MedChemComm

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this Accepted Manuscript with the edited and formatted Advance Article as soon as it is available.

You can find more information about *Accepted Manuscripts* in the **Information for Authors**.

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard <u>Terms & Conditions</u> and the <u>Ethical guidelines</u> still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.



www.rsc.org/medchemcomm



Curves showing two different trials of killing a MRSA strain with [Cp*IrCl(cis-1,2-diaminocyclohexane]Cl 85x40mm (300 x 300 DPI)

MedChemComm

Concise Article

Cite this: DOI: 10.1039/x0xx00000x

Received ooth January 2012, Accepted ooth January 2012

DOI: 10.1039/x0xx00000x

www.rsc.org/

Transition Metal Diamine Complexes with Antimicrobial Activity Against *Staphylococcus aureus* and Methicillin-Resistant *S. aureus* (MRSA).

G. W. Karpin,^{a,b} D. M. Morris,^{a,b} M. T. Ngo,^{a,b} J.S. Merola^{*a,b} and J. O.

Falkinham, III^{a,c}

Pentaalkylcyclopentadienyl (Cp*^R) Iridium (Ir) and cobalt (Co) 1,2-diamine complexes were synthesized. Susceptibility of *Staphylococcus aureus* and recent patient methicillin-resistant *S. aureus* (MRSA) isolates to the transition metal-diamine complexes were measured by broth microdilution and reported as the MIC and MBC. Hemolytic activities of the transition metal-complexes as well as toxicity toward Vero cells were also measured. The transition metal complex of Cp^{*R}Ir with cis-1,2-diaminocyclohexane, had strong antibiotic activity against *S. aureus* and MRSA (MIC = 4 μ M, MBC = 8 μ M) strains and killed 99% of *S. aureus* cells in 6 hours. Stronger antibiotic activity was associated with the presence of octyl linked to the cyclopentadienyl group and cyclohexane as the diamine backbone. Activity was greatly diminished by tri- or tetramethylation of the nitrogen of the diamine. A cyclopentadienylcobalt complex of cis-1,2-diaminocyclohexane also showed significant antimicrobial activity against both *S. aureus* and MRSA strains. The absence of hemolytic activity, Vero cell cytotoxicity and the significant anti-microbial activity of several members of the family of compounds reported suggest this is an area worth further development.

Introduction

Nosocomial infections involving Staphylococcus aureus and methicillin-resistant S. aureus (MRSA) continue to present major challenges in the United States and Europe.¹⁻³ There is also evidence of an alarming increase in S. aureus skin infections in children.^{4, 5} While both S. aureus and MRSA infections occur more frequently amongst persons with weakened defences against infection in hospitals and healthcare facilities, MRSA infections are also of increasing concern in the community at large.⁶ Many S. aureus and MRSA infections are intractable due to antibiotic resistance and their tendency to be localized in high densities, such as in abscesses or biofilms.⁷ Colonization of in-dwelling catheters leads to biofilm formation and results in catheters serving as sources of continual infection in patients.⁷ One strategy to combat these infections is to develop novel and effective anti-infective agents.

There has been an extensive body of work reporting on transition metal compounds with anti-cancer activity.⁸ Sadler and co-workers have synthesized and investigated the cytotoxic properties of a number of half-sandwich type organometallic compounds^{9, 10}. Others have also reported on the cytotoxic behavior of different variants of half-sandwich compounds.^{11, 12}

By comparison, the work on small inorganic/organometallic compounds as antimicrobials has been sparse.¹³ The antimicrobial properties of the Ag⁺ ion have long been known ^{14, 15}. More recently new compounds incorporating the silver ion with ligands such as N-heterocyclic carbenes have been shown to possess anti-microbial activity as well.¹⁶⁻¹⁸ In many other cases, anti-bacterial activity

stems from either making a metal complex of a known organic antibacterial^{19, 20} or making an organometallic analog of a known antibiotic by replacing an aryl group with an organometallic moiety such as ferrocene.^{21, 22} Recently, an intriguing polypeptide with three organometallic groups has been shown to be a potent antimicrobial. ²³ A review on small organometallic compounds as antibacterial agents appeared recently.²⁴

Previously, we have reported that compounds based on simple organometallic complexes comprised of either iridium (Ir) or rhodium (Rh) chelated by amino acids were potent in vitro inhibitors of members of the genus Mycobacterium.²⁵ Herein we extend our documentation of the strong antibiotic properties of a different family of transition metal complexes containing 1,2-diamine ligands to S. aureus and MRSA. Based on the encouraging initial results we believe that this work could lead to the identification of new therapeutic targets and development of a new class of urgently needed drugs to address the growing problem of difficult-to-treat S. aureus and MRSA infections.

Results and Discussion

Chemistry. In a previous paper, we reported on cyclopentadienyl rhodium and iridium amino acid complexes that exhibited significant anti-mycobacterial activity.²⁵ None of the amino acid compounds displayed any significant anti-staphylococcal activity. As part of that mycobacterial study, we also made complexes of two well-known drugs used in the treatment of tuberculosis (TB): isoniazid and ethambutol ²⁶. Neither [Cp*Ir(isoniazid)Cl]Cl nor

RSCPublishing

[Cp*Ir(ethambutol)Cl]Cl were active against an extensive panel of mycobacteria with MIC > 125 μ g/mL. However, our practice is to screen all compounds first against a broad panel of bacteria, yeast and fungi and we noted that the ethambutol compound had moderate activity against *S. aureus* (MIC = 35 μ g/mL) Noting that ethambutol is a substituted1,2-diaminoethane, we turned our attention to complexes of other diamine ligands. That replacement of the chelating amino acids with chelating 1,2-diamines was successful in producing the anti-staphylococcal compounds described in this work.

Complexes of rhodium and iridium were synthesized according to the general procedure outlined in Scheme 1. The penta-alkyl substituted Cp rings shown in the scheme may be pentamethyl (Cp*) or tetramethylalkyl (Cp*^R) the starting cyclopentadienes being synthesized as described previously²⁷. Using this route, an extensive series of compounds were prepared, fourteen of which are listed in table 1, **1-Ir** through **14-Ir**. No significant difference in activity was found between Rh and Ir compounds, so only Ir compounds are reported here.



Scheme 1. Outline of the 2-step synthesis of iridium complexes with 1,2-diamines as the organic ligand. Ring and diamine substitution is delineated in table 1.



Scheme 2. Outline of the synthesis of cobalt-containing complexes with 1,2-diaminocyclohexane.

It was also decided that the cobalt congeners should be investigated since the lighter, first-row transition elements would be less likely to have general cytotoxic behaviors. The route shown in scheme 1 is not applicable for cobalt, so a new synthesis based on cyclopentadienylcobaltdicarbonyl, CpCo(CO)₂, was developed²⁸ (Scheme 2). Only the 1,2-diaminocyclohexane compounds of cobalt, **15-Co** and **16-Co**, are discussed herein.

Characterization of Compound Structure and Purity. All compounds reported in this paper were characterized by standard analytical techniques: ¹H NMR spectroscopy, ¹³C NMR spectroscopy and High Resolution Mass Spectrometry. All spectroscopies were completely consistent with the proposed structures. In addition, to assure the purity of the complexes by examining the NMR spectroscopies for the presence of any

All complexes described in this paper are quite water soluble, even complex 12-Ir containing a decyl chain on the Cp ligand. This is not surprising since the compounds are cationic with a chloride counterion. Complexes that had been dissolved in D_2O were monitored by ¹H NMR spectroscopy and show no changes in physical appearance of the solution nor any change in the NMR spectra over months of monitoring. An indicator that, in general, these diamine complexes are stable in these *in vitro* conditions is that there was no anti-mycobacterial activity of the ethambutol complex. At the highest concentrations used, even a few percent decomposition of the ethambutol complex would release a sufficient amount of free ethambutol to show anti-mycobacterial activity and none was observed.

More to the, a standard solution of complex **5-Ir** in the growth medium was prepared and tested for activity against *S. aureus* at the time of preparation, at 6 months following preparation and at one year following preparation. The standard solution was maintained at ambient temperature during that period of time. The MIC of **5-Ir** against *S. aureus* was reproducibly 4 μ g/mL over the one year testing period showing no decomposition of the complex in the culture medium over a 12 month period of time

Anti-staphylococcal Activities and Complex Structures. Sixteen different complexes that were tested against *S. aureus* and various laboratory and patient strains of MRSA are listed in Table 1. A partial listing of the antimicrobial activities are listed in Table 2 while a more complete table can be found in the supplementary material. While a number of iridium complexes showed activity against *S. aureus*, two of those compounds, **4-Ir** and **5-Ir** were notable for their low MICs against both *S. aureus* and MRSA isolates (Table 2). Further, the MBCs for **5-Ir** were almost the same as the MICs, demonstrating its bactericidal activity.

Table 1. General structure and numbering for complexes in this paper. The table lists the various diamine ligand substitutions $(R_1, R_2, R_2 \text{ and } R_4)$, the C₅Me₄R substitution as well as the metal.

$\begin{array}{c} R_5 \\ R_5 \\ R_5 \\ R_5 \\ R_5 \\ R_5 \\ R_6 \\ R_5 \\ R_6 \\ R_7 \\ R_1 \\ R_2 \\ R_4 \\ R_3 \\ \end{array}$	x

Cpd	R_1, R_2, R_3	R_4	R ₅	R ₆	Х	Ligand
1-Ir	Н	Н	CH3	CH3	Cl	-CH ₂ -CH ₂ -
2-Ir	Н	Н	CH ₃	C_8H_1	Cl	-CH ₂ -CH ₂ -
3-Ir	Н	Н	CH ₃	CH ₃	Cl	cis-cyclohexane
4-Ir	Н	Н	CH ₃	C_8H_1	Cl	cis-cyclohexane
5-Ir	Н	Н	CH ₃	C_6H_1	Cl	cis-cyclohexane
6-Ir	Н	Н	CH ₃	CH ₃	Cl	trans-cyclohexane
7-Ir	Н	Н	CH,	C_8H_1	Cl	trans-cyclohexane
8-Ir	Н	Н	CH,	$C_{12}H_{2}$	Cl	cis-cyclohexane
9-Ir	CH ₃	CH ₃	CH ₃	CH3	Cl	-CH ₂ -CH ₂ -
10-Ir	CH,	Н	CH,	CH,	Cl	-CH ₂ -CH ₂ -
11-Ir	Н	C_7H_7	CH ₃	CH ₃	Cl	-CH ₂ -CH ₂ -
12-Ir	Н	C ₁₀ H	CH ₃	C_8H_1	Cl	-CH ₂ -CH ₂ -
13-Ir	Н	Н	CH ₃	CH ₃	Cl	(1S,2S)-1,2-bis
			-	-		(4-fluorophenvl)

14-Ir	Н	Н	CH3	CH3	Cl	(
						(
15-Co	Н	Н	Н	Н	Ι	С
16-Co	Н	Н	Н	Н	Ι	t

1S,2S)-1,2-bis 4-methoxypheny may be at play. Evidence for this is that 8-Ir with the significantly longer dodecyl chain on the Cp* ring has little activity. is-cyclohexane rans-cyclohexaneAryl groups on the ethylenediamine backbone also resulted in

Table 2. MICs(MBCs) of 1,2-diamine complexes of Cp complexes of iridium and cobalt agains S. aureus and MRSA strains in uM.

Compo	S. aureus	MRSA	MRSA	MRSA
und	6538	patient	43300	34380
1-Ir	296(600)	296(250)	296(600)	600(600)
2-Ir	124(124)	63(63)	124(124)	124 (124)
3-Ir	34(34)	34(34)	34(68)	34(34)
4-Ir	7(16)	7 (7)	7 (8)	7 (7)
5-Ir	4(4)	4(4)	4(4)	4 (4)
6-Ir	262 (524)	262 (524)	262 (524)	262 (524)
7-Ir	108(108)	108(108)	216 (432)	432 (432)
8-Ir	198 (400)	198 (400)	198 (250)	400 (400)
9-Ir	> 500	> 500	> 500	> 500
10-Ir	> 500	> 500	> 500	> 500
11-Ir	> 500	> 500	> 500	> 500
12-Ir	97(97)	97(97)	97(97)	97(97)
13-Ir	26(26)	26(26)	26(52)	26(26)
14-Ir	62.5(62.5)	52(52)	104(104)	104(104)
15-Co	11 (22)	22 (22)	44 (44)	44 (44)
16-Co	> 500	> 500	> 500	> 500
Vanco	0.7 (0.7)	o.7 (o.7)	o.7 (o.7)	o.7 (o.7)

The data in Table 2 not only demonstrate that these complexes exhibit potent activity against both S. aureus and MRSA but also provide preliminary insight into the structure-activity relationships (SARs) for these complexes. Our results thus far have suggested the following conclusions:

Substitutions to the amino groups with either four (9-Ir) or three (10-Ir) methyl groups on the nitrogen atoms leads to a loss of activity (Table 2). We speculate that the hydrogen bond contributions of the NH-functionalities on opposite sides of the diamine core are important to activity.

Several observations support the conclusion that the active species in these experiments is the organometallic complex and not the metalfree ligand nor the ligand-free metal. First, all starting materials and complex intermediates were tested against the S. aureus panel and showed no activity. The fact that only some of these complexes display any activity against staph strains is good evidence that the complexes are not decomposing to form the metal ion or some other indiscriminate metal fragment - if that were the case all of the compounds would show activity. These observations lead us to conclude that these complexes are robust and do not dissociate under conditions of the assay to yield a common iridium or rhodium species that would account for activity.

Comparison between 1-Ir and 2-Ir where the only difference is the substitution on the Cp* ring of an octyl group for one of the methyl groups shows an increase in activity of **2-Ir**, the octyl substituted ring complex. Similarly, comparison between 3-Ir, 4-Ir and 5-Ir, all complexes with the cis-1,2-diaminocyclohexane ligand, shows that there is an increase in activity for **4-Ir** and **5-Ir** where one methyl group of the Cp* ligand is substituted with either a hexyl or an octyl group. It is tempting to ascribe this effect to a simple increase in

activity but the range of compounds is too small to draw any significant conclusions. However, greater activity is observed for 4fluorophenyl groups, 13-Ir, than for 4-methoxyphenyl groups, 14-Ir.

hydrophobicity of the complexes, but a more specific interaction

The most dramatic difference in activity is observed for both iridium (3-Ir vs 6-Ir) and cobalt (15-Co vs 16-Co) where the sole difference is the use of the cis- vs the trans- isomer of 1,2-diaminocyclohexane. Both isomers complex strongly with the iridium and cobalt, but only the complexes with the cis-isomer shows significant activity against S. aureus.

Time-kill measurements. Survival of S. aureus strain ATCC 6358 and MRSA strain 34380 when exposed to 8 mg 4-Ir/L (i.e., MIC/MBC) after 1, 2, 3, and 6 h exposure at 37° C were measured in two independent experiments (Figure 1). 4-Ir killed both the antibiotic-susceptible S. aureus and the methicillin-resistant S. aureus strains (Figure 2).

Hemolytic activities. The complexes air 10 with strong antibiotic activity (i.e., 3-Ir, 4-Ir, and 5-Ir) were tested for hemolytic activity (< 10% hemolysis of sheep red blood cells). These complexes showed no hemolytic activity at their MICs or MBCs and hemolytic activity at the 10% level was not observed until concentrations of greater than 250 µg/mL were reached.

Mammalian toxicity. While the cobalt complexes discussed above were not tested, the iridium complexes that showed significant activity against S. aureus and MRSA strains showed no evidence of toxicity against Vero cell line ATCC CCL-81²⁹ at the highest concentrations tested of 500 µM. In addition, compound 4-Ir was tested for toxicity in mice and it was determined that no detrimental toxicological effects were present at doses of 5 mg/kg.²⁵

Experimental

Materials. Unless otherwise stated, synthetic work was carried out in air with untreated solvents. Commercially available reagents including all diamines were obtained from Sigma-Aldrich, St. Louis, MO 63103. Deuterated solvents for NMR spectroscopy were obtained from Cambridge Isotope Laboratories.



Figure 1. Killing S. aureus strain ATCC 6358 with 4-Ir.

MedChemComm



Figure 2. Killing of MRSA strain 34380 with 4-Ir.

General Procedure. A round bottom flask was charged with appropriate amounts of the respective $[Cp^{xR}IrCl_2]_2^{9,27,30}$ in dichloromethane with magnetic stirring. After 10 minutes the 1,2-diamine was added in a 2.1 molar excess drop wise to the stirring solution. After the addition of the 1,2-diamine the solution was allowed to react for an additional 30 minutes. The complexes were recrystallized from dichloromethane and ether or hexanes and collected on a fine fritted filter as yellow crystalline powders. ¹H NMR and ¹³C NMR spectra were collected on a Varian MR-400 NMR spectrometer. High-resolution mass spectrometry data was collected on Agilent 6220 Accurate Mass TOF LC-MS. Elemental analyses were performed by Atlantic Microlabs, Norcross, GA.

Synthesis of $[(η^5-C_5Me_5)Ir(ethylenediamine)CI]CI (1-Ir).$ Following the general procedure: 100. mg (0.125 mmol) of [Cp^{*}IrCl₂]₂ was combined with 15.9 mg (0.264 mmol) of ethylenediamine in dichloromethane. Yield: 91.7 mg (0.217 mmol) (86%) HRMS/ESI+ (*m/z*): Calcd for C₁₂H₂₃N₂[¹⁹³Ir]Cl 423.1257; Found 423.1265 Anal calc. C₁₂H₂₃Cl₂ N₂Ir C: 31.44 H: 5.06; Found C: 31.65 H: 5.10. ¹H NMR (400 MHz, CDCl₃) δ 6.81 (br s, 2H), 5.74 (br s, 2H), 3.27 (m, 2H), 3.07 (d, 2H) 1.89 (s, 15H, 5CpMe) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 93.2 (CpC) 92.8 (CpC) 92.2 (CpC), 54.1 (CH₂) ppm.

<u>Synthesis of $[(\eta^5-C_4Me_4C_8H_{17})Ir(ethylendiamine)Cl]Cl (2-Ir)</u>.$ $Following the general procedure, 100. mg (0.112 mmol) of <math>[Cp^{*(octyl)}IrCl_2]_2$ was combined with 14.2 mg (0.235 mmol) of ethylenediamine in dichloromethane. Yield: 85.1 mg (0.152 mmol) (68%) HRMS/ESI+ (*m*/*z*): Calcd for $C_{19}H_{37}N_2[^{193}Ir]Cl_2$ 556.1963; Found 556.1959 ¹H NMR (400 MHz, CDCl₃) δ 5.41 (br s, 2H, NH), 4.65 (br s, 2H, NH), 3.35 (d, 2H, CH₂), 3.07 (d, 2H, CH₂), 2.14 (m, 2H), 1.96 (m, 2H), 1.84 (s, 15H, 5CpMe), 1.43 (m, 2H), 1.25 (br m, 4H) 1.05-0.96, (m, 4H), 0.84 (m, 3H, CH₃) ppm.</u>

Synthesis of $[(\eta^5-C_5Me_5)Ir(cis-1,2-diaminocyclohexane)Cl]Cl ($ **3-Ir**).Following the general procedure, 87.1 mg (0.109 mmol) of[IrCp*Cl₂]₂ was combined with 26.2 mg (0.229 mmol) of*cis*-1,2diaminocyclohexane. Yield: 100.7 mg (0.211 mmol) (96%).HRMS/ESI+ (*m/z*): Calcd for C₁₆H₂₉N₂[¹⁹³Ir]Cl 477.1747; Found477.1752. Anal calc. C₁₆H₂₈Cl₂ N₂Ir · (H₂O) C: 36.19 H: 5.71; $Found C 35.83 H: 5.73. ¹H NMR (400 MHz, CDCl₃) <math>\delta$ 5.72 (s, 2H, NH₂) 3.27 (m, 1H, CH), 3.09 (m, 1H, CH), 2.34 – 2.14 (m, 2H), 2.08 (d, *J* = 12.6 Hz, 2H), 1.91 (s, 15H CpMe), 1.65 (q, *J*=12.9 Hz, 2H) CH₂), 1.43 (q, *J* = 12.9 Hz, 2H) 1.26 (m, 2H), 1.22 (m, 2H, CH₂) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 93.53 (CpC), 92.25(CpC), 84.75(CpC), 84.04(CpC), 34.9, 32.05, 31.72, 28.14, 26.93, 21.41, 10.44 , 9.74 ppm.

Synthesis of $[(\eta^5-C_4Me_4C_8H_{17})Ir(cis-1.2-diaminocyclohexane)Cl]Cl$ (4-Ir). Following the general procedure, 50.0 mg (0.056 mmol) of $[Cp^{*(octyl)}IrCl_2]_2$ was combined with 24.3 mg (0.212mmol) of *cis*-1,2-diaminocyclohexane. Yield: 53.1 mg (0.092 mmol) (83 %). HRMS/ESI+ (*m/z*): Calcd for $C_{23}H_{43}N_2[^{193}Ir]Cl 575.2751$; Found 575.2764. Anal calc. $C_{23}H_{43}Cl_2 N_2Ir C: 45.23 H: 7.10$; Found C: 45.19 H: 7.15. ¹H NMR (400 MHz, CDCl₃) δ 3.38 (2, H), 3.10 (m, 2H), 2.24 (t, *J* = 8.0 Hz, 4H), 2.08 (m, 2H), 1.84(s, CpMe), 1.81 (s, CpMe), 1.78 (s, CpMe), 1.76 (s, CpMe) 1.64 (t, *J* = 8.0 Hz, 2H, CH₂), 1.43 (p, *J* = 8.1 Hz, 4H), 1.32 (m, 4H, 2CH₂), 1.16 (m, 4H. 0.86 (t, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 88.2 (CpC) 86.2 (CpC) 86.1 (CpC), 51.39 (CH) 49.34 (CH), 31.88(CH₂), 31.87(CH₂), 30.59(CH₂), 29.69 (CH₂), 29.50(CH₂), 29.28(CH₂), 29.07(CH₂), 27.61(CH₂),26.44 (CH₂), 26.22(CH₂), 25.17 (CH₂), 24.12 (CH2), 15.25 (CH₃) ppm.

Synthesis of $[(η^5-C_4Me_4C_6H_{13})$ **Ir**(cis-1,2-diaminocyclohexane)CI]Cl (**5-Ir**). Following the general procedure, 50 mg (0.057 mmol) of [IrCp^{*hexyl}Cl₂]₂ was combined with 15.9 mg (0.139 mmol) of *cis*-1,2diaminocyclohexane. Yield: 52.0 mg (0.095 mmol) (85%).HRMS/ESI+ (m/z): Calcd for C₂₁H₃₉N₂[193Ir]Cl Calc 547.2509; Found 547.2541 . Anal calc. C: Calc 43.29 H: 6.75 Found C: 43.078 H: 6.74. ¹H NMR (400 MHz, CDCl₃) δ 6.51 (s, 1H, NH), 5.71 (s, 1H, NH), 4.03 (m. 1H, CH), 3.31 (m, 1H, CH), 2.24 (t, J = 8.5, 2H), 1.89 – 1.81 (2s,15H, CpMe), 1.78 (m, 4H), 1.47 – 1.34 (m, 4H), 1.34 – 1.12 (m, 2H), 0.92 – 0.76 (m, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 87.41 (CpC) 86.14 (CpC), 47.6 (CH) 46.31 (CH), 35.55 (CH₂), 32.64(CH₂), 29.69 (CH₂), 29.50 (CH₂), 27.61(CH₂), 26.44 (CH₂), 26.22, 24.12 (CH₂), 12.45 (CH₃), 10.4 (CpMe) ppm.

<u>Synthesis of $[(\eta^5-C_5Me_5)Ir(\pm-trans-1,2-diaminocyclohexane)Cl]Cl</u>$ (<u>6-Ir</u>). Following the general procedure, 47.1 mg (0.059 mmol) of[Cp^{*}IrCl₂]₂ was combined with 15.2 mg (0.133 mmol) of trans-1,2diaminocyclohexane. Yield 48.1 mg (0.101 mmol) (81%). Calcd forC₁₆H₂₉N₂[¹⁹³Ir]Cl 477.1765; Found 477.1761. ¹H NMR (400 MHz, $CDCl₃) <math>\delta$ 5.68 (s, 2H, NH₂) 3.27 (m, 1H, CH), 3.15 (m, 1H, CH), 2.25 (m, 2H), 2.08 (d, J = 11.6 Hz, 2H), 1.84 (2s, 15H CpMe), 1.65 (q, J = 11.6 Hz, 2H CH₂), 1.43 (q, J = 12.9 Hz, 2H) 1.26 (m, 2H), 1.22 (m, 2H, CH₂) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 94.85 (CpC), 93.25(CpC), 84.75(CpC), 84.04(CpC), 34.9, 32.05, 30.82, 28.14, 27.33, 21.41, (cyclohexane) 10.18 (CpMe) ppm.</u>

Synthesis of $[(\eta^5-C_4Me_4C_8H_{17})Ir(trans-\pm-1,2-$

 $\frac{\text{diaminocyclohexane}(CI]Cl (7-Ir).}{\text{forward}} Following the general procedure,}$ $75.0 mg (0.083 mmol) of [Cp*(octyl)IrCl_2]_2 was combined with 36.4 mg (0.318 mmol) of trans-1,2-diaminocyclohexane. Yield: 71.0 mg (0.119 mmol) (65%). HRMS/ESI+ ($ *m*/*z*): Calcd for $C_{23}H_{43}N_2[^{193}Ir]Cl 575.2755; Found 575.2771.). ¹H NMR (400 MHz, CDCl_3) & 3.56 (2, NH_2), 3.36 (m, 2H), 2.75 (t,$ *J*= 9.5 Hz, 4H), 2.17 (m, 2H), 1.86(s, CpMe), 1.84 (s, CpMe), 1.81 (s, CpMe), 1.76 (s, CpMe) 1.64 (t,*J*= 9.5 Hz, 2H, CH₂), 1.43 (p,*J*= 8.1 Hz, 4H), 1.37 (m, 4H, 2CH₂), 1.16 (m, 4H. 0.91 (t, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) & 88.7 (CpC) 88.5 (CpC) 84.2 (CpC), 51.56 (CH) 49.65(CH), 32.88(CH₂), 30.77(CH₂), 30.59(CH₂), 29.69 (CH₂), 29.50(CH₂), 29.28(CH₂), 29.07(CH₂), 26.81CH₂),26.44 (CH₂), 25.44(CH₂), 25.17 (CH₂), 24.12 (CH₂), 15.18(CH₃) ppm.

 $\begin{array}{l} \underline{Synthesis of [(\eta^5-C_4Me_4C_{12}H_{25})Ir(cis-1,2-diaminocyclohexane)Cl]Cl} \\ \underline{(8-Ir)}. Following the general procedure, 50.0 mg (0.051 mmol) of [Cp*(^{dodecyl})IrCl_2]_2 was combined with 12.0 (0.105 mmol) mg of cis-1,2-diaminocyclohexane. Yield: 39.2 mg (0.062 mmol) (60%). \\ HRMS/ESI+ (m/z): Calcd for C_{27}H_{51}N_2[193Ir]Cl Calc 631.3448 ; \end{array}$

Page 6 of 8

ARTICLE

Found 631.3475. ¹H NMR (400 MHz, CDCl₃) δ 5.69 (s, 1H, NH), 4.24 (s, 1H, NH), 3.24 (m. 1H, CH), 3.05 (m, 1H, CH), 2.39 – 2.19 (bm, 6H), 1.85 (s, 15H, CpMe), 1.71 – 1.62 (m, 4H), 1.61 – 1.52 (m, 4H), 1.51-1.45 (m, 4H), 1.40 (m. 6H), 1.35-1.15 (m, 10H), 0.84 (m, 3H, CH₃) ppm.

Synthesis of $[(\eta^5 - C_5 Me_5)Ir(N,N,N',N'-$

tetramethyl)ethylenediamine)Cl]Cl (9-Ir). Following the general procedure, 50.0 mg (0.062 mmol) of $[Cp^*IrCl_2]_2$ was combined in dichloromethane with 18.1 mg (0.155 mmol) of N,N,N',N'-tetramethylethylenediamine. Yield: 42.2 mg (0.088 mmol). HRMS/ESI+ (*m/z*): Calcd for C₁₆H₃₁N₂[¹⁹³Ir]Cl 479.1805; Found 479.1804. ¹H NMR (400 MHz, CDCl₃) δ 3.62 (m, 3H, CH₃), 3.23 (s, 3H, CH₃), 2.88 (m, 3H, CH₃), 2.42 (m, 2H, CH₂) 1.89 (s, 15H, CpMe) ppm.

Synthesis of $[(\eta^5-C_5Me_5)Ir(N,N,N'-trimethyl)ethylenediamine)Cl]Cl]$ (10-Ir). Following the general procedure, 50.0 mg (0.062 mmol) of $[Cp^*IrCl_2]_2$ was combined in dichloromethane with 17.8 mg (0.174 mmol) of N,N,N'-trimethylethylenediamine dissolved in 10 mL of methanol. Yield: 36.2 mg (0.078 mmol). HRMS/ESI+ (*m/z*): Calcd $C_{15}H_{29}N_2[^{193}Ir]Cl$ for 465.1648; Found 465.1650. ¹H NMR (400 MHz, CDCl₃) δ 4.65 (br s, 1H, NH), 3.42 (m, 3H, CH₃), 3.12 (m, 2H, CH₂), 2.65 (m, 3H, CH₃), 1.87 (s, 15H, CpMe) ppm.

Synthesis of $[(\eta^5-C_5Me_5)Ir(N-(1-Benzyl)ethylenediamine)CI][CI]$ (**11-Ir**). Following the general procedure: 50.0 mg (0.062 mmol) of [Cp^{*}IrCl₂]₂ was combined in dichloromethane with 19.8 mg (0.171 mmol) of N-(1-benzyl)ethylenediamine dissolved in 10 mL of methanol. Yield: 42.3 mg (0.082 mmol). HRMS/ESI+ (*m/z*): Calcd $C_{19}H_{29}N_2[^{193}Ir]Cl$ for 513.1649; Found 513.1648. ¹H NMR (400 MHz, CDCl₃) δ 7.39 (m, 4H, CH), 7.26 (d, 1H, CH), 5.68 (d, 2H, NH), 5.52 (s, 1H, NH), 4.57 (s, 2H), 2.1 (br s, 2H), 1.73 (s, 15H, CpMe), 1.35 (s, 2H) ppm. ¹³C NMR (101 MHz, CDCl₃) δ 125.6 (aryl), 124.9 (aryl), 88.2 (CpC), 87.1 (CpC), 55.2 (CH₂), 51.4 (CH₂), 12.5 (CH₃) ppm.

Synthesis of $[(\eta^5 - C_5 Me_4 C_8 H_{17}) Ir(N-(1$ naphthyl)ethylenediamine)Cl]Cl (12-Ir). Following the general procedure, 50.0 mg (0.056 mmol) of [Cp*(octyl)]rCl₂]₂ was combined in dichloromethane with 32.1 mg (0.171 mmol) of N-(1napthyl)ethylenediamine dissolved in 10 mL of methanol. Yield: 40.6mg (0.062 mmol) (55%). HRMS/ESI+ (*m/z*): Calcd for $C_{29}H_{43}N_2[^{193}Ir]$ 647.2321 Found 647.2394. Anal calc. $C_{29}H_{43}Cl_2$ N₂Ir) C: 49.71 H: 6.46 Found C: 48.86 H: 5.96. ¹H NMR (400 MHz, CDCl₃) δ 7.55 (d, J=6.9 Hz, 1H, ArH), 7.41 (d, J = 6.9 Hz, 1H, ArH), 7.14 (m, 4H, ArH), 4.88 (m, 2H), 3.72 (d, J=7.5 Hz, 1H), 3.60 (d, J = 7.5 Hz, 1H), 3.11 (s, 2H), 2.36 (t, J = 11.5 Hz, 2H), 2.02 (s, 3H, CpMe), 1.99 (s, 3H, CpMe) 1.93 (s, 3H, CpMe), 1.89 (s, 3H, CpMe), 1.79 (t, J=12.6, 2H, CH₂), 1.72 (d, 1H, CH) 1.52 – 1.28 (m, 4H), 1.16 (m, J = 11.5 Hz, 3H, CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) & 129.9 (aryl), 127.9 (aryl), 127.8 (aryl), 86.2 (CpC), 86.1 (CpC), 60.39 (CH₂), 58.2 (CH₂), 31.88(CH₂), 31.87(CH₂), 30.59(CH₂), 35.69 (CH₂), 32.50(CH₂), 27.28(CH₂), 26.07(CH₂), 27.55(CH₂), 22.30 (CH₂), 14.15 (CH₃) ppm.

Synthesis of $[(\eta^5-C_5Me_5)Ir(1S, 2S)-1, 2-Bis(4-$

<u>fluorophenyl)ethylenediamineCl]Cl (13-Ir)</u>. Following the general procedure, 50.0 mg (0.062 mmol) of [Cp*IrCl₂]₂ was combined with 42.3 mg (0.131 mmol) (1S, 2S)-1, 2-bis(4-fluorophenyl)ethylenediamine dihydrochloride in a 50/50 mix of dichloromethane and methanol. 15 mg of sodium bicarbonate was added to the solution mixture. Yield: 67.5 mg (0.110 mmol) (88%) HRMS/ESI+ (m/z): Calcd for C₂₄H₂₉N₂F₂[193Ir]Cl 611.1622; Found 611.1636. Anal calc. C: Calc 44.51 H: 4.58. Found C: 44.58 H: 4.67. ¹H NMR (400 MHz, DMSO-d6) 7.29 (dd, J = 8.5, 5.5 Hz, 2H), 7.22 (dd, J = 8.5, 5.5 Hz, 2H), 7.10 – 7.02 (m, 4H), 6.81 (m, 1H), 6.24 – 6.12 (m, 1H, NH), 5.58 (m, 1H, NH), 4.61 (t, J = 11.5 Hz,

1H), 4.40 (t, J = 11.5, 1H), 1.72 (s, 15H, CpMe) ppm. ¹³C NMR (101 MHz, DMSO-d6) 144.8 (aryl), 135.1 (aryl), 132.5 (aryl), 129.7 (aryl), 85.8 (CpC), 26.7 (CH), 8.95 (CpMe) ppm.

<u>Synthesis of $[(\eta^5 - C_5 Me_4 C_8 H_{17}) Ir(1S, 2S) - 1, 2 - Bis(4-</u></u>$

<u>methoxyphenyl)ethylenediamine]Cl (14-Ir)</u>. Following the general procedure, 50.0 mg (0.056 mmol) of [IrCp^{*}Cl₂]₂ was combined with 46.2 mg (0.145 mmol of (1S, 2S)-1,2-Bis(4-

methoxyphenyl)ethylenediamine dihydrochloride in a 50/50 mix of dichloromethane and methanol. 17 mg of sodium bicarbonate was added to the solution mixture. Yield 48.3 mg (0.076 mmol (67%). HRMS/ESI+ (*m/z*): Calcd for $C_{26}H_{35}N_2O_2[1931r]Cl calc: 635.2008 Found: 635.2005 ¹H NMR (400 MHz, CDCl₃) & 7.49 (d, J = 8.5 Hz, 1H), 7.38 (d, J = 8.5 Hz, 1H), 6.71 (d, J = 8.6 Hz, 1H), 6.62 (d, J = 8.6 Hz, 1H), 4.79 (m, 1H, NH), 3.67 (m, 2H), 3.55 (m, 2H), 2.01 (s, 2H), 1.88 (s, 15H, CpMe), 1.59 (s, 6H, 2CH₃) ppm. ¹³C NMR (101 MHz, CDCl₃) & 159.56 (aryl), 130.24 (aryl), 129.78 (aryl), 129.60 (aryl), 114.32 (aryl), 114.16 (aryl), 86.68 (CpC), 77.19 (CpC), 63.27 (CH), 55.11 (CH), 54.99 (CH), 9.52 (CpMe), 9.31 (CpMe) ppm.$

Synthesis of $[(\eta^5 - C_5 Me_5)Ir(\text{ethambutol})Cl][Cl]$.

Following the general procedure, 100. mg (0.125 mmol) of $[Cp*IrCl_2]_2$ was combined with 72.8 mg (0.262 mmol) Ethambutol dihydrochloride in a 50/50 mix of dichloromethane and methanol. 20 mg of sodium bicarbonate was added to the solution mixture. Yield: 67.7 mg (0.113 mmol) (45%). HRMS/ESI+ (m/z): Calcd for $C_{20}H_{39}N_2O_2[193Ir]Cl_2$ Calc 602.2018; Found 602.2022. ¹H NMR (400 MHz, D₂O) δ 4.13, (m, 4H) 3.23 (m, 2H), 2.21 (m, 2H, 2CH), 1.86 (s, 15H, CpMe), 1.21 (m, 3H) ppm.

Synthesis of $[(\eta^5-C_5H_5)Co(cis-1,2-diaminocylohexane)I]I$ (15-Co). 100. mg (0.240 mmol)of CpCo(CO)I₂ was dissolved in dichloromethane and allowed to stir for 10 minutes under nitrogen. 2.1 molar equivalents of *cis*-1,2-diaminocyclohexane was added dropwise via syringe to the solution and allowed to stir for 20 minutes under nitrogen. The solution immediately evolved CO gas from the reaction. The solution turned a clear red-brown color. The solvent and excess *cis*-1,2-diaminocyclohexane were removed using a rotary evaporator leaving a dark red-brown powder. The powder was then dissolved in deionized water and filtered on a fine frit. The filtrate was collected and the water was removed to yield 76. mg (0.208 mmol) 86%. HRMS/ESI+ (*m*/*z*): Calcd for C₁₁H₁₉N₂[Co]I 365.0033; Found 364.9924. ¹H NMR (400 MHz, methanol-*d*) δ 4.8 (s, H, Cp), 3.29 (m, 2H, 2CH), 1.79 (m, 4H, 2CH₂), 1.35 (m, 4H, 2CH₂), 1.6 (m, 4H), 1.35 (m, 4H, 2CH₂) ppm.

Following the same synthetic route as the previous cobalt complex, ±-trans-1,2-diaminocyclohexane was used to synthesize $[(n^{5}-C_{5}H_{5})Co(\pm-trans-1,2-diaminocylohexane)I]I$ (16-Co). Yield 67mg (0.184 mmol) 76% HRMS/ESI+ (*m/z*): Calcd for C₁₁H₁₉N₂[Co]I 365.0041; Found 365.0052 ¹H NMR (400 MHz, methanol-*d*) δ 5.18 (s, H, Cp), 3.41 (m, 2H, 2CH), 1.82 (m, 4H, 2CH₂), 1.42 (m, 4H, 2CH₂), 1.53 (m, 4H), 1.44 (m, 4H, 2CH₂) ppm.

Staphylococcal strains and measurement of MIC and MBC.

Staphylococcus aureus strains ATCC 6358 and ATCC 29213 were obtained from the American Type Culture Collection (ATCC), the unrelated methicillin-resistant *S. aureus* strain 43330 was obtained from Danville Community Hospital, and the recent patient isolates of MRSA (523000, 522870, 34864, 36361, 53016, and 34380) were obtained from Georgetown University Medical Center.³¹ MICs were measured by broth microdilution of fresh overnight cultures according to the Clinical and Laboratory Standards Institute (CCLI) guidelines with cation-adjusted Mueller-Hinton broth and an inoculum of 10^5 CFU/mL.³¹ Stocks of the compounds were dissolved in Mueller-Hinton broth. The MIC (μ M) was defined as the lowest concentration of compound completely inhibiting the appearance of turbidity by eye and confirmed by absorbance 540

8.

9.

11.

nm. The MBC (μ M) was defined as the lowest concentration of compound reducing the colony count by 99.9 % of the colony count in the initial, compound-free, inoculated well after 24 hr incubation. All results represent the average of three independent measurements. Both MIC and MBC experiments were also performed using brain heart infusion broth (BHIB), resulting in similar data for both MIC and MBC³².

Time - kill measurements. A single colony of the strain was inoculated into 2 mL of Mueller-Hinton broth in a 16 x 125 mm screw cap tube and incubated overnight at 37° C without aeration. One mL of that culture was inoculated into 9 mL of cation-adjusted Mueller-Hinton broth in a 125 mL flask and the culture incubated at 37° C with aeration (60 rpm) until reaching mid-log phase (2 x 10^{8} CFU/mL). A dilution of that culture was then added to 20 mL of Mueller-Hinton broth to reach a final density of 10⁵ CFU/mL. A volume of the complex stock in Mueller-Hinton broth was added to the S. aureus or MRSA suspension to reach its MIC and immediately and at 1, 3, and 6 hr, a 1 mL sample was removed and surviving CFU/mL were measured by spreading 0.1 mL (in triplicate) of undiluted, 10^{-1} , 10^{-2} , and 10^{-3} fold diluted suspensions on Mueller-Hinton agar and incubating plates at 37° C for 1 day. The results are reported as percent (%) surviving cells at each time point from two independent measurements³².

Toxicity Studies. Adapting the protocol of Babiceanu et al.³³, the iridium complexes were tested for toxicity against Vero cell line ATC CCL-81. While the cobalt complexes were not tested, the iridium compounds studied showed no toxicity at the maximum level tested of 500 μ M.

Hemolysis measurements. Hemolysis measurements were performed as described in Maisuria et al.³¹ and results reported at the lowest concentration of compound yielding 10 % hemolysis ($IC_{10\%}$).

Conclusions

The transition metal complexes described here show specific activity against S. aureus strains and lack activity against other microorganisms (i.e., Escherichia coli, Mycobacterium smegmatis, Candida albicans, and Aspergillus niger). In addition, the specificity of the complexes that show activity indicates that there are specific structure/activity relationships that must be met. This is most dramatically noted in the high activity observed for complexes of the cis isomer of 1,2-diaminocyclohexane (3-Ir and 15-Co) and the total lacke of activity of the complexes made with the trans isomer of 1,2-diaminocyclohexane (6-Ir and 16-Co). We have begun work on elucidating the mechanism of action of this novel class of anti-microbial agents and hope to report on those findings in the near future. The combination of high and specific activity for certain complexes, the low cytotoxicity as tested with Vero cells and the low hemolytic activity all suggest that this transition metal platform may prove to be useful in overcoming antibiotic resistance in S. aureus.

Acknowledgements

The authors acknowledge the guidance and technical assistance of Ms. Myra D. Williams with the preparation of cultures and measurement of MICs and MBCs and Ms. Chelsea Dollarhite with the measurement of killing and David DePena for aiding in the synthesis of compounds. They also acknowledge a grant of syringes from the Hamilton Company.

- ^a Virginia Tech Center for Drug Discovery and Development.
- ^b Department of Chemistry, Virginia Tech, Blacksburg, VA 24061, USA.

^c Department of Biological Sciences, Virginia Tech, Blacksburg, VA 24061, USA.

[†] Electronic Supplementary Information (ESI) available: A. Proton NMR spectra of the most active compounds reported in this paper, complexes **3-Ir**, **4-Ir** and **5-Ir**. B. Complete table of MICs and MBCs for all compounds and all strains of *S. aureus* and MRSA tested. See DOI: 10.1039/b000000x/

Notes and references

- R. M. Klevens, M. A. Morrison, J. Nadle, S. Petit, K. Gershman, S. Ray, L. H. Harrison, R. Lynfield, G. Dumyati and J. M. Townes, *Jama*, 2007, **298**, 1763-1771.
- M. E. De Kraker, P. G. Davey, H. Grundmann and B. S. Group, *PLoS medicine*, 2011, 8, e1001104.
- J. L. Hadler, S. Petit, M. Mandour and M. L. Cartter, *Emerging* infectious diseases, 2012, 18, 917.
- I. Naseri, R. C. Jerris and S. E. Sobol, Archives of Otolaryngology-Head & Neck Surgery, 2009, 135, 14-16.
- S. Saxena, P. Thompson, R. Birger, A. Bottle, N. Spyridis, I. Wong, A. P. Johnson, R. Gilbert and M. Sharland, *Emerging* infectious diseases, 2010, 16, 530-533.
- 6. R. Kouyos, E. Klein and B. Grenfell, *PLoS pathogens*, 2013, 9, e1003134.
- N. Balaban, O. Cirioni, A. Giacometti, R. Ghiselli, J. B. Braunstein, C. Silvestri, F. Mocchegiani, V. Saba and G. Scalise, *Antimicrobial agents and chemotherapy*, 2007, 51, 2226-2229.
 - J. C. Dabrowiak, *Metals in medicine*, John Wiley & Sons2009. Z. Liu, A. Habtemariam, A. M. Pizarro, S. A. Fletcher, A. Kisova,
 - Z. Liu, A. Habtemariam, A. M. Pizarro, S. A. Fletcher, A. Kisova, O. Vrana, L. Salassa, P. C. Bruijnincx, G. J. Clarkson, V. Brabec and P. J. Sadler, *Journal of medicinal chemistry*, 2011, **54**, 3011-3026.
- 10. Z. Liu and P. J. Sadler, Accounts of chemical research, 2014, 47, 1174-1185.
 - R. Payne, P. Govender, B. Therrien, C. M. Clavel, P. J. Dyson and G. S. Smith, *Journal of Organometallic Chemistry*, 2013, **729**, 20-27.
- A. A. Nazarov, C. G. Hartinger and P. J. Dyson, *Journal of Organometallic Chemistry*, 2014, 751, 251-260.
- T. B. Hadda, M. Akkurt, M. F. Baba, M. Daoudi, B. Bennani, A. Kerbal and Z. H. Chohan, *Journal of enzyme inhibition and medicinal chemistry*, 2009, 24, 457-463.
- 14. P. Bragg and D. Rainnie, *Canadian journal of microbiology*, 1974, **20**, 883-889.
- 15. R. Richards, *Microbios*, 1980, **31**, 83-91.
- A. Kascatan-Nebioglu, M. J. Panzner, C. A. Tessier, C. L. Cannon and W. J. Youngs, *Coordination Chemistry Reviews*, 2007, 251, 884-895.
- B. D. Wright, P. N. Shah, L. J. McDonald, M. L. Shaeffer, P. O. Wagers, M. J. Panzner, J. Smolen, J. Tagaev, C. A. Tessier, C. L. Cannon and W. J. Youngs, *Dalton transactions*, 2012, 41, 6500-6506.
- L. Oehninger, R. Rubbiani and I. Ott, *Dalton transactions*, 2013, 42, 3269-3284.
- J. R. Anacona, Journal of Coordination Chemistry, 2001, 54, 355-365.
- N. E. A. El-Gamel, Journal of Coordination Chemistry, 2009, 63, 534-543.
- D. Scutaru, L. Tataru, I. Mazilu, M. Vata, T. Lixandru and C. Simionescu, *Applied Organometallic Chemistry*, 1993, 7, 225-231.
- M. Patra, G. Gasser, M. Wenzel, K. Merz, J. E. Bandow and N. Metzler-Nolte, *Organometallics*, 2010, 29, 4312-4319.
- M. Wenzel, M. Patra, C. H. Senges, I. Ott, J. J. Stepanek, A. Pinto, P. Prochnow, C. Vuong, S. Langklotz, N. Metzler-Nolte and J. E. Bandow, ACS chemical biology, 2013, 8, 1442-1450.

- 24. M. Patra, G. Gasser and N. Metzler-Nolte, *Dalton transactions*, 2012, **41**, 6350-6358.
- G. Karpin, J. S. Merola and J. O. Falkinham, *Antimicrobial agents and chemotherapy*, 2013, AAC. 00452-00413.
- L. B. Heifets, M. Iseman and P. Lindholm-Levy, Antimicrobial agents and chemotherapy, 1986, 30, 927-932.
- D. M. Morris, M. McGeagh, D. De Peña and J. S. Merola, *Polyhedron*, 2014, 84, 120-135.
- 28. U. Koelle, B. Fuss, M. Belting and E. Raabe, *Organometallics*, 1986, **5**, 980-987.
- 29. M. Ehrich, personal communication.
- 30. J. S. Merola, D. Morris and N. De Weerd, *Acta Crystallographica Section E: Structure Reports Online*, 2013, **69**, m176-m176.
- B. B. Maisuria, M. L. Actis, S. N. Hardrict, J. O. Falkinham Iii, M. F. Cole, R. L. Cihlar, S. M. Peters, R. V. Macri, E. W. Sugandhi, A. A. Williams, M. A. Poppe, A. R. Esker and R. D. Gandour, *Bioorganic & Medicinal Chemistry*, 2011, 19, 2918-2926.
- K. Li, L. A. Schurig-Briccio, X. Feng, A. Upadhyay, V. Pujari, B. Lechartier, F. L. Fontes, H. Yang, G. Rao, W. Zhu, A. Gulati, J. H. No, G. Cintra, S. Bogue, Y. L. Liu, K. Molohon, P. Orlean, D. A. Mitchell, L. Freitas-Junior, F. Ren, H. Sun, T. Jiang, Y. Li, R. T. Guo, S. T. Cole, R. B. Gennis, D. C. Crick and E. Oldfield, *Journal of medicinal chemistry*, 2014, 57, 3126-3139.
- M. C. Babiceanu, B. A. Howard, A. C. Rumore, H. Kita and C. B. Lawrence, *Frontiers in Microbiology*, 2013, 4, 196.