

PCCP

Accepted Manuscript



This is an *Accepted Manuscript*, which has been through the Royal Society of Chemistry peer review process and has been accepted for publication.

Accepted Manuscripts are published online shortly after acceptance, before technical editing, formatting and proof reading. Using this free service, authors can make their results available to the community, in citable form, before we publish the edited article. We will replace this *Accepted Manuscript* with the edited and formatted *Advance Article* as soon as it is available.

You can find more information about *Accepted Manuscripts* in the [Information for Authors](#).

Please note that technical editing may introduce minor changes to the text and/or graphics, which may alter content. The journal's standard [Terms & Conditions](#) and the [Ethical guidelines](#) still apply. In no event shall the Royal Society of Chemistry be held responsible for any errors or omissions in this *Accepted Manuscript* or any consequences arising from the use of any information it contains.

ARTICLE

Fourier Transform Microwave Spectroscopy of Ac-Ser-NH₂: The Role of the Side Chain Interactions in Peptide Folding

Cite this: DOI: 10.1039/x0xx00000x

Received 00th January 2012,
Accepted 00th January 2012

DOI: 10.1039/x0xx00000x

www.rsc.org/

Carlos Cabezas,^a Martinus A.T. Robben,^b Anouk M. Rijs,^b Isabel Peña^a and J. L. Alonso^{a*}

The serine capped dipeptide *N*-acetyl-L-serinamide (Ac-Ser-NH₂) has been investigated using Fourier transform microwave spectroscopic techniques combined with laser ablation sources. Spectral signatures originating from one dominant species have been detected in the supersonic expansion. Rotational and nuclear quadrupole coupling constants of the two ¹⁴N nuclei have been used in the characterization of a C₇^{eq}/γ-turn structure, which is stabilized by a CO⋯HN intramolecular hydrogen bond closing a seven-membered ring. Two extra hydrogen bonds involving the polar side chain (–CH₂OH) further stabilize the structure. The non-observation of C₅ species, attributed to the presence of the polar side chain, is in contrast with the previous gas phase observation of the related dipeptides containing glycine or alanine residues. The *A-E* splitting pattern arising from the internal rotation of the methyl group has been analyzed and the internal rotation barrier has been determined.

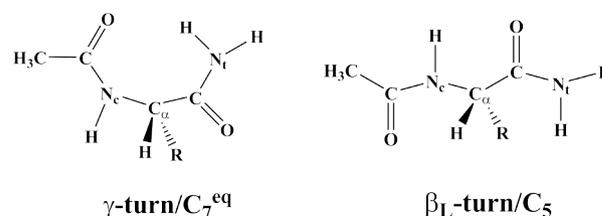
Introduction

Detailed knowledge of the mechanisms of protein folding is important in order to unravel structure-function relationships and improve our understanding of the way numerous processes in living cells function.¹ In solution or the solid state, the protein folding process is a complex interplay of both intra- and intermolecular interactions with the solvent or surrounding amino acids of the protein. Although all these interactions might be present in their biological environment, in order to fully understand the intramolecular folding mechanisms it is also necessary to be able to investigate the intrinsic interactions within the proteins, in absence of any solvent or crystal phase. In particular non-covalent interactions such as intramolecular hydrogen bonds are interesting, since they define the present secondary structure. Gas phase experiments can provide conditions required for studying only the inherent properties of the molecules under investigation, free of interactions with the environment.²⁻³

The first step towards fully understand protein folding requires detailed information on the intrinsic conformational preferences of amino acids, small peptides and peptide mimics. In particular, dipeptide mimics (containing two peptide linkages, –CO–NH–) have received much attention because they represent the smallest realistic and representative systems for designing local conformational effects in peptides and proteins. These molecules are also named capped dipeptides. The vast majority of this work has been done in molecular beam IR/UV double resonance experiments.³⁻⁴ A disadvantage of those techniques however, is that they require the molecule under study to possess a UV chromophore. Roughly speaking, this restricts this type of spectroscopy to molecules containing one or more aromatic rings, thereby excluding the majority of

the amino acids and limiting strongly the different peptides that can be studied.

In contrast, Fourier transform microwave (FTMW) spectroscopy does not require any chromophore; it only needs the molecule to be studied to have a permanent dipole moment.⁵ Therefore, it is particularly well suited for studying dipeptides containing amino acids as relevant as glycine, alanine or proline, elusive to IR/UV double resonance experiments. Lavrich *et al.*⁶ investigated the alanine dipeptide *N*-acetyl-alanine *N*'-methylamide (Ac-Ala-NHMe) and observed only a C₇^{eq} conformation using heating methods to bring molecules into the gas phase. Very recently, the combination of FTMW spectroscopic techniques with laser ablation methods⁷⁻¹⁰ has been successfully applied in the investigation of the conformational preferences of isolated protected dipeptides such as *N*-acetyl-glycinamide (Ac-Gly-NH₂),¹¹ *N*-acetyl-alaninamide (Ac-Ala-NH₂)¹² and *N*-acetyl-prolinamide (Ac-Pro-NH₂).¹³ For both Ac-Gly-NH₂ and Ac-Ala-NH₂, the molecules were found to exist as both the C₇^{eq} (γ-turn) as well as the C₅ (β_L-turn) conformation (See Scheme 1), in which the backbones are stabilized by a CO⋯HN intramolecular hydrogen bond closing a seven- or five-



Scheme 1. Chemical structures of the C₇^{eq} (left) and C₅ (right) configurations for Ac-XX-NH₂ (XX = Gly, Ala) derivatives. N_c and N_t indicate central and terminal nitrogen atoms, respectively.

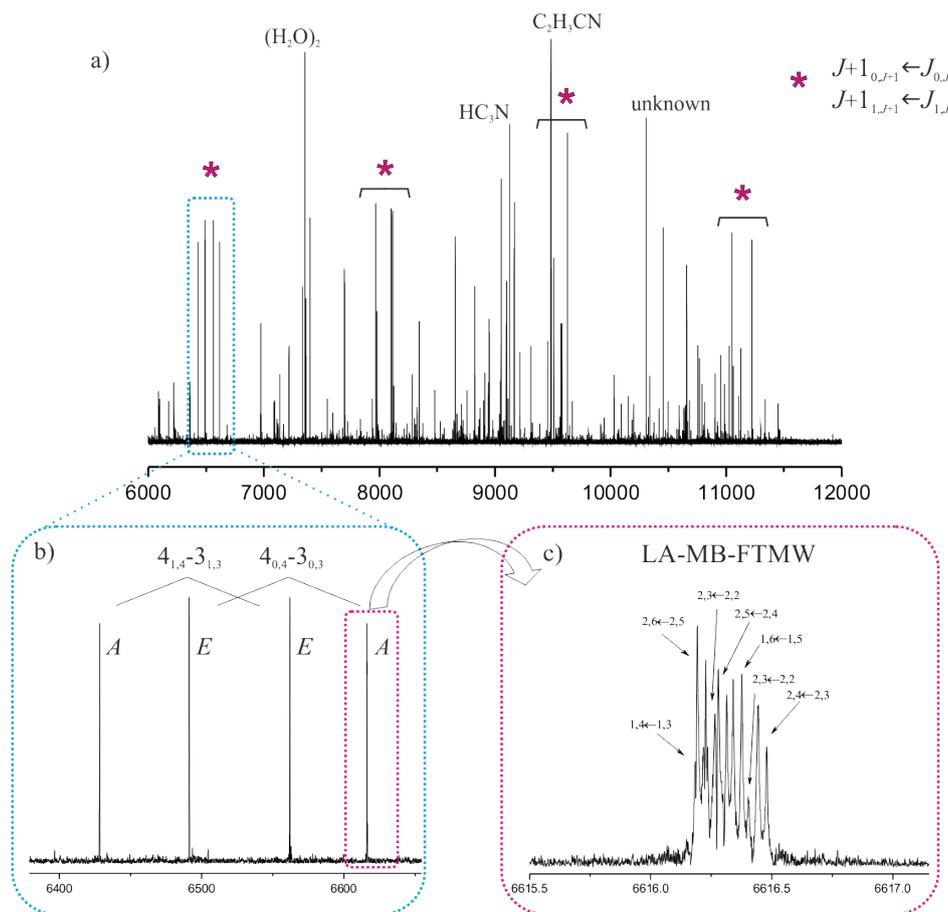


Figure 1. a) Broadband spectrum of Ac-Ser-NH₂ in the 6–12 GHz frequency region showing the most intense rotational transitions for the observed rotamer together with some of the photofragmentation products lines. b) A section of the broadband spectrum showing the *A-E* states of the $4_{1,4}-3_{1,3}$ and $4_{0,4}-3_{0,3}$ rotational transitions. c) The unperturbed *A*-state of the $4_{0,4}-3_{0,3}$ rotational transition of Ac-Ser-NH₂ showing its hyperfine structure completely resolved by LA-MB-FTMW spectrometer. Each hyperfine component labeled with the corresponding values of $I', F' \leftarrow I'', F'''$ quantum numbers, is split by Doppler effect.

membered ring, respectively. In contrast, in the investigation of the Ac-Pro-NH₂ only the C₇ (γ -turn) conformer was detected. The presence of the pyrrolidine ring provides rigidity to the peptide backbone and the formation of other configurations, possible in the other model peptides, it is not observed for the Ac-Pro-NH₂ dipeptide.

The present work reports the first conformational study using Fourier transform microwave techniques of a dipeptide analogue containing an amino acid with a polar side chain, *N*-acetyl-L-serinamide (Ac-Ser-NH₂). The introduction of a polar side chain is an important step because amino acids with polar functional groups are very relevant to protein function and structure.¹⁴ In fact, serine residues have been found to play determinant effects on the stability of transmembrane helices since they are involved in interhelical hydrogen bonds.¹⁵ Moreover, the hydrogen bonds established by the polar group of serine residues have been shown to be at the origin of the activation processes of β_2 -adrenergic receptors.^{16,17} From the conformational point of view, the presence of a polar functional group, a -CH₂OH, in the side chain of serine allows the establishment of additional intramolecular hydrogen bonds, which dramatically increase the number of low-energy conformers.¹⁸ However, sometimes these extra intramolecular interactions are the motif of the selective stabilization of a determined conformation, as it has been shown for the natural

α -amino acid asparagine.¹⁹ These additional interactions do not occur in the aliphatic amino acids whose dipeptides have been previously studied. Consequently, the main goal of our research is to elucidate how this polar side chain of serine affects the conformational preferences of the dipeptide Ac-Ser-NH₂. Will a rich conformational space be observed? Or will the polar side chain favor one of the C₇^{eq} or C₅ conformations?

Experimental

A commercial sample of Ac-Ser-NH₂ (GeneCust, ~99%, m.p.~183°C) was used without any further purification. A solid rod was prepared by pressing the compound's fine powder mixed with a small amount of commercial binder and was placed in the ablation nozzle. A picosecond Nd:YAG laser (10 mJ per pulse, 20 ps pulse width) was used as a vaporization tool. Products of the laser ablation were supersonically expanded using the flow of carrier gas (Ne, 15 bar) and characterized by both chirped pulse Fourier transform microwave (CP-FTMW)¹⁰ and laser ablation molecular beam Fourier transform microwave (LA-MB-FTMW)²⁰ spectroscopy. *N*-acetyl-L-serinamide was first investigated using the CP-FTMW spectrometer to sample swiftly the rotational spectra of the different conformers present in the gas-phase mixture. Details of the experimental setup have been given elsewhere.²¹

Chirped-pulses of 4 μs directly generated by the 24 Gs/s AWG were amplified to about 300W peak power using a traveling wave tube amplifier. A parabolic reflector system composed of dual ridge horns and two parabolic reflectors in a paraxial beam configuration²¹ was used to broadcast the excitation pulse and receive the broadband molecular emission. At a repetition rate of 2 Hz, a total of 70,000 free induction decays (4 FID emissions per gas pulse) each with 10 μs length duration, were averaged and digitized using a 50 Gs/s digital oscilloscope. The frequency domain spectrum in the 6-12 GHz frequency range was obtained by taking a fast Fourier transform (FFT) following the application of a Kaiser-Bessel window to improve baseline resolution.

The sub-Doppler resolution LA-MB-FTMW technique,²⁰ operating from 4 to 10 GHz, was used to record the rotational spectrum with the resolution necessary to analyze the hyperfine structure due to the presence of two ^{14}N nuclei in the molecule. A short microwave radiation pulse of 0.3 μs duration was applied to polarize all the vaporized molecules. The registered free induction decay was then converted to the frequency domain by Fourier transformation. All the transitions appeared as Doppler doublets due to the parallel configuration of the molecular beam and the microwave radiation. The resonance frequency was determined as the arithmetic mean of the two Doppler components. Frequency accuracy better than 3 kHz and an estimated resolution of 5 kHz are achieved in the experiment. From 50 to 100 averages were phase-coherently coadded to achieve reasonable signal to noise ratios (S/N).

Table 1. Experimental and calculated spectroscopic parameters for the observed rotamer and the five lowest energy conformers of Ac-Ser-NH₂. Ab initio energies are included for the predicted species.

	Experimental	$C_7^{\text{eq-I}}^{\text{b}}$	$C_5\text{-I}$	$C_7^{\text{eq-II}}$	$C_5\text{-II}$	$C_5\text{-III}$
A^{a}	1879.8183(73) ^f	1855	1957	2113	2017	1851
B	1001.544607(93)	1007	899	869	857	953
C	750.65138(13)	754	660	708	743	679
Δ_J	0.0573 (25)	-	-	-	-	-
$ \mu_a $	Y ^g	1.9	1.0	3.4	1.2	2.0
$ \mu_b $	Y	0.4	0.3	0.7	0.3	1.8
$ \mu_c $	N	0.1	0.5	2.2	0.2	0.3
N_c/χ_{aa}	1.9538(28)	2.00	2.38	2.16	2.25	2.19
N_c/χ_{bb}	-0.5099 (41)	-0.43	0.86	-2.17	-1.04	1.20
N_c/χ_{cc}	-1.4439 (41)	-1.56	-3.25	0.01	-1.21	-3.39
N_t/χ_{aa}	0.5880(31)	0.56	2.27	0.05	1.99	2.09
N_t/χ_{bb}	1.4018(48)	1.42	1.46	2.15	0.26	0.88
N_t/χ_{cc}	-1.9898(48)	-1.98	-3.73	-2.20	-2.22	-2.97
σ^{b}	1.7	-	-	-	-	-
N^{c}	89	-	-	-	-	-
ΔE^{d}	-	0	1108	1221	1894	1617
ΔG^{e}	-	0	659	892	1174	1187

^a A , B , and C represent the rotational constants (in MHz); Δ_J is the quartic centrifugal distortion constant (in kHz); χ_{aa} , χ_{bb} and χ_{cc} are the diagonal elements of the ^{14}N nuclear quadrupole coupling tensor (in MHz); N_c and N_t correspond to the central and terminal ^{14}N nuclei respectively; μ_a , μ_b and μ_c are the electric dipole moment components (in D). ^b rms deviation of the fit (in kHz). ^c Number of measured transitions. ^d Relative energies (in cm^{-1}) with respect to the global minimum calculated at MP2/6-311++G(d,p) level of theory. ^e Gibbs energies (in cm^{-1}) calculated at 298 K at MP2/6-311++G(d,p) level of theory. ^f Standard error in parentheses in units of the last digit. ^g “Yes” or “No” to observation of a-, b-, and c-type transitions. ^h The conformers are labeled by the size of the ring closed by $\text{CO}\cdots\text{NH}$ hydrogen bond and an index going up with increasing energy.

Results and discussion

The recorded broadband rotational spectrum of laser ablated *N*-acetyl-L-serinamide from 6 to 12 GHz is shown in Figure 1a. Decomposition product lines common to other studies of biomolecules^{22,23} attributable to cyanoderivatives, water clusters, etc. were easily identified. After excluding the aforementioned signals from the spectral analysis, the identification of rotational transitions belonging to a single species, was accomplished. Assignments were based on the identification of a-type $J+1_{0,J+1}\leftarrow J_{0,J}$ and $J+1_{1,J+1}\leftarrow J_{1,J}$ (with J ranging from 3 to 6) pairs of rotational progressions which appear splitted in two components as shown in Figure 1b for $4_{14-3_{13}}$ and $4_{04-3_{03}}$ transitions. We attributed the splittings to the internal rotation of the methyl group attached to N-terminal amide end of the observed rotamer. The V_3 torsional barriers for these methyl groups are low, as shown, for example, for the related molecule Ac-Ala-NH₂¹² causing the occurrence of the $A-E$ splittings due to the coupling of the torsional vibration to the overall rotational angular momentum. Once the analysis of the μ_a -type spectrum was completed, μ_b -type transitions were subsequently predicted and measured with no μ_c -type spectrum observed. Although several lines remained unassigned in the spectrum, identification of further rotamers could not be achieved.

Both A and E components showed partially resolved hyperfine structure as corresponding to a compound with ^{14}N nuclei. This is because the ^{14}N nuclei have a non-zero quadrupole moment ($I=1$) owing to a non-spherical distribution of the nuclear charge, which interacts with the electric field gradient created by the rest of the molecule at the site of these nuclei. The nuclear spin of ^{14}N nuclei couple to the rotational angular momentum resulting in a hyperfine structure in the rotational spectrum.⁵ However, the spectral resolution attainable in the CP-FTMW experiments is not enough to completely resolve these hyperfine effects. For this reason, in a second stage of the investigation *N*-acetyl-L-serinamide was probed using our LA-MB-FTMW technique, which provides the high resolution needed to fully resolve this complicated hyperfine structure (as shown in Figure 1c). Hence, a total of 89 hyperfine components from twelve ^a R - and three ^b R -branch transitions for the unperturbed A state (see Table S2 of the ESI†) were analyzed²⁴ using a Watson's Hamiltonian $H = H_R + H_Q$, where H_R is the semirigid rotor Hamiltonian and H_Q describes the nuclear quadrupole coupling interaction.²⁵ The quadrupole coupling Hamiltonian was set up in the coupled

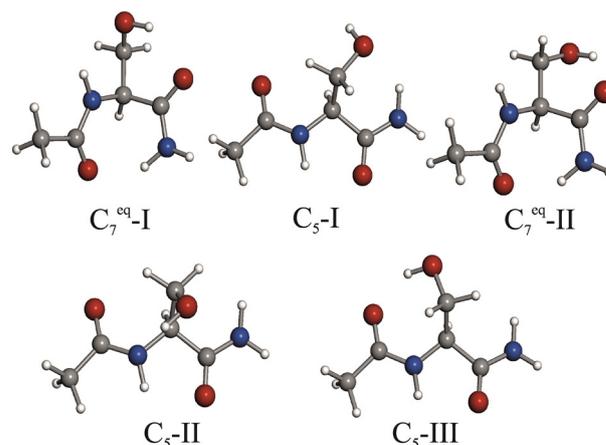


Figure 2. The predicted five low-energy conformers of Ac-Ser-NH₂.

basis set ($I_1 I_2 I J F$), $I_1+I_2=I$, $I+J=F$.²⁶ The energy levels involved in each transition were thus labelled with the quantum numbers J , K_{-1} , K_{+1} , I , F . The analysis rendered the accurate experimental rotational and nuclear quadrupole coupling constants shown in the left column of Table 1. The internal rotation barrier V_3 has been determined using the internal axis method in the form given by Woods.²⁷ The $A-E$ splittings (See Table S3 in the Supporting Information) were fit using the program XIAM²⁸ yielding the internal rotation parameters summarized in Table 2.

Table 2. Methyl internal rotation experimental parameters for the identified rotamer of Ac-Ser-NH₂.

V_3^a	97.2 (3) ^c [127]
$\langle(i,a)^b$	43.40 (2) [46]
$\langle(i,b)$	59.68 (2) [59]
$\langle(i,c)$	62.23 (2) [60]

^a V_3 is the internal rotation barrier in cm⁻¹. ^b Experimentally determined values for the angles between the top rotational axis and the principal axes system, in degrees. In squared brackets, the theoretical values predicted for conformer C₇^{eq}-I. ^c Standard error in parentheses in units of the last digit. ^d

In order to be able to identify the conformation corresponding to the observed rotamers in the spectrum, the conformational landscape of Ac-Ser-NH₂ was explored with the help of quantum chemical calculations. In a first step, semiempirical calculations²⁹ were performed to search for all possible energetic minima of the Ac-Ser-NH₂ molecular system. The resulting molecular geometries were further optimized with the Gaussian suite of programs,³⁰ using a computationally effective B3LYP density functional model with Pople's 6-311++G(d,p) basis set and afterward by second-order Møller-Plesset (MP2) perturbation theory in the frozen core approximation with the same basis set. Frequency calculations were performed with both methods to compute the Gibbs free energies, which should be more representative of the relative populations of each structure. The derived rotational and ¹⁴N nuclear quadrupole coupling constants together with the dipole moment components for the lowest-lying energy conformers (See Figure 2) at the MP2/6-311++G(d,p) level of theory are collected in Table 1. This level of theory has been found to behave satisfactorily in previous studies of several biomolecules.^{7-10,12,13} The results of the calculations at the B3LYP/6-311++G(d,p) level of theory are collected in Table S1 of the ESI†.

The conformational assignment of the observed rotamer was achieved by comparing the experimental spectroscopic constants with those predicted *ab initio*. The experimentally determined rotational constants are similar to those for the C₇^{eq}-I conformer. As we have recently shown,¹¹⁻¹³ nuclear quadrupole coupling constants can be used as fingerprints in conformational analysis of related dipeptides. These parameters are sensitive to the chemical environment of the nitrogen nuclei and to the orientation of the amino group with respect to the principal inertial axis system. Thus, a final comparison between the experimental and theoretical values for those constants clearly serves to discriminate between all the conformers and allows the unequivocal identification of the observed rotamer as conformer C₇^{eq}-I. The observed rotamer exhibited an intense μ_a -type spectrum and weak μ_b -type transitions which is also in agreement with the identification of the C₇^{eq}-I conformer when considering the respective calculated values of the electric dipole moment components. Additionally, the methyl internal

rotation experimental parameters are in good agreement with those estimated theoretically (Table 2) for conformer C₇^{eq}-I, which also supports the achieved assignment.

The experimental determination of the ¹⁴N nuclear quadrupole coupling constants constitutes an exceptional tool that allows the unequivocal establishment of the orientation of the side chain -NH₂ and -NH groups with respect to the molecular frame. Those constants can be used to deduce the nature of the intramolecular interactions in which this functional group is involved. Hence, in the C₇^{eq}-I conformer structure the acetyl carbonyl oxygen is hydrogen bonded to one of the terminal amide hydrogens (C=O...H-N_i), closing a seven-membered cycle in which the serine side chain is oriented equatorially. Moreover, the -CH₂OH group of the serine side chain is participating in two additional hydrogen bonds: one N_c-H...O-H and one O-H...O=C. As can be seen in Figure 3, the estimated distances of the hydrogen bonds show that the O-H...O=C interaction is stronger than the N_c-H...O-H. This fact can be attributed to the dominant donor character of the OH group over its acceptor propensity, leading to a quite unbalanced H-bonding network. The distance for the C=O...H-N_i bond (γ -turn) is analogous to those related for other γ -turns of aliphatic dipeptides such as Ac-Gly-NH₂¹¹ and Ac-Ala-NH₂¹² for which intramolecular bond distances of 2.03 and 2.07 Å were found, respectively. This fact points to that the intramolecular interactions of the side chain does not affect to the strength of the γ -turn bond. However, the side chain extra interactions, which cannot take place in any other possible conformation of the Ac-Ser-NH₂, seem to be the factor which accounts for the overstabilization of this species and, thus, the non observation of C₅ species. In contrast, for the related Ac-Gly-NH₂¹¹ and Ac-Ala-NH₂¹² dipeptides both C₇^{eq} and C₅ species were detected with the approximate population C₇^{eq}/C₅ ratio around 2 and 3, respectively. Because no intramolecular interactions involving the lateral side chain can occur in Gly and Ala dipeptides, the stability/abundance difference between C₇^{eq} and C₅ species is exclusively determined by the strength of the C=O...H-N interactions (seven- or five-membered ring). On this basis, we can infer that the presence of polar side chains alter significantly the conformational preferences of dipeptides containing aliphatic amino acids.

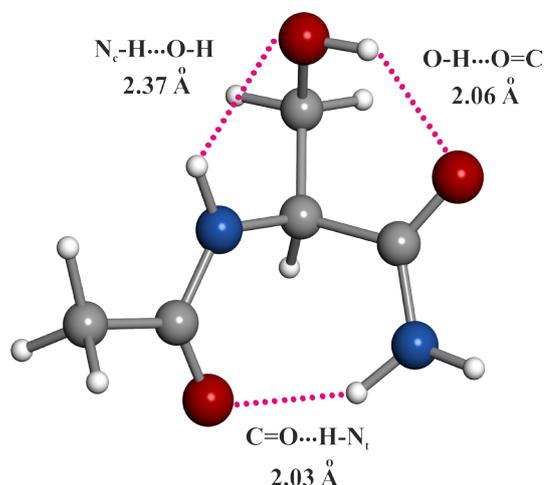


Figure 3. 3D *ab initio* structure (Cartesian coordinates in Table S4 of the ESI†) of observed conformer of Ac-Ser-NH₂ showing the intramolecular interactions and the estimated distances which stabilize the structure.

The results obtained here for Ac-Ser-NH₂ can be compared with those reported previously for the analogous tripeptide Ac-Phe-Ser-NH₂ using IR-UV ion dip spectroscopy.^{31,32} For the Ac-Phe-Ser-NH₂ molecule it was observed only the C₇/γ-turn conformer where the C₇ ring established by the serine residue is structurally similar to that of the detected conformer of Ac-Ser-NH₂. Additionally, Yan et al.³¹ found the same C₇ structure to be the most stable for the tripeptide Ac-Phe-Cys-NH₂. This peptide only differs by one atom with respect to Ac-Phe-Ser-NH₂, where the –OH side chain of serine is replaced by the –SH group in cysteine. For the Ac-Phe-Cys-NH₂ an additional structure which exhibits C₁₀/β-turn geometry was observed. The presence of the second structure was attributed to the weaker SH...O hydrogen bond strength, allowing the competition with other type of interactions such as SH-π interactions resulting in the presence of both γ-turn as β-turn conformations. This result confirms our conclusion that for Ac-Phe-Ser-NH₂ which exhibits a strong hydrogen bond, only one conformer is present, while when weaker H-bond interactions are formed such as for Cys or for the previously studied Gly¹¹ and Ala¹² capped peptides, other conformers besides this C₇ structure will be present.

Conclusions

The present investigation of the Ac-Ser-NH₂ system together with the previous work on the Ac-Gly-NH₂, Ac-Ala-NH₂ and Ac-Pro-NH₂ illustrate the capabilities of the Fourier transform microwave techniques to investigate the conformational preferences of biologically relevant peptides isolated in gas phase. The structural information derived for these aliphatic dipeptides, which are elusive to other high resolution spectroscopic techniques, is of utmost importance not only to gain knowledge about their intrinsic conformational properties but also to serve as a benchmark for theoretical investigations.

Our results for the serine dipeptide Ac-Ser-NH₂ show that its conformational landscape in gas phase is dominated by a single C₇^{eq}/γ-turn species. Now, the initial research question about whether the weaker polar side chain favors one of the C₇ and C₅ conformations through the formation of intramolecular hydrogen bonds can be answered. The additional intramolecular interactions formed by the presence of a polar group in the side chain of serine have been shown to be at the origin of the conformational locking to a C₇^{eq} species observed for the dipeptide Ac-Ser-NH₂. Hence, it has been demonstrated that the presence of a polar side chain increases the plausible number of conformations but on the contrary imparts restrictions on the amount of conformers observed.

Acknowledgements

This research was supported by the Ministerio de Ciencia e Innovación (grant numbers CTQ 2010-19008, CTQ 2013-40717-P and Consolider Ingenio 2010 CSD 2009-00038), Junta de Castilla y León (grant number VA175U13) and ERC (grant number 610256 “Nanocosmos”). C.C. thanks to the Junta de Castilla y León for the postdoctoral contract (grant number CIP13/01).

Notes and references

^a Grupo de Espectroscopia Molecular (GEM). Edificio Quifima. Laboratorios de Espectroscopia y Bioespectroscopia. Unidad Asociada CSIC, Parque Científico Uva, Universidad de Valladolid, Paseo de Belén 5, 47011 Valladolid, Spain

E-mail: jalonso@gf.uva.es; Phone: +34 983186345

^b Radboud University, Institute for Molecules and Materials, FELIX Laboratory, Toernooiveld 7-c, 6525 ED Nijmegen, The Netherlands

† Electronic Supplementary Information (ESI) available: [Measured frequencies for the nuclear quadrupole coupling hyperfine components and splittings for the A-E internal rotation components of the C₇^{eq}-I conformer of Ac-Ser-NH₂ together with the cartesian coordinates for the *ab initio* predicted geometry of theory for the observed conformer of Ac-Ser-NH₂]. See DOI: 10.1039/b000000x/

1. Y. Park, V. Helms, On the Derivation of Propensity Scales for Predicting Exposed Transmembrane Residues of Helical Membrane Proteins, *Bioinformatics*, 2007, **23**, 701-708.
2. E. G. Robertson, John P. Simons, Getting into Shape: Conformational and Supramolecular Landscapes in Small Biomolecules and their Hydrated Clusters, *Phys. Chem. Chem. Phys.*, 2006, **8**, 1033-1048.
3. M.S. de Vries, P. Hobza, Gas-phase Spectroscopy of Biomolecular Building Blocks, *Ann. Rev. Phys. Chem.*, 2007, **58**, 585-612.
4. a) W. Chin, F. Piuze, I. Dimicoli, M. Mons, Probing the competition between secondary structures and local preferences in gas phase isolated peptide backbones, *Phys. Chem. Chem. Phys.*, 2006, **8**, 1033-1048. b) S. Jaeqx, W. Du, E. J. Meijer, J. Oomens, A. M. Rijs, Conformational Study of Z-Glu-OH and Z-Arg-OH: Dispersion Interactions versus Conventional Hydrogen Bonding, *J. Phys. Chem. A*, 2012, **117**, 1216-1227 c) S. Jaeqx, J. Oomens, A.M. Rijs, Gas-phase Salt Bridge Interactions between Glutamic Acid and Arginine, *Phys. Chem. Chem. Phys.*, 2013, **15**, 16341-16352. d) S. Jaeqx, J. Oomens, A. Cimas, M.-P. Gaigeot, A.M. Rijs, Gas-Phase Peptide Structures Unraveled by Far-IR Spectroscopy: Combining IR-UV Ion-Dip Experiments with Born-Oppenheimer Molecular Dynamics Simulations, *Angew. Chem. Int. Ed.*, 2014, **53**, 3663-3666. e) E. Gloaguen, M. Mons in Isolated Neutral Peptides, Topics in Current Chemistry “Gas-Phase IR Spectroscopy and Structure of Biological Molecules”, (A. M Rijs & J. Oomens Ed.) Springer. In press. 2014. DOI: 10.1007/128_2014_580.
5. W. Gordy and R. L. Cook, Microwave Molecular Spectra, Wiley, New York, 1984.
6. R. J. Lavrich, D. F. Plusquellic, R. D. Suenram, G. T. Fraser, A. R. Hight Walker, M. J. Tubergen, Experimental studies of peptide bonds: Identification of the C₇^{eq} conformation of the alanine dipeptide analog N-acetyl-alanine N'-methylamide from torsion-rotation interactions, *J. Chem. Phys.* 2003, **118**, 1253-1265.
7. J. L. Alonso, J. C. Lopéz, Topics in Current Chemistry “Microwave Spectroscopy of Biomolecular Building Blocks”, (A. M Rijs & J. Oomens Ed.) Springer. In press. 2014. DOI: 10.1007/128_2014_601.
8. J. L. Alonso, C. Pérez, M. E. Sanz, J. C. Lopéz, S. Blanco, Seven Conformers of L-Threonine in the Gas Phase: a LA-MB-FTMW Study, *Phys. Chem. Chem. Phys.* 2009, **11**, 617-627.
9. I. Peña, M. E. Sanz, J. C. López, J. L. Alonso, Preferred Conformers of Proteinogenic Glutamic Acid, *J. Am. Chem. Soc.* 2011, **134**, 2305-2312.
10. S. Mata, I. Peña, C. Cabezas, J. C. López, J. L. Alonso, A Broadband Fourier-transform Microwave Spectrometer with Laser Ablation Source: The Rotational Spectrum of Nicotinic Acid, *J. Mol. Spectrosc.*, 2012, **280**, 91-96.
11. C. Puzzarini, M. Biczysko, V. Barone, L. Largo, I. Peña, C. Cabezas, J. L. Alonso, Accurate Characterization of the Peptide Linkage in the

- Gas Phase: A Joint Quantum-Chemical and Rotational Spectroscopy Study of the Glycine Dipeptide Analogue, *J. Phys. Chem. Lett.*, 2014, **5**, 534-540.
12. C. Cabezas, M. Varela, V. Cortijo, A.I. Jimenez, I. Pena, A.M. Daly, J.C. Lopez, C. Cativiela, J.L. Alonso, The Alanine Model Dipeptide Ac-Ala-NH₂ Exists as a Mixture of C₇^{eq} and C₅ Conformers, *Phys. Chem. Chem. Phys.*, 2013, **15**, 2580-2585.
13. C. Cabezas, M. Varela, J. L. Alonso, Probing the γ -Turn in a Short Proline Dipeptide Chain, *ChemPhysChem*, 2013, **14**, 2539-2543.
14. C. Budiman, T. Tadokoro, C. Angkawidjaja, Y. Koga, S. Kanaya, Role of Polar and Nonpolar Residues at the Active Site for PPLase Activity of FKBP22 from *Shewanella sp. SIB1*, *The FEBS Journal*, 2012, **279**, 976-986.
15. A. Senes, I. Ubarretxena-Belandia, D. M. Engelman, The Ca-H...O Hydrogen Bond: A Determinant of Stability and Specificity in Transmembrane Helix Interactions, *Proc. Natl. Acad. Sci. U.S.A.*, 2001, **98**, 9056-9061.
16. K. Wieland, H. M. Zuurmond, C. Krasel, A. P. Ijzerman, M. J. Lohse, Involvement of Asn-293 in Stereospecific Agonist Recognition and in Activation of the β_2 -Adrenergic Receptor, *Proc. Natl. Acad. Sci. U.S.A.*, 1996, **93**, 9276-9281.
17. G. Liapakis, J. A. Ballesteros, S. Papachristou, W. C. Chan, X. Chen, J. A. Javitch, The Forgotten Serine, *J. Biol. Chem.*, 2000, **275**, 37779-37788.
18. S. Blanco, M. E. Sanz, J. C. López, J. L. Alonso, Revealing the Multiples Structures of Serine, *Proc. Natl. Acad. Sci. U.S.A.*, 2007, **104**, 20183-20188.
19. C. Cabezas, M. Varela, S. Mata, I. Peña, J.C. López, J. L. Alonso, The Conformational Locking of Asparagine, *Chem. Commun.*, 2012, **48**, 5934-5936.
20. C. Bermúdez, S. Mata, C. Cabezas, J. L. Alonso, Tautomerism in Neutral Histidine, *Angew. Chem. Int. Ed.* 2014, **53**, 11015-11018.
21. I. Peña, S. Mata, A. Martín, C. Cabezas, A. M. Daly, J. L. Alonso, Conformations of D-xylose: The Pivotal Role of the Intramolecular Hydrogen-Bonding, *Phys. Chem. Chem. Phys.* 2013, **15**, 18243-18248.
22. I. Peña, L. Kolesníková, C. Cabezas, C. Bermúdez, M. Berdakin, A. Simão, J. L. Alonso, The shape of D-glucosamine, *Phys. Chem. Chem. Phys.*, 2014, **16**, 23244-23250.
23. I. Peña, C. Cabezas, J. L. Alonso, The Nucleoside Uridine Isolated in the Gas Phase, *Angew. Chem. Int. Ed.*, 2015, **54**, 2991-2994.
24. H. M. Pickett, *J. Mol. Spectrosc.* 1991, **148**, 371-377.
25. J. K. G. Watson, *Vibrational Spectra and Structure*, Vol. 6, Elsevier, Amsterdam, 1977.
26. H. M. Foley, *Phys. Rev.* 1947, **71**, 747-751.
27. a) R. C. Woods, *J. Mol. Spectrosc.*, 1996, **21**, 4-24. b) R. C. Woods, *J. Mol. Spectrosc.*, 1997, **22**, 49-59.
28. H. Hartwig, H. Dreizler, *Z. Naturforsch.* 1996, **51a**, 923-932.
29. J. J. P. Stewart, Optimization of Parameters for Semiempirical Methods I. Method, *J. Comput. Chem.*, 1989, **10**, 209-220.
30. G.W.T. M. J. Frisch, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G. A. Petersson, H. Nakatsuji, M. Caricato, X. Li, H. P. Hratchian, A. F. Izmaylov, J. Bloino, G. Zheng, J. L. Sonnenberg, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. Bearpark, J. J. Heyd, E. Brothers, K. N. Kudin, V. N. Staroverov, T. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, N. Rega, J. M. Millam, M. Klene, J. E. Knox, J. B. Cross, V. Bakken, C. Adamo, J. Jaramillo, R. Gomperts, R. E. Stratmann, O. Yazyev, A. J. Austin, R. Cammi, C. Pomelli, J. W. Ochterski, R. L. Martin, K. Morokuma, V. G. Zakrzewski, G. A. Voth, P. Salvador, J. J. Dannenberg, S. Dapprich, A. D. Daniels, O. Farkas, J. B. Foresman, J. V. Ortiz, J. Cioslowski, D. J. Fox, Gaussian 09, Revision B.01, in, Gaussian, Inc., Wallingford CT, 2010.
31. B. Yan, S. Jaeqx, W. J. van der Zande, A. M. Rijs, A Conformation-Selective IR-UV Study of the Dipeptides Ac-Phe-Ser-NH₂ and Ac-Phe-Cys-NH₂: Probing the SH...O and OH...O Hydrogen Bond Interactions, *Phys. Chem. Chem. Phys.*, 2014, **16**, 10770-10778.
32. M. Alauddin, H. S. Biswal, E. Gloaguen, M. Mons, Intra-Residue Interactions in Proteins: Interplay between Serine or Cysteine Side Chains and Backbone Conformations, Revealed by Laser Spectroscopy of Isolated Model Peptides, *Phys. Chem. Chem. Phys.*, 2015, **17**, 2169-2178.