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1 Valle Agricola lentil, an unknown lentil (*Lens culinaris* Medik.) seed from Southern
2 Italy as a novel antioxidant and prebiotic source

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18

19 **Abstract**

20

21 In order to promote 'Valle Agricola' lentil, an autochthonous lentil of the Campania Region,
22 a thorough investigation on its biochemical and nutritional properties has been carried out.
23 Macronutrient content (proteins, carbohydrates and lipids), free and total amino acids, and
24 unsaturated fatty acids, were determined. The antioxidant capability of raw 'Valle Agricola' lentils,
25 as well as of boiled ones, was estimated in terms of their total phenol content (TPC), ORAC value,
26 and free radical scavenging capacities using DPPH and ABTS assays. Data obtained evidenced that
27 boiling process slightly decreased Valle Agricola lentils antioxidant power. Furthermore, when
28 trypsin and chymotrypsin inhibitory activities were measured, an high decrease of the levels of anti-
29 nutritional factors, were estimated. In order to have a phytochemical overview of this
30 autochthonous lentil seed, LC-ESI-MS/MS analyses was applied on raw and boiled lentil extracts.
31 Flavonols glycosides and free flavanols, as well as typical seed prebiotic saccharides, were the most
32 representative constituents.

33

34 **Keywords:** Amino acids, Anti-nutritional factors, Antioxidant activity, *Lens culinaris* Medik,
35 Nutritional value

36

37 1. Introduction

38

39 Exponential growth of the human population has led to serious environmental changes with
40 progressive alteration of the natural environment and biodiversity loss. The development of
41 chemical-industrial agriculture, and the tendency to uniform farming on productive and rewarding
42 species have condemned the provision favouring the subsequent extinction of all landraces not
43 producing adequate income ¹. Mediterranean area seems to be only partially affected. In fact, it still
44 represents one of the largest sources of wild plant cultivars, many of which are locally cultivated
45 and consumed as traditional food ². In different Italian regions, wild plants are expression of their
46 cultures and are traditionally used for the preparation of “homemade” products, essentially destined
47 to the local market ³⁻⁶. In order to preserve the precious genetic variability of Italian local cultivars,
48 their biochemical and nutritional valorisation has to be pursued. The knowledge of nutritional value
49 of local plant species, and of the beneficial effects on human health arising from their consumption,
50 could be a valid approach to support their cultivation and marketing ⁷.

51 In particular, it is interesting preserve the production of local legume species as the lentils. Lentil
52 (*Lens culinaris* Medik.) is a nutritious pulse food whose cultivars are characterized by seeds that differ
53 for color (range from yellow to red-orange to green, brown and black), size (*microsperma* or
54 *macrosperma*) and form ⁸. Generally consumed boiled as soup or processed into flour ⁹, lentils are
55 reported to contain a large amount of inexpensive and accessible high quality proteins and is a well-
56 balanced source of essential amino acids, carbohydrates, fibre, minerals, and vitamins ¹⁰. Furthermore,
57 as they are poorly rich in fats, and contain bioactive natural antioxidants, they are recognized an ideal
58 complement to cereals in vegetarian diet and a functional food with a positive healthy impact ^{11, 12}.

59 Italy is broadly known for its production of some lentil cultivars which are mainly cultivated in
60 marginal areas of Central and Southern Italy. Among them, ‘Castelluccio di Norcia’ (Protected
61 Geographic Indication; EC Reg. n°. 1065/97) and ‘Colfiorito’ (Umbria Region), ‘Onano’ (Lazio),
62 ‘Fucino’ (Abruzzo), ‘Altamura’ (Puglia); ‘Mormanno’ (Calabria), ‘Pantelleria’ and ‘Villalta’

63 (Sicilia) are renowned at international level¹³. Indeed, several other lentil cultivars are grown in
64 Italy. In Campania region, for instance, lentil cultivars as ‘San Geraldo’ (Avellino), ‘Colliano’
65 (Salerno) and ‘Valle Agricola’ (Caserta) are locally produced and consumed.

66 ‘Valle Agricola’ lentil is so called as it is from Valle Agricola, a small town localized, near Caserta,
67 in the piedmont part of the Matese Mountain chain. It is a local ecotype, belonging to microsperma
68 varieties, characterized by medium dimensions and a rather dark colour, with a thin skin and a
69 characteristic flavour. Produced in low quantities through family-run terraces cultivation, these
70 high-quality lentils are stored in large weave canvas bags, and sold mainly on the local market.

71 In order to valorize ‘Valle Agricola’ lentil and to encourage its consume at local and international
72 levels, an extensive nutritional and biochemical investigation aimed to define its macronutrient
73 content has been carried out (**Fig. 1**). Antioxidant capability of raw and boiled ‘Valle Agricola’
74 lentil material was estimated in terms of total phenol content and ORAC value of extracts properly
75 prepared. As lentils are known to contain some anti-nutritional factors which could limit their
76 consumption, trypsin and chymotrypsin inhibitory activities were also performed. LC-MS-based
77 metabolic profile analyses were also performed on extracts from raw and boiled lentil seeds.

78

79

80 **2. Experimental**

81

82 **2.1. Chemicals and reagents**

83 2,2'-azobis-(2-amidinopropane)-dihydrochloride (AAPH), *nor*-leucine (nor-Leu), fluorescein,
84 Folin-Ciocalteu reagent, gallic acid, hydrogen peroxide, peroxidase from horseradish, salts and 6-
85 hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (Trolox[®]) were purchased from Sigma-
86 Aldrich (Milan, Italy). Solvents were from Sigma-Aldrich. Chemicals and solvents for the Kjeldahl
87 method were from Carlo Erba Reagents (Milan, Italy), whereas those for automated amino acid
88 analysis were provided from Biochrom (Cambridge, U.K.). Fatty acid methyl esters (Supelco[™] 37
89 component FAME mix) were obtained from Supelco (Park-Bellefonte, PA, USA).

90

91 **2.2. Plant material and sampling**

92 *Lens culinaris* seeds were grown under typical soil, and illumination conditions in Valle Agricola,
93 71 km Northwest from Caserta, Italy (geographical coordinates: 41°25'33"60 N, 14°15'21"24 E).
94 Lentil plants (~40 cm tall), manually and randomly harvested across the reference field on July
95 2013, were sun-dried for one week. Dried lentil seeds were powdered with the Cyclone Sample Mill
96 Instrument (PBI International, Milan, Italy), until flour of homogeneous size was obtained. All
97 measurements were performed by triplicate.

98

99 **2.3. Ash and moisture content**

100 Ash content and moisture level were determined according to the AOAC official method ¹⁴.

101

102 **2.4. Macronutrient content.**

103 *Total protein content:* Nitrogen concentration was obtained by the Kjeldahl method ¹⁴ and total
104 protein content was estimated using a nitrogen factor of 6.25. Samples (~1.0 g each) were analysed

105 using a Mineral Six digester and an Auto Disteam semi-automatic distilling unit (VWR
106 International PBI, Milan, Italy). *Total lipid content*: Aliquots (2.0 g) of frozen lentil flour sample
107 were lyophilized using a FTS-System Flex-Dry™ instrument (SP Scientific, Stone Ridge, NY
108 USA). Thus, the freeze-dried materials were extracted for 4 h through a Soxhlet apparatus using
109 CHCl₃ as extracting solvent. The solutions obtained were dried by a rotary evaporator in order to
110 provide the crude fat extracts. The lipid content was determined gravimetrically. *Total carbohydrate*
111 *content*: Total carbohydrates were obtained by subtracting (moisture + crude protein + crude fat +
112 ash + total polyphenol + free sugar) from 100¹⁵.

113

114 **2.5. Amino acids composition**

115 For the analysis of free amino acid composition, three aliquots of ~200 mg of seed flour were
116 precipitated with 80% cold ethanol (1.0 mL), in the presence of *nor*-Leu (50.0 nmol) as internal
117 standard, homogenized with a teflon pestle and centrifuged at 14000 ×g, at 4 °C. The supernatants
118 were lyophilized, treated with 3% sulfosalicylic acid (500 µL) to precipitate any protein fraction
119 still present, and centrifuged again¹⁶. Thus, the supernatants obtained were analysed. For the
120 analysis of total (free and protein) amino acids, ~10 mg of seed flour were hydrolysed with 0.5 mL
121 of 6 N HCl containing 0.02% phenol and *nor*-Leu as internal standard at 110 °C for 20 h⁴.
122 Following hydrolysis, HCl was removed under vacuum and samples were re-suspended in 0.5 mL
123 of 0.2 M lithium citrate buffer (pH 2.2). Aliquots of hydrolysed and non-hydrolysed samples were
124 directly analysed on a Biochrom 30 amino acid analyser (Biochrom, Cambridge, U.K.), equipped
125 with a post-column ninhydrin derivatization system.

126

127 **2.6. Gas chromatographic analysis of fatty acid methyl esters**

128 The analysis of fatty acid methyl esters content were performed as previously reported⁴. Briefly,
129 each CHCl₃ crude extract (1.0 mg; see “Macronutrient content” section) was dissolved in 2.0 mL of
130 2 N KOH in MeOH. The solution was stirred for 30 min at room temperature, added with heptanes

131 (0.8 mL), vortexed and then centrifuged, using a Beckman GS-15R centrifuge (Beckman Coulter
132 Italia), for 10 min at 4200 $\times g$. Organic upper phase (1.0 μL) was analysed by GC (100 m \times 0.25
133 mm i.d., 0.2 μm SP2380, fused silica capillary column; Supelco, Sigma-Aldrich). The fatty acid
134 methyl esters were identified by comparing their retention times with those of the standard fatty
135 acid methyl esters (Grain Fatty Acid Methyl Ester mix 47801, Supelco).

136

137 **2.7. Free sugar (pentoses and hexoses) content**

138 For the analysis of monosaccharide composition, three aliquots of about 50 mg of seed flour were
139 precipitated with 80% cold ethanol (1.0 mL) homogenized with a teflon pestle and centrifuged at
140 14000 $\times g$, at 4 $^{\circ}\text{C}$. Thus, the supernatants obtained were determined using a Bio-LC[®] (Dionex
141 Corp., Sunnyvale, CA, USA) equipped with a CarboPac[®] PA10 column (2 \times 250 mm, Dionex
142 Milan, Italy) and a guard column Amino Trap[™] (2 \times 50 mm, Dionex). Protocols employed were as
143 suggested by the manufacturer (Technical Note 40 by Dionex Corp.).

144

145 **2.8. Evaluation of antioxidant capability of ‘Valle Agricola’ lentil flour**

146 Total phenol content (TPC) and antioxidant capability of ‘Valle Agricola’ lentil flour were
147 measured on methanol extracts, previously obtained through ultrasound-assisted maceration.

148 To this purpose three replicate lentil flour samples (2.0 g each) underwent ultrasound assisted
149 extraction (Dr. Hielesher UP 200S, Germany) using pure methanol as extracting solvent (50 mL for
150 each sample). Four sonication cycles were performed (30 min each) in order to achieve the
151 maximum recovery of the lentil flour metabolic content. At the end of each sonication cycle,
152 samples were centrifuged at 2,044 $\times g$ for 10 min in a Beckman GS-15R centrifuge (Beckman
153 Coulter, Milano, Italy) fitted with rotor S4180. Obtained supernatants were dried under vacuum by
154 a rotary evaporator (Heidolph Hei-VAP Advantage, Germany) to yield crude extracts, which were
155 stored at -20°C until use.

156 Furthermore, in order to evaluate the effect of boiling (the most used cooking method for lentils
157 consume) on phenol content and antioxidant capacity, three replicate samples (2.0 g each) of raw
158 material was first boiled in water (50 mL, each). After 30 min, the extracts were centrifuged at 4500
159 $\times g$. Cooking water (supernatant) was directly investigated for its TPC value and antioxidant power,
160 whereas the pellet underwent an extraction with methanol, as described above. Folin-Ciocalteu,
161 ABTS, DPPH, and ORAC assays were carried out performing three replicate measurements for
162 three samples (n=3) of each extract. Recorded activities were compared to a blank. Results are the
163 mean \pm SD values. Student *t*-test was applied in order to determine statistical significance
164 (significance level was set at $P < 0.05$).

165

166 **2.9. Total phenol content determination**

167 Total phenol amount of 'Valle Agricola' lentil flour was determined according to the Folin-
168 Ciocalteu procedure on aliquots of extracts in methanol. The content of total phenols (TPC value)
169 of the samples was expressed as mg gallic acid equivalents (GAE) per g of seed flour.

170

171 **2.10. Determination of ABTS radical cation scavenging capacity**

172 The determination of ABTS⁺ solution scavenging capacity was estimated as previously reported⁵.
173 The results were expressed in terms of TEAC values ($\mu\text{mol Trolox}^{\text{®}}$ Equivalents per g of extract).

174

175 **2.11. Determination of DPPH radical scavenging capacity**

176 The determination of DPPH' (2,2-diphenyl-1-picrylhydrazyl) scavenging capability was estimated
177 as previously reported⁵. The results were expressed in terms of TEAC values ($\mu\text{mol Trolox}^{\text{®}}$
178 Equivalents per g of extract).

179

180

181

182 **2.12. Oxygen radical absorbance capacity assay (ORAC)**

183 The antioxidant potential of 'Valle Agricola' lentil flour was measured by Oxygen Radical
184 Absorbance capacity (ORAC) assay. The analysis was performed using 96-well microplates in
185 which 25 μL of appropriately diluted methanolic extract from raw and cooked lentils, as well as of
186 extract from lentil boiling water, were mixed with fluorescein (FL) probe [150.0 μL (80 nM, final
187 concentration) in 7.5 mM Na-phosphate buffer, pH 7.0 (Na/P)] and preincubated for 10 minutes at
188 37 $^{\circ}\text{C}$. Then, 2,2'-azobis-(2-amidinopropane)-dihydrochloride (AAPH) solution [25.0 μL (18.4
189 mM, final concentration)] was rapidly added. In parallel with the test samples, a blank (25 μL Na/P,
190 150 μL FL and 25 μL AAPH) and solutions of the reference antioxidant Trolox[®] (6.25-50 μM , final
191 concentration levels) were properly prepared in Na/P with 150 μL FL and 25 μL AAPH. The
192 fluorescence decay ($\lambda_{\text{ex}} = 485 \text{ nm}$, $\lambda_{\text{em}} = 525 \text{ nm}$) was recorded every minute for 75 min using a
193 Tecan SpectraFluor fluorescence and absorbance reader. Antioxidant curves (fluorescence vs. time)
194 were normalized to the curve of the blank. From the normalized curves, the area under the
195 fluorescence decay curve (AUC) was calculated. Linear regression equations between net AUC
196 ($\text{AUC}_{\text{antioxidant}} - \text{AUC}_{\text{blank}}$) and antioxidant concentration were calculated. The antioxidant activity
197 (ORAC value) was calculated by using the Trolox[®] calibration curve. The ORAC values were
198 expressed as Trolox[®] equivalents ($\mu\text{mol per g}$).

199

200 **2.13. Determination of Relative Antioxidant Activity Index (RACI)**

201 As the units and the scale of the data from various chemical methods are different, the data in each
202 data set should be transformed into standard scores, a dimensionless quantity derived by subtracting
203 the mean from the raw data divided by the standard deviation, according to the equation standard
204 score = $(x - \mu)/\sigma$, where x represents the raw data, μ the mean, and σ the standard deviation. The
205 standard scores of a sample for different assays when averaged give a single unit less value termed
206 as RACI, which is a specific combination of data from different chemical methods with no unit
207 limitation and no variance among methods¹⁷.

208

209 2.14. Determination of trypsin and chymotrypsin inhibitory activities

210 Three replicate samples of 'Valle Agricola' lentil flour (1.0 g each) were extracted overnight under
211 magnetic stirring at 4 °C in 80 mM Tris•Cl, pH 7.8, containing 0.1 M CaCl₂ (1:4; w/v). The
212 reaction mixtures were centrifuged at 24000 ×g (Centrifuge Avant J-25, Beckman Coulter, CA,
213 USA) at 4 °C for 60 min. The supernatants were first filtered on Miracloth (pore size: 22-25 μm)
214 and then freeze-dried.

215 The protein concentration was determined with the Bio-Rad Protein Assay kit following the
216 manufacturer's instructions and using bovine serum albumin (BSA) as standard.

217 Trypsin and anti-trypsin activities, as well as chymotrypsin and anti-chymotrypsin activities, were
218 determined following a procedure already described¹⁸, using TAME (*p*-toluenesulfonyl-L-arginine
219 methyl ester) and BTEE (N-benzoyl-L-Tyrosine ethyl ester) as substrates, respectively. One
220 inhibitory unit is defined as the amount of inhibitor that, under the assay conditions, yields a 30%
221 decrease of the assayed enzymatic activity. Increasing concentration of lentil protein extract was
222 added to a fixed concentration of enzyme that in our condition corresponding to 0.05
223 absorbance/min monitoring for 5 min the reaction. Residual enzyme activities are expressed as
224 percentage of the control samples (enzyme activity in the absence of protein extract extracted). The
225 IC₅₀ values on trypsin and α-chymotrypsin activities (i.e. the half maximal inhibitory concentration)
226 of the lentil protein extract were calculated as previously reported¹⁸.

227

228 2.15. LC and LC-MS/MS analyses

229 In order to provide a preliminary phytochemical overview of 'Valle Agricola' lentil seeds, LC-ESI-
230 MS/MS analysis were applied on raw and boiled lentil methanolic extracts (see section 2.8).

231 For the LC/MS analysis, a Waters 2695 LC system, equipped with a Waters 2487 dual λ absorbance
232 detector, was coupled to a Quattro Micro (Micromass, Manchester, UK) triple-quadrupole
233 instrument.

234 A Phenomenex[®] Gemini C18 column (3.0 μm particle size, 150 mm \times 2.00 mm) together with a
235 pre-column was used for chromatographic separation. The mobile phase consisted of A: 0.1%
236 formic acid in water and B: acetonitrile. Starting with 95% A, a linear gradient was followed to
237 85% A in 10 min, then decreasing to 75% A at 30 min, and to 20% A at 40 min, then returning to
238 starting conditions and allowing to re-equilibrate for 5 min. The total analysis time was 46 min, the
239 flow rate was 0.3 ml/min, the detection wavelength was set at 280 nm and the injection volume was
240 5.0 μL . Electrospray ionization (ESI) mass spectra were acquired in the negative ion mode.
241 Nitrogen was used as the nebulization and desolvation gas at flow rates of 50 and 500 l/h,
242 respectively. The source and desolvation temperatures were set at 120 and 500°C. The applied
243 potentials on the electrospray capillary and on the cone ranged from 2.5 to 3.5 kV and from 15 to 55
244 V, respectively, and were optimized for each molecule. CAD mass spectra were obtained by
245 introducing argon as a collision gas into the radiofrequency (Rf)-only quadrupole. The Ar pressure
246 was kept low ($\sim 3.0 \times 10^{-3}$ mbar) to minimize multiple collisions. Collision energy was varied from
247 10 to 45 eV and optimized for each molecule. Data acquisition and processing were carried out
248 using the software MassLynx[™] version 4.0 supplied with the instrument. Full scan mass spectra
249 were acquired from m/z 50 to 1500 in MS and from m/z 20 to the m/z value of the precursor ion in
250 MS/MS experiments, with a scan time and an interscan time of 0.2 and 0.1 s, respectively.

251

252 **2.16. Statistical analysis**

253 Analyses were repeated three times for each sample; mean and standard deviation (SD) of
254 experimental values are reported. Data analysis was carried out with Excel Office 2010 (Microsoft
255 Corporation, Redmond, WA, USA). The IC_{50} values were calculated on the basis of inhibition
256 curves by plotting the residual enzyme activities versus different concentrations of protein extract
257 by fitting data with a non-linear regression analysis on a semi-logarithm scale by using the
258 GraphPad Prism 5 software (GraphPad software Inc., California, USA). All measurements were
259 performed in triplicate with standard deviations always below 5% for each experiment

260 **3. Results and discussion**

261

262 **3.1. Nutritional values**

263 Nutritional values of *L. culinaris* seeds from ‘Valle Agricola’ are reported in **Table 1** and compared
264 with values reported by the Istituto Nazionale di Ricerca per gli Alimenti e la Nutrizione (INRAN)
265 for Italian raw lentil (hereafter INRAN lentil). The protein content of ‘Valle Agricola’ lentil (26.3
266 g/100 g) was higher than INRAN lentil (22.7 g/100 g) while lipids (2.5 g/100 g) was 2.5-fold than
267 INRAN lentil (1.0 g/100 g). Our results showed that protein content of ‘Valle Agricola’ lentil are
268 similar to Colfiorito (27.1 g/100 g), Colliano (26.0 g/100 g) and Altamira (25.5 g/100 g) cultivars¹⁹.
269 In addition, the lipids content was higher than that the above cited cultivars. However, it was in
270 accordance with the previously reported range 0.3-3.5 g/100 g²⁰ confirming that lentil are a low
271 source of lipids. The content of carbohydrates is 56.72% (range 43.3-74.9 g/100 g published for
272 other lentils²⁰), slightly higher than that reported for INRAN lentil (about 1.11-fold). The amounts
273 of saccharose, glucose and fructose were 81.6 mg/100 g, 11.4 mg/100 g and 1.0 mg/100 g,
274 respectively, with a glucose/fructose ratio of 11.63. The low rate of free sucrose, as well as of its
275 hexose components, allowed us to hypothesize that the carbohydrate amount consisted mainly in
276 resistant starch (RS), dietary fiber (DF) and oligosaccharides, which are reported as the main pulse
277 carbohydrate fractions²¹. Peculiarly, lentils are known to contain galactooligosaccharide (GOS),
278 with prebiotic effects, which, primarily via bifidogenesis, include increased calcium absorption and
279 pathogen reduction²².

280

281 **3.2. Amino acid content**

282 The content of free and total (free plus protein) amino acids is reported in **Table 2**. Comparing the
283 total amino acid content of ‘Valle Agricola’ lentil with INRAN one, both qualitative and

284 quantitative differences were found. The total amino acid content for 100 g product of ‘Valle
285 Agricola’ was about 1.17-fold that that of INRAN lentil (24.8 and 21.2 g, respectively).

286 Glx (glutamic acid + glutamine) was the most abundant among total amino acids (about 18% for
287 both ‘Valle Agricola’ lentil and INRAN lentil). Among the other amino acids in ‘Valle Agricola’
288 lentil, abundant, in decreasing order, were Asx (aspartic acid + asparagine) (12.5%), arginine
289 (8.17%), leucine (8.05%) and serine (7.12%), overall accounting for about 35% of the total. In
290 addition, the amount of essential amino acids [His, Ile, Leu, Lys, Met, Phe, Thr, Val; Trp
291 (tryptophan is not included as it was not determined in the total hydrolysed samples – see Table 2)]
292 in ‘Valle Agricola’ lentil was 8.9 g/100 g vs. 8.0 g/100 g in INRAN lentil. The amount of both
293 methionine and cysteine (as cysteic acids) in ‘Valle Agricola’ lentil did not exceed 0.45 g/100 g of
294 product represented 1.81% of total amino acids confirming the low content of sulphur containing
295 amino acids as reported for other legumes.

296 Considering free amino acid, the total amount in ‘Valle Agricola’ lentil was 147.6 mg/100 g. The
297 analysis evidenced the presence of several non-protein amino acids. Asparagine was by far the most
298 abundant among free amino acids (about 30%). Furthermore, α -amino adipic acid (15.8 mg/100g),
299 glutamic acid (8.9 mg/100g) and serine (8.1 mg/100g) were among the free amino acids the most
300 abundant in ‘Valle Agricola’ lentil, whereas the amount of each of the other non-protein amino
301 acids (carnitine, γ -amino-*n*-butyric acid, hydroxylysine, hydroxyproline, 1-methylhistidine, *o*-
302 phospho-serine, sarcosine and taurine) did not exceed 23 mg/100 g of product (13.6% total content
303 of free amino acid). Furthermore, the amount of free essential amino acids, His, Ile, Leu, Lys, Met,
304 Phe, Thr, Trp, and Val, in ‘Valle Agricola’ lentil was 12.6 mg/100 g vs. 8.9 g/100 g (not significant
305 as contribution in a human diet).

306

307 **3.3. Fatty acid composition analysis**

308 GC-MS analysis showed that ‘Valle Agricola’ lentils have relatively low fat. In particular, the fatty
309 acid composition was more rich in unsaturated acid content (301 g/100 g; 73.6%) than in saturated

310 one (108 g/100 g; 26.4%; **Table 3**). The most abundant unsaturated acids were oleic (C18:1 c) and
311 eicosenoic (C20:1) acids, which represented 34.3% and 17.4% respectively, of total fatty acids,
312 whereas no traces of linolenic acid were detected. Indeed, the presence of α -linolenic acid was
313 reported to be related to lentil germination and it achieved high value in lentil sprout²³. Erucic acid,
314 a monosaturated ω -9 fatty acid, was also identified in appreciable level (34.5 mg/100 g). The
315 presence of this latter fatty acid, was previously reported as minor fatty acid constituent of red lentil
316 seeds²⁴. Dietary erucic acid (13-*cis*-docosenoic acid) is used in the management of the
317 adrenoleukodystrophy as a glyceryl trierucate from which is commonly called as Lorenzo's oil²⁵.

318

319 **3.4. Anti-proteinase inhibitor activity**

320 Lentils are a significant dietary source of protease inhibitors such as trypsin- and chymotrypsin
321 inhibitors that are present at higher concentrations in lentils seeds which certainly interfere with
322 protein digestion²⁶. In this framework, the anti-trypsin as well as anti-chymotrypsin assays were
323 carried out on protein extract (~22.5 mg/ g of seeds) obtained from 'Valle Agricola' lentil flour and
324 the inhibitory activity was reported as IC₅₀ value. The anti-chymotrypsin activities of 'Valle
325 Agricola' lentil (IC₅₀ 4.6) were 2-fold higher than the anti-trypsin activities (IC₅₀ 8.9), while not
326 protease activities were detected in same experimental conditions. On the other hand, to confirm the
327 decreasing effect of boiling process on trypsin inhibitor content²⁷, aliquots of 'Valle Agricola'
328 lentil seeds were boiled (30 min) and the anti-protease activity re-assayed on protein extract (~2.0
329 mg/ g of boiled seeds). Overall, these results indicated that the anti-chymotrypsin as well as anti-
330 trypsin activities were 10.45-fold (IC₅₀ 48.3) and 6.12-fold (IC₅₀ 54.4) lower than the detected in
331 raw lentil seeds, respectively. Therefore, boiling process, as previously reported for other legumes,
332 reduces the antiprotease activities of 'Valle Agricola' lentil seeds and any their adverse effects in
333 humans²⁸.

334

335 **3.5. Total phenol content and antioxidant capacity of raw and boiled 'Valle Agricola' lentils**

336 In order to determine the total polyphenol content (TPC) of ‘Valle Agricola’ lentil seeds, they were
337 extracted through ultrasound assisted maceration using methanol as extractant. The calculated TPC
338 value was compared to that obtained from an equal amount of lentils, previously boiled in water for
339 30 min, as in the common culinary tradition. To this purpose, lentils, once cooked, were filtered and
340 extracted by ultrasound-assisted maceration in methanol. The polyphenol content of boiling water
341 was also measured. TPC value of raw lentils, 23.1 ± 0.4 mg GAE per g, was slightly higher than that
342 of cooked lentils (20.3 ± 0.5 mg GAE per g), whereas a poor amount of phenol substances was
343 released in boiling water samples (2.8 ± 0.2 mg GAE per g). Thus, the effect of conventional boiling
344 treatment seemed to influence only weakly the ‘Valle Agricola’ lentil polyphenol content. This
345 finding was in line with Aguilera and co-authors²⁹, who evidenced that thermal processing of the
346 ‘Pardina’ lentil flour apparently did not affect its total phenol content and antioxidant activity.
347 Indeed, performing HPLC-PAC and HPLC-MS (ESI) analysis on raw and processed materials, they
348 found that catechins and procyanidins, flavonols, flavones, and flavanones decreased, while
349 hydroxybenzoic compounds exhibited an important increase. On the contrary, Xu and coauthors³⁰
350 reported that different four thermal processing methods (conventional boiling, conventional
351 steaming, pressure boiling, and pressure steaming) caused significant reduction in total phenolic
352 content, antioxidant capacities, and antiproliferation properties of lentils and other commonly
353 consumed cool-season food legumes. Han and Baik⁹ reported that TPC of lentils was higher than
354 that of chickpeas, peas and soybeans; it was only ~ 12 mg per g for cv. ‘Pardina’ and ‘Crimson’.
355 According to Kalogeropoulos and coauthors³¹, who reported that lentils score the highest TPC
356 values among 14 different types and varieties of pulses, ‘Valle Agricola’ raw lentil seeds had a TPC
357 value of about 23.1 mg GAE per g of material.

358 The evaluation of antiradical properties of the three extracts, carried out performing DPPH,
359 ABTS methods, as well as ORAC one, revealed that boiling treatment slightly decreased ‘Valle
360 Agricola’ lentils free radical scavenging efficacy (**Fig. 2**). Data obtained from ABTS, DPPH,
361 ORAC, and Folin-Ciocalteu reagent (FCR) tests were used to calculate RACI (*Relative*

362 *Antioxidant Capacity Index*) value for each extract. The data obtained by applying ABTS method
363 positively correlated to those from DPPH method (0.977), ORAC assay (R = 0.997), and to TPC
364 value (0.907). Similarly, a positive correlation (**Table 4A**) was observed between the DPPH data
365 and those of ORAC (R = 0.991) and TPC data, R = 0.976). RACI calculation represents the average
366 of the standard scores obtained from the raw data for the various methods (**Table 4B**). The results
367 emphasized that the raw lentils were able to display an antioxidant capability (RACI 0.716) slightly
368 stronger than cooked lentils (RACI 0.617), whereas cooking water did not show antioxidant effects
369 (RACI -1.151). As polyphenols are considered the main secondary metabolites affecting the
370 antioxidant power of plant extracts, data obtained was explained assuming the great amount of these
371 compounds in the vacuoles. In fact, although flavonoid glycosides are drastically reduced by
372 thermal processing, this latter could trigger the release of more bound phenolic acids during
373 breakdown of cellular constituents and membranes¹², and promote the conversion of glycosides
374 constituents, mainly present at lentil seed coat level, in their relative free aglycones. The high
375 carbohydrate content in “Valle Agricola” lentil seeds could be also involved because these
376 compounds, commonly referred as dietary fiber constituents, are able to act as carrier of dietary
377 antioxidants³². The previously reported increase in metal chelating activity of cooked lentils seeds³³
378 was, for instance, attributed to the formation of Maillard browning pigments by interaction of
379 carbohydrates (especially reducing sugars) and proteins during cooking. Furthermore, Agil *et al.*
380 (2013) found that the polysaccharide lentil fraction exhibited a strong antioxidant potential³⁴.

381

382 **3.6. LC-MS-based metabolite profile of ‘Valle Agricola’ lentil seed extracts**

383 To identify the main constituents in ‘Valle Agricola’ lentil seeds, LC-ESI-MS/MS analyses of raw
384 and boiled extracts, prepared by ultrasound assisted maceration, were carried out. The identified
385 metabolites are listed in **Table 5**. Compounds **1-5** were tentatively identified as oligosaccharides. In
386 particular, compound **1** showed the $[M-H]^-$ ion at m/z 827 and MS^2 fragments ions at m/z 665, 503,
387 341 and 179, which were formed by cleavage of the glycosidic bond with the loss of one, two, three

388 and four hexose moieties (-162 Da), respectively. Compound **2**, characterized by a $[M-H]^-$ ion at
389 m/z 517, was hypothesized to be a dihexosyl cyclitol, likely ciceritol³⁵. Its MS² spectrum provided
390 the fragment ion at m/z 193, due to the neutral loss of two hexose residues (-324 Da). This
391 compound, together with other galactosides of myo-inositol isomers, are broadly accumulated in
392 legume species seeds³⁶. The accumulation of galatosyl cyclitols in seeds starts at the stage of pod
393 filling and increases during seed maturation and desiccation steps³⁷. Compounds **3-5** were
394 putatively identified as 6-carbon tetrasaccharide, trisaccharide and disaccharide, respectively. In
395 particular they could be α -Galactosides of the raffinose family (RFO), which were found as the
396 most abundant oligosaccharides in lentil seeds. The raffinose series oligosaccharides contents in
397 legumes are known to increase with maturation progress³⁸ and they have been also related to the
398 ability of legumes to survive dry periods³⁹. All these compounds are referred as prebiotic
399 carbohydrates that are important components of healthy diets, supporting healthful hindgut
400 microflora. Prebiotic concentrations in lentils appear to be related to genetic and environmental
401 factors⁴⁰. When boiling process, it was found that cooking water was particularly enriched in these
402 compounds, on the basis of their huge solubility in water⁴¹.

403 Flavonol glycosides were abundantly present. Compounds **6**, **9**, **11**, **13** and **14** were identified as
404 kaempferol glycosides. In particular, the MSⁿ spectra of compound **6**, which showed its $[M-H]^-$ ion
405 at m/z 901, were in accordance with the presence of a kaempferol tetraglycoside. This compound,
406 together with both the detected kaempferol triglycoside isomers (**9** and **11**) were previously
407 identified as a component of lentil seeds and aerial parts^{42, 43}. The MS² spectrum of the deprotonated
408 metabolites **9** and **11** at m/z 755 showed fragment ions at m/z 593 $[M-H-162]^-$, 285 (kaempferol
409 aglycone) and 284 (radical kaempferol)⁴⁴. Compounds **13** and **14** were tentatively identified as
410 hexosyl and pentosyl kaempferol, respectively. The collision of the $[M-H]^-$ ion (m/z 447 for **13** and
411 417 for **14**) provided a radical aglycone at m/z 284, as well as the $[aglycone-H]^-$ ion at m/z 285,
412 attributable to kaempferol. Metabolites **7** and **8** were identified as quercetin derivatives. The
413 MS/MS analyses allowed us to detect neutral losses of 162 and 176 Da, which were characteristic

414 of the presence of a residue of hexose, and hexuronic acid, respectively. The ion at m/z 301 in both
415 the cases was found to be the base peak. In raw lentil extract, pure quercetin (**17**) was also identified
416 on the basis of the comparison of its retention time and mass spectra with those of a standard pure
417 compound. ‘Valle Agricola’ flavanol constituents were represented by catechin (**14**) and epicatechin
418 (**15**), as well as by their derivatives characterized by a galloyl B-ring (**21** and **22**). Compounds **10**
419 and **12** likely corresponded to procyanidin dimers. In fact, both the metabolites showed a
420 deprotonated molecular ion at m/z 577, which after collision-activated fragmentation provided the
421 fragment ion at m/z 289, which, in turn, could correspond to either catechin or epicatechin⁴⁵. The
422 MS/MS spectra of compound **18** gave the fragment ion at m/z 179, suggesting the presence of a
423 caffeoyl moiety in this compound. The neutral loss of 162 Da allowed us to tentatively identify it as
424 a caffeoyl hexose. Compound **20** likely corresponded to ferulic acid, whereas compound **19** ($[M-$
425 $H]^-$ at m/z 339) could be its deoxyhexosyl derivative.

426

427

428 **Conclusions and perspectives**

429

430 'Valle Agricola' lentil was particularly rich in proteins and low in lipids. These nutritional
431 features make it a good food for diets low in energy content. All protein amino acids were
432 identified, and the quali-quantitative analysis of free amino acids was reported for the first time.
433 Raw lentil seeds contained an appreciable amount of polyphenol compounds and were able to exert
434 a significant antioxidant capability. When boiling process was applied, 'Valle Agricola' lentil
435 antioxidant power was retained suggesting that the regular consumption of this product could
436 maximize the dietary intake of health beneficial compounds with protective or disease preventive
437 properties. Indeed, thermal process applied increased the wholesomeness of the 'Valle Agricola'
438 lentils as a strong decrease of trypsin and trypsin-chymotrypsin inhibitors, common legumes anti-
439 nutrients, was detected. Overall, our data showed that 'Valle Agricola' microsperma lentils are a
440 good source of nutrients and bioactive compounds able to provide health benefits.

441

442

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444

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449 *monitoraggio del rischio ambientale*', AMRA (P. O. R. 2000–2006, misura 3.16).

450

451 **Conflict of interest**

452 The authors have no conflicts of interest

453

454

455 **References**

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533

534 **Figure captions**

535

536 **Fig. 1.** Schematic workflow of the experimental strategy for the biochemical and nutritional
537 characterization of Valle Agricola lentils. SE = Soxhlet Extraction; LLE = Liquid Liquid Extraction

538

539 **Fig. 2.** Antioxidant Capacity in both raw and cooked lentils and cooking water expressed as μmol
540 Trolox Equivalents (TE) per g of each sample \pm SD from A) ABTS method data; B) DPPH method
541 data; C) ORAC method data.

1 **Table 1.** A; nutritional value of Valle Agricola lentil seeds. B, Values are means (\pm SD) of triplicate
2 analyses (n = 3) and are expressed on weight basis.

3

	<i>L. culinaris</i> Valle Agricola (g/100g)	<i>L. culinaris</i> INRAN (g/100g)
Ash	3.5 \pm 0.1	n.r.
Moisture	8.6 \pm 0.2	11.2
Proteins	26.3 \pm 0.5	22.7
Lipids	2.5 \pm 0.2	1.0
Carbohydrates	56.7	51.1

n.r., not reported

4

5

6

7

8 **Table 2.** Total and free amino acid composition of Valle Agricola lentil seeds. Protein amino acids
 9 are reported in bold. Values are means (\pm SD) of triplicate analyses ($n = 3$) and are
 10 expressed on weight basis.
 11

Amino acid ^a	<i>Valle Agricola lentil</i>		<i>Lentil INRAN</i>
	Total amino acids ^b (g/100g)	Free amino acid content ^c mg/100g	Total amino acids (g/100g)
<i>essential amino acids</i>			
His	0.25 \pm 0.01	1.3 \pm 0.2	0.56
Ile	1.1 \pm 0.1	0.90 \pm 0.04	1.04
Leu	2.0 \pm 0.1	1.0 \pm 0.02	1.35
Lys	1.9 \pm 0.1	0.26 \pm 0.02	1.60
Met	0.21 \pm 0.01	0.05 \pm 0.02	0.19
Phe	1.3 \pm 0.1	2.2 \pm 0.1	1.19
Thr	1.1 \pm 0.05	4.1 \pm 0.1	0.89
Trp	n.d.	1.1 \pm 0.5	0.02
Val	1.0 \pm 0.05	1.6 \pm 0.2	1.19
<i>non-essential amino acids</i>			
AAAA	-	15.8 \pm 0.2	-
Ala	1.3 \pm 0.1	6.8 \pm 0.1	0.99
Arg	2.0 \pm 0.1	8.8 \pm 0.2	1.74
Asn	-	50.8 \pm 0.7	-
Asx	3.0 \pm 0.5	-	2.67
Asp	-	2.4 \pm 0.1	-
Car	-	4.3 \pm 0.3	-
Cys	-	-	0.21
Cysteic acid	0.24 \pm 0.04	-	-
GABA	-	0.43 \pm 0.02	-
Gln	-	1.2 \pm 0.1	-
Glx	4.5 \pm 0.3	-	3.87
Glu	-	8.9 \pm 0.2	-
Gly	1.3 \pm 0.1	1.5 \pm 0.1	0.94
Hyllys	-	0.84 \pm 0.09	-
Hypro	-	4.8 \pm 0.3	-
1-mhis	-	1.9 \pm 0.1	-
Phser	-	5.9 \pm 0.2	-
Pro	1.2 \pm 0.2	4.1 \pm 0.4	0.94
Sarc	-	5.0 \pm 0.5	-
Ser	1.8 \pm 0.1	8.1 \pm 0.1	1.26
Taur	-	1.7 \pm 0.1	-
Tyr	0.71 \pm 0.03	1.2 \pm 0.1	0.70
Total (g)	24.85	0.148	21.23

12 ^a free and protein amino acids. Three letter code has been used: 1-mhis, L-1-methylhistidine; AAAA, L- α -aminoadipic
 13 acid; Ala, L-alanine; Asn, L-asparagine; Asx, L-asparagine + L-aspartic acid; Arg, L-arginine; Asp, L-aspartic acid;
 14 Carn, carnitine; Cys, L-half cystine; GABA, γ -amino-n-butyric acid; Gln, L-glutamine; Glu, L-glutamic acid; Glx, L-
 15 glutamine + L-glutamic acid; Gly, glycine; Hyllys, L-Hydroxylysine, Hypro, L-Hydroxyproline; His, L-Histidine; Ile,
 16 L-isoleucine; Leu, L-leucine; Lys, L-lysine; Met, L-methionine; Phe, L-phenylalanine; Phser, o-phospho-L-serine;
 17 Pro, L-proline; Sarc, sarcosine; Ser, L-serine; Taur, taurine; Thr, L-threonine; Trp, L-tryptophan; Tyr, L-tyrosine;
 18 Val, L-valine; n.d., not determined.

19 ^b free plus protein-derived amino acids (see text).

20 ^c free amino acids content for lentil INRAN and other cultivars is not previously published.
 21
 22
 23

24 **Table 3.** Fatty acid constituents of Valle Agricola and INRAN lentil seeds. Values are means (\pm SD)
 25 of triplicate analyses (n = 3) and are expressed on weight basis.

26

	Valla Agricola lentils		<i>Lentil INRAN</i>
Fatty acid	Fatty acid content (mg/100g)		Fatty acid content (mg/100g)
<i>Saturated</i>			
palmitic	C16:0	<i>traces</i>	120.0
heptadecanoic	C17:0	37.5 \pm 2.1	-
stearic	C18:0	<i>traces</i>	10
docosanoic acid	C22:0	70.5 \pm 7.8	-
<i>Unsaturated</i>			
elaidic	C18:1 t	29.5 \pm 2.1	-
oleic	C18:1 c	140 \pm 3	170
eicosenoic	C20:1	71 \pm 3	-
erucic	C22:1	35 \pm 2	-
linoleic	C18:2	25.5 \pm 7.8	360
α -linolenic	C18:3	-	100
Total fatty acids		409	760

27

28

29 **Table 4.** A, correlation coefficients between the different antioxidant methods applied; B, standard
 30 scores of the antioxidant capability of each extract for each applied method and relative
 31 RACI values.

32

33

A

	ABTS	DPPH	ORAC	TPC
ABTS	1.000	0.977	0.997	0.907
DPPH		1.000	0.991	0.976
ORAC			1.000	0.938
TPC				1.000

34

35

B

	ABTS	DPPH	ORAC	TPC	RACI
raw lentils	0.670	0.870	0.680	0.644	0.716
cooked lentils	0.186	0.862	0.358	1.061	0.617
cooking water	-1.465	-0.852	-1.486	-0.801	-1.151

36

37

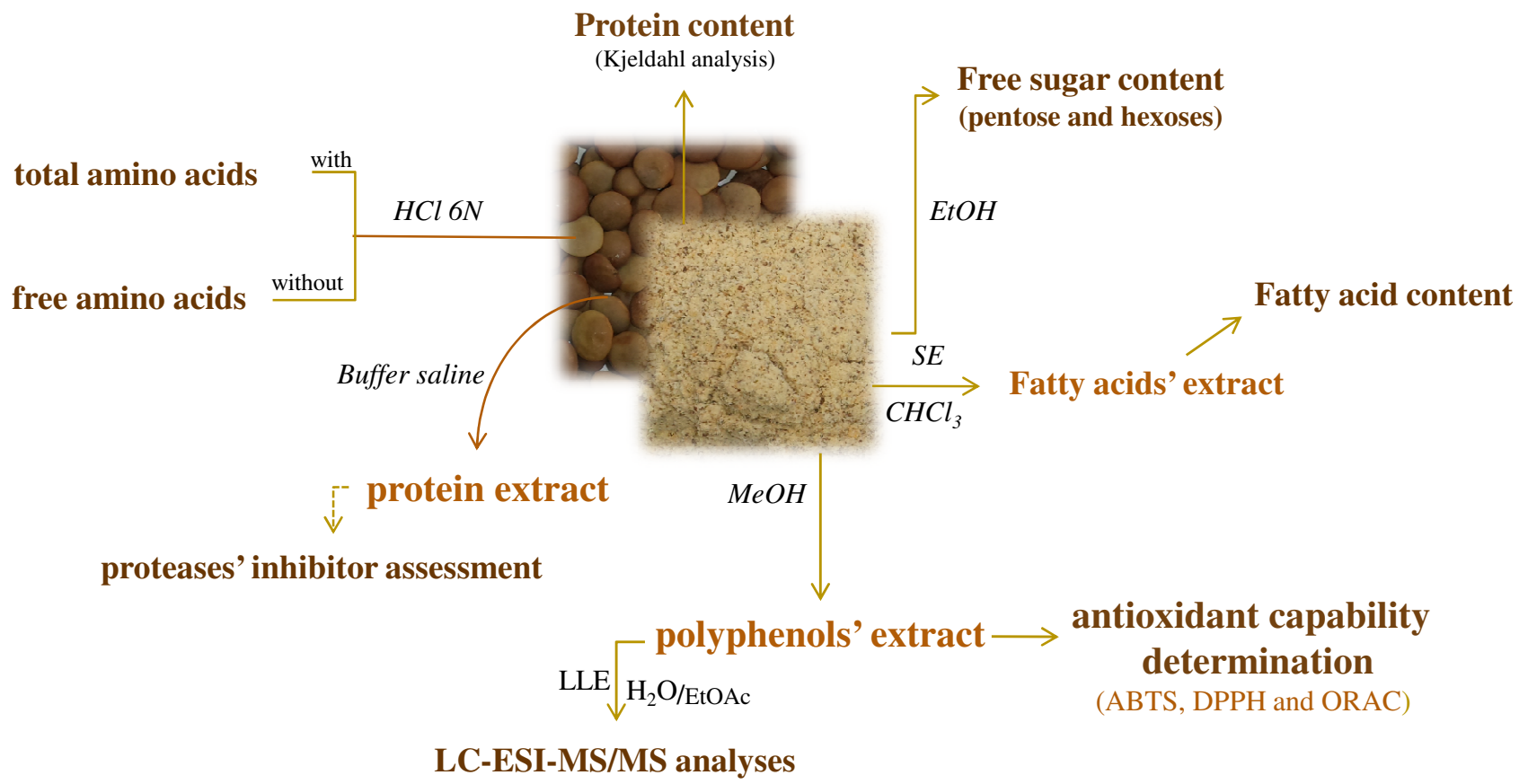
38 **Table 5.** LC-MS/MS data of the main constituents of ‘Valle Agricola’ lentil seed extracts. LSE = raw
 39 Lentil Seed Extract; BLSE = Boiled Lentil Seed Extract; BH₂O = Boiling water

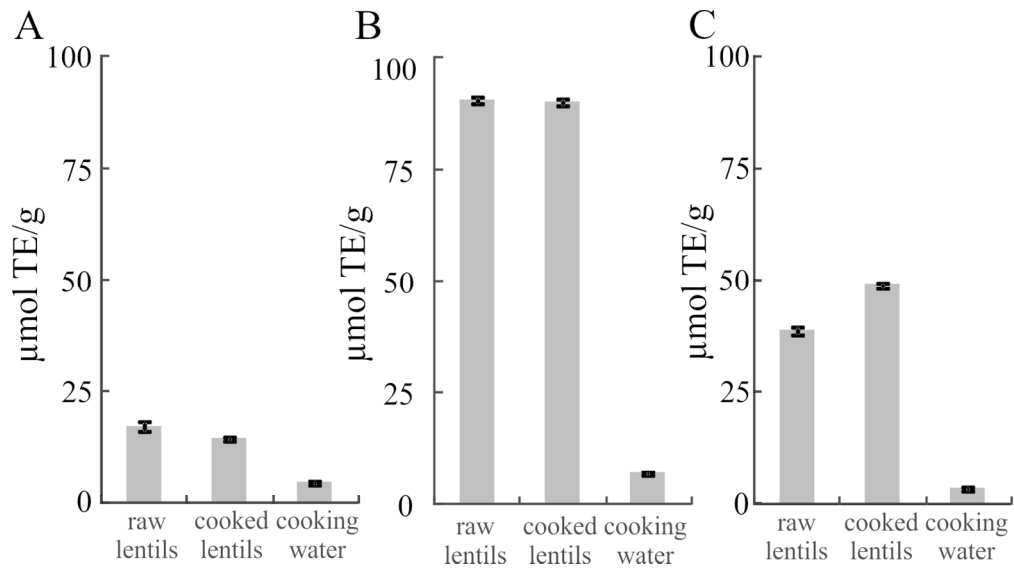
40

Peak	tR (min)	[M-H] ⁻ (m/z)	MS/MS (m/z)	Tentative assignment	LSE	BLSE	BH ₂ O
1	8.0	827	665, 503, 341, 179	pentahexoside	+	-	+
2	8.1	517	193, 179, 161	dihexosyl cyclitol (e.g. ciceritol)	+	-	+
3	8.2	665	503, 383, 341, 283, 231, 179	tetrasaccharide	+	-	+
4	8.5	503	341, 203, 163	trisaccharide (e.g. raffinose)	+	-	+
5	8.6	341	203, 163	disaccharide (e.g. galactinol)	+	-	+
6	22.6	901	755, 417, 285	kaempferol tetraglycoside	+	-	+
8	23.4	463	301	quercetin hexoside	+	+	-
7	25.9	477	301	quercetin hexuronide	+	+	+
9	26.0	755	593, 285, 284	kaempferol triglycoside	+	-	+
10	26.5	577	289, 225, 203, 163	procyanidin dimer	+	+	+
11	26.9	755	285, 284	kaempferol triglycoside	+	-	+
12	27.2	577	289, 225, 203, 163	procyanidin dimer	+	+	+
13	27.6	447	285	kaempferol hexoside	+	+	-
14	28.2	417	285, 241, 151	kaempferol pentoside	+	+	-
15	28.9	289	245, 137	catechin	+	-	-
16	29.5	289	245, 161, 137	epicatechin	+	-	-
17	30.2	301	273, 245, 229, 179, 173, 163, 151, 149, 121, 107	quercetin	+	-	-
18	30.8	341	179, 163	caffeoyl hexose	+	-	-
19	31.8	339	193, 163	feruloyl deoxyhexose	+	-	-
20	32.9	193	177, 163	ferulic acid	+	-	-
21	36.8	305	289, 249, 219, 175, 147, 145, 131, 119, 103	gallocatechin	+	-	-
22	37.7	305	289, 249, 219, 175, 147, 145, 131, 119, 103	epigallocatechin	+	-	-

41

42





138x76mm (300 x 300 DPI)

Graphical abstract:

Valle Agricola lentil, an unknown lentil (*Lens culinaris* Medik.) seed from Southern Italy as a novel antioxidant and prebiotic source

