorescence enhancement (11-folds) at 460 nm. In contrast, other metal ions such as Mn²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Hg²⁺, Na⁺, K⁺, Mg²⁺, Ca²⁺, Pb²⁺ and Cr³⁺ caused almost no fluorescence increase. Importantly, the addition of Ga³⁺ or In³⁺, which belong to the same group with Al³⁺ in the periodic table, had no significant effect on the fluorescence, demonstrating that sensor **1** was even highly selective for Al³⁺ over Ga³⁺ and In³⁺. So far, only a few selective sensors which can discriminate Al³⁺ from both Ga³⁺ and In³⁺ have been reported.^{20-22,26} The selectivity of **1** toward Al³⁺ in bis-tris buffer alone was also observed, but fluorescence enhancement was not high enough (Fig. S1).



Fig. 1. Fluorescence spectra of sensor **1** (10 μ M) upon addition of metal nitrate salts (18 equiv) of Mn²⁺, Fe³⁺, Co²⁺, Ni²⁺, Cu²⁺, Zn²⁺, Cd²⁺, Hg²⁺, Na⁺, K⁺, Mg²⁺, Ca²⁺, Al³⁺, Ga³⁺, In³⁺, Pb²⁺ and Cr³⁺ with an excitation of 410 nm in bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)). Inset: Picture of the fluorescence corresponded with **1** and **1**-Al³⁺ ($\lambda_{ex} = 365$ nm)

To further study the fluorescence sensing behavior of **1**, a quantitative investigation of the binding affinity of **1** with Al^{3+} was studied by fluorescence titration (Fig. 2). When the sensor **1** was titrated with Al^{3+} , the fluorescence intensity increased up to 18 equiv and then no change was observed, which might be due to a combinational effect of the inhibition of C=N isomerization and the activation of chelation enhanced fluorescence (CHEF).^{45,74-76} Imines are poorly fluorescent in part due to isomerization of the C=N

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double bond in the excited state. The C=N isomerization might be inhibited upon the complexation with certain metal ions. Therefore, the stable chelation of **1** with AI^{3+} ions made the C=N isomerization of **1** inhibited, resulting in fluorescence enhancement. Moreover, the complexation of **1** with AI^{3+} ion induced rigidity in the resulting molecule and tended to produce a chelation-enhanced fluorescence detection. The fluorescence quantum yields of sensor **1** without and with AI^{3+} were 0.0062 and 0.4240, respectively.⁷⁷



Fig. 2. Fluorescent titration spectra of sensor **1** (10 μ M) in the presence of different concentrations of Al³⁺ in bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)). Inset: the fluorescence at 460 nm ($\lambda_{ex} = 410$ nm) of **1** as a function of the Al³⁺ concentration.

The binding properties of **1** with Al^{3+} were further studied by UV-vis titration experiments (Fig. 3). On the treatment with Al^{3+} to the solution of **1**, the absorption peak

at 425 nm gradually decreased while the peak at 352 nm increased. Two clear isosbestic points were observed at 395 and 468 nm, indicative of a clean conversion of **1** into the **1**- Al^{3+} complex.



Fig. 3. Absorption spectral changes of **1** (10 μ M) in the presence of different concentrations of Al³⁺ ions (5, 10, 15, 20, 25, 30, 35, 40, 50, 60 and 70 μ M) in bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)). Inset: absorbance of **1** at 425 nm as a function of the Al³⁺ concentration.

The Job plot for the binding between **1** and Al^{3+} exhibited a 2:1 stoichiometry (Fig. 4).⁷⁸ Also, the negative-ion mass spectrum for the reaction of **1** with Al^{3+} showed the formation of the $[2 \cdot 1(-4H^+)+Al^{3+}]^-$ complex [m/z: 647.27; calcd, 647.24] (Fig. S2). From the fluorescence titration, the association constant was calculated to be 3.1 x 10⁸ M⁻¹ from

Li's equations (see the supplementary materials and Fig. S3).⁷⁹ This value is one of the highest binding affinities among such simple Schiff bases for Al³⁺ in buffer solution, indicating that the carboxylic group showed the most effective binding property among other functional groups such as imine, amine, quinolone, etc (Table S1).



Fig. 4. Job plot of sensor 1 and Al^{3+} , where the intensity at 460 nm was plotted against the mole fraction of aluminum ions. The total concentration of aluminum ions with sensor 1 was 10 μ M in bis-tris buffer (10 mM, pH 7.0, water:CH₃OH=1:1 (v/v)).

By using the above-mentioned fluorescence titration results, the detection limit of **1** for the analysis of Al^{3+} ions on the basis of $3\sigma/K$ was calculated to be 2.9×10^{-7} M (Fig. S4).⁸³ The detection limit of **1** was quite lower than the WHO limit of Al^{3+} (7.41 × 10^{-6} M) acceptable in drinking water, which means that sensor **1** is a powerful tool for the detection of Al^{3+} in aqueous solution. The analytical performance characteristics of the proposed Al³⁺-sensitive chemosensor were investigated. Fig. 5 shows a calibration curve obtained from the plot of fluorescence intensity with the added Al³⁺ concentration. The curve equation was $I = 0.08063C_{Al} + 13.924$ that C_{Al} unit was nM (R² = 0.996). The linear range of quantitative detection for aluminum ion was determined to be 40-1000 nM with a detection limit of 290 nM (3 σ). The detection limit appeared within the range of quantitative detection of aluminum ion. Therefore, the sensor **1** was not only selective towards aluminum (III) ion but also possessed a linear response range with a low limit of detection.



Fig. 5. Calibration curve based on fluorescence intensity with the added Al^{3+} concentration.

The ¹H NMR titration experiments of sensor $\mathbf{1}$ with Al^{3+} ion were studied to further

examine the binding mode (Fig. 6). The protons of carboxyl and hydroxyl groups did not appear maybe due to the intra- or inter-molecular hydrogen bonds. Upon addition of 0.5 equiv of Al^{3+} , the H_a proton of the imine moiety and the H_c and H_d protons of the ethyl groups were shifted to downfield by 0.32, 0.10, and 0.06 ppm, respectively. Also, the protons of the two benzene rings were more complicated and shifted to downfield except for H_b which was shifted to upfield by 0.15 ppm. Further addition of Al^{3+} showed no spectral change. These results suggested that the two oxygen atoms of the carboxyl and hydroxyl groups and the nitrogen atom of the imine moiety might be involved in Al^{3+} coordination. Based on the ¹H NMR titration, Job plot, and ESI-mass spectrometry analysis, we proposed the structure of a 2:1 complex of **1** and Al^{3+} , as shown in Scheme 2.



Fig. 6. ¹H NMR titrations of **1** with Al(NO₃)₃: (I) **1**; (II) **1** with 0.2 equiv of Al³⁺; (III) **1** with 0.5 equiv of Al³⁺; (IV) **1** with 1 equiv of Al³⁺.



Scheme 2. Proposed structure of a 2:1 complex of 1 and Al³⁺.

To further check the practical applicability of sensor **1** as AI^{3+} selective fluorescent sensor, we carried out competition experiments. When **1** was treated with 18 equiv of AI^{3+} in the presence of the same concentration of other metal ions, the fluorescence enhancement caused by AI^{3+} was retained with Mn^{2+} , Co^{2+} , Ni^{2+} , Zn^{2+} , Cd^{2+} , Hg^{2+} , Na^+ , K^+ , Mg^{2+} , Ca^{2+} , Ga^{3+} , In^{3+} , and Pb^{2+} (Fig. 7). Instead, Fe^{3+} , Cu^{2+} and Cr^{3+} inhibited about 80%, 80% and 100 % of the interaction between **1** and AI^{3+} , which are well known as the strong quenchers of fluorescence.^{84,85} Thus, sensor **1** can be used as a selective fluorescent sensor for AI^{3+} in the presence of most competing metal ions.





Fig. 7. (a) Competitive selectivity of **1** toward Al^{3+} in the presence of other metal ions (18 equiv) with an excitation of 410 nm in bis-tris buffer (10 mM, water:CH₃OH=1:1 (v/v)).

(b) The fluorescence images of **1** upon addition of Al^{3+} (18 equiv) in the absence and presence of various metal ions (18 equiv) with an excitation of 365 nm in bis-tris buffer (10 mM, water:CH₃OH=1:1 (v/v)).

For biological applications, the pH dependence of the $Al^{3+}-2\cdot 1$ complex was examined in bis-tris buffer (10 mM, water:CH₃OH=1:1 (v/v)) solution (Fig. 8). The fluorescence intensity of the $Al^{3+}-2\cdot 1$ complex displayed a significant response between pH 5 and 10, which includes the environmentally relevant range of pH 7.0-8.4.⁸⁶ These results indicated that Al^{3+} could be clearly detected by the fluorescence spectra measurement using **1** over the environmentally relevant pH range (pH 7.0-8.4).



Fig. 8. Fluorescence intensity (at 460 nm) of $Al^{3+}-2\cdot 1$ complex at different pH values (3-12) in bis-tris buffer (10 mM, water:CH₃OH=1:1 (v/v)).

3.2. Application for cell imaging

Further experiments were conducted to test whether intracellular Al^{3+} could be monitored by fluorimetry. Adult human dermal fibroblasts were first incubated with various concentrations of Al^{3+} (0, 30, 50, 100, 150 and 200 µM) for 4 h and then exposed to 1 for 30 min before imaging, so that 1 could permeate easily through the living cells without any harm. The fibroblasts that were cultured with both Al^{3+} and 1 exhibited fluorescence (Fig. 9), whereas the cells cultured without Al^{3+} or 1 did not exhibit fluorescence. The intensity and region of the fluorescence within the cell containing 1 gradually increased with an increase in the Al^{3+} concentration form 0 to 200 µM and the fluorescence intensity of the cells persisted even after 5 h from exposure to 1. Moreover, the biocompatibility of 1 was also examined with the living cells (Fig. S5). All the fibroblasts were still alive for 24 h. These results showed that the new sensor 1 was efficient to image Al^{3+} in cells, and could be useful in the determination of the exposure level of cells to Al^{3+} .



Fig. 9. Fluorescence images of fibroblasts cultured with Al^{3+} and **1**. Cells were exposed to 0 (A and G), 30 (B and H), 50 (C and I), 100 (D and J), 150 (E and K) and 200 μ M (F and L) $Al(NO_3)_3$ for 4 h and then later with **1** for 30 min. The images (A-F) were observed with the light microscope and the images (G-L) were taken with a fluorescence microscope. The scale bar is 50 μ m.

3.3. Theoretical calculations for 1 and Al³⁺-2·1 complex

To gain an insight into fluorescent sensing mechanism for $Al^{3+}-2\cdot 1$ complex, timedependent density functional theory (TD-DFT) calculations were performed at the optimized geometries (S₀) of **1** and $Al^{3+}-2\cdot 1$ complex (Fig. S6). The optimized geometries were calculated based on Job plot, ESI-mass and ¹H NMR titration. In case of **1**, the main molecular orbital (MO) contribution of the first lowest excited state was determined for HOMO \rightarrow LUMO transition (394.61 nm, Fig. S7), which indicated intramolecular charge transfer (ICT) transition from the julolidine moiety to benzoic acid moiety. For Al³⁺-2·1 complex, the first lowest excited state was determined for HOMO \rightarrow LUMO transition (499.43 nm, Fig. S8), which indicated the forbidden transition (oscillator strength: 0.0045). The main molecular orbital (MO) contributions of the fifth lowest excited states were determined for HOMO \rightarrow LUMO+1 and HOMO-1 \rightarrow LUMO+1 transitions (367.42 nm, Fig. S8), which showed allowed transition and was similar to intramolecular charge transfer (ICT) transition of 1. There were no obvious changes in the electronic transitions between 1 and Al³⁺-2·1 complex. Only, hypochromic shift (394.61 to 367.42 nm) was observed upon chelating of 1 with Al³⁺. These results suggested that the sensing mechanism of 1 toward Al³⁺ was originated by the rigid structure of Al³⁺-2·1 complex, which might inhibit the non-radiative process such as C=N isomerization. Thus, the CHEF effect and the inhibition of C=N isomerization plays an important role in fluorescence enhancement.^{87,88}

4. Conclusion

We have developed a simple, cost-effective, fluorescent aminobenzoic acid-based chemosensor **1** for selective determination of Al^{3+} in aqueous solution. Sensor **1** displayed a remarkable fluorescence enhancement in the presence of Al^{3+} ion and can be successfully applied to living cells for detecting Al^{3+} . The large enhancement of fluorescence was explained by the inhibition of C=N isomerization and the activation of chelation enhanced fluorescence (CHEF), which were explained by theoretical calculations. On the basis of Job plot, ¹H NMR titration and ESI-mass spectrometry

analysis, the formation of 2:1 (1:M) complex was proposed. Moreover, the sensor 1 showed one of the highest binding constants (3.1×10^8) among such simple Schiff bases in aqueous solution and displayed a linear response range with a low limit of detection (290 nM) for the analytical performance of Al³⁺ ion.

Acknowledgements

Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (NRF-2014R1A2A1A11051794 and NRF-2015R1A2A2A09001301) are gratefully acknowledged.

Supplementary data

Supplementary materials associated with this article can be found, in the online version.

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Graphical abstract



A highly selective and sensitive fluorescent chemosensor exhibited an enhanced fluorescence in the presence of Al^{3+} and in living cells.