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The phosphaguanylation reaction of phosphines with carbodiimides catalyzed by half-sandwich yttrium tris(trimethylsilylmethyl) ate complex is achieved for the first time to prepare efficiently phosphaguanidines. The catalyst system of yttrium ate complex displays better catalytic activity than those neutral yttrium complexes.

Introduction

Catalytic guanylation reaction of amines with carbodiimides (CGAC reaction) has become a hot area because it provides a straightforward and atom-economical route to prepare guanidines (RN=C(NR'R")NHR).1-3 In contrast, catalytic phosphaguanylation reaction of phosphines with carbodiimides (hereafter as CPPC reaction), which is also known as catalytic addition of phosphines to carbodiimides or hydrophosphination of carbodiimides has received much less attention, although this reaction provides an atom-economical synthesis of phosphaguanidines $(R'_2PC(=NR)NHR).$ These phosphaguanidines, as analogues of guanidines, are widely used as building blocks of ancillary ligands for many transition species.4,5 metal Initially, this catalyst-free direct phosphaguanylation reaction of diphenylphosphine with an N,N'-diaryl substituted carbodiimide was reported to afford the phosphaguanidine in a low yield under rather harsh conditions.⁶ Recently, the direct phosphaguanylation reaction of the active hydrogen heptaphosphide dianion $[HP_7]^{2-}$ with RN=C=NR (R = 2,6-diisopropylphenyl (Dipp), iPr, and Cy) was reported to afford the functionalized cluster anions $[P_7C(=NR)NHR]^{2.7}$ Verv recently. the stoichiometric NaH-mediated phosphaguanylation of phosphine boranes with carbodiimides provided an alternative synthesis of phosphaguanidines by

removing the borane using 1,4-diazabicyclo[2.2.2]octane (DABCO).⁸ The phosphaguanylation reaction of phosphines with the less electrophilic N,N'-dialkyl or N-alky-N'-aryl carbodiimides cannot be achieved without suitable catalysts, therefore, the search and design of suitable catalyst systems for the CPPC reaction are in high demand.



Fig. 1 The reported catalyst precursors for the CPPC reaction

In 2006, Hou *et al.* reported the first CPPC reaction catalyzed via alkali metal bis(trimethylsilyl)amides [MN(SiMe₃)₂] (1: M = Li, Na, K) (**Fig. 1** for the structure of complexes 1-8).⁹ The heavier group 2 catalysts 2 and 3 were tested by Hill and Barrett. The homoleptic alkaline earth (Ca, Sr and Ba) amides 2 displayed higher activity than β -diketiminato calcium complex 3.¹⁰ The triamidoamine-supported zirconium complex 4 showed the medium activity for the CPPC reaction.¹¹ In the attempt to test the rare-earth metals, the half-sandwich lanthanum aminobenzyl complex 5-La was found by

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Hou et al. to display the highest activity among the rare-earth metals because of the largest ionic radius (1.032 Å for La^{3+}), however, the corresponding yttrium complexes 5-Y and 6-Y $(0.900 \text{ Å for } \text{Y}^{3+})$ showed the obviously lower activity.¹² Schmidt *et al.* reported the CPPC reaction using homoleptic α metalated N,N-dimethylbenzylamine lanthanum complex 7-La, in which three N,N-dimethylbenzylamine ligands were released via protonolysis with phosphines to yield the active La-P species.¹³ All these reported catalyst systems involved only the neutral metal amides or alkyl complexes. We reported the synthesis and structural characterization of half-sandwich rareearth metal tris(trimethylsilylmethyl) ate complexes bearing one 1-phenyl-2,3,4,5-tetrapropylcyclopentadienyl ligand and catalytic application in CGAC reaction to construct guanidines.¹⁴ We envision that these ate complexes, which might have the potential advantages of the neutral metal amides and alkyl complexes, could serve as good catalyst precursors for the CPPC reaction to construct phosphaguanidines. Here we report these results.





 0.20 mmol.^{b} determined by ³¹P NMR.

Results and discussion

Condition screening

As a control experiment, barely mixing of diphenylphosphine and N,N-diisopropylcarbodiimide (DIC) in the absence of catalysts resulted in no phosphaguanidine product even at 140 °C for 24 h in C₆D₅Cl (Table 1, entry 1). However, when a small amount of **8-Y** was added, the corresponding

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phosphaguanidine 9a can be detected by ³¹P NMR (Table 1, entries 2-6). Both polar solvent (THF) and non-polar solvents (benzene and toluene) were tested, and it revealed that THF was the best option probably owing to the excellent solubility for the rare-earth complex (Table 1, entries 2-4). Low yields were found when the reaction was carried out at room temperature (Table 1, entry 5) or under the low catalyst loading (1 mol%) (Table 1, entry 6). All of these complexes with different metal centers (Er, Tm and Lu) had catalytic activities (Table 1, entries 7-9). The ate complex 8-Y showed the better catalytic activity than the neutral yttrium complexes, such as the half-sandwich constrained-geometry complex 6-Y, mono-Cp complex 10-Y, and tris(trimethylsilylmethyl) complex Y(CH₂TMS)₃(thf)₂. The comparison clearly shows the combining significance of cyclopentadiene-based ligands and ate complex (Table 1, entries 4 and 11-13).

Substrate scope

The anionic rare-earth complex 8-Y was chosen for the CPPC reaction and representative results were summarized in Table 2. Various carbodiimides could be applied to the present conditions. When symmetric N,N'-dialkylcarbodiimides, such as DIC, N,N'-dicyclohexylcarbodiimide (DCC) were utilized, the corresponding phosphaguanidines (9a, 9b, 9g, 9h, and 9km) were obtained in excellent yields under the relatively milder conditions. In case of *N*-ethyl-*N'-tert*-butylcarbodiimides (9c), this CPPC reaction need higher catalyst loadings and long reaction time probably due to the steric hindrance of tert-butyl group. Unsymmetric N-aryl-N'-alkyl carbodiimides, such as Nphenyl-N'-cyclopentyl, N-phenyl-N'-cyclohexyl, N-p-tol-N'cyclohexyl, N-phenyl-N'-cycloheptyl, N-phenyl-N'-cyclooctyl and N-phenyl-N'-CH2-cyclohexyl carbodiimides could also be applied to construct phosphaguanidines (9d-f, 9i, 9j and 9n-r), but higher catalyst loadings and long reaction time were required owing to the low electrophilicity of N-alky-N'-aryl carbodiimides for accepting nucleophilic attack of the phosphine. The phosphaguanylation product originated from such carbodiimides N,N'-bis(2,6bulkv as diisopropylphenyl)carbodiimide and diphenylphosphine was not observed, probably owing to the steric hindrance at nitrogen atom, which prevents the coordination of the carbodiimide to the rare-earth metal center. Most diarylphosphines could be utilized in this reaction to afford the corresponding products (9a-r) in good vields. The dialkylphosphine, such as di-isobutylphosphine could not be applied, probably due to the decreased acidity compared to diarylphosphines. When the primary phosphine PhPH2 was treated with one equivalent of DIC. both monoand double-addition products $PhP{iPrN=C(NHiPr)}_{2}$ could be *i*PrN=C(PHPh)(NH*i*Pr), detected by ³¹P NMR (mono-addition product: -62.1 ppm double-addition product: -34.9 ppm). When the amount of DIC was increased to 2 equivalents, the double-addition product 9s can be obtained exclusively in 93% yield.

Although phosphaguanidines could, in principle, have eight possible isomers $E_{syn}(\alpha,\beta)$, $E_{anti}(\alpha,\beta)$, $Z_{syn}(\alpha,\beta)$, $Z_{anti}(\alpha,\beta)$,¹⁵ all of the ¹H, ¹³C and ³¹P NMR spectra of **9a-r** indicated only one isomer in solution. X-ray single crystal diffraction

analysis of phosphaguanidine **9i** reveals it an E_{syn} -(α) isomer with the phenyl group placed on the C=N double bond in the solid state (**Fig. 2**).

Table 2 Substrate scope a,b



^a Conditions: phosphine, 0.53 mmol; carbodiimide, 0.50 mmol, THF, 5 ml, unless otherwise noted. ^b Isolated yield. ^c 6 mol % catalyst, 24 h. ^d carbodiimide, 1 mmol.



Fig. 2 ORTEP drawing of 9i with 30% ellipsoids. Hydrogen atoms, except that on nitrogen atom N2, are omitted for clarity.

To gain further insight into the mechanism of the CPPC reaction, we conducted the stoichiometric reaction by in situ NMR monitoring. The original spectra of 8-Y in THF-d₈ without TMS internal standard was illustrated in Fig. 3a. The double peaks of -1.19 ppm and single peak of -0.13 ppm were ascribed to CH₂ and CH₃ of CH₂SiMe₃, respectively. When 8-Y was treated with 3 equivalents of DIC, the ¹H NMR spectra showed the peaks of DIC remained unchanged after 12 h at room temperature (Fig. 3b). This result shows that the insertion of DIC into the Y-CH2SiMe3 alkyl bond does not occur at room temperature. However, when 8-Y was treated with 3 equivalents of Ph2PH, the reaction mixture turned to light yellow immediately. After 12 h at room temperature, the ¹H NMR spectra showed that the integral of the CH₂SiMe₃ groups decreased and the peak of SiMe₄ was observed in 0 ppm (Fig. 3c). And after 24 h at room temperature, the ¹H NMR spectra showed that the peaks of CH₂SiMe₃ almost disappeared (Fig. 3d). The formation of SiMe₄ resulted from the acid-base reaction between phosphine P-H bond and Y-CH₂SiMe₃ alkyl bond. The comparative results show that the acid-base reaction between P-H bond and Y-CH₂SiMe₃ alkyl bond is faster than the insertion of the carbodiimide into the Y-CH₂SiMe₃ alkyl bond at room temperature. This result also indicated that all of the three CH₂SiMe₃ groups in 8-Y were involved in the acidbase reaction. Subsequent addition of 3 equivalents of DIC led apparently to the doublet peak in 1.16 ppm and multiple peak in 3.47 ppm for CH₃ and CH of DIC in the ¹H NMR spectrum, respectively (Fig. 3e). After heating this mixture at 80 °C for 1 h, no obvious change in ¹H NMR spectrum was observed, which indicated that DIC did not insert into the Y-P bond. Interestingly, when another 3 equivalents of Ph₂PH were added into the above solution, all the peaks belonged to iPr moiety of DIC changed within 15 min (Fig. 3f). The peaks of CH₃ and CH of DIC shifted to 0.93 ppm and 4.04 ppm, which were in consistent with the corresponding chemical shifts of iPr of phosphaguanidine 9a. After an excess of Ph₂PH and DIC in 1:1 ratio was added, the catalytic formation of phosphaguanidine 9a was observed obviously.



Fig. 3 In situ NMR spectra of stoichiometric reaction. (a) **8-Y**; (b) **8-Y** + 3 DIC at room temperature for 12 h; (c) **8-Y** + 3 Ph₂PH at room temperature for 12 h; (d) **8-Y** + 3 Ph₂PH at room temperature for 24 h; (e) **8-Y** + 3 Ph₂PH + 3 DIC at 80 °C for 1 h; (f) (**8-Y** + 3 Ph₂PH + 3 DIC) + 3 Ph₂PH at room temperature for 15 min.



Scheme 1 A plausible mechanism for the CPPC reaction.

A possible reaction mechanism for the CPPC reaction was proposed in Scheme 1. The formation of the active phosphide species **A** might undergo two possible pathways: i) The acidbase reaction between a phosphine and **8-Y**, and ii) The insertion of the carbodiimide into the Y-alkyl bond followed by protonolysis yielding an amidine and Y-phosphido complex **A**. The insertion pathway can be excluded because the acid-base reaction is faster than the insertion reaction according to the experimental results as shown in **Fig. 3***b* and **3***c*. The coordination of carbodiimides took place to form the coordinate species **B**. The fast coordination and disassociation of DIC to the yttrium center in THF was probably out of the NMR time scale, so the obvious chemical shifts of DIC were not observed, which was in consistent with the observation of **Fig. 3***e*. Then, the coordination of phosphine might yield the adduct **C**. The intramolecular nucleophilic attack of Y–P bond on the central carbon of the coordinate carbodiimide and the protonolysis with R_2P –H bond released the phosphaguanidine product **9** and regenerated **A** to finish the catalytic cycle.

Conclusions

In summary, we have reported a half-sandwich yttrium tris(trimethylsilylmethyl) ate complex catalyzed phosphaguanylation reaction of phosphines with carbodiimides to prepare efficiently phosphaguanidines. The present yttrium catalyst system is easier to prepare and cheaper than those reported neutral lanthanum complexes. It displays the best catalytic activity among these known yttrium complexes, probably owing to the cooperative effect between the anionic yttrium core and cationic lithium moiety. Further investigation on the mechanism and catalytic application is underway.

Experimental section

General considerations

All reagents were purchased from commercial sources and used without treatment, unless otherwise indicated. All reactions were conducted under a slightly positive pressure of dry nitrogen using standard Schlenk line techniques or under a nitrogen atmosphere in a Vigor (SG 1200/750TS-F) glovebox. The nitrogen in the glove box was constantly circulated through a copper/molecular sieves catalyst unit. The oxygen and moisture concentrations in the glovebox atmosphere were monitored by an O2/H2O Combi-Analyzer to ensure both were always below 1 ppm. Unless otherwise noted, all starting materials were commercially available and were used without further purification. Solvents were purified by an Mbraun SPS-800 Solvent Purification System and dried over fresh Na chips in the glovebox. Organometallic samples for NMR spectroscopic measurements were prepared in the glovebox by use of J. Young valve NMR tubes (Wilmad 528-JY). ¹H, ¹³C NMR and ³¹P spectra were recorded on Bruker-400 spectrometer (FT, 400 MHz for ¹H; 100MHz for ¹³C and 160 MHz for ³¹P) at room temperature. Highresolution mass spectra (HRMS) were recorded on a Bruker Apex IV FTMS mass spectrometer using ESI (electrospray ionization) source.

NMR tube reaction

In a glovebox, a J. Young valve NMR tube was charged with **8**-**Y** (5.7 mg, 0.006 mmol), THF (0.5 mL), diphenylphosphine (43 mg, 0.23 mmol), and *N*,*N'*-diisopropylcarbodiimide (25 mg, 0.20 mmol). The tube was taken out of the glovebox and then heated at 80 °C in an oil bath for 1 h. Formation of the phosphaguanidine was monitored by ³¹P NMR spectroscopy.

Preparative scale reaction

In a glovebox, the mixture of THF solution (1 mL) of **8-Y** (14.4 mg, 0.015 mmol) and THF solution (2 mL) of diphenylphosphine (99 mg, 0.53 mmol) was added to a Schlenk tube. Then *N*,*N*'-diisopropylcarbodiimide (63 mg, 0.5 mmol) was added to the above reaction mixture. The Schlenk tube was taken outside the glovebox, and the mixture was stirred at 80 °C for 1 h. After the stirring, the solvent was removed under reduced pressure. The residue was extracted with hexane and filtered to give a clean solution. After removal of the solvent under vacuum, the residue was recrystallized in hexane to provide the phosphaguanidine **9a**. Compounds **9b-m** were prepared in a similar procedure.

*i***PrN=C(PPh₂)(NH***i***Pr) (9a).⁹ Colorless solid, isolated yield 97% (151 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): \delta = 0.96 (d, J = 6.4 Hz, 6H; CH(CH₃)₂), 1.26 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 3.66 (d, ³J = 6.3 Hz, 1H; NH), 4.29-4.47 (m, 2H; CH), 7.01-7.08 (m, 6H; C₆H₅), 7.44-7.49 (m, 4H; C₆H₅) ppm; ¹H NMR (400 MHz, THF-d₈, Me₄Si): \delta = 0.92 (d, J = 2.7 Hz, 6H; CH(CH₃)₂), 0.94 (d, J = 3.0 Hz, 6H; CH(CH₃)₂), 3.52 (d, ³J = 6.4 Hz, 1H; NH), 3.98-4.08 (m, 2H; CH), 7.34-7.40 (m, 10H; C₆H₅) ppm.**

CyN=C(PPh₂)(NHCy) (9b).⁹ Colorless solid, isolated yield 95% (186 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.95-1.92 (m, 20H; Cy), 3.82 (d, ³*J* = 6.8 Hz, 1H; NH), 4.07-4.19 (m, 2H; CH), 7.02-7.09 (m, 6H; C₆H₅), 7.49-7.52 (m, 4H; C₆H₅) ppm.

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*t***BuN=C(PPh₂)(NHEt) (9c)**.⁹ Colorless solid, isolated yield 83% (130 mg);¹H NMR (400 MHz, C_6D_6 , Me_4Si): $\delta = 1.31$ (t, J = 7.2 Hz, 3H; CH_2CH_3), 1.36 (s, 9H; $C(CH_3)_3$), 3.77-3.83 (m, 3H, NH and CH_2CH_3), 7.01-7.06 (m, 6H; C_6H_5), 7.45-7.49 (m, 4H; C_6H_5) ppm.

PhN=C(PPh₂)(NHCy) (9d).⁹ Colorless solid, isolated yield 85% (164 mg); ¹H NMR (400 MHz, C_6D_6 , Me_4Si): $\delta = 0.88$ -1.94(m, 10H; Cy), 4.22-4.24(m, 1H; CH), 4.38 (d, ³*J* = 7.0 Hz, 1H; NH), 6.84 (t, *J* = 7.6 Hz, 1H; C_6H_5), 7.03-7.11 (m, 10H; C_6H_5), 7.44 (t, *J* = 6.3 Hz, 4H; C_6H_5) ppm.

PhN=C(PPh₂){NH-(CH₂)₅} (9e). Colorless solid, isolated yield 83% (155 mg);.¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.23-1.87 (m, 8H; -(CH₂)₄-), 4.38 (d, ³J = 6.3 Hz, 1H; NH), 4.55-4.61 (m, 1H; CH), 6.83-6.87 (m, 1H; C₆H₅), 7.00-7.12 (m, 10H; C₆H₅), 7.38-7.43 (m, 4H; C₆H₅) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 23.8, 33.2, 53.9, 122.3, 123.3 (d, ⁴J_{pc} = 1.8 Hz), 128.6, 128.9 (d, ³J_{pc} = 6.8 Hz), 129.4, 134.4 (d, ²J_{pc} = 20.1 Hz), 135.3 (d, ¹J_{pc} = 15.5 Hz), 151.9 (d, ³J_{pc} = 12.1 Hz), 156.8 (d, ¹J_{pc} = 36.7 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -14.6 ppm; HRMS: *m*/*z* calcd for C₂₄H₂₆N₂P: 373.1828 [M + H] ⁺; found: 373.1817.

p-tolN=C(PPh₂)(NHCy) (9f). Colorless solid, isolated yield 85% (170 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.96-1.95 (m, 10H; Cy), 2.07 (s, 3H; Me), 4.25-4.26 (m, 1H; CH), 4.35 (d, ³*J* = 6.6 Hz, 1H; NH), 6.89-7.05 (m, 10H; C₆H₅), 7.43-7.45 (m, 4H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 20.9, 24.6, 26.0, 32.6, 49.8, 123.1 (d, ⁴*J*_{pc} = 1.6 Hz), 128.9 (d, ³*J*_{pc} = 6.8 Hz), 129.2, 129.4, 131.1, 134.4 (d, ²*J*_{pc} = 20.0 Hz), 135.5 (d, ¹*J*_{pc} = 15.8 Hz), 149.4 (d, ³*J*_{pc} = 12.5 Hz), 156.1 (d, ¹*J*_{pc} = 36.3 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -14.9 ppm; HRMS: *m/z* calcd for C₂₆H₃₀N₂P: 401.2141 [M + H] ⁺; found: 401.2144.

*i***PrN=C{P(C₆H₄Me-2)₂}(NH***i***Pr) (9g)**.⁹ Colorless solid, isolated yield 88% (150 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.94 (d, J = 6.4 Hz, 6H; CH(CH₃)₂), 1.33 (d, J = 5.4 Hz, 6H; CH(CH₃)₂), 2.41 (s, 6H; Me), 3.72 (d, ³J = 6.4 Hz, 1H; NH), 4.34-4.47 (m, 2H; CH), 6.92-7.06 (m, 8H; C₆H₄) ppm.

CyN=C{P(C₆H₄Me-2)₂}(NHCy) (9h). Colorless solid, isolated yield 87% (183 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.89-1.94 (m, 20H; Cy), 2.45 (s, 6H; Me), 3.90 (d, ³J = 7.0 Hz, 1H; NH), 4.08-4.26 (m, 2H, CH), 6.98-7.07 (m, 8H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 21.3 (d, ³J_{pc} = 21.7 Hz), 24.7, 25.3, 26.1, 26.4, 32.7, 35.9, 49.1, 60.9 (d, ³J_{pc} = 35.2 Hz), 126.8, 129.6, 130.6 (d, ³J_{pc} = 4.5 Hz), 143.0 (d, ²J_{pc} = 25.9 Hz), 151.8 (d, ¹J_{pc} = 30.2 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -32.9 ppm; HRMS: *m/z* calcd for C₂₇H₃₈N₂P: 421.2767 [M + H]⁺; found: 421.2754.

PhN=C{P(C₆H₄Me-4)₂}{NH-(CH₂)₅} (9i). Colorless solid, isolated yield 84% (168 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.28-1.90 (m, 8H; -(CH₂)₄-), 1.98 (s, 6H; Me), 4.52 (d, ³J = 6.4 Hz, 1H; NH), 4.61-4.65 (m, 1H; CH), 6.84-6.86 (m, 1H; C₆H₅), 6.89 (d, J = 7.6 Hz, 4H; C₆H₄), 7.12 (d, J = 4.7 Hz, 4H; C₆H₅), 7.39 (t, J = 7.6 Hz, 4H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 21.1, 23.8, 33.3, 53.8, 122.2, 123.3 (d, ⁴J_{pc} = 1.8 Hz), 128.5, 129.8 (d, ³J_{pc} = 7.0 Hz), 132.1 (d, ¹J_{pc} = 14.3 Hz), 134.5 (d, ²J_{pc} = 20.3 Hz), 139.4, 152.1 (d, ³J_{pc} = 11.9

Hz), 157.4 (d, ${}^{1}J_{pc} = 37.1$ Hz) ppm; ${}^{31}P{}^{1}H$ NMR (160 MHz, C₆D₆): $\delta = -16.2$ ppm; HRMS: *m/z* calcd for C₂₆H₃₀N₂P: 401.2141 [M + H] ⁺; found: 401.2129. Single crystals of **9i** suitable for X-ray analysis were grown in THF/hexane for 1 day at room temperature.

PhN=C{P(C₆H₄Me-4)₂}(NHCy) (9j). Colorless solid, isolated yield 81% (168 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.88-1.42 (m, 10H; Cy), 2.00 (s, 6H; Me), 4.26-4.30 (m, 1H; CH), 4.50 (d, ³J = 7.2 Hz, 1H; NH), 6.83-6.88(m, 1H; C₆H₅), 6.91 (d, J = 7.6 Hz, 4H; C₆H₄), 7.12 (d, J = 4.5 Hz, 4H; C₆H₅), 7.42 (t, J = 7.6 Hz, 4H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 21.1, 24.8, 26.0, 32.7, 49.9, 122.1, 123.4 (d, ⁴J_{pc} = 1.7 Hz), 128.5, 129.8 (d, ³J_{pc} = 7.1 Hz), 132.1 (d, ¹J_{pc} = 14.4 Hz), 134.5 (d, ²J_{pc} = 20.3 Hz), 139.4, 152.1 (d, ³J_{pc} = 12.0 Hz), 156.8 (d, ¹J_{pc} = 36.7 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -15.9 ppm; HRMS: *m/z* calcd for C₂₇H₃₂N₂P: 415.2298 [M + H]⁺; found: 415.2287.

*i*PrN=C{P(C₆H₄Me-4)₂}(NH*i*Pr) (9k).⁹ Colorless solid, isolated yield 95% (162 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ =1.00 (d, J = 6.4 Hz, 6H; CH(CH₃)₂), 1.28 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 2.02 (s, 6H; Me), 3.79 (d, ³J = 6.6 Hz, 1H; NH), 4.34-4.51 (m, 2H; CH), 6.93 (d, J = 7.5 Hz, 4H; C₆H₄), 7.44 (t, J = 7.7 Hz, 4H; C₆H₄) ppm.

CyN=C{P(C₆H₄Me-4)₂}(NHCy) (9). Colorless solid, isolated yield 91% (191 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.93-1.98 (m, 20H; Cy), 2.02 (s, 6H; Me), 3.94 (d, ³J = 7.0 Hz, 1H; NH), 4.13-4.24 (m, 2H; CH), 6.96 (d, J = 7.6 Hz, 4H; C₆H₄), 7.49 (t, J = 7.6 Hz, 4H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 21.1, 24.7, 25.3, 26.2, 26.4, 32.7, 35.8, 49.2, 60.2 (d, ³J_{pc} = 33.6 Hz), 129.8 (d, ³J_{pc} = 7.0 Hz), 132.5 (d, ¹J_{pc} = 13.2 Hz), 134.4 (d, ²J_{pc} = 19.7 Hz), 139.2, 152.8 (d, ¹J_{pc} = 31.8 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -19.5 ppm; HRMS: *m/z* calcd for C₂₇H₃₈N₂P: 421.2767 [M + H] ⁺; found: 421.2760.

*i*PrN=C{P(C₆H₃Me₂-3,5)₂}(NH*i*Pr) (9m).⁹ Colorless solid, isolated yield 85% (157 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.03 (d, J = 6.4 Hz, 6H; CH(CH₃)₂), 1.32 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 2.03 (s, 12H; Me), 3.91 (d, ³J = 6.6 Hz, 1H; NH), 4.37-4.54 (m, 2H, CH), 6.75 (s, 2H; C₆H₃), 7.28 (d, J= 8.0 Hz, 4H; C₆H₃) ppm.

PhN=C{P(C₆H₄Me-4)₂}{NH-(CH₂)₇} (9n). Colorless solid, isolated yield 87% (186 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.23-1.97 (m, 12H; -(CH₂)₆-), 2.01 (s, 6H; Me), 4.44-4.45 (m, 1H; CH), 4.55 (d, ³J = 7.2 Hz, 1H; NH), 6.82-6.86(m, 1H; C₆H₅), 6.92 (d, J = 7.7 Hz, 4H; C₆H₄),7.09 (d, J = 4.6 Hz, 4H; C₆H₅), 7.41 (t, J = 7.7 Hz, 4H; C₆H₄) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 21.1, 24.3, 28.1, 34.5, 52.1, 122.0, 123.3 (d, ⁴J_{pc} = 1.7 Hz), 128.4, 129.8 (d, ³J_{pc} = 7.0 Hz), 132.0 (d, ¹J_{pc} = 14.3 Hz), 134.4 (d, ²J_{pc} = 20.3 Hz), 139.4, 152.1 (d, ³J_{pc} = 12.0 Hz), 156.6 (d, ¹J_{pc} = 36.7 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -15.9 ppm; HRMS: *m*/z calcd for C₂₈H₃₄N₂P: 429.2454 [M + H]⁺; found: 429.2444.

PhN=C(PPh₂){NH-(CH₂)₇} (90). Colorless solid, isolated yield 93% (186 mg);.¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.21-1.90 (m, 12H; -(CH₂)₆-), 4.43 (br, 2H; CH and NH), 6.82-6.86 (m, 1H; C₆H₅), 7.03-7.10 (m, 10H; C₆H₅), 7.42-7.45 (m, 4H; C₆H₅) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 24.2,

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= 1.7 Hz). 128.5. 128.9

28.1, 34.4, 52.1, 122.2, 123.3 (d, ${}^{4}J_{pc} = 1.7$ Hz), 128.5, 128.9 (d, ${}^{3}J_{pc} = 6.9$ Hz), 129.4, 134.4 (d, ${}^{2}J_{pc} = 20.1$ Hz), 135.2 (d, ${}^{1}J_{pc} = 15.6$ Hz), 151.9 (d, ${}^{3}J_{pc} = 12.2$ Hz), 156.1 (d, ${}^{1}J_{pc} = 36.9$ Hz) ppm; ${}^{31}P{}^{1}H{}$ NMR (160 MHz, C₆D₆): $\delta = -14.3$ ppm; HRMS: *m/z* calcd for C₂₆H₃₀N₂P: 401.2141 [M + H] ⁺; found: 401.2131.

PhN=C(PPh₂)(NHCH₂Cy) (9p). Colorless solid, isolated yield 81% (162 mg);.¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 0.65-1.58 (m, 11H; Cy), 3.31 (t, *J* = 5.8 Hz, 2H; CH₂), 4.46 (t, *J* = 5.0 Hz, 1H; NH), 6.81-6.85 (m, 1H; C₆H₅), 7.03-7.10 (m, 10H; C₆H₅), 7.40-7.44 (m, 4H; C₆H₅) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 26.1, 26.7, 30.9, 37.6, 48.5, 122.1, 123.2 (d, ⁴*J*_{pc} = 1.6 Hz), 128.5, 128.9 (d, ³*J*_{pc} = 6.8 Hz), 129.4, 134.4 (d, ²*J*_{pc} = 20.1 Hz), 135.0 (d, ¹*J*_{pc} = 15.4 Hz), 151.9 (d, ³*J*_{pc} = 11.9 Hz), 157.3 (d, ¹*J*_{pc} = 36.2 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -13.5 ppm; HRMS: *m/z* calcd for C₂₆H₃₀N₂P: 401.2141 [M + H]⁺; found: 401.2141.

PhN=C{P(C₆H₄Me-4)₂} NH-(CH₂)₈ (9q). Colorless solid, isolated yield 80% (177 mg); ¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.30-1.92 (m, 14H; -(CH₂)₇-), 2.00 (s, 6H; Me), 4.45 (br, 1H; CH), 4.56 (d, ³J = 7.1 Hz, 1H; NH), 6.82-6.87(m, 1H; C₆H₅), 6.92 (d, J = 7.6 Hz, 4H; C₆H₄),7.10 (d, J = 4.4 Hz, 4H; C₆H₅), 7.42 (t, J = 7.6 Hz, 4H; C₆H₄),7.10 (d, J = 4.4 Hz, 4H; C₆D₆,Me₄Si): δ = 21.1, 23.9, 25.7, 27.4, 32.1, 51.2, 122.1, 123.4 (d, ⁴J_{pc} = 1.5 Hz), 128.5, 129.8 (d, ³J_{pc} = 7.0 Hz), 132.1(d, ¹J_{pc} = 14.4Hz), 134.4 (d, ²J_{pc} = 20.3 Hz), 139.4, 152.1 (d, ³J_{pc} = 12.0 Hz), 156.5(d, ¹J_{pc} = 36.9Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -15.9 ppm; HRMS: *m/z* calcd for C₂₉H₃₆N₂P: 443.2611[M + H] ⁺; found: 443.2602

PhN=C(PPh₂){NH-(CH₂)₈} (9r). Colorless solid, isolated yield 93% (186 mg);.¹H NMR (400 MHz, C₆D₆, Me₄Si): δ = 1.28-1.85 (m, 14H; -(*CH*₂)₇-), 4.44 (br, 2H; CH and NH), 6.82-6.86 (m, 1H; C₆H₅), 7.03-7.11 (m, 10H; C₆H₅), 7.42-7.45 (m, 4H; C₆H₅) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): δ = 23.8, 25.7, 27.5, 31.9, 51.2, 122.2, 123.3 (d, ⁴J_{pc} = 1.2 Hz), 128.5, 128.9 (d, ³J_{pc} = 6.8 Hz), 129.4, 134.4 (d, ²J_{pc} = 20.1 Hz), 135.3 (d, ¹J_{pc} = 15.6 Hz), 151.9 (d, ³J_{pc} = 12.2 Hz), 156.1 (d, ¹J_{pc} = 37.5 Hz) ppm; ³¹P{¹H} NMR (160 MHz, C₆D₆): δ = -14.3 ppm; HRMS: *m*/z calcd for C₂₇H₃₂N₂P: 415.2298 [M + H] ⁺; found: 415.2289.

PhP{iPrN=C(NHiPr)}₂ (9s). Colorless oil, isolated yield 93% (168 mg);.¹H NMR (400 MHz, C₆D₆, Me₄Si): $\delta = 0.98$ (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 1.08 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 1.22 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 1.34 (d, J = 6.1 Hz, 6H; CH(CH₃)₂), 4.24-4.36 (m, 6H; CH and NH), 7.04-7.10 (m, 3H; C₆H₅), 7.51-7.55 (m, 2H; C₆H₅) ppm; ¹³C NMR (100 MHz, C₆D₆,Me₄Si): $\delta = 22.4$ (d, ⁴ $J_{pc} = 2.9$ Hz), 25.3 (d, ⁴ $J_{pc} = 8.0$ Hz), 42.9, 52.6 (d, ³ $J_{pc} = 36.9$ Hz), 129.2 (d, ³ $J_{pc} = 6.4$ Hz), 129.6, 133.3 (d, ¹ $J_{pc} = 17.6$ Hz), 134.0 (d, ² $J_{pc} = 19.1$ Hz), 151.4 (d, ¹ $J_{pc} = 35.4$ Hz); ³¹P{¹H} NMR (160 MHz, C₆D₆): $\delta =$ -34.9 ppm; HRMS: *m/z* calcd for C₂₀H₃₆N₄P: 363.2672 [M + H] ⁺; found: 363.2669.

X-ray diffraction analysis

Data collections for **9i** were performed at 180 K on a SuperNova diffractometer using graphite-monochromated Mo K α radiation ($\lambda = 0.71073$ Å). Using Olex2, the structure of **9i** was solved with the Superflip structure solution program using

Charge Flipping and refined with the XL refinement package using Least Squares minimization. Refinement was performed on F^2 anisotropically for all the non-hydrogen atoms by the full-matrix least-squares method. The hydrogen atoms were placed at the calculated positions and were included in the structure calculation without further refinement of the parameters. Crystallographic data (excluding structure factors) have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number: CCDC 1047716. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data_request/cif.

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